

SUPPLEMENTARY DATA

**Supplementary Table 1.** Primer sequences used for quantitative real time PCR analysis.

Gene	Forward Primer	Reverse Primer	Gene Bank
<i>Acadl</i>	GGCTTGCTTGGCATCAACA	CGCAGAGAAACATGG	NM_007381.3
<i>Acadm</i>	AGAGCTCTAGACGAAGCCACGA	GAGTTCAACCTTCATCGCCATT	NM_007382.4
<i>Agrp</i>	CCCAGGTCTAAGTCTGAATGGC	TTCTGTGGATCTAGCACCTCCG	NM_007427.2
<i>Cart</i>	AACGCATTCCGATCTACGAGA	ACAGTCACACAGCTTCCCGAT	NM_013732.6
<i>Ccl2</i>	TCTCTCTTCCTCCACCACCATG	GCGTTAACTGCATCTGGCTGA	NM_011333.3
<i>Ccl3</i>	GCGGCTGATGATTGGACAA	ATCTCCAGCTCGAGCAATGG	NM_011337.2
<i>Cnr1</i>	GGCACCTCTTTCTCAGTCACGT	GGTGATGGTACGGAAGGTGGTA	NM_007726.3
<i>Cd36</i>	GGACATACTTAGATGTGGAACCCATA	TGTTGACCTGCAGTCGTTTTG	NM_001159558.1
<i>Cpt1a</i>	CACCAACGGGCTCATCTTCT	CCTTCTATCGAATTTGCTCTGGTT	NM_013495.2
<i>Cpt1b</i>	TGGGACTGGTCGATTGCAT	AGTGGCCATACCTTTCCGG	NM_009948.2
<i>Crh</i>	ATTTACACACGCAGTCGGTAT	AAGCCCAGGAATGAAGTCCA	NM_205769.2
<i>Emr1</i>	GGATGGATAATGGCTGCTGGT	CCAGGCAAGGAGGACAGAGTTT	NM_010130.4
<i>Hprt</i>	CAGTCCCAGCGTCGTGATTA	AGCAAGTCTTTCAGTCCTGTG	NM_013556
<i>Il6</i>	TCTGCAAGAGACTTCCATCCAGT	TGTCACCAGCATCAGTCCCA	NM_031168.1
<i>Lep</i>	CGGAGAGCCACGCAACTT	CAGCCCCGGGCAGTTT	NM_146146.2
<i>Lpl</i>	GGACTGAGGATGGCAAGCAA	GCCACTGTGCCGTACAGAGA	NM_008509.2
<i>Mch</i>	TTCAAAGAACACAGGCTCCAAA	ACTCAGCATTCTGAACTCCATTCTC	NM_029971.2
<i>Npy</i>	ACTCCGCTCTGCGACACTACAT	GCGTTTTCTGTGCTTTCCTTCA	NM_023456.2
<i>Npy1r</i>	CTGCAGTATTTTCGGCCACTCT	ACTGTCCCAGATCTTGTCCATC	NM_010934
<i>Pik3ca</i>	ACCTCAGGCTTGAAGAGTGTCG	CCGTAAGTCGTGCCATTTTTA	NM_008839.2
<i>Pik3r1</i>	AACCGAAACAAAGCGGAGAA	TTGACTTCGCCGTCTACCACT	NM_001024955.1
<i>Pomc</i>	GCCACTGAACATCTTTGTCCC	AATCTCGGCATCTTCCACGT	NM_008895.3
<i>Ppara</i>	GTCACACAATGCAATTCGCTTT	TTTGCTTTTTTCAGATCTTGGA	NM_011144.6
<i>Pparg1c</i>	ACAGCCGTAGGCCAGGTAC	GCCTTTCGTGCTCATAGGCTT	NM_008904.2
<i>Stat3</i>	ATCTGTGTGACACCAACGACCT	TCAGCACCTTCACCGTTATTTT	NM_213659.2
<i>Tnf</i>	ATGAGAAGTTCCCAAATGGCCT	GGGTCTGGGCCATAGAACTGA	NM_013693.2
<i>Trh</i>	GTGCCAACCAAGACAAGGAT	TTCTTCCCAGCTTCTTTGGA	NM_009426.2

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**Supplementary Table 2.** Sequences of PCR primers used for EpiTYPER methylation analysis and product size of each amplicon.

Amplicon	Primer	Sequence	Product size (bp)
Ppara_#3	forward	aggaagagagGGTAGAGTTTTAGTGTTGAGTTGGA	491
	reverse	cagtaatac gactcactataggagaaggctATAAAAAA ACTACCCAAAATCACCC	
Ppara_#4	forward	aggaagagagGGGTGATTTTGGGTAGTTTTTTAT	445
	reverse	cagtaatac gactcactataggagaaggctCCAACCCCTAAACACCTAAA ACT	
Ppara_#7	forward	aggaagagagTG TAGTTTTAGGTGTTTAGGGGTTG	491
	reverse	cagtaatac gactcactataggagaaggctAAATCAATCTATCAAAAAACCTCCA	
Pik3r1_#1	forward	aggaagagagGTAGTTTTTGTTTTGGGAAGGAA	346
	reverse	cagtaatac gactcactataggagaaggctACCAA ACTAAACCATAATAATCCCC	
Pik3r1_#8	forward	aggaagagagGGGGATTATTATGGTTTAGTTGG	354
	reverse	cagtaatac gactcactataggagaaggctAACAA ACTACCAACTCCCAAT	
Pik3r1_#9	forward	aggaagagagGATTGGGAGTTGGTAGTTTGT	183
	reverse	cagtaatac gactcactataggagaaggctTAAAACCCAA ACTAAACAAAAAAA	
Pik3r1_#10	forward	aggaagagagTTTTTTTGTTTAGTTGGGTTTTA	358
	reverse	cagtaatac gactcactataggagaaggctAACTCCTAAACCTTAATAACCTCC	
Pik3r1_#11	forward	aggaagagagGGGAGGGTTATTAAGGTTTAGGAG	385
	reverse	cagtaatac gactcactataggagaaggctACAAACAAA ACCAAAAATTACAAAA	
Ppargc1a_#1	forward	aggaagagagTTTATGTTATTTTATATAGAGTTTTGGTTG	356
	reverse	cagtaatac gactcactataggagaaggctAACCAA ATTTTCCTTTTCTTTCTTC	
Ppargc1a_#3	forward	aggaagagagAAGTTATTA AAAAGTAGGTTGGGTTGTT	489
	reverse	cagtaatac gactcactataggagaaggctCCTCAAACACTCCTAATAAAAAAAA	
Ppargc1a_#4	forward	aggaagagagTTTTATTATTGTTTATGGTGTGGTT	373
	reverse	cagtaatac gactcactataggagaaggctAAAA TCCCTCCTTCAATAATTCTA	
Cpt1b_#1	forward	aggaagagagAGTGAATTGGAAGTTATTGTTTGG	408
	reverse	cagtaatac gactcactataggagaaggctATACTAAA ACCACTCCCTCCCTAA	
Cpt1b_#8	forward	aggaagagagTTAGGGAAGGGAGTGTTTATAGTAT	266
	reverse	cagtaatac gactcactataggagaaggctTTCTCCACCCCAATTTAAAAATAA	
Cpt1b_#10	forward	aggaagagagTTTGGTTTTTTTGGTTTATGTTTTT	462
	reverse	cagtaatac gactcactataggagaaggctAATCTCCTATCCCATAACTCCCTAA	
Cpt1b_#12	forward	aggaagagagAAGTAAATTTGAGTTGTGAGTTGGG	488
	reverse	cagtaatac gactcactataggagaaggctCCATCCTAAA ATTTATTCACACCT	
Cpt1b_#22	forward	aggaagagagGTTGGAGTAGTAGTGGTTTTTGAGG	416
	reverse	cagtaatac gactcactataggagaaggctCCTATACTAATCCCAACTCACAAC	
Cpt1a_#5	forward	aggaagagagGAAAGATGGAGGTAATAGGGTTTT	482
	reverse	cagtaatac gactcactataggagaaggctCCAAAA ACCAACACTCATAATC	
Cpt1a_#7	forward	aggaagagagGAGATTATGAGTGTGTTGTTTTTG	364
	reverse	cagtaatac gactcactataggagaaggctTTCTTACCCTAAA AACCTCAATTT	
Cpt1a_#15	forward	aggaagagagGGTTTTTAGGGTAAGGAATGTTGTT	383
	reverse	cagtaatac gactcactataggagaaggctAAAAAAA TACCCTCTACTTCTCCA	

