# Accepted Manuscript

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PII: S0003-9861(15)30068-0

DOI: 10.1016/j.abb.2015.09.017

Reference: YABBI 7077

To appear in: Archives of Biochemistry and Biophysics

Received Date: 5 May 2015

Revised Date: 1 September 2015

Accepted Date: 17 September 2015

Please cite this article as: R. van den Berg, D.O. Mook-Kanamori, E. Donga, M. van Dijk, J. Gert van Dijk, G.-J. Lammers, K.W. van Kralingen, C. Prehn, J. Adamski, J.A. Romijn, K. Willems van Dijk, E.P.M. Corssmit, P.C.N. Rensen, N.R. Biermasz, A single night of sleep curtailment increases plasma acylcarnitines: novel insights in the relationship between sleep and insulin resistance, *Archives of Biochemistry and Biophysics* (2015), doi: 10.1016/j.abb.2015.09.017.

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# A single night of sleep curtailment increases plasma acylcarnitines: novel insights in the relationship between sleep and insulin resistance

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23 Running title: Short sleep increases plasma acylcarnitines

- 24 Keywords: Sleep loss, sleep deprivation, insulin resistance, diabetes, metabolomics,
- 25 acylcarnitines

#### 27 Abstract

We have previously shown that acute sleep curtailment induces insulin resistance, both in 28 29 healthy individuals as well as in patients with type 1 diabetes, suggesting a causal role for 30 sleep disturbances in pathogenesis of insulin resistance, independent of endogenous insulin production. However, the underlying mechanisms remain unclear. This study aimed to 31 explore the metabolic pathways affected by sleep loss using targeted metabolomics in 32 33 human fasting plasma samples. Healthy individuals (n = 9) and patients with type 1 diabetes (n = 7) were studied after a single night of short sleep (4 hours) versus normal sleep (8 34 hours) in a cross-over design. Strikingly, one night of short sleep specifically increased the 35 plasma levels of acylcarnitines, essential intermediates in mitochondrial fatty acid oxidation 36 (FAO). Specifically, short sleep increased plasma levels of tetradecenoyl-L-carnitine (C14:1) 37 (+32%.  $p=2.67*10^{-4}$ ), octadecanoyl-L-carnitine (C18:1) (+22%,  $p=1.92*10^{-4}$ ) 38 and 39 octadecadienyl-L-carnitine (C18:2) (+27%, p=1.32\*10<sup>-4</sup>). Since increased plasma acylcarnitine 40 levels could be a sign of disturbed FAO, it is possible that sleep curtailment acutely induces inefficient mitochondrial function. Our observations provide a basis for further research into 41 the role of acylcarnitines as a potential mechanistic pathway by which sleep deprivation -42 even short term – causes adverse metabolic effects, such as insulin resistance. 43

#### 45 Introduction

Diabetes mellitus (DM) is characterized by either an absolute (type 1; DM1) or relative (type 46 47 2; DM2) deficiency of insulin. Both DM1 and DM2 are associated by increased morbidity and 48 increased cardiovascular risk (1;2). Peripheral insulin resistance precedes the development of DM2 and recently it has been recognized that a certain degree of insulin resistance is also 49 present in DM1 (3). Therefore, uncovering modifiable risk factors in an early stage of insulin 50 51 resistance development is of crucial importance to reduce the number of patients with DM2 52 and improve glycemic control in DM1. Interestingly, the DM2 epidemic coincides with a reduction in the average sleep duration, which has gradually declined with ~1.5 hours per 53 54 night (4) over the past decades. In fact, large epidemiological cohorts have documented an association between sleep duration and increased insulin resistance (5). Furthermore, short 55 sleep has been associated with poor glycemic control in DM1 (6). Both short and long 56 57 duration of sleep are associated with an increased risk for insulin resistance, implying that 58 there might be an optimal sleep duration of approximately 8 hours (7-10). Several human intervention studies showed that decreased sleep duration causes insulin resistance. 59 60 Repeated sleep curtailment during more than 6 nights increased insulin resistance in healthy individuals (11-13). Moreover, we previously published that even one single night 61 with partial sleep loss, i.e. 4 hours sleep allowed, a condition representative for incidental 62 63 daily life sleep habits, is sufficient to induce peripheral insulin resistance in both healthy 64 young individuals (14) as well as patients with DM1 (15).

The mechanism by which acute sleep curtailment induces insulin resistance has not been fully elucidated. Plasma metabolomics is considered a valuable approach to assess underlying biological processes, complementary to genomics and transcriptomics. Strikingly,

68 metabolite levels reflect biological activity of the encoded proteins and are thus closer to the clinical endpoints (16). Indeed, metabolomics has previously been demonstrated to be a 69 powerful tool in investigating insulin resistance and DM2 (17). Thus far, the effects of sleep 70 71 loss on the human metabolome are poorly characterized. Prolonged sleep deprivation during 5 days has been shown to induce metabolite changes in lipid, carbohydrate, amino 72 acid and protein pathways (18;19). In contrast, Davies et al. (20) subjected healthy 73 individuals to complete sleep restriction of 24 hours. This extreme sleep deprivation 74 75 resulted in increased plasma levels of glycerophospholipids, acylcarnitines, sphingolipids and amino acids. However, the sleep intervention and control sleep occurred on 76 consecutive days in all individuals. Differences between metabolite levels were also 77 observed between the wake periods, suggesting that the study conditions were not fully 78 comparable. In addition, none of these previous studies included measurements of insulin 79 80 resistance. Therefore, the aim of the present study was to use metabolomics to explore 81 pathways involved in the relationship between sleep and insulin resistance in a cohort with proven insulin resistance upon short sleep duration (14;15). To this end, we examined 163 82 metabolites in 16 individuals (healthy individuals and individuals with DM1) subjected to a 83 night of normal sleep duration (8 hours) and one night of short sleep duration (4 hours). 84 Here, we report that one night of sleep curtailment specifically increases the metabolic class 85 of acylcarnitines in plasma, suggesting that increased acylcarnitines are associated with the 86 observed relationship between sleep curtailment and induction of insulin resistance. 87

88

#### 89 RESEARCH DESIGN AND METHODS

90 Protocol

Two studies were previously performed, to study the effect of one night of short sleep 91 duration (4 hours) compared to normal sleep duration (8 hours) on peripheral insulin 92 resistance (14;15). The studies applied the same study design in two different populations, 93 94 namely healthy individuals and patients (14) with type 1 diabetes (DM1)(15). Healthy individuals were studied to determine the effects of a single night of short sleep duration on 95 insulin resistance. The second study assessed the effects of short sleep duration on insulin 96 resistance in DM1 patients on stable insulin pump therapy. DM1 patients do not have 97 endogenous insulin production and therefore cannot compensate for fluctuations in insulin 98 resistance. We hypothesized that variations in sleep duration could contribute the intra-99 individual variations in glucoregulation. In both healthy individuals and individuals with 100 DM1, decreased sleep duration induced insulin resistance. Therefore, we reasoned that a 101 single night of short sleep duration may increase peripheral insulin resistance via a common 102 103 metabolic pathway. To investigate which pathways could be involved, we analyzed metabolites from both studies and pooled the data. 104

105

#### 106 Subjects

107 The study was approved by the medical ethical committee of the Leiden University Medical 108 Center and all subjects gave written informed consent. We recruited a total of 18 109 individuals. Briefly, nine healthy individuals were recruited by advertisement and nine 110 individuals with DM1 with stable continuous subcutaneous insulin pump therapy were 111 included from our outpatient clinic. Exclusion criteria for all individuals were BMI>26 kg/m<sup>2</sup>, 112 history of sleep disorders, psychiatric disorders and use of sleep medication,  $\beta$ -blocking 113 drugs and prokinetic drugs. All individuals had a stable weight in the past 3 months and had

114 regular and non-extreme sleeping habits. Habitual sleep duration was assessed by 7 days of actigraphy (Actiwatch AW7; Cambridge Neurotechnology, Cambridge, UK) prior to both 115 study days and sleep questionnaires (Epworth Sleepiness Scale, Pittsburg Sleep Quality 116 117 Index and Berlin Questionnaire). Subjects were instructed to maintain a regular dietary, activity and sleep regiments 3 days prior to both study days, fitting their habits, which they 118 recorded in a diary. DM1 patients were instructed to keep a stable insulin pump setting. Of 119 120 the 18 recruited individuals, 2 individuals with DM1 were excluded from all analyses, one 121 due to previously undiagnosed sleep apnea and one due to nocturnal hypoglycemia.

122

#### 123 Experimental design

Subjects were subjected to in-hospital sleep registration for 3 days, of which study day 1 124 was for basal measurements and habituation to hospital conditions. Sleep duration and 125 126 quality (of parameters) was assessed by polysomnography as described previously (14;15). 127 All subjects underwent both a normal sleep night of at least 8 hours and one night of 4 hours sleep, the order of which was determined by balanced assignment, in a cross-over 128 design with at least 3 weeks interval between measurements. In both sleep conditions, 129 subjects spent 8.5 hours (from 23:00 to 7:30) in bed and were fasting from 22:00 onwards. 130 During sleep curtailment, subjects were allowed to sleep from 01:00 to 05:00, the remaining 131 132 time they were allowed to read or watch movies in upward position in dim light. Their wakefulness was monitored. After the night of normal or short sleep, a fasting plasma 133 sample was obtained at 8:30 am, after which a hyperinsulinemic euglycemic clamp was 134 performed as described in detail previously (14;15) to establish peripheral insulin sensitivity, 135 endogenous glucose production and hepatic insulin sensitivity. Briefly, a primed (17.6 136

μmol\*kg<sup>-1</sup>) continuous (0.22 μmol\*kg<sup>-1</sup>\*min<sup>-1</sup>) infusion of [6,6-<sup>2</sup>H<sub>2</sub>]glucose (Cambridge 137 Isotope laboratory, Andover, MA) was administered via a catheter. Infusion of insulin 138 (Actrapid, Novo Nordisk, Alphen a/d Rijn) occurred simultaneously according to DeFronzo 139 (21). Blood samples were obtained every 5 minutes from the contralateral arm for glucose 140 measurements to adjust variable infusion of 20% glucose with 3% [6,6-<sup>2</sup>H<sub>2</sub>]glucose to 141 maintain euglycemia (i.e. 5.0 mmol/l), which was started 4 min after start of insulin infusion. 142 Free fatty acids were determined in basal fasting plasma samples as by enzymatic 143 144 colorimetric assay (14;15).

