

# Oxytocin reverses osteoporosis in a sex-dependent manner

Guillaume E. Beranger<sup>1,2,3</sup>, Mansour Djedaini<sup>1,2,3</sup>, Séverine Battaglia<sup>4,5</sup>,  
Christian H. Roux<sup>1,2,3,6</sup>, Marcel Scheideler<sup>7,8,9</sup>, Dominique Heymann<sup>4,5</sup>,  
Ez-Zoubir Amri<sup>1,2,3\*</sup> and Didier F. Pisani<sup>1,2,3\*</sup>

<sup>1</sup>UMR 7277, Institut of Biology Valrose (iBV), Université de Nice-Sophia-Antipolis, Nice, France, <sup>2</sup>UMR 7277, Institut of Biology Valrose (iBV), CNRS, Nice, France, <sup>3</sup>U1091, INSERM, Institut of Biology Valrose (iBV), Nice, France, <sup>4</sup>Université de Nantes, Nantes, France, <sup>5</sup>UMR 957, INSERM, Equipe LIGUE Nationale Contre le Cancer 2012, Nantes, France, <sup>6</sup>Service de Rhumatologie, Hôpital l'Archet CHU, Nice, France, <sup>7</sup>Helmholtz Center Munich, Institute for Diabetes and Cancer, Neuherberg, Germany, <sup>8</sup>Heidelberg University Hospital, Heidelberg, Germany, <sup>9</sup>German Center for Diabetes Research (DZD), Neuherberg, Germany

## OPEN ACCESS

### Edited by:

Andrzej Bartke,  
Southern Illinois University School of  
Medicine, USA

### Reviewed by:

David George Monroe,  
Mayo Clinic College of Medicine,  
USA

James R. Edwards,  
University of Oxford, UK

### \*Correspondence:

Ez-Zoubir Amri and Didier F. Pisani,  
UMR 7277 CNRS – UMR 1091  
INSERM, Faculté de Médecine,  
Institut of Biology Valrose (iBV),  
Université de Nice-Sophia-Antipolis,  
28 Avenue de Valombrose,  
Nice Cedex 2 06107, France  
amri@unice.fr; pisani@unice.fr

### Specialty section:

This article was submitted to  
Endocrinology of Aging, a section of  
the journal *Frontiers in Endocrinology*

**Received:** 13 February 2015

**Accepted:** 04 May 2015

**Published:** 19 May 2015

### Citation:

Beranger GE, Djedaini M, Battaglia S,  
Roux CH, Scheideler M, Heymann D,  
Amri E-Z and Pisani DF (2015)  
Oxytocin reverses osteoporosis in a  
sex-dependent manner.  
*Front. Endocrinol.* 6:81.  
doi: 10.3389/fendo.2015.00081

The increase of life expectancy has led to the increase of age-related diseases such as osteoporosis. Osteoporosis is characterized by bone weakening promoting the occurrence of fractures with defective bone regeneration. Men aged over 50 have a prevalence for osteoporosis of 20%, which is related to a decline in sex hormones occurring during andropause or surgical orchidectomy. As we previously demonstrated in a mouse model for menopause in women that treatment with the neurohypophyseal peptide hormone oxytocin (OT) normalizes body weight and prevents the development of osteoporosis, herein we addressed the effects of OT in male osteoporosis. Thus, we treated orchidectomized mice, an animal model suitable for the study of male osteoporosis, for 8 weeks with OT and then analyzed trabecular and cortical bone parameters as well as fat mass using micro-computed tomography. Orchidectomized mice displayed severe bone loss, muscle atrophy accompanied by fat mass gain as expected in andropause. Interestingly, OT treatment in male mice normalized fat mass as it did in female mice. However, although OT treatment led to a normalization of bone parameters in ovariectomized mice, this did not happen in orchidectomized mice. Moreover, loss of muscle mass was not reversed in orchidectomized mice upon OT treatment. All of these observations indicate that OT acts on fat physiology in both sexes, but in a sex specific manner with regard to bone physiology.

**Keywords:** orchidectomy, bone, adipose tissue, male, female, mice

## Introduction

Life expectancy continually increases in industrialized countries and is linked to aging-associated phenotypes such as a decrease in hormonal levels, lean and bone mass, as well as gain in fat mass. Along with aging-associated diseases defined as a major cause of morbidity and mortality, osteoporosis can be found essentially associated to a severe decrease in circulating estrogens due to menopause in women (1). Although with lower frequency than in women, the osteoporosis in men aged over 50 has a prevalence of 20% and is a relatively common burden with treatments less defined than in women (2).

Osteoporosis, defined by the deterioration of bone density and microarchitecture leading to bone fragility (3), is a major public health problem with 6.3 million of cases expected in 2050 (4). Besides

substantial financial costs, this disease leads to a deleterious life quality with a significant decrease in mobility up to the loss of autonomy, a susceptibility to other diseases such as osteoarthritis and obesity, and finally a significant increase in morbidity and mortality, especially in the course of the year following a hip fracture. Osteoporosis in men is less well known, but seems to be also associated with a decrease in sex hormone levels (androgens), particularly testosterone, and is aggravated by environmental factors such as nutrition and physical inactivity (2). It is difficult to anticipate the onset of osteoporosis in men (5), except after surgical removal of the testis (orchidectomy), for example, in case of prostate cancer treatment to reduce testosterone level, which leads to a drop in the level of circulating androgens (6). In these cases, preventive postoperative treatments can be provided to limit the development of osteoporosis. As for women, a decrease of sex hormones can also cause weight gain, behavioral changes, and muscle wasting in men (5).

Osteoporosis may affect trabecular or cortical bone associated with different types of fractures (7, 8). The strength and integrity of the bone depend on the balance between resorption by osteoclasts and bone formation by osteoblasts (9, 10). The decrease of bone mass, which occurs during osteoporosis, implies an acceleration of the “bone turnover” with an imbalance between resorption and formation in favor of bone resorption (11, 12).