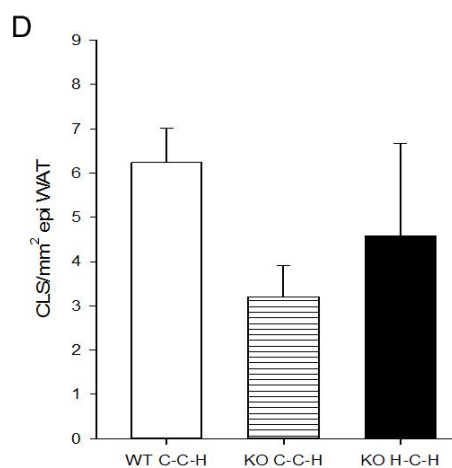
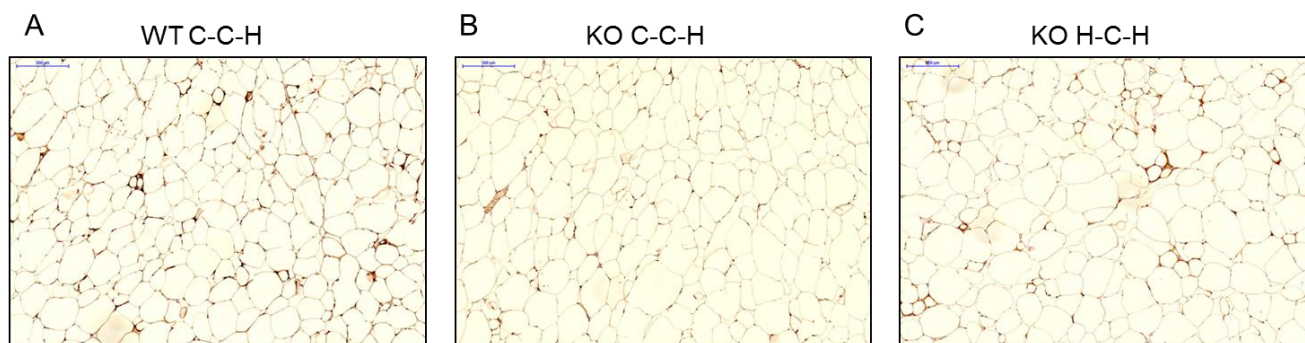
SUPPLEMENTARY DATA

**Supplementary Table 3.** Sequences of oligonucleotide used for EMSA

Name	Primer	Sequence
CG_Cpt1b+227,+224	forward	AAGCCTGGCCAACCGCCGCTGCCACCGAACC
CG_Cpt1b+227,+224	reverse	GGTTCGGTGGCAGCGGCGGTTGGCCAGGCTT
F_CG_Cpt1b+227,+224	competition	AAGCCTGGCCAACCGCCGCTGCCACCGAACC
(me)CG_Cpt1b+227,+224	forward	AAGCCTGGCCAAC-(me)CGC-(me)CGCTGCCACCGAACC
(me)CG_Cpt1b+227,+224	reverse	GGTTCGGTGGCAG-(me)CGG-(me)CGGTTGGCCAGGCTT
F_(me)CG_Cpt1b+227,+224	competition	AAGCCTGGCCAAC-(me)CGC-(me)CGCTGCCACCGAACC
CG_Cpt1b-72	forward	GGCCCATGTCCCCACGTCTTCAGGCCTGG
CG_Cpt1b-72	reverse	CCAGGCCTGAAGGACGTGGGGACATGGGCC
F_CG_Cpt1b-72	competition	GGCCCATGTCCCCACGTCTTCAGGCCTGG
(me)CG_Cpt1b-72	forward	GGCCCATGTCCCCA-(me)CGTCTTCAGGCCTGG
(me)CG_Cpt1b-72	reverse	CCAGGCCTGAAGGA-(me)CGTGGGGACATGGGCC
F_(me)CG_Cpt1b-72	competition	GGCCCATGTCCCCA-(me)CGTCTTCAGGCCTGG
CG_Cpt1b-202	forward	TCCTTTTGGGGGAGCGCCTAGGGAGGGTGG
CG_Cpt1b-202	reverse	CCACCCTCCCTAGGCGCTCCCCAAAAGGA
F_CG_Cpt1b-202	competition	TCCTTTTGGGGGAGCGCCTAGGGAGGGTGG
(me)CG_Cpt1b-202	forward	TCCTTTTGGGGGAG-(me)CGCCTAGGGAGGGTGG
(me)CG_Cpt1b-202	reverse	CCACCCTCCCTAGG-(me)CGCTCCCCAAAAGGA
F_(me)CG_Cpt1b-202	competition	TCCTTTTGGGGGAG-(me)CGCCTAGGGAGGGTGG
CG_Ppara-140	forward	TGGCCCTGCGGACCCGCAGGCGGAGTGCAG
CG_Ppara-140	reverse	CTGCACTCCGCTGCGGGTCCGCAGGGCCA
F_CG_Ppara-140	competition	TGGCCCTGCGGACCCGCAGGCGGAGTGCAG
(me)CG_Ppara-140	forward	TGGCCCTGCGGACC-(me)CGCAGGCGGAGTGCAG
(me)CG_Ppara-140	reverse	CTGCACTCCGCTG-(me)CGGGTCCGCAGGGCCA
F_(me)CG_Ppara-140	competition	TGGCCCTGCGGACC-(me)CGCAGGCGGAGTGCAG
Oct1	forward	TGTCGAATGCAAATCACTAGAA
Oct1	reverse	TTCTAGTGATTTGCATTGACA
NFkB	forward	AGTTGAGGGGACTTTCCAGGC
NFkB	reverse	GCCTGGGAAAGTCCCCTCAACT