145

#### 146 Metabolomics

Metabolomics analysis was performed on fasting plasma samples in all individuals using the 147 Biocrates Absolute*IDQ*<sup>™</sup> p150 kit (Biocrates, Life Science AG, Innsbruck, Austria) in the 148 149 Genome Analysis Center at the Helmholtz Zentrum, Munich, Germany. The assay procedures of the Absolute/DQ<sup>TM</sup> p150 kit as well as the metabolite nomenclature have 150 been described in detail previously (22;23). Briefly, 10 µL of each plasma sample was 151 pipetted into a 96 well sandwich plate containing an inserted filter with previously applied 152 stable isotope labeled internal standards. The filters in the wells were dried using a stream 153 of nitrogen. Amino acids were derivatized with 5% phenylisothiocyanate reagent (PITC) and 154 the filters were dried again. Metabolites as well as internal standards were extracted with 5 155 mM ammonium acetate in methanol and the solutions were centrifuged through the filter 156 157 membrane into the lower deep well plate. The extracts were diluted with MS running solvent and analyzed. Flow injection analysis (FIA) tandem mass spectrometry (MS/MS) 158 method was used to quantify 163 metabolites, including free carnitine, 40 acylcarnitines, 14 159

160 amino acids (13 proteinogenic + ornithine), hexoses (sum of hexoses), 92 glycerophospholipids (15 lysophosphatidylcholines (lysoPC) and 77 phosphatidylcholines 161 (PC), and 15 sphingolipids. Internal standards served as reference for the calculation of 162 metabolite concentrations ( $\mu$ M). The complete list of analyzed metabolites grouped by 163 metabolite class is presented in supplementary material (Table S3). 164

165

#### 166 Statistical analysis

For all metabolites, differences between short and normal sleep were calculated by 167 subtracting plasma levels obtained after short sleep from those obtained after normal sleep. 168 Paired Students T-tests for were performed comparing normal and short sleep (SPSS 169 statistical package edition 20) with Bonferroni post-hoc correction for multiple testing. P< 170  $3.07*10^{-4}$  (=0.05/163; after correction) was considered statistically significant. Calculations 171 172 for hyperinsulinemic euglycemic clamp analysis were described previously (14;15). Since we 173 aimed to investigate the effect of short sleep on metabolite levels, individuals of both groups (healthy individuals and individuals with DM1) were pooled to determine effects of 174 175 sleep duration. Two way repeated measure ANOVA was performed to analyze interaction effects of subgroup (healthy vs. DM1) with sleep duration. Data are presented as means ± 176 SD. Since baseline characteristics and insulin sensitivity data were published for healthy 177 individuals and individuals with DM1 separately, in this paper these data are shown for the 178 two groups together. To allow comparison between subgroups, the baseline characteristics, 179 180 sleep indices and insulin sensitivity data are included in the supplemental tables and were compared using Student's t-test. 181

#### 183 **RESULTS**

#### 184 Basal clinical characteristics

Metabolites were measured in sixteen individuals after a night of short sleep (4 hours) 185 versus after a night of normal sleep (8 hours) duration. Subjects had a mean age of 44 ± 14 186 years and included 8 women. Individuals were lean, with an average BMI of  $23.7 \pm 2.2 \text{ kg/m}^2$ 187 and a waist hip ratio of  $0.85 \pm 0.08$  (Table 1). The study population consisted of nine healthy 188 189 individuals (56%) and seven individuals with type 1 diabetes mellitus (DM1) (44%). Sleep 190 duration prior to the study days did not differ healthy individuals (mean recorded sleep duration prior to study day 1 and 2: 420 ± 20 min vs. 476 ± 11 min; p=0.19) nor in individuals 191 with DM1 (mean recorded sleep duration prior to study day 1 and 2: 475 ± 8 min vs. 490 ± 7 192 min; p=0.12). Results of healthy individuals and individuals with DM1 were reported 193 previously separately (14;15). Age, sex distribution, BMI and waist-hip ratio were 194 195 comparable between these two subgroups (Table S1).

196

### 197 Short sleep increases insulin resistance

Short sleep intervention was effective in reducing total sleep time (TST) by -51% (461 ± 25 vs 198 225  $\pm$  26 min, p < 0.001). The reduction of sleep duration was due to decreased sleep 199 duration of both non-REM (stage 2 and stage 3) and REM sleep (Table 2). Fasting plasma 200 free fatty acids did not differ between sleep conditions (Table 2) or between subgroups 201 (Table S2). Next, the effect of short sleep on insulin resistance was investigated by 202 203 hyperinsulinemic euglycemic clamp studies. Interestingly, a single night of short sleep increased peripheral insulin resistance, as indicated by a decreased glucose disposal rate 204 (GDR) (34.1  $\pm$  13.8 vs 27.9  $\pm$  9.8  $\mu$ mol\*kg LBM<sup>-1</sup>\*min<sup>-1</sup>, p = 0.001) and decreased glucose 205

infusion rate (GIR) (29.0 ± 14.7 vs 22.1 ± 10.7 µmol\*kg LBM<sup>-1</sup>\*min<sup>-1</sup>, p=0.001). Short sleep 206 tended to increase endogenous glucose production (EGP) by the liver in all subjects (4.7  $\pm$ 207 1.9 vs 5.5  $\pm$  1.6  $\mu$ mol\*kg LBM<sup>-1</sup>\*min<sup>-1</sup>, p=0.08; Table 2). This was mainly due to increased 208 endogenous glucose production in the subset of healthy individuals (Table S2; previously 209 published in (14)). Expectedly, individuals with DM1 displayed higher baseline insulin 210 resistance than in healthy individuals (3) (EGP 6.2  $\pm$  1.9 vs. 3.6  $\pm$  0.6, p=0.003; GDR 25.5  $\pm$  6.4 211 vs. 40.7 ± 14.3, p=0.028; GIR 19.0 ± 7.0 vs. 36.9 ± 14.4, p=0.014, Table S2). Moreover, short 212 213 sleep increased peripheral insulin resistance irrespective of this difference in baseline insulin sensitivity, suggesting a that short sleep may induce insulin resistance in healthy individuals 214 215 and individuals with DM1 via a common pathway. Therefore, the effect of short sleep was investigated for healthy individuals and individuals with DM1 together. 216

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#### 218 Short sleep specifically increases plasma acylcarnitines

To investigate possible pathways which could be involved in the increased of insulin 219 resistance by short sleep duration, we performed metabolomics analysis on fasting morning 220 plasma samples. A total of 163 metabolites representing 5 different classes were measured 221 (Table S3). Short sleep increased thirteen metabolites (p<0.05) (Table 3). Strikingly, all of 222 these are acylcarnitines. After stringent post-hoc correction, short sleep significantly 223 increased plasma levels of tetradecenoyl-L-carnitine (C14:1) by +32% (plasma level 224 difference: +0.017  $\mu$ M, p=2.67\*10<sup>-4</sup>), octadecenoyl-L-carnitine (C18:1) by +22% (plasma level 225 difference: +0.015  $\mu$ M, p=1.92\*10<sup>-4</sup>) and octadecadienyl-L-carnitine (C18:2) by +27% 226 (plasma level difference: +0.005  $\mu$ M, p=1.32\*10<sup>-4</sup>). Short sleep duration increased 227 acylcarnitines in both subgroups, indicating that the effect of short sleep on acylcarnitines 228

was not dependent on having DM1 or being healthy. There was no interaction effect of the subgroup (healthy vs. DM1) with the sleep duration (short vs. normal) for the 13 increased acylcarnitines. Baseline acylcarnitine levels (i.e. after normal sleep) did not differ between healthy individuals and DM1, except for a higher level of C:12-DC in DM1 (0.087  $\pm$  0.005 vs 0.101  $\pm$  0.005  $\mu$ M, p<0.0001) (Table S5). Acylcarnitines levels did not differ between healthy individuals and DM1 after short sleep (Table S6). We therefore conclude that a single night of short sleep specifically increased plasma acylcarnitines.

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#### 237 DISCUSSION

The present study aimed to explore the metabolic pathways affected by sleep curtailment 238 using targeted plasma metabolomics in individuals (healthy individuals and individuals with 239 type 1 diabetes (DM1)) subjected to both short sleep (4 hours) and normal sleep (8 hours). 240 241 As part of the same study, we previously reported that this short sleep intervention 242 increased peripheral insulin resistance in both study groups as determined by hyperinsulinemic euglycemic clamp analysis (14;15). We now show that one night of short 243 244 sleep specifically increases plasma levels of acylcarnitines, in both healthy individuals and DM1 patients. 245

Our study is the first to show that short sleep duration increased plasma acylcarnitines in concert with increased insulin resistance in both healthy individuals and individuals with DM1. This indicates that short sleep duration affects metabolism irrespective of pre-existing insulin producing capacity. The relationship between increased plasma acylcarnitine levels and increased insulin resistance is supported by association studies. Human studies showed increased plasma levels of acylcarnitines in individuals with

impaired fasting glucose and with type 2 diabetes (DM2), compared to healthy controls
(24;25). The significance of this association is still a matter of debate, since human
intervention studies are lacking (26).

It is interesting to speculate about the biological relevance of increased plasma levels 255 of acylcarnitines. Acylcarnitines are vital to energy homeostasis. They are esters of fatty 256 acids and carnitine, which are transported over the outer and inner mitochondrial 257 membranes by carnitine palmitoyl transferases (CPTs). Thus, acylcarnitines are essential to 258 259 shuttle fatty acids from the cytoplasm into mitochondria were they can be oxidized and enter the tricarboxylic acid (TCA) cycle to generate ATP. An excess of acylcarnitines is 260 generally viewed as a result from a mismatch between TCA flux and fatty acid oxidation 261 (FAO) (27). Previously reported causes of this mismatch include prolonged fasting and 262 excessive muscle activity (28-30). The present study, in which subjects participated in a 263 264 protocol that controlled for food intake and physical activity, adds sleep deprivation as a 265 provoking event. A mismatch between FAO and TCA flux has been related to mitochondrial dysfunction. Patients with inborn errors of FAO have increased plasma levels of especially 266 long chain acylcarnitines (31). Interestingly, altered mitochondrial parameters have been 267 frequently linked to insulin resistance in the context of both DM1 and DM2 (32-38). 268 Moreover, mitochondrial dysfunction in mice induces skeletal muscle insulin resistance (27) 269 while TCA-FAO mismatch predisposes mice to diet-induced obesity and insulin resistance 270 (39). It is therefore tempting to speculate that in our model of insulin resistance due to 271 272 short sleep deprivation, the increased plasma acylcarnitine levels are a sign of inefficient mitochondrial function. 273