In addition to bone loss, osteoporosis is associated with a gain in bone marrow adiposity due to the formation of adipocytes, at the expense of osteoblasts, both derived from a common progenitor: the mesenchymal stem cell. Adiposity is also connected to the prevalence of bone fracture in osteoporosis (13). The adipocyte/osteoblast balance is essential for bone homeostasis, as it determines (1) the amount of essential osteoblasts for bone formation and (2) the secretion of adipokines involved in bone metabolism. We previously characterized the major role of this balance in the pathophysiology of osteoporosis, which is controlled by a circulating hormone, namely oxytocin (OT). *In vitro*, the treatment of mesenchymal stem cells with O promotes the formation of osteoblasts and inhibits adipogenesis (14). In addition, we showed an inverse correlation between OT circulating levels and the occurrence of osteoporosis (14–16) on a small cohort of post-menopausal women first, and then on a larger cohort from a European prospective multicenter study (OPUS). Finally, we demonstrated that treatment of ovariectomized mice, an animal model mimicking menopause in women, by daily injections of OT (1 mg/kg/day) prevented and reversed the osteoporotic phenotype. Indeed, both OT preventive and curative treatments led to normalization of bone parameters (morphometric and mechanical), to a decrease of bone marrow adiposity and, interestingly, to a normalization in weight gain induced by ovariectomy (17).

Oxytocin belongs to the family of pituitary hormones and regulates the function of peripheral organs including the mammary glands and smooth muscle of the uterus. Due to this property, it has commonly been used in other medical obstetrics without significant side effects (18). Thus, OT emerges as a promising molecule in the treatment of osteoporosis.

Herein, we daily injected orchidectomized male mice with 1 mg/kg of OT for 8 weeks to analyze its effect on male osteoporosis. We previously used this protocol with success to reverse

the bone phenotype in ovariectomized female mice (17, 19). Analysis of treated mice demonstrated that OT did not restore the bone loss and muscle atrophy in orchidectomized male mice. These results demonstrated a sex-dependent efficiency of OT on bone remodeling following gonadectomy that might be linked to the difference in sexual hormones between male and female.

## Materials and Methods

### Animals

The experiments were conducted in accordance with the French and European regulations for the care and use of research animals and were approved by the local experimentation committee. Animals were maintained under constant temperature ( $21 \pm 2^\circ\text{C}$ ) and 12:12-hour light–dark cycles, with *ad libitum* access to standard chow diet and water. Ten-week-old C57Bl/6J mice were randomized and subjected to bilateral orchidectomy, ovariectomy or sham surgery by the manufacturer (Charles River Laboratories). For sham-operated mice, the testis or ovaries were exteriorized but replaced intact. Two weeks after surgery, orchidectomized (ORX), ovariectomized (OVX), or sham groups of mice ( $n = 12$ ) were intra-peritoneally injected with vehicle (Ve, NaCl 0.9%) or OT (1 mg/kg) on a daily basis and over 8 weeks.

### Micro-Computed Tomography

Trabecular bone microarchitecture of the distal femoral metaphysis was analyzed using the high-resolution SkyScan-1076 X-ray micro-computed tomography system (SkyScan). Femora were scanned after necropsy using the same parameters: 9  $\mu\text{m}$  of pixel size, 49 kV, 0.5 mm thick aluminum filter, and  $0.5^\circ$  of rotation step. Calculation of femur trabecular bone parameters (Bone Volume/Total Volume: BV/TV) following 3D morphometric parameters (Bone ASBMR nomenclature) were performed on 0.9 mm of femur length from the distal growth plate excluding the cortical bone. Analysis of cortical bone was performed using the same parameters. Cortical cross-sectional area (CSA) was determined on a transversal section localized at the mid-length of the femoral shaft.

Adipose tissue quantification was carried out using a SkyScan-1178 X-ray micro-computed tomography system. Mice were anesthetized and scanned using the same parameters: 104  $\mu\text{m}$  of pixel size, 49 kV, 0.5 mm thick aluminum filter, and  $0.9^\circ$  of rotation step. Total adipose tissue volume was determined between the lumbar vertebra 1 (L1) and the caudal vertebra 4 (C4), whereas intra-abdominal and subcutaneous adipose tissues areas were measured on one section at the lumbar 5 (L5) level. 3D reconstructions and analysis of bone parameters and adipose tissue areas or volumes were performed using NRecon and CTAn software (SkyScan).

### Plasma Measurements

Blood was sampled by intracardiac puncture from anesthetized animals and plasma was stored at  $-80^\circ\text{C}$ . Procollagen I N-Terminal propeptide (PINP, USCN) or Cross Linked C-Telopeptide of Type I Collagen (CTXI, USCN) plasma levels was measured using an ELISA kit as per the manufacturer's instructions.

## Isolation and Analysis of RNA

Total RNA was extracted using a TRI-Reagent kit as per the manufacturer's instructions. For total bone RNA extraction, humeri were homogenized using an Ultra Turrax homogenizer and samples were then processed as per the TRI-Reagent extraction protocol. Two micrograms of total RNA, digested with DNase I (Promega), were subjected to reverse transcription-polymerase chain reaction (RT-PCR) analysis as previously described (20). Quantitative PCR assays were run using an ABI Prism 7,000 real-time PCR machine (PerkinElmer Life and Analytical Sciences). The final reaction volume was 20  $\mu$ l, including 100 nM of specific primers, 20 ng of reverse-transcribed RNA, and 10  $\mu$ l of SYBR Green master mix (Applied Biosystems). Quantitative PCR conditions were as follows: 2 min at 50°C; 10 min at 95°C; and 40 cycles of 15 s at 95°C, 1 min at 60°C. The expression of genes was normalized to the expression of two housekeeping genes, 36B4 and hprt. Gene expression was quantified using the comparative  $-\Delta$ Ct method. The oligonucleotides for each target of interest, designed using Primer Express software (PerkinElmer Life and Analytical Sciences), are shown (forward and reverse) in **Table 1**.

## Statistical Analyses

Data are expressed as mean values  $\pm$ SEM and was analyzed using the two-tailed Student's *t*-test. Differences were considered statistically significant at  $p \leq 0.05$ .