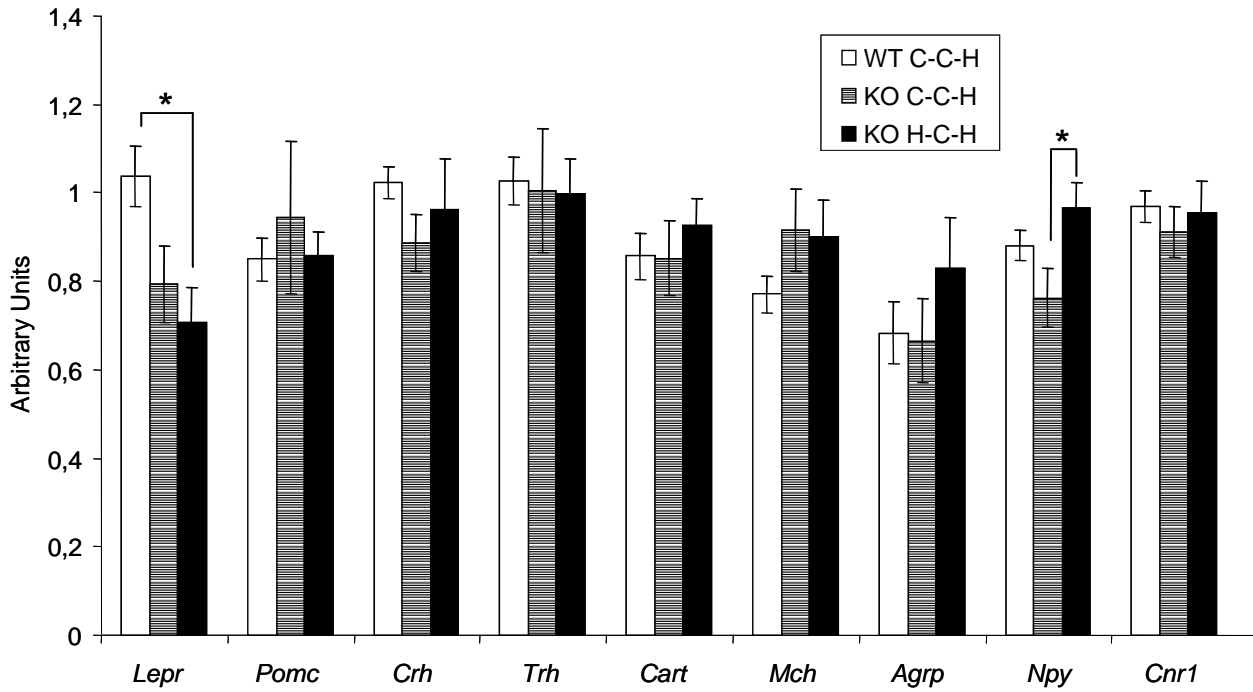
## SUPPLEMENTARY DATA

**Supplementary Figure 1.** Crown-like structures (CLS) in white adipose tissue. Tissues were stained with an F4/80 anti-mouse antibody. Crown-like structures were determined in three randomly chosen areas within the slides. Slightly fewer CLS were observed in KO C-C-H compared to WT C-C-H and KO H-C-H, respectively. However, this was not statistically significant (**D**). n=4-13 per group. CLS: Crown-like structure, WAT: White adipose tissue.



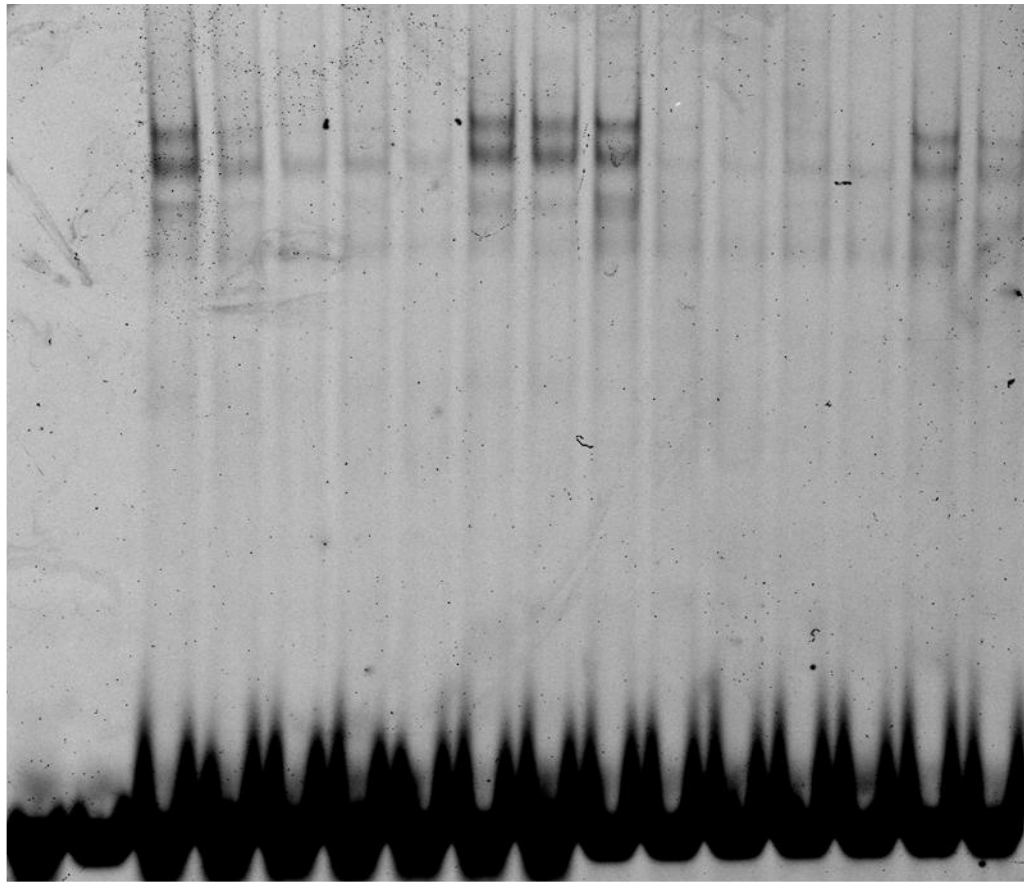
SUPPLEMENTARY DATA

**Supplementary Figure 2.** Gene expression levels in hypothalamus of leptin receptor – long form (LEPR), proopiomelanocortin (POMC), corticotropin releasing hormone (CRH), thyrotropin-releasing hormone (TRH), cocaine and amphetamine-regulated transcript (CART), melanin-concentrating hormone (MCH), agouti-related protein (AgRP), neuropeptide Y (NPY), cannabinoid receptor 1 (CNR1) and suppressor of cytokine signaling 3 (SOCS3). \*P<0.05, n=4-13 per group.