274 The tissue distribution of acylcarnitines coincides with important targets of insulin, i.e. muscle and liver. The majority of the body's L-carnitine is stored in muscle (~ 97% of the 275 body's L-carnitine), followed by liver which contains 1% of the total L-carnitine pool (40). 276 Acylcarnitine results from the acylation of L-carnitine, and is therefore dependent on the 277 fatty acid pool of the tissue. Interestingly, animal studies demonstrate the distribution of 278 acylcarnitines is different between metabolic organs. In mice, the muscle tissue contains 279 relatively more long-chain acylcarnitines, including C14:1 and C18:1, while liver is richer in 280 free carnitines and short-chain carnitines (41). Collectively, these data suggest that plasma 281 short-chain acyl- and free carnitines are mainly derived from the liver, as indeed 282 demonstrated in pigs (42), while plasma long-chain acylcarnitines in plasma presumably 283 originate from muscle tissue. These data thus suggest that the increase in long-chain 284 acylcarnitine that we observe after a single night of short sleep is likely derived from muscle. 285 286 Mechanistically, increased acylcarnitine levels after short sleep duration could be a 287 marker of altered metabolic processes: increased fatty acid oxidation (FAO), inefficient mitochondrial function or a disturbed metabolism of the branched-chain amino acids 288 (BCAA) valine, isoleucine or leucine. Although disturbed BCAA metabolism has been 289 associated with insulin resistance in humans (43), our data do not support a role of BCAA 290 metabolism as short sleep duration did not increase BCAA plasma levels or short-chain 291 acylcarnitines. Increased acylcarnitine levels due to increased FAO can be caused by either 292 increased energy demand and/or prolonged fasting. In the present study, the length of 293 fasting was equal; however energy expenditure was not measured. Therefore, we cannot 294 exclude that the increased acylcarnitines after short sleep are due to increased FAO. Sleep is 295 296 accompanied by lower resting energy expenditure than wakefulness (44) and therefore

297 short sleep duration may increase energy demand. In fact, complete (24 h) sleep deprivation increases energy demand by 7% (45). However, the effects of short sleep duration on 298 energy expenditure are inconclusive (46). A recent study shows that short sleep intervention 299 300 for five consecutive days increased long-chain plasma acylcarnitines (19). Interestingly, after one night of recovery sleep, plasma acylcarnitines did not normalize. Likely, the increased 301 acylcarnitines were not due to differences in overnight energy expenditure. Besides being a 302 marker of insulin resistance and/or mitochondrial processes, acylcarnitines could also play a 303 304 causal role in development of insulin resistance. In vitro studies have shown that acylcarnitines have bioactive properties and indeed have pro-inflammatory effects (47;48). 305 Of note, treatment of both rodent and human myotubes with acylcarnitines in a 306 physiological concentration caused decreased insulin signaling and glucose uptake in 307 response to insulin (49). Although this finding needs to be confirmed in vivo, it provides a 308 309 putative causal link between acylcarnitines and insulin resistance.

310 Taken all these data together, it is interesting to speculate on a mechanistic model for the relationship between sleep curtailment and insulin resistance. Upon sleep 311 312 curtailment, the energy demands of peripheral tissues increases at a time conflicting with the physiological circadian rhythm. The energy homeostasis is adapted to anticipate the 313 changing energy need and availability throughout the day. Indeed, muscle tissue is also 314 under circadian control (50). These clock genes are also important in driving rhythmicity in 315 energy producing capacity of the mitochondria, as evidenced by mice studies (51). We 316 hypothesize that the mismatch in energy producing capacity and demand could be the 317 cause of incomplete FAO, leading to accumulation of intermediates of FAO. Acylcarnitine 318

levels increase, which may increase insulin resistance either through direct interaction withinsulin signaling or through increased inflammatory pathways.

Our findings are supported by three studies which have investigated the effects of 321 322 sleep on the human metabolome. Davies et al. (20) subjected 12 healthy individuals to an extreme sleep deprivation of 24 hours and reported nine increased short and medium-chain 323 acylcarnitines, including tetradecenoyl-L-carnitine. Bell et al. (18) reported a trend towards 324 increased acylcarnitines after prolonged mild sleep curtailment of 8 consecutive nights of 325 326 5.5 hours sleep in 11 young individuals with family history of DM2. Weljie et al (19) also reported increased C18:1, C10:0 and C12:0 acylcarnitines upon five consecutive nights of 4 327 328 hours sleep. Strikingly, despite the difference in study populations and sleep curtailment protocols of the present and previous studies used, the acylcarnitines invariably increase 329 after sleep curtailment. 330

In conclusion, the present study shows that a single night of 4 hours short sleep, which induces insulin resistance (14;15), also increases plasma levels of acylcarnitines, in particular tetradecenoyl-L-carnitine, octadecenoyl-L-carnitine and octadecadienyl-Lcarnitine. We propose that sleep curtailment impairs mitochondrial function, which coincides with insulin resistance. Our findings provide a basis for mechanistic studies to further elucidate the role of acylcarnitines in the complex relationship between short sleep and increased insulin resistance.

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#### 339 Acknowledgements

340 We thank Julia Scarpa, Werner Römisch-Margl and Katharina Faschinger for support with 341 the metabolomics measurements performed at the Helmholtz Centrum München, Genome

Analysis Center. Author contributions. R.v.d.B. wrote manuscript and performed analysis, 342 D.O.M.K. and K.W.v.D. analyzed data and supervised metabolome analysis, E.D., M.v.D., and 343 J.G.v.D. performed the initial study, G-J.L. and K.W.v.K. performed polysomnography, C.P. 344 and J.A. performed metabolome analysis and quality assurance, and E.P.M.C, J.A.R, P.C.N.R. 345 and N.R.B designed and supervised the study. All authors have approved final version of the 346 manuscript. Disclosure statement. Authors declare no conflict of interest. Funding. This 347 study was supported by a pilot grant from the Dutch Diabetes Research Foundation and by 348 grants from the European Foundation for the Study of Diabetes (J.A.R), the Netherlands 349 Organization for Scientific Research (NWO-VENI grant 016.136.125 to N.R.B.) and the 350 German Federal Ministry of Education and Research (BMBF) to the German Center Diabetes 351 Research (DZD e.V.) (J.A.). P.C.N.R. is Established Investigator of the Dutch Heart Foundation 352 (NHS2009T038). D.O.M.K. was supported by Dutch Science Organization (ZonMW-VENI 353 354 Grant 916.14.023).

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		ACCEPTED MANUSCRIPT
357		References
358		
359	1.	Chillaron, JJ, Flores Le-Roux, JA, Benaiges, D, Pedro-Botet, J: Type 1 diabetes, metabolic
360		syndrome and cardiovascular risk. Metabolism 63:181-187, 2014
361	2.	Fox,KM, Wu,Y, Kim,J, Grandy,S: Cardiovascular event rates and healthcare resource
362		utilisation among high-risk adults with type 2 diabetes mellitus in a large population-
363		based study. Int J Clin Pract 69:218-227, 2015
364	3.	Donga, E, Dekkers, OM, Corssmit, EP, Romijn, JA: Insulin resistance in patients with type
365		1 diabetes assessed by glucose clamp studies: systematic review and meta-analysis.
366		Eur J Endocrinol 173:101-109, 2015
367	4.	Leproult, R, Van, CE: Role of sleep and sleep loss in hormonal release and metabolism.
368		Endocr Dev 17:11-21, 2010
369	5.	Cappuccio, FP, D'Elia, L, Strazzullo, P, Miller, MA: Quantity and quality of sleep and
370		incidence of type 2 diabetes: a systematic review and meta-analysis. Diabetes Care
371		33:414-420, 2010
372	6.	Borel,AL, Pepin,JL, Nasse,L, Baguet,JP, Netter,S, Benhamou,PY: Short sleep duration
373		measured by wrist actimetry is associated with deteriorated glycemic control in type 1
374		diabetes. Diabetes Care 36:2902-2908, 2013

375	7. Buxton,OM, Marcelli,E: Short and long sleep are positively associated with obesity
376	diabetes, hypertension, and cardiovascular disease among adults in the United States
377	Soc Sci Med 71:1027-1036, 2010

Liu,Y, Croft,JB, Wheaton,AG, Perry,GS, Chapman,DP, Strine,TW, McKnight-Eily,LR,
 Presley-Cantrell,L: Association between perceived insufficient sleep, frequent mental
 distress, obesity and chronic diseases among US adults, 2009 behavioral risk factor
 surveillance system. *BMC Public Health* 13:84, 2013

Najafian, J, Mohamadifard, N, Siadat, ZD, Sadri, G, Rahmati, MR: Association between
 sleep duration and diabetes mellitus: Isfahan Healthy Heart Program. *Niger J Clin Pract* 16:59-62, 2013

385 10. Ohkuma,T, Fujii,H, Iwase,M, Kikuchi,Y, Ogata,S, Idewaki,Y, Ide,H, Doi,Y, Hirakawa,Y,
 386 Nakamura,U, Kitazono,T: Impact of sleep duration on obesity and the glycemic level in
 387 patientswith type 2diabetes. *Diabetes Care* 36:611-617, 2013

11. Nedeltcheva,AV, Kessler,L, Imperial,J, Penev,PD: Exposure to recurrent sleep
 restriction in the setting of high caloric intake and physical inactivity results in
 increased insulin resistance and reduced glucose tolerance. *J Clin Endocrinol Metab* 94:3242-3250, 2009

392 12. Spiegel,K, Leproult,R, Van,CE: Impact of sleep debt on metabolic and endocrine
 393 function. *Lancet* 354:1435-1439, 1999

- Buxton,OM, Pavlova,M, Reid,EW, Wang,W, Simonson,DC, Adler,GK: Sleep restriction
   for 1 week reduces insulin sensitivity in healthy men. *Diabetes* 59:2126-2133, 2010
- 14. Donga, E, van, DM, van Dijk, JG, Biermasz, NR, Lammers, GJ, van Kralingen, KW,
   Corssmit, EP, Romijn, JA: A Single Night of Partial Sleep Deprivation Induces Insulin
   Resistance in Multiple Metabolic Pathways in Healthy Subjects. *J Clin Endocrinol Metab* 2010
- 400 15. Donga, E, van, DM, van Dijk, JG, Biermasz, NR, Lammers, GJ, van, KK, Hoogma, RP,
  401 Corssmit, EP, Romijn, JA: Partial sleep restriction decreases insulin sensitivity in type 1
  402 diabetes. *Diabetes Care* 33:1573-1577, 2010
- 403 16. Gieger, C, Geistlinger, L, Altmaier, E, Hrabe de, AM, Kronenberg, F, Meitinger, T,
  404 Mewes, HW, Wichmann, HE, Weinberger, KM, Adamski, J, Illig, T, Suhre, K: Genetics
  405 meets metabolomics: a genome-wide association study of metabolite profiles in
  406 human serum. *PLoS Genet* 4:e1000282, 2008

407 17. Suhre, K: Metabolic profiling in diabetes. J Endocrinol 221:R75-R85, 2014

408 18. Bell,LN, Kilkus,JM, Booth,JN, III, Bromley,LE, Imperial,JG, Penev,PD: Effects of sleep
409 restriction on the human plasma metabolome. *Physiol Behav* 122:25-31, 2013

410	19.	Weljie,AM, Meerlo,P, Goel,N, Sengupta,A, Kayser,MS, Abel,T, Birnbaum,MJ, Dinges,DF,
411		Sehgal,A: Oxalic acid and diacylglycerol 36:3 are cross-species markers of sleep debt.