**TABLE 1 | Sequence of primers used for gene expression analysis.**

| Target gene        | Forward                | Reverse                      |
|--------------------|------------------------|------------------------------|
| RANKL              | GGAAGTCAACACATTGTGGG   | GCCTCCATCATAGCTGGAGC         |
| OPG                | TCCGGCGTGGTGCAA        | AGAAGCCCATCTGGACATTTTTG      |
| Tracp              | TGCCTACCTGTGTGGACATGA  | CACATAGCCCCACACCGTTCTC       |
| Collagen 1 alpha 1 | GCGAAGGCAACAGTCGCT     | CTTGTTGGTTTTGTATTGATGAC      |
| Osteocalcin        | CCACCCGGGAGCAGTGT      | CTAAATAGTGATACCGTAGATGCGTTTG |
| Hprt               | GCCTAAGATGAGCGCAAGTTGA | AGGCAGATGGCCACAGGACTA        |
| 36B4               | TCCAGGCTTTGGGCATCA     | CTTTATCAGCTGCACATCACTCAGA    |

**TABLE 2 | MicroCT analysis of femora trabecular and cortical bone parameters.**

|                           | Mice group                 | Sham-Ve<br>(n = 12) | Sham OT<br>1 mg/kg (n = 12)  | ORX Ve<br>(n = 12)           | ORX OT<br>1 mg/kg (n = 12)   |
|---------------------------|----------------------------|---------------------|------------------------------|------------------------------|------------------------------|
| <b>Trabecular bone</b>    |                            |                     |                              |                              |                              |
| Distal femoral metaphysis | BV/TV (%)                  | 15.4 $\pm$ 1.2      | 11.9 $\pm$ 1.2               | 3.2 $\pm$ 0.3 <sup>a</sup>   | 3.1 $\pm$ 0.3 <sup>a</sup>   |
|                           | Tb. Th. ( $\mu$ m)         | 69 $\pm$ 1          | 63 $\pm$ 2 <sup>a</sup>      | 65 $\pm$ 1 <sup>a</sup>      | 62 $\pm$ 1 <sup>a</sup>      |
|                           | Tb. N. (mm <sup>-1</sup> ) | 2.2 $\pm$ 0.1       | 1.9 $\pm$ 0.1                | 0.5 $\pm$ 0.04 <sup>a</sup>  | 0.5 $\pm$ 0.04 <sup>a</sup>  |
|                           | Tb. Sp. ( $\mu$ m)         | 238 $\pm$ 9         | 247 $\pm$ 6                  | 532 $\pm$ 23 <sup>a</sup>    | 528 $\pm$ 16 <sup>a</sup>    |
| <b>Cortical bone</b>      |                            |                     |                              |                              |                              |
| Mid-femur diaphysis       | CSA (mm <sup>2</sup> )     | 1.04 $\pm$ 0.08     | 0.97 $\pm$ 0.06 <sup>a</sup> | 0.83 $\pm$ 0.04 <sup>a</sup> | 0.83 $\pm$ 0.04 <sup>a</sup> |

Control (Sham) and ORX mice were submitted to daily injections of OT or Ve for 8 weeks starting 2 weeks after surgery. Detailed analysis of trabecular parameters was performed on the distal metaphysis of femora and on the mid-femur diaphysis for cortical bone parameters. Bone volume/total volume (BV/TV), trabecular thickness (Tb. Th), trabecular spacing (Tb. Sp), trabecular number (Tb. N), and cortical cross-sectional area (CSA) are reported in the **Table 1**.

<sup>a</sup> $p < 0.05$  vs. Sham-Ve (n = 12 mice/group). Data are represented as mean  $\pm$  SEM.

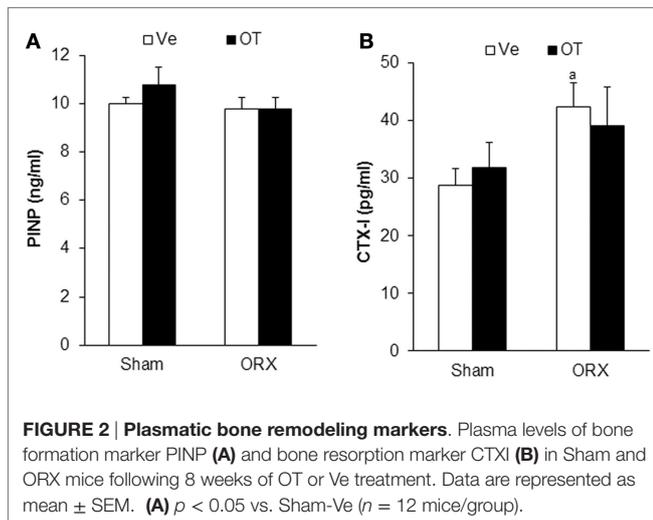
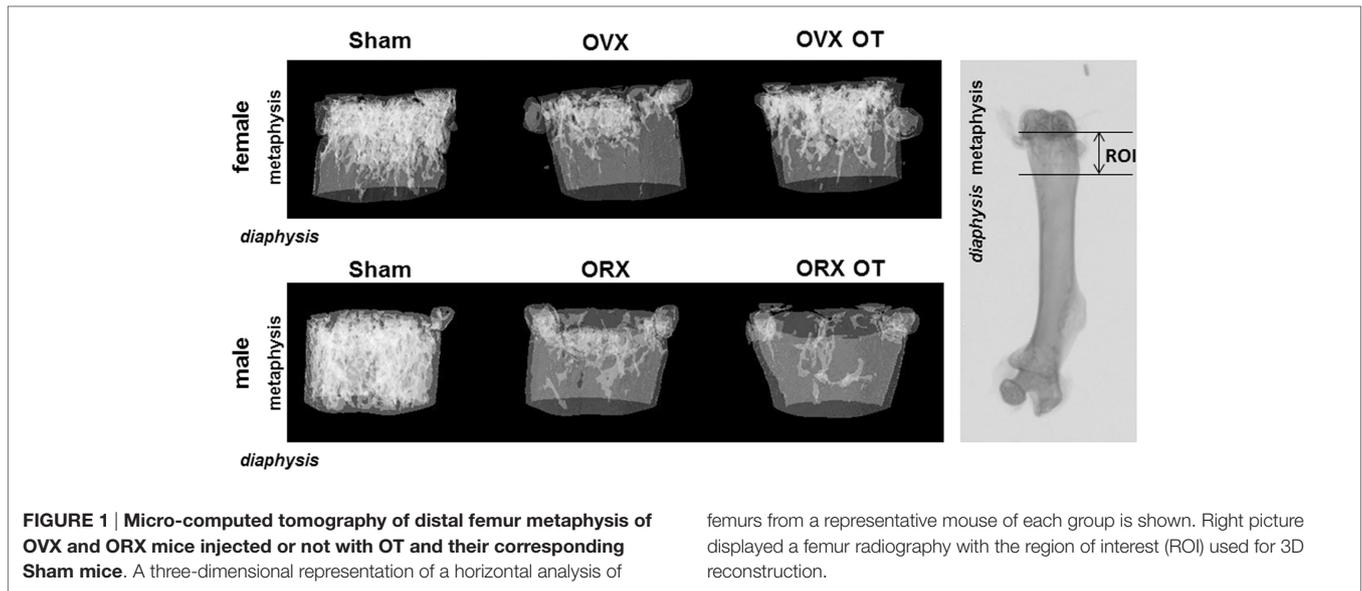
## Results