SUPPLEMENTARY DATA

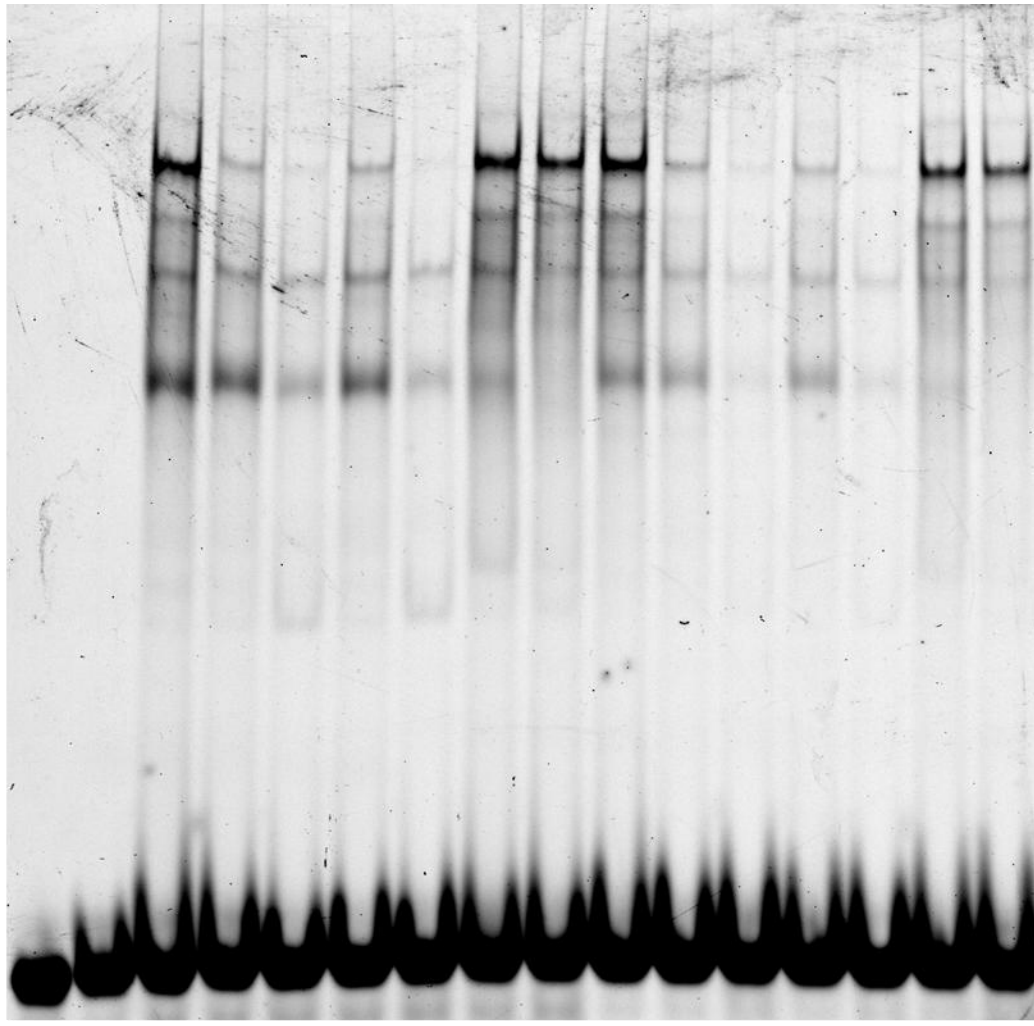
**Supplementary Figure 3.** No methylation specific formation of a protein-DNA complex at the Cpt1b CpG<sub>-72</sub> site in mouse C2C12 myoblasts. Methylated (m) and unmethylated (um) Cy5-labelled probes carrying the CpG<sub>-72</sub> site, were investigated in competition EMSAs using C2C12 mouse myoblast nuclear extracts. Lane 1 and 2 represent oligonucleotides without incubation with nuclear extract. Lane 3 and 10 represent protein-DNA complex formation at the unmethylated and methylated CpG. In lane 4, 5, 13, 14 competition with unlabeled methylated oligonucleotides was performed, whereas in lane 6, 7, 11, 12 competition with unlabeled unmethylated oligonucleotides was performed. Specificity was assured by competition experiments with unlabeled Oct1/NfκB consensus oligonucleotides (lanes 8, 9, 15, 16).



	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
Labelled Oligo	um	m	um								m					
Competitor	-	-	-	10x m	50x m	10x um	50x um	10x NfκB	50x NfκB	-	10x um	50x um	10x m	50x m	10x NfκB	50x NfκB
NE	-	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+