412 Proc Natl Acad Sci U S A 112:2569-2574, 2015

- 413 20. Davies, SK, Ang, JE, Revell, VL, Holmes, B, Mann, A, Robertson, FP, Cui, N, Middleton, B,
- 414 Ackermann,K, Kayser,M, Thumser,AE, Raynaud,FI, Skene,DJ: Effect of sleep deprivation
- 415 on the human metabolome. *Proc Natl Acad Sci U S A* 2014
- 416 21. DeFronzo,RA, Tobin,JD, Andres,R: Glucose clamp technique: a method for quantifying
- 417 insulin secretion and resistance. *Am J Physiol* 237:E214-E223, 1979
- 22. Römisch-Margl,W, Prehn,C, Bogumil,R, Röhring,C, Suhre,K, Adamski,J: Procedure for
  tissue sample preparation and metabolite extraction for high-throughput targeted
  metabolomics. *Metabolomics* 8:133-142, 2012
- 23. Illig,T, Gieger,C, Zhai,G, Romisch-Margl,W, Wang-Sattler,R, Prehn,C, Altmaier,E,
  Kastenmuller,G, Kato,BS, Mewes,HW, Meitinger,T, de Angelis,MH, Kronenberg,F,
  Soranzo,N, Wichmann,HE, Spector,TD, Adamski,J, Suhre,K: A genome-wide perspective
  of genetic variation in human metabolism. *Nat Genet* 42:137-141, 2010
- 425 24. Mai,M, Tonjes,A, Kovacs,P, Stumvoll,M, Fiedler,GM, Leichtle,AB: Serum levels of
  426 acylcarnitines are altered in prediabetic conditions. *PLoS One* 8:e82459, 2013

- 427 25. Mihalik,SJ, Goodpaster,BH, Kelley,DE, Chace,DH, Vockley,J, Toledo,FG, DeLany,JP:
  428 Increased levels of plasma acylcarnitines in obesity and type 2 diabetes and
  429 identification of a marker of glucolipotoxicity. *Obesity (Silver Spring)* 18:1695-1700,
  430 2010
- 431 26. Schooneman,MG, Vaz,FM, Houten,SM, Soeters,MR: Acylcarnitines: reflecting or
  432 inflicting insulin resistance? *Diabetes* 62:1-8, 2013

433 27. Koves,TR, Ussher,JR, Noland,RC, Slentz,D, Mosedale,M, Ilkayeva,O, Bain,J, Stevens,R,
434 Dyck,JR, Newgard,CB, Lopaschuk,GD, Muoio,DM: Mitochondrial overload and
435 incomplete fatty acid oxidation contribute to skeletal muscle insulin resistance. *Cell*436 *Metab* 7:45-56, 2008

28. Chen,C, Krausz,KW, Shah,YM, Idle,JR, Gonzalez,FJ: Serum metabolomics reveals
irreversible inhibition of fatty acid beta-oxidation through the suppression of
PPARalpha activation as a contributing mechanism of acetaminophen-induced
hepatotoxicity. *Chem Res Toxicol* 22:699-707, 2009

441 29. Xu,Q, Vu,H, Liu,L, Wang,TC, Schaefer,WH: Metabolic profiles show specific
442 mitochondrial toxicities in vitro in myotube cells. *J Biomol NMR* 49:207-219, 2011

30. Soeters,MR, Sauerwein,HP, Duran,M, Wanders,RJ, Ackermans,MT, Fliers,E,
Houten,SM, Serlie,MJ: Muscle acylcarnitines during short-term fasting in lean healthy
men. *Clin Sci (Lond)* 116:585-592, 2009

- 31. Rinaldo, P, Cowan, TM, Matern, D: Acylcarnitine profile analysis. *Genet Med* 10:151-156,
  2008
- Razak,F, Anand,SS: Impaired mitochondrial activity in the insulin-resistant offspring of
  patients with type 2 diabetes. Petersen KF, Dufour S, Befroy D, Garcia R, Shulman GI. N
- 450 Engl J Med 2004; 350: 664-71. Vasc Med 9:223-224, 2004
- 451 33. Morino,K, Petersen,KF, Dufour,S, Befroy,D, Frattini,J, Shatzkes,N, Neschen,S,
- 452 White,MF, Bilz,S, Sono,S, Pypaert,M, Shulman,GI: Reduced mitochondrial density and
- 453 increased IRS-1 serine phosphorylation in muscle of insulin-resistant offspring of type
- 454 2 diabetic parents. *J Clin Invest* 115:3587-3593, 2005
- 34. Befroy,DE, Petersen,KF, Dufour,S, Mason,GF, de Graaf,RA, Rothman,DL, Shulman,GI:
  Impaired mitochondrial substrate oxidation in muscle of insulin-resistant offspring of
  type 2 diabetic patients. *Diabetes* 56:1376-1381, 2007
- 458 35. Kelley, DE, He, J, Menshikova, EV, Ritov, VB: Dysfunction of mitochondria in human
  459 skeletal muscle in type 2 diabetes. *Diabetes* 51:2944-2950, 2002
- 36. Ritov,VB, Menshikova,EV, He,J, Ferrell,RE, Goodpaster,BH, Kelley,DE: Deficiency of
  subsarcolemmal mitochondria in obesity and type 2 diabetes. *Diabetes* 54:8-14, 2005

462	37.	Mogensen, M, Sahlin, K, Fernstrom, M, Glintborg, D, Vind, BF, Beck-Nielsen, H, Hojlund, K:
463		Mitochondrial respiration is decreased in skeletal muscle of patients with type 2
464		diabetes. <i>Diabetes</i> 56:1592-1599, 2007

38. Cree-Green,M, Newcomer,BR, Brown,MS, Baumgartner,AD, Bergman,B, Drew,B,
Regensteiner,JG, Pyle,L, Reusch,JE, Nadeau,KJ: Delayed skeletal muscle mitochondrial
ADP recovery in youth with type 1 diabetes relates to muscle insulin resistance. *Diabetes* 64:383-392, 2015

39. Muoio,DM, Noland,RC, Kovalik,JP, Seiler,SE, Davies,MN, DeBalsi,KL, Ilkayeva,OR,
Stevens,RD, Kheterpal,I, Zhang,J, Covington,JD, Bajpeyi,S, Ravussin,E, Kraus,W,
Koves,TR, Mynatt,RL: Muscle-specific deletion of carnitine acetyltransferase
compromises glucose tolerance and metabolic flexibility. *Cell Metab* 15:764-777, 2012

- 473 40. Reuter,SE, Evans,AM: Carnitine and acylcarnitines: pharmacokinetic, pharmacological
  474 and clinical aspects. *Clin Pharmacokinet* 51:553-572, 2012
- 475 41. Schooneman,MG, Achterkamp,N, Argmann,CA, Soeters,MR, Houten,SM: Plasma
  476 acylcarnitines inadequately reflect tissue acylcarnitine metabolism. *Biochim Biophys*477 Acta 1841:987-994, 2014
- 478 42. Schooneman,MG, Ten Have,GA, Van,VN, Houten,SM, Deutz,NE, Soeters,MR:
  479 Transorgan fluxes in a porcine model reveal a central role for liver in acylcarnitine
  480 metabolism. *Am J Physiol Endocrinol Metab*ajpendo, 2015

481	43. Newgard,CB, An,J, Bain,JR, Muehlbauer,MJ, Stevens,RD, Lien,LF, Haqq,AM, Shah,SH,
482	Arlotto,M, Slentz,CA, Rochon,J, Gallup,D, Ilkayeva,O, Wenner,BR, Yancy,WS, Jr.,
483	Eisenson, H, Musante, G, Surwit, RS, Millington, DS, Butler, MD, Svetkey, LP: A branched-
484	chain amino acid-related metabolic signature that differentiates obese and lean
485	humans and contributes to insulin resistance. Cell Metab 9:311-326, 2009

- 486 44. Thearle,MS, Pannacciulli,N, Bonfiglio,S, Pacak,K, Krakoff,J: Extent and determinants of
  487 thermogenic responses to 24 hours of fasting, energy balance, and five different
  488 overfeeding diets in humans. *J Clin Endocrinol Metab* 98:2791-2799, 2013
- 489 45. Jung,CM, Melanson,EL, Frydendall,EJ, Perreault,L, Eckel,RH, Wright,KP: Energy
  490 expenditure during sleep, sleep deprivation and sleep following sleep deprivation in
  491 adult humans. *J Physiol* 589:235-244, 2011
- 492 46. Klingenberg, L, Sjodin, A, Holmback, U, Astrup, A, Chaput, JP: Short sleep duration and its
  493 association with energy metabolism. *Obes Rev* 13:565-577, 2012
- 494 47. Adams,SH, Hoppel,CL, Lok,KH, Zhao,L, Wong,SW, Minkler,PE, Hwang,DH, Newman,JW,
  495 Garvey,WT: Plasma acylcarnitine profiles suggest incomplete long-chain fatty acid
  496 beta-oxidation and altered tricarboxylic acid cycle activity in type 2 diabetic African497 American women. J Nutr 139:1073-1081, 2009

- 498 48. Rutkowsky, JM, Knotts, TA, Ono-Moore, KD, McCoin, CS, Huang, S, Schneider, D, Singh, S,
- 499 Adams,SH, Hwang,DH: Acylcarnitines activate proinflammatory signaling pathways.
- 500 Am J Physiol Endocrinol Metab 306:E1378-E1387, 2014
- 49. Aguer,C, McCoin,CS, Knotts,TA, Thrush,AB, Ono-Moore,K, McPherson,R, Dent,R,
  Hwang,DH, Adams,SH, Harper,ME: Acylcarnitines: potential implications for skeletal
  muscle insulin resistance. *FASEB J* 29:336-345, 2015
- 50. Harfmann,BD, Schroder,EA, Esser,KA: Circadian Rhythms, the Molecular Clock, and 505 Skeletal Muscle. *J Biol Rhythms* 30:84-94, 2015
- 506 51. Peek,CB, Affinati,AH, Ramsey,KM, Kuo,HY, Yu,W, Sena,LA, Ilkayeva,O, Marcheva,B,
  507 Kobayashi,Y, Omura,C, Levine,DC, Bacsik,DJ, Gius,D, Newgard,CB, Goetzman,E,
  508 Chandel,NS, Denu,JM, Mrksich,M, Bass,J: Circadian clock NAD+ cycle drives
  509 mitochondrial oxidative metabolism in mice. *Science* 342:1243417, 2013
- 510
- 511

## **TABLES**

# **Table 1: Study population characteristics<sup>1</sup>.**

	Subjects (n = 16)	
Females (%)	8 (50%)	
Age (years)	44 ± 14	
BMI (kg/m²)	23.7 ± 2.2	
WHR	0.85 ± 0.08	

- 515 BMI = body mass index. WHR = waist hip ratio. Data is presented as mean (SD or percentage).
- <sup>1</sup> Data are pooled from two previously published studies (14;15).