### Oxytocin Treatment does not Restore Bone Parameters in Orchidectomized Mice

We analyzed bone parameters upon OT treatment in orchidectomized (ORX) and ovariectomized (OVX) mice, animal models for male and female osteoporosis, respectively. Two weeks post-surgery, controls (male or female Sham) and ORX or OVX mice were daily injected with OT at a 1 mg/kg dose or vehicle (Ve, NaCl 0.9%). After 8 weeks of treatment, trabecular and cortical bone parameters were analyzed using high-resolution micro-computed tomography (MicroCT). We confirmed our previous observations showing that OT treatment restored bone architecture of OVX mice (14, 17). As expected in ORX mice, orchidectomy induced a dramatic trabecular bone loss evidenced by the significant decline in BV/TV, trabecule thickness (Tb. Th.), and trabecule number (Tb. N.) as well as by an increase in the trabecule spacing (Tb. Sp.) compared to the Sham group (**Table 2**). In a similar way, cortical bone was also affected upon orchidectomy as depicted by the significant decrease of mid-diaphysis cortical CSA as shown in **Table 2**. In contrast to OVX mice, none of both cortical and trabecular parameters were restored in ORX mice upon OT treatment for 8 weeks as observed in **Figure 1**. In parallel, we quantified various bone remodeling markers in plasma from Sham and ORX mice treated with OT or vehicle. Regarding bone formation, neither orchidectomy nor OT injection modified its activity according to the plasma level of PINP (**Figure 2A**). In terms of bone resorption, we measured a significant increase in the level of CTXI in ORX mice, which was not significantly modified upon OT treatment (**Figure 2B**).

### OT Treatment does not Significantly Impact the Expression of Bone Molecular Markers *In vivo*

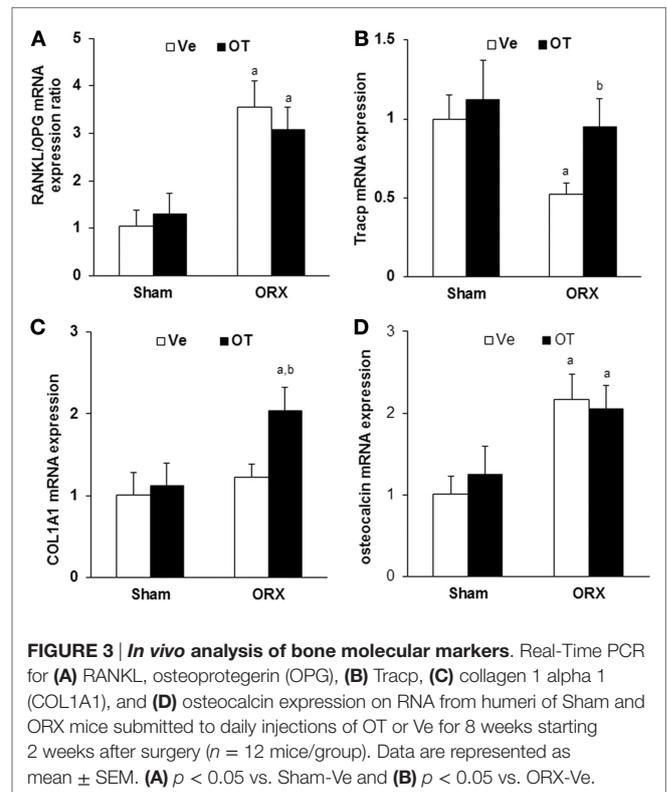
Next, we analyzed the expression of bone remodeling markers *in vivo* to address the effect of OT at the molecular level. We quantified mRNA expression using real-time PCR and mRNAs isolated from total long bones of Sham and ORX mice treated with vehicle or OT. We observed that orchidectomy induced a threefold increase of RANKL/OPG mRNA ratio, indicating an increase of pro-osteoclastogenic signaling. RANKL/OPG ratio was not affected by OT treatment and was maintained at a high level compared to the Sham group (**Figure 3A**). Regarding bone