SUPPLEMENTARY DATA

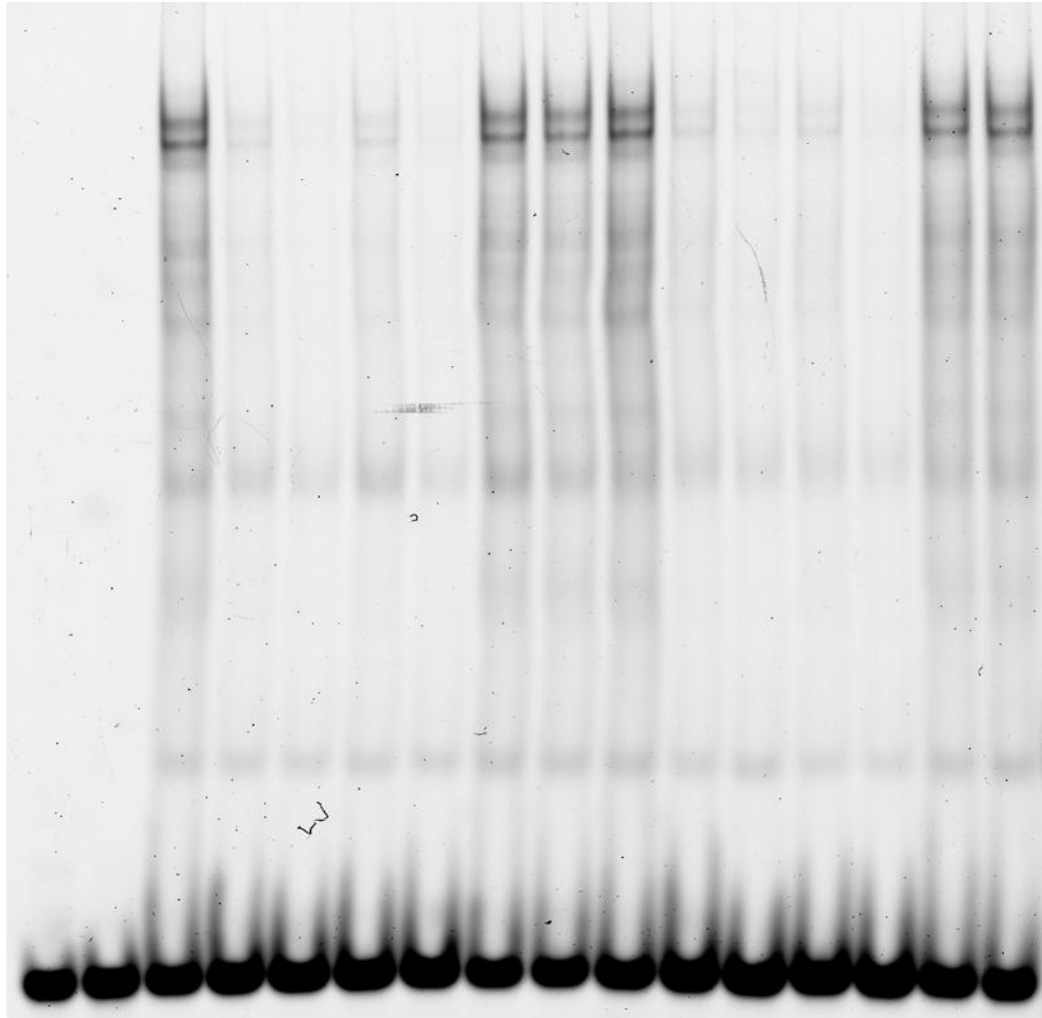
**Supplementary Figure 4.** No methylation specific formation of a protein-DNA complex at the Cpt1b CpG<sub>+227/+224</sub> site in mouse C2C12 myoblasts. Methylated (m) and unmethylated (um) Cy5-labelled probes carrying the CpG<sub>+227/+224</sub> site, were investigated in competition EMSAs using C2C12 mouse myoblast nuclear extracts. Lane 1 and 2 represent oligonucleotides without incubation with nuclear extract. Lane 3 and 10 represent protein-DNA complex formation at the unmethylated and methylated CpG. In lane 4, 5, 13, 14 competition with unlabeled methylated oligonucleotides was performed, whereas in lane 6, 7, 11, 12 competition with unlabeled unmethylated oligonucleotides was performed. Specificity was assured by competition experiments with unlabeled Oct1/NfKB consensus oligonucleotides (lanes 8, 9, 15, 16).



	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
Labelled Oligo	um	m	um						m							
Competitor	-	-	-	10x m	50x m	10x um	50x um	10x Oct1	50x Oct1	-	10x um	50x um	10x m	50x m	10x Oct1	50x Oct1
NE	-	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+

SUPPLEMENTARY DATA

**Supplementary Figure 5.** No methylation specific formation of a protein-DNA complex at the Cpt1b CpG<sub>-202</sub> site in mouse C2C12 myoblasts. Methylated (m) and unmethylated (um) Cy5-labelled probes carrying the CpG<sub>-202</sub> site, were investigated in competition EMSAs using C2C12 mouse myoblast nuclear extracts. Lane 1 and 2 represent oligonucleotides without incubation with nuclear extract. Lane 3 and 10 represent protein-DNA complex formation at the unmethylated and methylated CpG. In lane 4, 5, 13, 14 competition with unlabeled methylated oligonucleotides was performed, whereas in lane 6, 7, 11, 12 competition with unlabeled unmethylated oligonucleotides was performed. Specificity was assured by competition experiments with unlabeled Oct1/NfKB consensus oligonucleotides (lanes 8, 9, 15, 16).



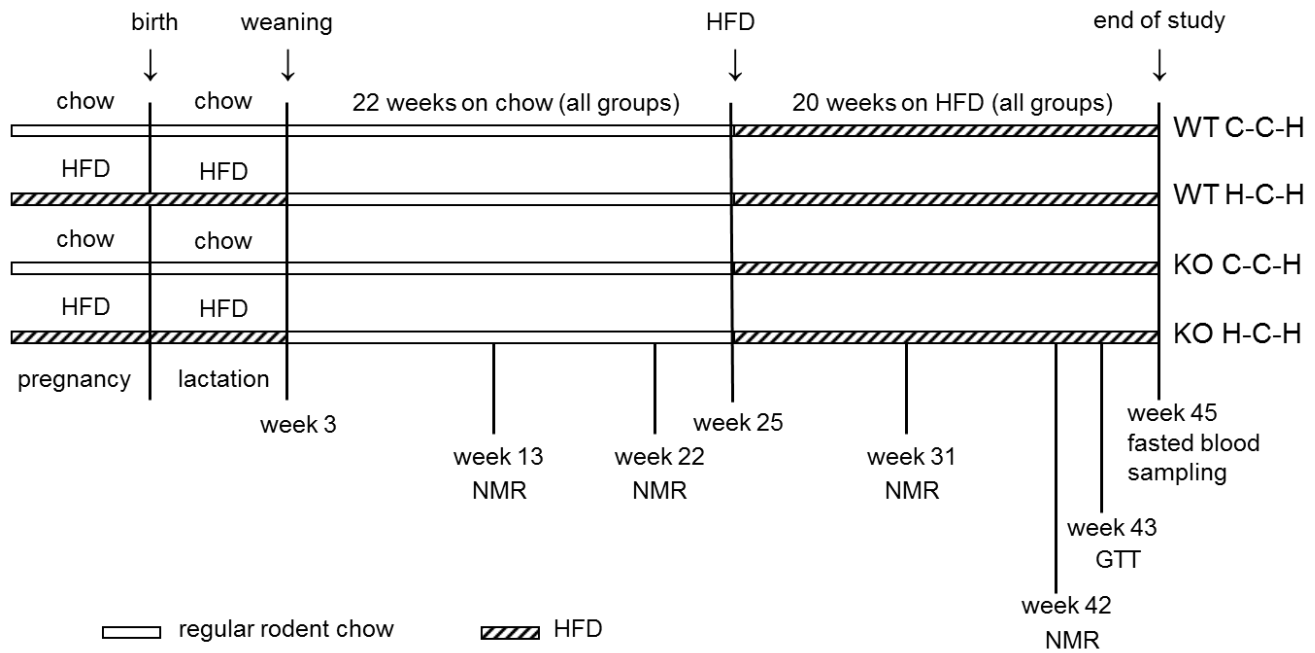
	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
Labelled Oligo	um	m	um						m							
Competitor	-	-	-	10x m	50x m	10x um	50x um	10x Oct1	50x Oct1	-	10x um	50x um	10x m	50x m	10x Oct1	50x Oct1
NE	-	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+