519	Table 2: Effects of short sleep on sleep parameters and insulin sensitivity	1
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	Subjects (n = 16)		
	Normal sleep	Short sleep	р
Sleep parameters			
TST (min)	461 ± 25	225 ± 24	<0.001
Stage 1 (% of TST)	10 ± 3	10 ± 6	0.798
Stage 2 (% of TST)	43 ± 7	37 ± 9	0.002
Stage 3 (% of TST) (SWS)	24 ± 7	34 ± 10	<0.001
REM sleep (% of TST)	23 ± 4	18±8	0.025
Sleep efficiency (%)	93 ± 4	91 ± 7	0.418
Plasma parameter			
Free fatty acids (mmol/l)	0.65 ± 0.24	0.61 ± 0.19	0.24
Insulin sensitivity parameters			
EGP ( $\mu$ mol * kg LBM <sup>-1</sup> * min <sup>-1</sup> )	4.7 ± 1.9	5.5 ± 1.7	0.087
GDR ( $\mu$ mol * kg LBM <sup>-1</sup> * min <sup>-1</sup> )	34.1 ± 13.8	27.9 ± 9.8	0.001
GIR ( $\mu$ mol * kg LBM <sup>-1</sup> * min <sup>-1</sup> )	29.0 ± 14.7	22.1 ± 10.7	0.001

520 Insulin sensitivity parameters were determined by hyperinsulinemic euglycemic clamp.

521 EGP = endogenous glucose production, GDR = glucose disposal rate (glucose Rd), GIR = glucose infusion rate.

522 LBM = lean body mass. Sleep characteristics were determined by polysomnography. TST = total sleep time.

523 SWS = slow wave sleep. Data is presented as means (SD). Effect of sleep intervention was tested with paired

524 Students T-test, significant differences shown in bold.

<sup>1</sup> Data are pooled from two previously published studies (14;15).

		All subjects			Healthy		~	DM1	
Metabolite	Mean Difference <sup>1</sup>	Change (%) <sup>2</sup>	P-Value	Mean Difference <sup>1</sup>	Change (%) <sup>2</sup>	P-Value	Mean Difference <sup>1</sup>	Change (%) <sup>2</sup>	P-Value
C0	0.191	0.7%	8.81*10 <sup>-1</sup>	0.398	1.3%	8.47*10 <sup>-1</sup>	-0.075	-0.3%	9.61*10 <sup>-1</sup>
C2	<b>0.662</b> <sup>\$</sup>	16.8%	<b>2.28*10</b> <sup>-2</sup>	0.184	4.4%	5.40*10 <sup>-1</sup>	1.276 <sup>\$</sup>	35.6%	1.33*10 <sup>-2</sup>
C3	-0.006	-2.6%	6.76*10 <sup>-1</sup>	-0.002	-0.7%	9.33*10 <sup>-1</sup>	-0.012	-6.2%	6.03*10 <sup>-1</sup>
C3:1	-0.001	-5.9%	4.16*10 <sup>-1</sup>	0.000	-0.1%	9.93*10 <sup>-1</sup>	-0.001	-12.3%	1.58*10 <sup>-1</sup>
C3-DC (C4-OH)	0.007	11.3%	4.55*10 <sup>-1</sup>	0.006	8.0%	7.20*10 <sup>-1</sup>	0.009	17.7%	2.87*10 <sup>-1</sup>
C3-OH	0.001	3.3%	5.03*10 <sup>-1</sup>	0.002	8.5%	5.91*10 <sup>-2</sup>	-0.001	-2.1%	8.24*10 <sup>-1</sup>
C4	0.011	10.3%	1.22*10 <sup>-1</sup>	0.008	7.2%	4.95*10 <sup>-1</sup>	0.015	14.4%	8.68*10 <sup>-2</sup>
C4:1	0.002	6.7%	2.90*10 <sup>-1</sup>	0.002	8.7%	2.55*10 <sup>-1</sup>	0.001	4.8%	6.58*10 <sup>-1</sup>
C5	0.011	10.2%	2.80*10 <sup>-1</sup>	0.012	10.4%	4.59*10 <sup>-1</sup>	0.010	9.8%	4.30*10 <sup>-1</sup>
C5:1	0.000	1.0%	8.74*10 <sup>-1</sup>	-0.001	-2.5%	7.86*10 <sup>-1</sup>	0.001	5.0%	6.20*10 <sup>-1</sup>
C5:1-DC	0.000	2.6%	6.75*10 <sup>-1</sup>	-0.001	-4.8%	5.96*10 <sup>-1</sup>	0.002	12.5%	1.27*10 <sup>-1</sup>
C5-DC (C6-OH)	-0.001	-4.1%	4.87*10 <sup>-1</sup>	-0.001	-8.3%	2.56*10 <sup>-1</sup>	0.000	0.7%	9.45*10 <sup>-1</sup>
C5-M-DC	0.000	0.3%	9.32*10 <sup>-1</sup>	0.000	-1.1%	8.45*10 <sup>-1</sup>	0.001	1.9%	7.11*10 <sup>-1</sup>
C5-OH (C3-DC-M)	0.000	0.3%	9.40*10 <sup>-1</sup>	0,000	-1.5%	6.89*10 <sup>-1</sup>	0.001	2.4%	7.40*10 <sup>-1</sup>
C6 (C4:1-DC)	0.005	8.7%	1.22*10 <sup>-1</sup>	0.001	1.9%	7.82*10 <sup>-1</sup>	0.011	18.8%	6.20*10 <sup>-2</sup>
C6:1	0.001	2.6%	$3.99*10^{-1}$	0.001	3.5%	3.35*10 <sup>-1</sup>	0.000	1.5%	8.05*10 <sup>-1</sup>
C7-DC	0.005 <sup>\$</sup>	20.0%	<b>5.41*10</b> <sup>-4</sup>	0.002	8.4%	8.50*10 <sup>-2</sup>	<b>0.008</b> <sup>\$</sup>	34.8%	<b>3.21*10</b> <sup>-4</sup>
C8	0.003	1.9%	$6.06*10^{-1}$	-0.005	-3.1%	5.56*10 <sup>-1</sup>	0.013 <sup>\$</sup>	11.0%	4.38*10 <sup>-3</sup>
C8:1	0.012	15.4%	$1.62*10^{-1}$	0.008	11.8%	3.93*10 <sup>-1</sup>	0.017	18.9%	3.09*10 <sup>-1</sup>
C9	0.000	-0.4%	9.47*10 <sup>-1</sup>	-0.001	-4.7%	5.73*10 <sup>-1</sup>	0.001	5.4%	5.51*10 <sup>-1</sup>

**Table 3:** Difference between short sleep and normal sleep duration in acylcarnitine levels.

C10	0.014	5.8%	1.84*10 <sup>-1</sup>	0.003	1.1%	8.62*10 <sup>-1</sup>	0.027*	15.1%	4.54*10 <sup>-3</sup>
C10:1	0.008	6.9%	$2.68*10^{-1}$	-0.004	-3.1%	$6.68*10^{-1}$	0.022*	23.9%	2.72*10 <sup>-2</sup>
C10:2	0.000	-0.7%	8.65*10 <sup>-1</sup>	0.000	-0.1%	9.79*10 <sup>-1</sup>	0.000	-1.3%	$8.51^{*}10^{-1}$
C12	0.012 <sup>\$</sup>	17.2%	<b>1.70*10</b> <sup>-3</sup>	0.010	12.0%	9.00*10 <sup>-2</sup>	0.015 <sup>\$</sup>	27.5%	<b>3.30*10</b> <sup>-3</sup>
C12:1	0.017 <sup>\$</sup>	23.6%	1.74*10 <sup>-3</sup>	0.014	17.0%	8.32*10 <sup>-2</sup>	0.021 <sup>\$</sup>	35.2%	5.85*10 <sup>-3</sup>
C12-DC	0.002	2.4%	1.53*10 <sup>-1</sup>	0.002	2.1%	3.78*10 <sup>-1</sup>	0.003	2.8%	$3.01^{*}10^{-1}$
C14	0.004 <sup>\$</sup>	15.6%	<b>1.21*10<sup>-2</sup></b>	0.002	8.5%	<b>2.42*10</b> <sup>-1</sup>	<b>0.006</b> <sup>\$</sup>	28.0%	1.97*10 <sup>-2</sup>
C14:1	0.020 <sup>#</sup>	32.4%	<b>2.67*10</b> <sup>-4</sup>	<b>0.017</b> <sup>\$</sup>	23.5%	2.34*10 <sup>-2</sup>	0.024 <sup>\$</sup>	49.1%	6.55*10 <sup>-3</sup>
C14:1-OH	0.001	6.2%	3.79*10 <sup>-1</sup>	0.000	-1.7%	$8.10^{+10^{-1}}$	0.001	16.9%	$2.36*10^{-1}$
C14:2	0.006 <sup>\$</sup>	26.1%	<b>5.48*10</b> <sup>-4</sup>	0.005	17.3%	5.01*10 <sup>-2</sup>	0.008 <sup>\$</sup>	41.5%	<b>4.17*10</b> <sup>-3</sup>
C14:2-OH	0.000	5.0%	3.64*10 <sup>-1</sup>	0.000	6.4%	2.91*10 <sup>-1</sup>	0.000	3.3%	7.48*10 <sup>-1</sup>
C16	0.007 <sup>\$</sup>	10.8%	1.94*10 <sup>-2</sup>	0.005	6.7%	<b>2.05*10</b> <sup>-1</sup>	0.010	17.1%	5.91*10 <sup>-2</sup>
C16:1	0.005 <sup>\$</sup>	8.8%	9.60*10 <sup>-3</sup>	0.005 <sup>\$</sup>	8.3%	3.06*10 <sup>-2</sup>	0.005	9.6%	<b>1.55*10</b> <sup>-1</sup>
C16:1-OH	0.001 <sup>\$</sup>	20.8%	5.71*10 <sup>-3</sup>	0.001	11.8%	<b>1.90*10</b> <sup>-1</sup>	0.002 <sup>\$</sup>	33.8%	1.10*10 <sup>-2</sup>
C16:2	0.001 <sup>\$</sup>	21.3%	9.72*10 <sup>-3</sup>	0.001	9.6%	<b>2.39*10</b> <sup>-1</sup>	0.002 <sup>\$</sup>	39.5%	<b>1.91*10<sup>-2</sup></b>
C16:2-OH	0.000	2.1%	6.29*10 <sup>-1</sup>	0.000	3.8%	5.45*10 <sup>-1</sup>	0.000	-0.1%	9.83*10 <sup>-1</sup>
C16-OH	0.000	0.9%	8.89*10 <sup>-1</sup>	0.000	-3.4%	5.84*10 <sup>-1</sup>	0.000	6.1%	$6.49^{*}10^{-1}$
C18	0.003	10.8%	7.34*10 <sup>-2</sup>	0.002	9.6%	3.04*10 <sup>-1</sup>	0.003	12.5%	$1.18^{*}10^{-1}$
C18:1	0.016 <sup>#</sup>	22.3%	<b>1.92*10</b> <sup>-4</sup>	0.015*	20.1%	<b>2.96*10</b> <sup>-3</sup>	0.019 <sup>\$</sup>	25.1%	<b>2.84*10<sup>-2</sup></b>
C18:1-OH	0.001	12.8%	6.02*10 <sup>-2</sup>	0.001	5.7%	3.93*10 <sup>-1</sup>	0.002	22.3%	1.07*10 <sup>-1</sup>
C18:2	0.007 <sup>#</sup>	27.0%	1.32*10 <sup>-4</sup>	0.005 <sup>\$</sup>	20.7%	<b>9.01*10</b> <sup>-4</sup>	0.010 <sup>\$</sup>	34.4%	<b>1.38*10<sup>-2</sup></b>