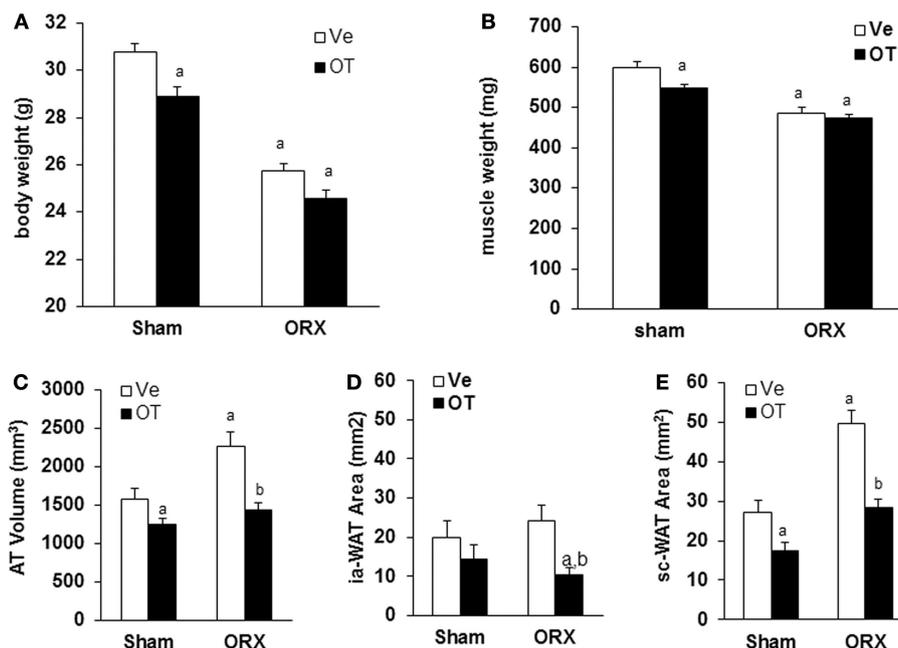
resorption, mRNA expression of the osteoclastogenic marker Tracp decreased upon orchidectomy and was normalized after OT treatment (Figure 3B). As expected, similar observations were obtained for OVX mice [data not shown (17)]. We measured the expression of osteoblast markers such as collagen 1 alpha 1 (COL1A1) and osteocalcin. COL1A1 mRNA expression was not affected by orchidectomy but was induced by OT treatment in the ORX group (Figure 3C). Osteocalcin expression was induced upon orchidectomy and was not restored in ORX-OT mice group (Figure 3D). Opposite data were obtained for female mice, a decrease in COL1A and osteocalcin mRNA upon ovariectomy that were restored upon OT treatment [data not shown (17)].

### Oxytocin Treatment does not Normalize Orchidectomy-Induced Body Weight Loss

Then, we analyzed body weight and body composition in mice upon OT or Ve treatment for 8 weeks. We previously observed



that, in OVX mice displaying an increase of both body weight and fat mass, OT was able to restore both parameters to levels of the control Sham group [data not shown (17)]. As previously described (21), ORX mice displayed a dramatic loss of total body weight ( $\approx 5$  g) mainly due to a decrease in lean mass, especially in muscle mass, in the absence of testosterone (Figure 4). However, daily OT injections for 8 weeks did not restore body weight and muscle mass compared to Sham-Ve mice (Figure 4).



**FIGURE 4 | Effects of OT on body weight, muscle weight, and fat mass in ORX mice.** Sham and ORX mice were submitted to daily injections of OT or Ve for 8 weeks starting 2 weeks after surgery. **(A)** Body and **(B)** muscle weights were measured at the end of treatment. **(C)** The volume of white adipose tissue between lumbar vertebra 1 (L1) and the caudal vertebra 4 (C4)

was measured using micro-computed tomography. **(D,E)** Intra-abdominal and subcutaneous adipose tissue areas were measured on one section at the L4/L5 (lumbar vertebra 4 and 5) junction level ( $n = 12$  mice/group). Data are represented as mean  $\pm$  SEM. **(A)**  $p < 0.05$  vs. Sham-Ve and **(B)**  $p < 0.05$  vs. ORX-Ve.

## Oxytocin Treatment Normalizes the Increase of Orchidectomy-Induced Fat Mass

As we showed that the OT treatment reversed ovariectomy-induced fat mass gain, we aimed to know whether this is also the case for male mice. Using micro-computed tomography, we quantified adipose tissue (AT) volume between the lumbar vertebra 1 (L1) and the caudal vertebra 4 (C4). We found a 50% increase in fat volume within this area in ORX compared to Sham mice (**Figure 4C**). Interestingly, we observed that OT restored the adipose tissue volume in ORX mice to the level of that observed in Sham-Ve mice, and a significant decrease of AT volume in Sham-OT treated mice was also observed. We next quantified the intra-abdominal and subcutaneous white adipose tissue (WAT) by measuring their respective areas on a MicroCT section at the L4/L5 junction. We observed that orchidectomy induced a significant increase of the subcutaneous WAT depot compared to the intra-abdominal depot (**Figures 4D,E**). Finally, OT treatment decreased both depots and more specifically restored the surface of subcutaneous WAT to the level observed in Sham-Ve mice. Thus, we observed that OT rescues the increase of the orchidectomy-induced WAT depot in a similar manner as that observed in female OVX mice (**Figure 5**).