SUPPLEMENTARY DATA

**Supplementary Figure 6.** Schematic representation of the study including an additional group of wild type mice that received a high fat diet during pregnancy and lactation and between week 25 and 45 of life (WT H-C-H). Open bars indicate periods of food exposition with a regular rodent chow, hatched bars indicate periods of food exposition with a high fat diet. HFD: High fat diet; NMR: Nuclear magnetic resonance spectroscopy; GTT: Intra peritoneal glucose tolerance test; WT: Wild type offspring; KO: *Gipr*<sup>-/-</sup> offspring. For further details see also Figure 1 in the main text.

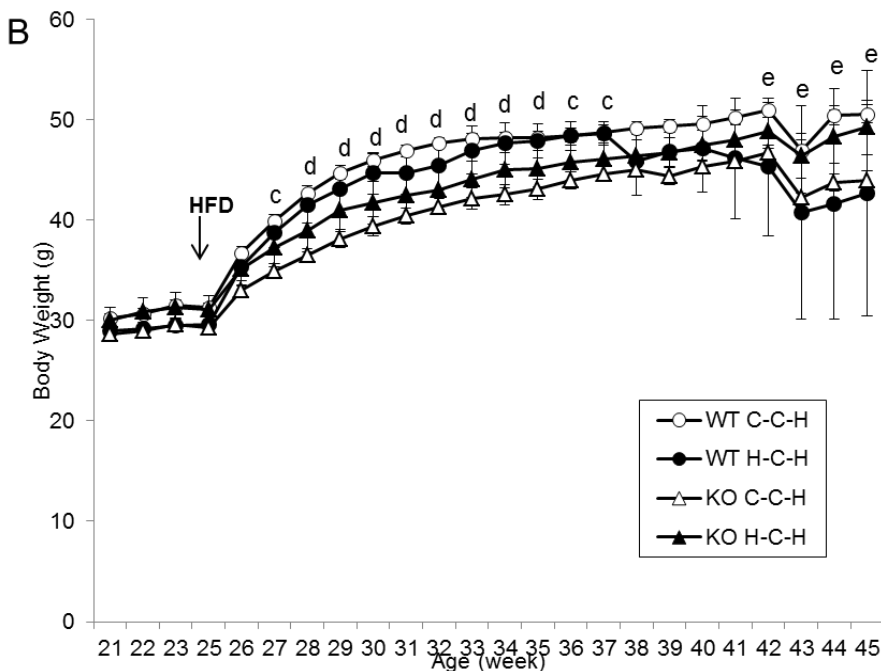
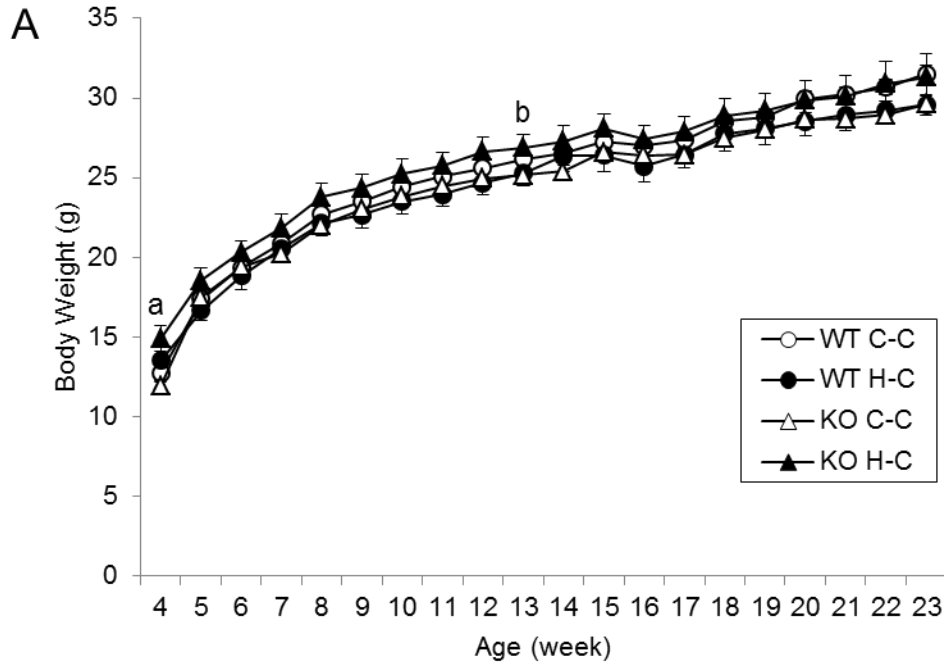
Results of key experiments of the study with these for groups are shown in Supplementary Figures 7-11. A two-way ANOVA analysis with genotype and maternal diet as the independent variables was performed. Significance was accepted at P<0.05.