<sup>1</sup>Difference in metabolite levels (μM) as measured by Biocrates/DQ<sup>TM</sup> p150 kit between short and normal sleep duration. Positive mean difference indicates an increase after short sleep duration. Negative mean difference indicates a decrease after short sleep duration.

<sup>2</sup>Change (%) represents percentage of change in metabolite level in short compared to normal sleep (metabolite level (short sleep) – metabolite level (normal sleep)) / metabolite level (normal sleep).

DM1 = individuals with type 1 diabetes. P-values are based on paired Students t-tests. N= 16 (healthy: n=9, DM1: n=7). Full results table is shown in Supplemental Table S2. Abbreviations of acylcarnitines are shown in Supplemental Table S3.

\$: Significant difference (p<0.05). #: Significant difference after Bonferroni correction (p<3.0\*10<sup>-4</sup> (=0.05/163)). Significant differences metabolites in all subjects are displayed in bold.

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Metabolite	Interaction P-value	Effect of sleep P-value	Effect of DM1 status P-value
C2	3.24 * 10 <sup>-2</sup>	6.70 * 10 <sup>-3\$</sup>	8.33 * 10 <sup>-1</sup>
C7-DC	1.10 * 10 <sup>-3</sup>	<1.0 * 10 <sup>-4#</sup>	4.53 * 10 <sup>-1</sup>
C12	<b>4.48 * 10</b> <sup>-1</sup>	1.80 * 10 <sup>-3#</sup>	2.35 * 10 <sup>-1</sup>
C12:1	<b>4.43</b> * 10 <sup>-1</sup>	<b>1.90</b> * 10 <sup>-3#</sup>	<b>2.25</b> * 10 <sup>-1</sup>
C14	<b>2.24 * 10</b> <sup>-1</sup>	8.70 * 10 <sup>-3#</sup>	1.01 * 10 <sup>-1</sup>
C14:1	4.05 * 10 <sup>-1</sup>	<b>3.00</b> * 10 <sup>-4#</sup>	<b>1.68 * 10</b> <sup>-1</sup>
C14:2	<b>2.30 * 10</b> <sup>-1</sup>	<b>4.00</b> * 10 <sup>-4#</sup>	<b>4.22</b> * 10 <sup>-1</sup>
C16	<b>3.48 * 10</b> <sup>-1</sup>	<b>1.68</b> * 10 <sup>-2\$</sup>	<b>2.09</b> * 10 <sup>-1</sup>
C16:1	8.94 * 10 <sup>-1</sup>	1.25 * 10 <sup>-2\$</sup>	<b>3.82</b> * 10 <sup>-1</sup>
С16:1-ОН	<b>1.42 * 10</b> <sup>-1</sup>	<b>3.30</b> * 10 <sup>-3#</sup>	8.79 * 10 <sup>-1</sup>
C16:2	8.34 * 10 <sup>-2</sup>	<b>4.30</b> * 10 <sup>-3\$</sup>	6.62 * 10 <sup>-1</sup>
C18:1	5.97 * 10 <sup>-1</sup>	<b>3.00</b> * 10 <sup>-4#</sup>	<b>7.64</b> * 10 <sup>-1</sup>
C18:2	1.35 * 10 <sup>-1</sup>	<1.00 * 10 <sup>-4#</sup>	1.81 * 10 <sup>-1</sup>

**Table 4.** Interaction effects of diabetes status and short sleep on increased acylcarnitine levels

DM1 = individuals with type 1 diabetes. Abbreviations of acylcarnitines are shown in Supplemental Table S3.

<sup>\$</sup>p<0.05, #p<0.004 (0.05/13) (two way repeated measure ANOVA).

#### SUPPLEMENTARY TABLES

Table S1. General population characteristics of healthy individuals and patients with type 1 diabetes<sup>1</sup>.

	Healthy	DM1
Females (%)	4 (44%)	4 (57%)
Age (years)	45 ± 14	43 ± 16
BMI (kg/m²)	23.8 ± 2.2	23.5 ± 2.2
WHR	0.88 ± 0.05	0.81 ± 0.09

DM1 = individuals with type 1 diabetes. BMI = body mass index. WHR = waist hip ratio. Healthy individuals n= 9, DM n= 7. Data are represented as mean  $\pm$  SD (percentage). <sup>1</sup>Data previously published separately (14;15).

	Healthy			DM1			Healthy vs. DM1	
	Normal sleep	Short sleep	Effect of sleep	Normal sleep	Short sleep	Effect of sleep	Normal sleep	Short sleep
Sleep characteristics			P-value			P-value	P-value	P-value
TST (min)	454 ± 26	228 ± 32	<0.0001	469 ± 22	222 ± 19	<0.0001	0.237	0.761
Stage 1 (% of TST)	10 ± 3	11 ± 6	0.490	11 ± 3	10 ± 6	0.868	0.577	0.664
Stage 2 (% of TST)	41 ± 7	35 ± 10	0.006	44 ± 7	41 ± 6	0.160	0.533	0.327
Stage 3 (% of TST) (SWS)	25 ± 5	33 ± 9	0.007	23 ± 9	35 ± 12	0.006	0.570	0.447
REM sleep (% of TST)	24 ± 5	21±8	0.364	22 ± 4	14 ± 6	0.038	0.684	0.069
Sleep efficiency (%)	91 ± 5	90 ± 7	0.699	94 ± 2	93 ± 8	0.484	0.199	0.464
Plasma parameter				5				
Free fatty acids (mmol/l)	0.63 ± 0.15	$0.62 \pm 0.14$	0.76	0.67 ± 0.31	0.59 ± 0.24	0.23	0.74	0.81
Insulin sensitivity parameters								
EGP (µmol*kg LBM <sup>-1</sup> *min <sup>-1</sup> )	3.57 ± 0.6	4.43 ± 0.8	0.017	6.21 ± 1.9	6.88 ± 1.4	0.505	0.003	0.001
GDR (µmol*kg LBM <sup>-1</sup> *min <sup>-1</sup> )	40.7 ± 14.3	32.5 ± 10.2	0.009	25.5 ± 6.4	22.1 ± 5.1	0.039	0.028	0.035
GIR (µmol*kg LBM <sup>-1</sup> *min <sup>-1</sup> )	36.9 ± 14.4	27.8 ± 10.5	0.006	19.0 ± 7.0	$14.9 \pm 5.0$	0.041	0.014	0.014

## Table S2. Sleep and insulin sensitivity parameters of healthy individuals and patients with type 1 diabetes<sup>1</sup>.

DM1= individuals with type 1 diabetes. Insulin sensitivity parameters were determined by hyperinsulinemic euglycemic clamp. EGP = Endogenous glucose production, GDR = glucose disposal rate (glucose Rd), GIR = Glucose infusion rate. Sleep characteristics were determined by polysomnography. TST = total sleep time. SWS = slow wave sleep. Free fatty acids were measured in basal fasting plasma samples. Effect of sleep intervention was tested with paired Students T-test, significant differences shown in bold. Healthy individuals n= 9, DM1 n = 7. Data is presented as means ± SD.

<sup>1</sup>Data previously published in separately (14;15).

# Table S3. Metabolites determined by Biocrates*IDQ*<sup>™</sup> p150 kit.

Metabolite Class	Short name	Biochemical Name					
	СО	DL-Carnitine					
	C2	Acetyl-L-carnitine					
	С3	Propionyl-L-carnitine					
	C3:1	Propenyl-L-carnitine					
	C3-DC / C4-OH	Malonyl-L-carnitine / hydroxybutyryl-L-carnitine					
	СЗ-DС-М / С5-ОН	Methylmalonyl-L-carnitine / hydroxyvaleryl-L-carnitine					
	СЗ-ОН	Hydroxypropionyl-L-carnitine					
	C4	Butyryl-L-carnitine					
	C4:1	Butenyl-L-carnitine					
	C4:1-DC / C6	Fumaryl-L-carnitine/Hexanoyl-L-carnitine					
	C5	Valeryl-L-carnitine					
ines	C5:1	TiglyI-L-carnitine					
arnit	C5:1-DC	Glutaconyl-L-carnitine					
cylc	С5-DС / С6-ОН	Glutaryl-L-carnitine/Hydroxyhexanoyl-L-carnitine					
4	C5-M-DC	Methylglutaryl-L-carnitine					
	C6:1	Hexenoyl-L-carnitine					
	C7-DC	Pimelyl-L-carnitine					
	C8	Octanoyl-L-carnitine					
	C8:1	Octenoyl-L-carnitine					
	С9	Nonayl-L-carnitine					
	C10	Decanoyl-L-carnitine					
	C10:1	Decenoyl-L-carnitine					
	C10:2	Decadienyl-L-carnitine					
	C12	Dodecanoyl-L-carnitine					
	C12:1	Dodecenoyl-L-carnitine					