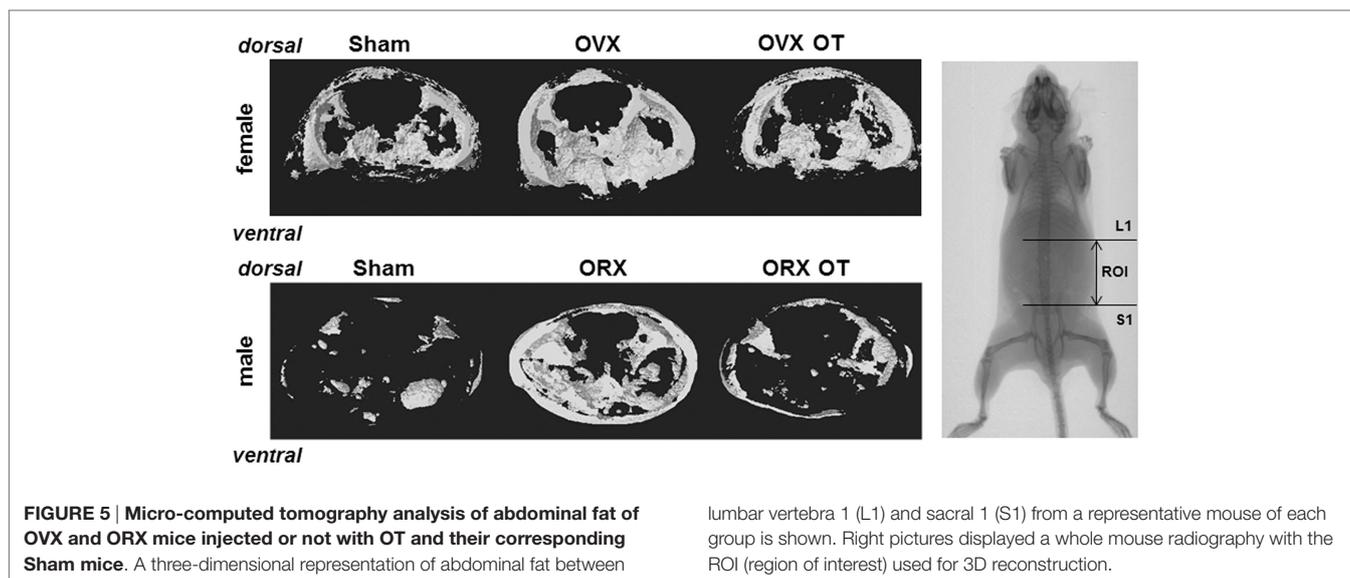
## Discussion

As for post-menopausal women or female animal models of osteoporosis (ovariectomized mice, OVX), severe bone loss occurs in men suffering from testicular dysfunction or in the corresponding

male animal models (orchidectomized mice, ORX). Osteoporosis in hypogonadal men is characterized by an elevated bone turnover, resulting in a sustained loss of bone mass (22), which is paralleled by decreased muscle mass and increased fat mass (21, 23). Orchidectomy in male rodents induces trabecular and cortical bone loss, a lowered muscle mass – assessed by lean body mass – with a concomitant gain in fat mass (24, 25). Furthermore, as observed for hypogonadal men (21), testosterone treatment prevents bone loss and body composition changes in these rodents (25).

We and others demonstrated that OT plays a key role in the physiological regulation of bone, muscle, and adipose tissue (14, 17, 19, 26). Of note, it has been established that OT pathway was involved in the physiology of bone remodeling for both genders by the analysis of OT and OTR knock-out mice (27). OT and OTR knock-out mice display an osteoporotic phenotype without any sex difference according to the authors. Using OVX mice as an animal model mimicking post-menopausal osteoporosis, we demonstrated that OT prevented and reversed osteoporosis in mice and also normalized body weight. The later effect was mainly due to a decrease in fat mass (14, 17). In this study, we addressed the role of OT on bone tissue and body composition using ORX mice, *in fine* to investigate if OT could be considered as a therapeutic treatment option for male osteoporosis at the expense of testosterone.

Our study demonstrates that daily injection of OT in ORX mice did not restore the osteoporotic phenotype. We assume that this difference regarding the regulation of bone remodeling and



homeostasis upon gonadectomy in male and female, could be due to differences in the physiopathological process of osteoporosis induced either by testis or ovary removal. Actually, if the phenotypical outcome remains the same for both genders, it is known that the decline of the hormonal level displays some common and some different features depending if it occurs in male or female. On one hand, it is known that one of the first consequences of gonads removal is an increase of the osteoclastogenic potential of bone marrow cells leading to an increase of bone resorption (28). But on the other hand, if the periosteal bone formation rate is increased in OVX rats, there is a decrease in this periosteal bone formation rate in ORX rats (29).

Estrogens (i.e., estradiol) and androgens (i.e., testosterone) are sex steroid hormones displayed by the two genders, but with different levels, i.e., higher testosterone levels found in male and higher estradiol levels found in female. A proportion of estradiol originates from testosterone through its conversion by cytochrome p450 aromatase (30). Moreover, estradiol as well as testosterone treatments restored bone loss in case of hypogonadism by suppressing bone resorption (31). Despite these common features, estradiol and testosterone do not regulate bone remodeling through the same cellular pathways. It has been demonstrated that testosterone was able to regulate bone formation and resorption activity through both direct and indirect inhibitory effects on osteoblasts and osteoclasts. By contrast, estradiol targets only osteoclast formation using a non-direct pathway through osteoblastic cells (32). We observed in our previous work that OT regulates bone remodeling in OVX mice through an effect on osteoblasts (14, 17). This observation might explain that OT can replace estrogens in OVX mice and rescue the osteoporotic phenotype but not in ORX mice for which androgens are missing. However, estrogens are able to rescue the osteoporotic phenotype of ORX mice (33) whereas OT that can substitute for estrogen in OVX mice is unable to do so in male mice. This last discrepancy could be explained by the fact that OT and its receptor are regulated by estrogens and thus are located downstream in the estrogen signaling pathway. As a

whole, these observations may lead to understand why OT can compensate for the estrogen defect in OVX regarding the bone phenotype whereas it is inefficient in the case of ORX mice that are lacking testosterone.

Androgens may not only impact muscle size or weight but also functional aspects such as muscle fiber type composition. In this way, testosterone treatment promotes lean mass as well as myofiber hypertrophy (34), and prevents the orchidectomy-induced shift of type I slow oxidative toward type IIa fast oxidative muscle fibers in slow muscle (35). Thus, testosterone regulates both the mass and function of skeletal muscle in adults. OT plasma levels and muscle stem cells OTR levels dramatically decline with age and demonstrate that OT is required for skeletal muscle tissue regeneration and homeostatic maintenance (19). In this recent work, Elabd and collaborators showed that acute OT treatment restored muscle regeneration in old mice, while OT signaling attenuation altered muscle regeneration in young mice (19). This effect seems to be triggered directly through muscle stem cell function. In other words, old mice deficient for OT displayed sarcopenia features. It is interesting to note that muscle mass declines with age in OT *knock-out* mice. The resistance of young mice could be related to high-testosterone levels, and the phenotype may be associated to the decrease in testosterone levels found in older mice (19). In our study, ORX mice displayed an expected loss of muscle mass, which was not rescued by OT treatment. This could be due to the interrelation between OT and testosterone pathways and can explain the inability of OT to rescue muscle mass decrease in ORX mice where testosterone level is very low.