SUPPLEMENTARY DATA

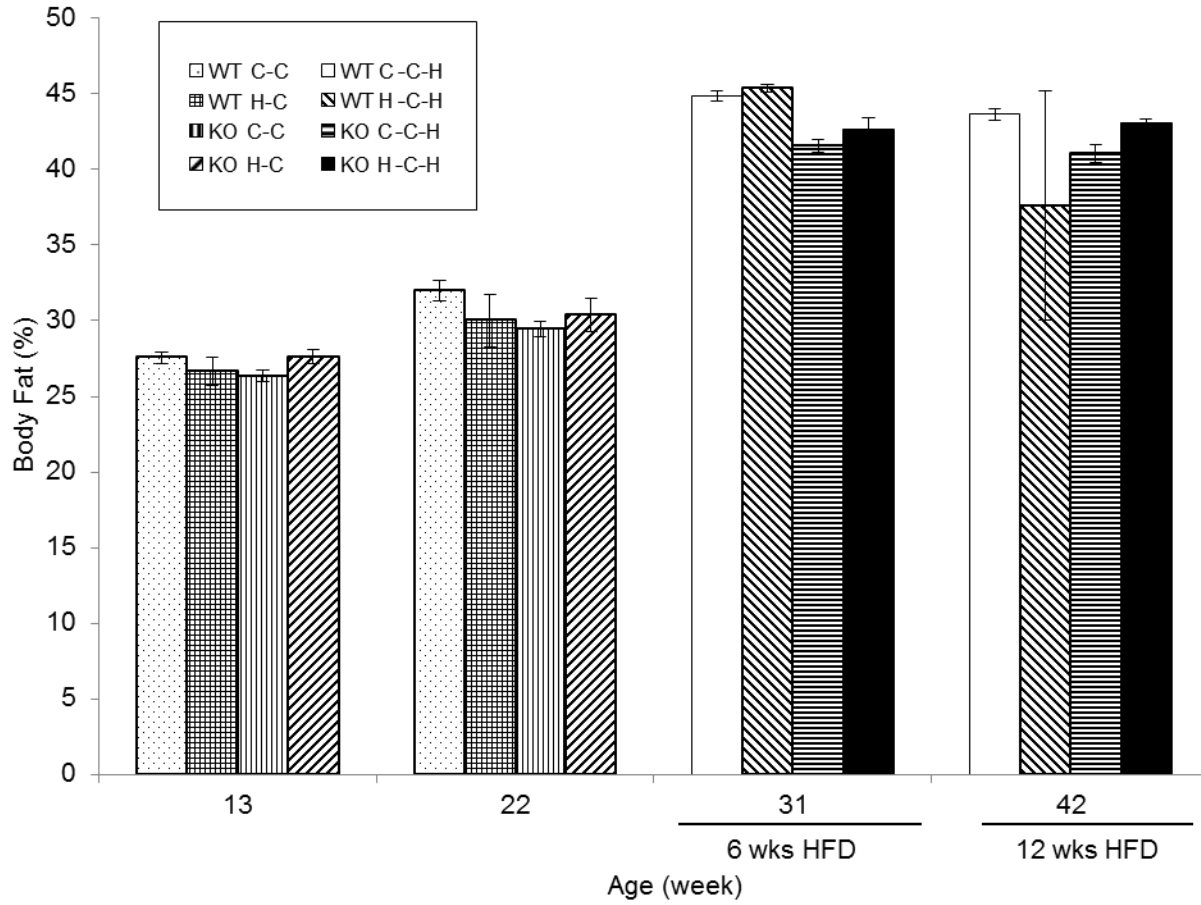
**Supplementary Figure 7. A:** Growth curve of offspring from 4 weeks until 23 weeks of age on a standard rodent chow. WT C-C: Wild type offspring exposed to a regular chow during pregnancy/ lactation and post weaning; WT H-C: Wild type offspring exposed to HFD during pregnancy/ lactation and post weaning; KO C-C: *Gipr*<sup>-/-</sup> offspring exposed to a regular chow during pregnancy/ lactation and post weaning; KO H-C: *Gipr*<sup>-/-</sup> offspring exposed to a HFD during pregnancy/ lactation and switched to a regular diet post weaning; n= 3-13 per group, a = two-way ANOVA P<0.05 for maternal diet, b = two-way ANOVA P<0.05 for interaction genotype x maternal diet.

**B:** Growth curve of offspring after a HFD was started at 25 weeks of age; n= 3-13 per group, for WT H-C-H n= 2 for week 42-45, c = two-way ANOVA P<0.05 for genotype, d = two-way ANOVA P<0.01 for genotype, e = two-way ANOVA P<0.05 for interaction genotype x maternal diet.



SUPPLEMENTARY DATA

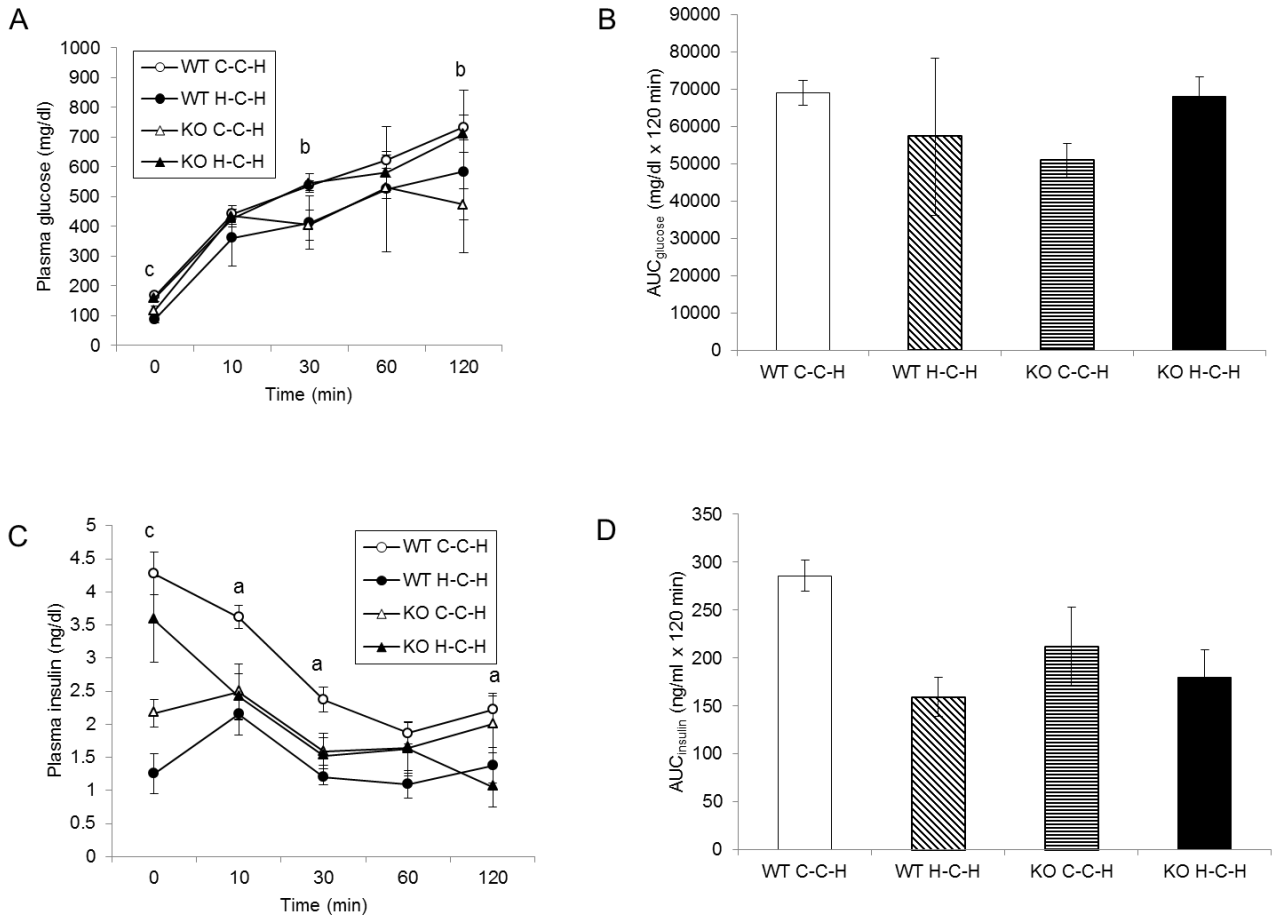
**Supplementary Figure 8.** Adiposity was measured by nuclear magnetic resonance spectroscopy, n= 3-13 per group, for WT H-C-H n=2 at week 42. Two-way ANOVA: P<0.001 for genotype at 31 weeks and P<0.01 for interaction genotype x maternal diet at 42 weeks.