	C12-DC	Dodecanedioyl-L-carnitine					
	C14	Tetradecanoyl-L-carnitine					
	C14:1	Tetradecenoyl-L-carnitine					
	C14:1-OH	Hydroxytetradecenoyl-L-carnitine					
	C14:2	Tetradecadienyl-L-carnitine					
	С14:2-ОН	Hydroxytetradecadienyl-L-carnitine					
10	C16	Hexadecanoyl-L-carnitine					
tines	C16:1	Hexadecenoyl-L-carnitine					
arni	С16:1-ОН	Hydroxyhexadecenoyl-L-carnitine					
vcylc	C16:2	Hexadecadienyl-L-carnitine					
4	С16:2-ОН	Hydroxyhexadecadienyl-L-carnitine					
	С16-ОН	Hydroxyhexadecanoyl-L-carnitine					
	C18	Octadecanoyl-L-carnitine					
	C18:1	Octadecenoyl-L-carnitine					
	С18:1-ОН	Hydroxyoctadecenoyl-L-carnitine					
	C18:2	Octadecadienyl-L-carnitine					
Sugars	н1	Hexose					
	Arg	Arginine					
	Gln	Glutamine					
	Gly	Glycine					
st	His	Histidine					
) acic	Met	Methionine					
nino	Orn	Ornithine					
An	Phe	Phenylalanine					
	Pro	Proline					
	Ser	Serine					
	Thr	Threonine					

	Тгр	Tryptophan					
•	Tyr	Tyrosine					
minc	Val	Valine					
a A	xLeu	xLeucine					
	lysoPC a C14:0	lysoPhosphatidylcholine acyl C14:0					
	lysoPC a C16:0	lysoPhosphatidylcholine acyl C16:0					
	lysoPC a C16:1	lysoPhosphatidylcholine acyl C16:1					
	lysoPC a C17:0	lysoPhosphatidylcholine acyl C17:0					
	lysoPC a C18:0	lysoPhosphatidylcholine acyl C18:0					
	lysoPC a C18:1	lysoPhosphatidylcholine acyl C18:1					
	lysoPC a C18:2	lysoPhosphatidylcholine acyl C18:2					
	lysoPC a C20:3	lysoPhosphatidylcholine acyl C20:3					
	lysoPC a C20:4	lysoPhosphatidylcholine acyl C20:4					
pids	lysoPC a C24:0	lysoPhosphatidylcholine acyl C24:0					
holi	lysoPC a C26:0	lysoPhosphatidylcholine acyl C26:0					
dsor	lysoPC a C26:1	lysoPhosphatidylcholine acyl C26:1					
sropl	lysoPC a C28:0	lysoPhosphatidylcholine acyl C28:0					
slyce	lysoPC a C28:1	lysoPhosphatidylcholine acyl C28:1					
	lysoPC a C6:0	lysoPhosphatidylcholine acyl C6:0					
	PC aa C24:0	Phosphatidylcholine diacyl C 24:0					
	PC aa C26:0	Phosphatidylcholine diacyl C 26:0					
	PC aa C28:1	Phosphatidylcholine diacyl C 28:1					
	PC aa C30:0	Phosphatidylcholine diacyl C 30:0					
Y	PC aa C30:2	Phosphatidylcholine diacyl C 30:2					
	PC aa C32:0	Phosphatidylcholine diacyl C 32:0					
	PC aa C32:1	Phosphatidylcholine diacyl C 32:1					
	PC aa C32:2	Phosphatidylcholine diacyl C 32:2					

	PC aa C32:3	Phosphatidylcholine diacyl C 32:3
	PC aa C34:1	Phosphatidylcholine diacyl C 34:1
	PC aa C34:2	Phosphatidylcholine diacyl C 34:2
	PC aa C34:3	Phosphatidylcholine diacyl C 34:3
	PC aa C34:4	Phosphatidylcholine diacyl C 34:4
	PC aa C36:0	Phosphatidylcholine diacyl C 36:0
	PC aa C36:1	Phosphatidylcholine diacyl C 36:1
	PC aa C36:2	Phosphatidylcholine diacyl C 36:2
	PC aa C36:3	Phosphatidylcholine diacyl C 36:3
	PC aa C36:4	Phosphatidylcholine diacyl C 36:4
	PC aa C36:5	Phosphatidylcholine diacyl C 36:5
	PC aa C36:6	Phosphatidylcholine diacyl C 36:6
pids	PC aa C38:0	Phosphatidylcholine diacyl C 38:0
holi	PC aa C38:1	Phosphatidylcholine diacyl C 38:1
dsor	PC aa C38:3	Phosphatidylcholine diacyl C 38:3
ropł	PC aa C38:4	Phosphatidylcholine diacyl C 38:4
ilyce	PC aa C38:5	Phosphatidylcholine diacyl C 38:5
0	PC aa C38:6	Phosphatidylcholine diacyl C 38:6
	PC aa C40:1	Phosphatidylcholine diacyl C 40:1
	PC aa C40:2	Phosphatidylcholine diacyl C 40:2
	PC aa C40:3	Phosphatidylcholine diacyl C 40:3
	PC aa C40:4	Phosphatidylcholine diacyl C 40:4
	PC aa C40:5	Phosphatidylcholine diacyl C 40:5
$\rightarrow$	PC aa C40:6	Phosphatidylcholine diacyl C 40:6
	PC aa C42:0	Phosphatidylcholine diacyl C 42:0
	PC aa C42:1	Phosphatidylcholine diacyl C 42:1
	PC aa C42:2	Phosphatidylcholine diacyl C 42:2

PC aa C42:4	Phosphatidylcholine diacyl C 42:4
PC aa C42:5	Phosphatidylcholine diacyl C 42:5
PC aa C42:6	Phosphatidylcholine diacyl C 42:6
PC ae C30:0	Phosphatidylcholine acyl-alkyl C 30:0
PC ae C30:1	Phosphatidylcholine acyl-alkyl C 30:1
PC ae C30:2	Phosphatidylcholine acyl-alkyl C 30:2
PC ae C32:1	Phosphatidylcholine acyl-alkyl C 32:1
PC ae C32:2	Phosphatidylcholine acyl-alkyl C 32:2
PC ae C34:0	Phosphatidylcholine acyl-alkyl C 34:0
PC ae C34:1	Phosphatidylcholine acyl-alkyl C 34:1
PC ae C34:2	Phosphatidylcholine acyl-alkyl C 34:2
PC ae C34:3	Phosphatidylcholine acyl-alkyl C 34:3
PC ae C36:0	Phosphatidylcholine acyl-alkyl C 36:0
PC ae C36:1	Phosphatidylcholine acyl-alkyl C 36:1
PC ae C36:2	Phosphatidylcholine acyl-alkyl C 36:2
PC ae C36:3	Phosphatidylcholine acyl-alkyl C 36:3
PC ae C36:4	Phosphatidylcholine acyl-alkyl C 36:4
PC ae C36:5	Phosphatidylcholine acyl-alkyl C 36:5
PC ae C38:0	Phosphatidylcholine acyl-alkyl C 38:0
PC ae C38:1	Phosphatidylcholine acyl-alkyl C 38:1
PC ae C38:2	Phosphatidylcholine acyl-alkyl C 38:2
PC ae C38:3	Phosphatidylcholine acyl-alkyl C 38:3
PC ae C38:4	Phosphatidylcholine acyl-alkyl C 38:4
PC ae C38:5	Phosphatidylcholine acyl-alkyl C 38:5
PC ae C38:6	Phosphatidylcholine acyl-alkyl C 38:6
PC ae C40:0	Phosphatidylcholine acyl-alkyl C 40:0
PC ae C40:1	Phosphatidylcholine acyl-alkyl C 40:1
1	

Glycerophospholipids

	PC ae C40:2	Phosphatidylcholine acyl-alkyl C 40:2					
	PC ae C40:3	Phosphatidylcholine acyl-alkyl C 40:3					
	PC ae C40:4	Phosphatidylcholine acyl-alkyl C 40:4					
	PC ae C40:5	Phosphatidylcholine acyl-alkyl C 40:5					
	PC ae C40:6	Phosphatidylcholine acyl-alkyl C 40:6					
	PC ae C42:0	Phosphatidylcholine acyl-alkyl C 42:0					
	PC ae C42:1	Phosphatidylcholine acyl-alkyl C 42:1					
pids	PC ae C42:2	Phosphatidylcholine acyl-alkyl C 42:2					
holi	PC ae C42:3	Phosphatidylcholine acyl-alkyl C 42:3					
dsor	PC ae C42:4	Phosphatidylcholine acyl-alkyl C 42:4					
ropt	PC ae C42:5	Phosphatidylcholine acyl-alkyl C 42:5					
ilyce	PC ae C44:3	Phosphatidylcholine acyl-alkyl C 44:3					
0	PC ae C44:4	Phosphatidylcholine acyl-alkyl C 44:4					
	PC ae C44:5	Phosphatidylcholine acyl-alkyl C 44:5					
	PC ae C44:6	Phosphatidylcholine acyl-alkyl C 44:6					
	SM (OH) C14:1	Hydroxysphingomyeline C 14:1					
	SM (OH) C16:0	Hydroxysphingomyeline C 16:0					
	SM (OH) C22:1	Hydroxysphingomyeline C 22:1					
	SM (OH) C22:2	Hydroxysphingomyeline C 22:2					
s	SM (OH) C24:1	Hydroxysphingomyeline C 24:1					
olipid	SM C16:0	Sphingomyeline C 16:0					
lingo	SM C16:1	Sphingomyeline C 16:1					
sph	SM C18:0	Sphingomyeline C 18:0					
	SM C18:1	Sphingomyeline C 18:1					
	SM C20:2	Sphingomyeline C 20:2					
	SM C22:3	Sphingomyeline C 22:3					
	SM C24:0	Sphingomyeline C 24:0					