In contrast to bone and muscle, OT treatment reversed orchidectomy-induced fat gain. It is accepted that hypogonadism induces adiposity in male, but the specific relation with visceral (intra-abdominal) adiposity is less documented compared to female mice (36). Indeed, visceral adiposity is a well-known risk factor for hypogonadism in male, due to the fact that this tissue is a key location for conversion of testosterone into estradiol, but the direct involvement of androgens on visceral fat development is

not clear yet (37). Under physiological conditions, visceral adipose tissue mass is two times higher in male compared to female, independently of hypogonadism but correlated with physiological levels of estradiol (38). After menopause, and not andropause, this visceral fat mass increases dramatically (38). These data clearly link estrogens, and not androgens, to the development of this adipose tissue. OT treatment induces a decrease in subcutaneous and intra-abdominal adipose tissue of OVX and ORX mice, with a higher effect on intra-abdominal adipose tissue in female due to the fact that this tissue is specifically impacted after ovariectomy. Thus, it is tempting to hypothesize that this OT effect is mediated via an estrogen dependent pathway.

In conclusion, our present work associated with our previous published studies, demonstrated that OT is able to reverse features of hypogonadism directly related to estrogen pathway, but OT is not able to compensate the decline in androgen levels. OT seems to be an interesting alternative to estrogens treatment in post-menopausal women, and due to its important function in adiposity and in muscle regeneration, OT could be proposed in association with efficient testosterone treatment in men upon

hypogonadism. Moreover, this kind of associated treatment could be very interesting in men displaying a defect in estrogen synthesis due to aromatase activity deficiency (39).

## Author Contributions

EA and DP designed the work. GB, MD, SB, and DP performed experiments. GB, CR, MS, DH, EA, and DP analyzed and interpreted data. All the authors were involved in manuscript writing and have approved the final version.

## Acknowledgments

The authors kindly acknowledge the IRCAN Animal core facilities, and Octalia Technologies for edition of the manuscript. This work was supported by CNRS (Centre National de la Recherche Scientifique), Fondation pour la Recherche Médicale (grant DVO20081013470), and the Conseil Général des Alpes Maritimes (CG06). GB was supported by “CNRS service Partenariat et Valorisation.”

## References

1. Cauley JA. Estrogen and bone health in men and women. *Steroids* (2014). doi:10.1016/j.steroids.2014.12.010
2. Giusti A, Bianchi G. Treatment of primary osteoporosis in men. *Clin Interv Aging* (2014) **10**:105–15. doi:10.2147/CIA.S44057
3. Raisz LG, Rodan AG. Pathogenesis of osteoporosis. *Endocrinol Metab Clin North Am* (2003) **32**(1):15–24. doi:10.1016/S0889-8529(02)00055-5
4. Cooper C, Campion G, Melton JL III. Hip fractures in the elderly: a world-wide projection. *Osteoporos Int* (1992) **2**(6):285–9. doi:10.1007/BF01623184
5. Cunningham GR. Andropause or male menopause? Rationale for testosterone replacement therapy in older men with low testosterone levels. *Endocr Pract* (2013) **19**(5):847–52. doi:10.4158/EP13217.RA
6. Daniell HW. Osteoporosis after orchiectomy for prostate cancer. *J Urol* (1997) **157**(2):439–44. doi:10.1016/S0022-5347(01)65165-6
7. Chan GK, Duque G. Age-related bone loss: old bone, new facts. *Gerontology* (2002) **48**(2):62–71. doi:10.1159/000048929
8. Raisz LG. Pathogenesis of osteoporosis: concepts, conflicts, and prospects. *J Clin Invest* (2005) **115**(12):3318–25. doi:10.1172/JCI27071
9. Karsenty G, Wagner FE. Reaching a genetic and molecular understanding of skeletal development. *Dev Cell* (2002) **2**(4):389–406. doi:10.1016/S1534-5807(02)00157-0
10. Seeman E, Delmas DP. Bone quality – the material and structural basis of bone strength and fragility. *N Engl J Med* (2006) **354**(21):2250–61. doi:10.1056/NEJMra053077
11. Rodan GA, Martin JT. Therapeutic approaches to bone diseases. *Science* (2000) **289**(5484):1508–14. doi:10.1126/science.289.5484.1508
12. Teitelbaum SL. Bone resorption by osteoclasts. *Science* (2000) **289**(5484):1504–8. doi:10.1126/science.289.5484.1504
13. Rosen CJ, Bouxsein LM. Mechanisms of disease: is osteoporosis the obesity of bone? *Nat Clin Pract Rheumatol* (2006) **2**(1):35–43. doi:10.1038/nprheum0070
14. Elabd C, Basillais A, Beaupied H, Breuil V, Wagner N, Scheideler M, et al. Oxytocin controls differentiation of human mesenchymal stem cells and reverses osteoporosis. *Stem Cells* (2008) **26**(9):2399–407. doi:10.1634/stemcells.2008-0127
15. Breuil V, Amri EZ, Panaia-Ferrari P, Testa J, Elabd C, Albert-Sabonnadière C, et al. Oxytocin and bone remodelling: relationships with neurohypothalamic hormones, bone status and body composition. *Joint Bone Spine* (2011) **78**(6):611–5. doi:10.1016/j.jbspin.2011.02.002
16. Breuil V, Panaia-Ferrari P, Fontas E, Roux C, Kolta S, Eastell R, et al. Oxytocin, a new determinant of bone mineral density in post-menopausal women: analysis of the OPUS cohort. *J Clin Endocrinol Metab* (2014) **99**(4):E634–41. doi:10.1210/jc.2013-4126
17. Beranger GE, Pisani DF, Castel J, Djedaini M, Battaglia S, Amiaud J, et al. Oxytocin reverses ovariectomy-induced osteopenia and body fat gain. *Endocrinology* (2014) **155**(4):1340–52. doi:10.1210/en.2013-1688
18. Smith JG, Merrill CD. Oxytocin for induction of labor. *Clin Obstet Gynecol* (2006) **49**(3):594–608. doi:10.1097/00003081-200609000-00019
19. Elabd C, Cousin W, Upadhyayula P, Chen RY, Chooljian MS, Li J, et al. Oxytocin is an age-specific circulating hormone that is necessary for muscle maintenance and regeneration. *Nat Commun* (2014) **5**:4082. doi:10.1038/ncomms5082
20. Pisani DF, Djedaini M, Beranger GE, Elabd C, Scheideler M, Ailhaud G, et al. Differentiation of human adipose-derived stem cells into “brite” (brown-in-white) adipocytes. *Front Endocrinol* (2011) **2**:87. doi:10.3389/fendo.2011.00087
21. Katznelson L, Finkelstein JS, Schoenfeld DA, Rosenthal DI, Anderson EJ, Klubanski A. Increase in bone density and lean body mass during testosterone administration in men with acquired hypogonadism. *J Clin Endocrinol Metab* (1996) **81**(12):4358–65. doi:10.1210/jc.81.12.4358
22. Riggs BL, Parfitt MA. Drugs used to treat osteoporosis: the critical need for a uniform nomenclature based on their action on bone remodeling. *J Bone Miner Res* (2005) **20**(2):177–84. doi:10.1359/JBMR.041114
23. Bhasin S, Storer TW, Berman N, Yarasheski KE, Clevenger B, Phillips J, et al. Testosterone replacement increases fat-free mass and muscle size in hypogonadal men. *J Clin Endocrinol Metab* (1997) **82**(2):407–13. doi:10.1210/jcem.82.2.3733
24. Erben RG, Eberle J, Stahr K, Goldberg M. Androgen deficiency induces high turnover osteopenia in aged male rats: a sequential histomorphometric study. *J Bone Miner Res* (2000) **15**(6):1085–98. doi:10.1359/jbmr.2000.15.6.1085
25. Vanderschueren D, Vandenput L, Boonen S, Van Herck E, Swinnen JV, Bouillon R. An aged rat model of partial androgen deficiency: prevention of both loss of bone and lean body mass by low-dose androgen replacement. *Endocrinology* (2000) **141**(5):1642–7. doi:10.1210/en.141.5.1642
26. Maejima Y, Iwasaki Y, Yamahara Y, Kodaira M, Sedbazar U, Yada T. Peripheral oxytocin treatment ameliorates obesity by reducing food intake and visceral fat mass. *Aging (Albany NY)* (2011) **3**(12):1169–77.
27. Tamma R, Colaizzi G, Zhu LL, DiBenedetto A, Greco G, Montemurro G, et al. Oxytocin is an anabolic bone hormone. *Proc Natl Acad Sci U S A* (2009) **106**(17):7149–54. doi:10.1073/pnas.0901890106
28. Most W, van der Wee-Pals L, Ederveen A, Papapoulos S, Löwik C. Ovariectomy and orchidectomy induce a transient increase in the osteoclastogenic potential of bone marrow cells in the mouse. *Bone* (1997) **20**(1):27–30. doi:10.1016/S8756-3282(96)00309-2
29. Turner RT, Wakley KG, Hannon SK. Differential effects of androgens on cortical bone histomorphometry in gonadectomized male and female rats. *J Orthop Res* (1990) **8**(4):612–7. doi:10.1002/jor.1100080418