SUPPLEMENTARY DATA

**Supplementary Figure 9.** Glucose tolerance test at 43 weeks of age (18 weeks on HFD). After an overnight fast, mice received an i.p. injection of 2.0 g glucose per kg body weight. **A:** Plasma glucose levels. **B:** AUC: Area under the curve for glucose, two-way ANOVA:  $P < 0.05$  for interaction genotype x maternal diet. **C:** Plasma insulin levels. **D:** AUC: Area under the curve for insulin, two-way ANOVA:  $P < 0.05$  for maternal diet.

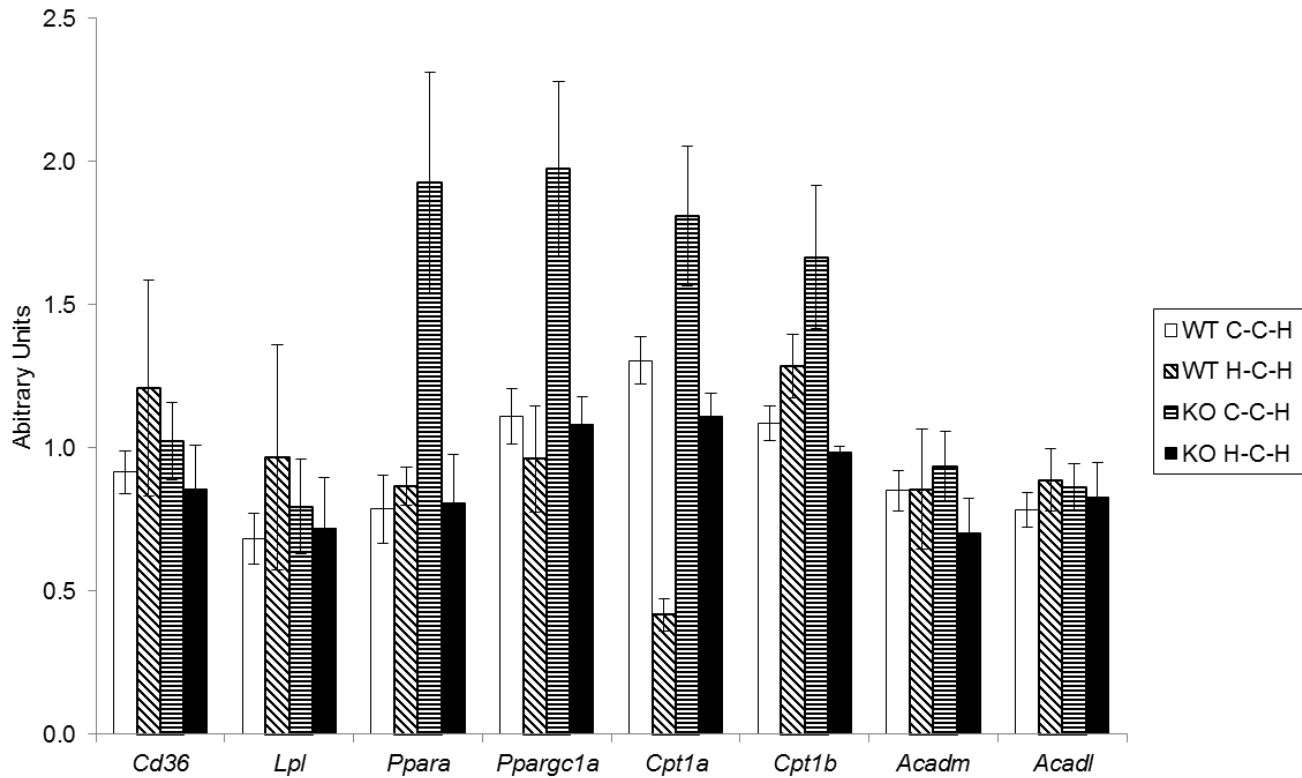
$n = 3-13$  per group, a = two-way ANOVA  $P < 0.05$  for maternal diet, b = two-way ANOVA  $P < 0.05$  for interaction genotype x maternal diet, c = two-way ANOVA  $P < 0.01$  for interaction genotype x maternal diet.



SUPPLEMENTARY DATA

**Supplementary Figure 10.** Gene expression levels in gastrocnemius muscle. *Cd36*: Cluster of differentiation 36 fatty acid transporter, *Lpl*: Lipoprotein lipase, *Ppara*: Peroxisome proliferator-activated receptor  $\alpha$ , *Ppargc1a*: Peroxisome proliferator-activated receptor gamma coactivator 1- $\alpha$ , *Cpt1a*: Carnitine palmitoyltransferase-1  $\alpha$ , *Cpt1b*: Carnitine palmitoyltransferase-1  $\beta$ , *Acadm*: Medium-chain acyl-CoA dehydrogenase, *Acadl*: Long-chain acyl-CoA dehydrogenase. n=4-13 per group; n=2 for WT H-C-H.

Two-way ANOVA: *Ppara*: P<0.05 for interaction genotype x maternal diet. *Ppargc1a*: P<0.05 for genotype and for maternal diet. *Cpt1a*: P<0.01 for genotype, P<0.001 for maternal diet and P<0.01 for interaction genotype x maternal diet. *Cpt1b*: P<0.01 for interaction genotype x maternal diet.



SUPPLEMENTARY DATA

**Supplementary Figure 11.** Gene expression levels in hypothalamus of phosphatidylinositol 3-kinase catalytic subunit p110 (*Pik3ca*), phosphatidylinositol 3-kinase regulatory subunit p85 $\alpha$  (*Pik3r1*), signal transducer and activator of transcription 3 (*Stat3*) and neuropeptide Y receptor 1 (*Npy1r*). n=4-13 per group; n=2 for WT H-C-H.

Two-way ANOVA: *Pik3r1*: P<0.05 for genotype and for maternal diet.