30. Simpson ER, Mahendroo MS, Means GD, Kilgore MW, Hinshelwood MM, Graham-Lorence S, et al. Aromatase cytochrome P450, the enzyme responsible for estrogen biosynthesis. *Endocr Rev* (1994) **15**(3):342–55. doi:10.1210/er.15.3.342
31. Matsumoto C, Inada M, Toda K, Miyaura C. Estrogen and androgen play distinct roles in bone turnover in male mice before and after reaching sexual maturity. *Bone* (2006) **38**(2):220–6. doi:10.1016/j.bone.2005.08.019
32. Michael H, Härkönen PL, Väänänen HK, Hentunen TA. Estrogen and testosterone use different cellular pathways to inhibit osteoclastogenesis and bone resorption. *J Bone Miner Res* (2005) **20**(12):2224–32. doi:10.1359/JBMR.050803
33. Onoe Y, Miyaura C, Ito M, Ohta H, Nozawa S, Suda T. Comparative effects of estrogen and raloxifene on B lymphopoiesis and bone loss induced by sex steroid deficiency in mice. *J Bone Miner Res* (2000) **15**(3):541–9. doi:10.1359/jbmr.2000.15.3.541
34. Sinha-Hikim I, Artaza J, Woodhouse L, Gonzalez-Cadavid N, Singh AB, Lee MI, et al. Testosterone-induced increase in muscle size in healthy young men is associated with muscle fiber hypertrophy. *Am J Physiol Endocrinol Metab* (2002) **283**(1):E154–64. doi:10.1152/ajpendo.00502.2001
35. Chambon C, Duteil D, Vignaud A, Ferry A, Messaddeq N, Malivindi R, et al. Myocytic androgen receptor controls the strength but not the mass of limb muscles. *Proc Natl Acad Sci U S A* (2010) **107**(32):14327–32. doi:10.1073/pnas.1009536107
36. Katznelson L, Rosenthal DI, Rosol MS, Anderson EJ, Hayden DL, Schoenfeld DA, et al. Using quantitative CT to assess adipose distribution in adult men with acquired hypogonadism. *AJR Am J Roentgenol* (1998) **170**(2):423–7. doi:10.2214/ajr.170.2.9456958
37. Mammi C, Calanchini M, Antelmi A, Cinti F, Rosano GM, Lenzi A, et al. Androgens and adipose tissue in males: a complex and reciprocal interplay. *Int J Endocrinol* (2012) **2012**:789653. doi:10.1155/2012/789653
38. Blouin K, Boivin A, Tchernof A. Androgens and body fat distribution. *J Steroid Biochem Mol Biol* (2008) **108**(3–5):272–80. doi:10.1016/j.jsbmb.2007.09.001
39. Carani C, Qin K, Simoni M, Faustini-Fustini M, Serpente S, Boyd J, et al. Effect of testosterone and estradiol in a man with aromatase deficiency. *N Engl J Med* (1997) **337**(2):91–5. doi:10.1056/NEJM199707103370204

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2015 Beranger, Djedaini, Battaglia, Roux, Scheideler, Heymann, Amri and Pisani. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.