SM C24-1	Sphingomyeline C 24:1
SIVI U24.1	Springoniyenne C 24.1
SM C26:0	Sphingomyeline C 26:0
SM C26:1	Sphingomyeline C 26:1
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	All subjects			Healthy			DM1		
Metabolite	Mean	Change	P-value	Mean	Change (%) <sup>2</sup>	P-value	Mean	Change (%) <sup>2</sup>	P-value
C0	0.191	0.6%	8.81*10 <sup>-1</sup>	0.398	1.2%	8.47*10 <sup>-1</sup>	-0.075	-0.3%	9.61*10 <sup>-1</sup>
C2	0.662	16.8%	2.28*10 <sup>-2</sup>	0.184	4.3%	5.40*10 <sup>-1</sup>	1.276	35.6%	1.33*10 <sup>-2</sup>
C3	-0.006	-2.6%	6.76*10 <sup>-1</sup>	-0.002	-0.7%	9.33*10 <sup>-1</sup>	-0.012	-6.2%	6.03*10 <sup>-1</sup>
C3-DC (C4-OH)	0.007	11.3%	4.55*10 <sup>-1</sup>	0.006	8.0%	7.20*10 <sup>-1</sup>	0.009	17.7%	2.87*10 <sup>-1</sup>
C3-OH	0.001	3.3%	5.03*10 <sup>-1</sup>	0.002	8.5%	5.91*10 <sup>-2</sup>	-0.001	-2.1%	8.24*10 <sup>-1</sup>
C3:1	-0.001	-5.9%	4.16*10 <sup>-1</sup>	0.000	-0.1%	9.93*10 <sup>-1</sup>	-0.001	-12.3%	1.58*10 <sup>-1</sup>
C4	0.011	10.3%	1.22*10 <sup>-1</sup>	0.008	7.2%	4.95*10 <sup>-1</sup>	0.015	14.4%	8.68*10 <sup>-2</sup>
C4:1	0.002	6.7%	2.90*10 <sup>-1</sup>	0.002	8.7%	2.55*10 <sup>-1</sup>	0.001	4.8%	6.58*10 <sup>-1</sup>
C5	0.011	10.2%	2.80*10 <sup>-1</sup>	0.012	10.4%	4.59*10 <sup>-1</sup>	0.010	9.8%	4.30*10 <sup>-1</sup>
C5-DC (C6-OH)	-0.001	-4.1%	4.87*10 <sup>-1</sup>	-0.001	-8.3%	2.56*10 <sup>-1</sup>	0.000	0.7%	9.45*10 <sup>-1</sup>
C5-M-DC	0.000	0.3%	9.32*10 <sup>-1</sup>	0.000	-1.0%	8.45*10 <sup>-1</sup>	0.001	1.8%	7.11*10 <sup>-1</sup>
C5-OH (C3-DC-M)	0.000	0.3%	9.40*10 <sup>-1</sup>	0.000	-1.5%	6.89*10 <sup>-1</sup>	0.001	2.4%	7.40*10 <sup>-1</sup>
C5:1	0.000	1.0%	8.74*10 <sup>-1</sup>	-0.001	-2.5%	7.86*10 <sup>-1</sup>	0.001	5.0%	6.20*10 <sup>-1</sup>
C5:1-DC	0.000	2.6%	6.75*10 <sup>-1</sup>	-0.001	-4.8%	5.96*10 <sup>-1</sup>	0.002	12.5%	1.27*10 <sup>-1</sup>
C6 (C4:1-DC)	0.005	8.7%	1.22*10 <sup>-1</sup>	0.001	1.9%	7.82*10 <sup>-1</sup>	0.011	18.8%	6.20*10 <sup>-2</sup>
C6:1	0.001	2.6%	3.99*10 <sup>-1</sup>	0.001	3.5%	3.35*10 <sup>-1</sup>	0.000	1.5%	8.05*10 <sup>-1</sup>
C7-DC	0.005	19.9%	5.41*10 <sup>-4</sup>	0.002	8.4%	8.50*10 <sup>-2</sup>	0.008	34.8%	3.21*10 <sup>-4</sup>
C8	0.003	1.9%	6.06*10 <sup>-1</sup>	-0.005	-3.1%	5.56*10 <sup>-1</sup>	0.013	11.0%	4.38*10 <sup>-3</sup>
C8:1	0.012	15.4%	1.62*10 <sup>-1</sup>	0.008	11.8%	3.93*10 <sup>-1</sup>	0.017	18.9%	3.09*10 <sup>-1</sup>
C9	0.000	-0.4%	9.47*10 <sup>-1</sup>	-0.001	-4.7%	5.73*10 <sup>-1</sup>	0.001	5.3%	5.51*10 <sup>-1</sup>
C10	0.014	5.8%	1.84*10 <sup>-1</sup>	0.003	1.1%	8.62*10 <sup>-1</sup>	0.027	15.1%	4.54*10 <sup>-3</sup>
C10:1	0.008	6.9%	2. <del>6</del> 8*10 <sup>-1</sup>	-0.004	-3.1%	6. <del>6</del> 8*10 <sup>-1</sup>	0.022	23.9%	2.72*10 <sup>-2</sup>

 Table S4. Metabolite changes after one night of short sleep duration.

C10:2	0.000	-0.7%	8.65*10 <sup>-1</sup>	0.000	-0.1%	9.79*10 <sup>-1</sup>	0.000	-1.3%	8.51*10 <sup>-1</sup>
C12	0.012	17.2%	1.70*10 <sup>-3</sup>	0.010	12.0%	9.00*10 <sup>-2</sup>	0.015	27.5%	3.30*10 <sup>-3</sup>
C12-DC	0.002	2.4%	1.53*10 <sup>-1</sup>	0.002	2.1%	3.78*10 <sup>-1</sup>	0.003	2.8%	3.01*10 <sup>-1</sup>
C12:1	0.017	23.6%	1.74*10 <sup>-3</sup>	0.014	17.0%	8.32*10 <sup>-2</sup>	0.021	35.2%	5.85*10 <sup>-3</sup>
C14	0.004	15.6%	1.21*10 <sup>-2</sup>	0.002	8.5%	2.42*10 <sup>-1</sup>	0.006	28.0%	1.97*10 <sup>-2</sup>
C14:1	0.020	32.4%	2.67*10 <sup>-4</sup>	0.017	23.5%	2.34*10 <sup>-2</sup>	0.024	49.1%	6.55*10 <sup>-3</sup>
C14:1-OH	0.001	6.2%	3.79*10 <sup>-1</sup>	0.000	-1.7%	8.10*10 <sup>-1</sup>	0.001	16.9%	2.36*10 <sup>-1</sup>
C14:2	0.006	26.1%	5.48*10 <sup>-4</sup>	0.005	17.3%	5.01*10 <sup>-2</sup>	0.008	41.5%	4.17*10 <sup>-3</sup>
C14:2-OH	0.000	5.0%	3.64*10 <sup>-1</sup>	0.000	6.4%	2.91*10 <sup>-1</sup>	0.000	3.3%	7.48*10 <sup>-1</sup>
C16	0.007	10.8%	1.94*10 <sup>-2</sup>	0.005	6.7%	2.05*10 <sup>-1</sup>	0.010	17.1%	5.91*10 <sup>-2</sup>
C16-OH	0.000	0.9%	8.89*10 <sup>-1</sup>	0.000	-3.4%	5.84*10 <sup>-1</sup>	0.000	6.0%	6.49*10 <sup>-1</sup>
C16:1	0.005	8.8%	9.60*10 <sup>-3</sup>	0.005	8.3%	3.06*10 <sup>-2</sup>	0.005	9.6%	1.55*10 <sup>-1</sup>
C16:1-OH	0.001	20.8%	5.71*10 <sup>-3</sup>	0.001	11.7%	1.90*10 <sup>-1</sup>	0.002	33.8%	1.10*10 <sup>-2</sup>
C16:2	0.001	21.3%	9.72*10 <sup>-3</sup>	0.001	9.6%	2.39*10 <sup>-1</sup>	0.002	39.5%	1.91*10 <sup>-2</sup>
C16:2-OH	0.000	2.1%	6.29*10 <sup>-1</sup>	0.000	3.8%	5.45*10 <sup>-1</sup>	0.000	-0.1%	9.83*10 <sup>-1</sup>
C18	0.003	10.8%	7.34*10 <sup>-2</sup>	0.002	9.6%	3.04*10 <sup>-1</sup>	0.003	12.5%	1.18*10 <sup>-1</sup>
C18:1	0.016	22.3%	1.92*10 <sup>-4</sup>	0.015	20.1%	2.96*10 <sup>-3</sup>	0.019	25.1%	2.84*10 <sup>-2</sup>
C18:1-OH	0.001	12.8%	6.02*10 <sup>-2</sup>	0.001	5.7%	3.93*10 <sup>-1</sup>	0.002	22.3%	1.07*10 <sup>-1</sup>
C18:2	0.007	27.0%	1.32*10 <sup>-4</sup>	0.005	20.7%	9.01*10 <sup>-4</sup>	0.010	34.4%	1.38*10 <sup>-2</sup>
Arg	4.297	6.0%	2.74*10 <sup>-1</sup>	6.988	10.0%	2.66*10 <sup>-1</sup>	0.838	1.1%	8.57*10 <sup>-1</sup>
Gln	13.186	4.5%	2.52*10 <sup>-1</sup>	10.500	3.6%	4.88*10 <sup>-1</sup>	16.641	5.7%	4.00*10 <sup>-1</sup>
Gly	-9.002	-4.7%	2.84*10 <sup>-1</sup>	-3.264	-1.9%	7.57*10 <sup>-1</sup>	-16.380	-7.4%	2.69*10 <sup>-1</sup>
His	0.533	1.0%	8.27*10 <sup>-1</sup>	-0.174	-0.3%	9.57*10 <sup>-1</sup>	1.441	3.0%	7.32*10 <sup>-1</sup>
Met	0.264	1.3%	7.98*10 <sup>-1</sup>	0.557	2.6%	6.91*10 <sup>-1</sup>	-0.113	-0.6%	9.48*10 <sup>-1</sup>
Orn	3.038	8.3%	1.48*10 <sup>-1</sup>	5.466	15.2%	1.05*10 <sup>-1</sup>	-0.083	-0.2%	9.70*10 <sup>-1</sup>

Phe	2.588	6.2%	1.06*10 <sup>-1</sup>	3.131	7.6%	1.16*10 <sup>-1</sup>	1.890	4.5%	5.13*10 <sup>-1</sup>
Pro	-3.161	-1.9%	6.20*10 <sup>-1</sup>	-0.909	-0.5%	9.10*10 <sup>-1</sup>	-6.057	-4.0%	5.93*10 <sup>-1</sup>
Ser	3.273	3.6%	4.27*10 <sup>-1</sup>	2.253	2.7%	7.18*10 <sup>-1</sup>	4.585	4.6%	4.29*10 <sup>-1</sup>
Thr	4.527	4.7%	5.04*10 <sup>-1</sup>	4.930	5.1%	5.55*10 <sup>-1</sup>	4.010	4.1%	7.47*10 <sup>-1</sup>
Trp	1.178	1.9%	5.56*10 <sup>-1</sup>	2.563	4.2%	3.30*10 <sup>-1</sup>	-0.602	-1.0%	8.58*10 <sup>-1</sup>
Tyr	4.638	9.8%	1.05*10 <sup>-1</sup>	6.966	14.7%	1.30*10 <sup>-1</sup>	1.644	3.5%	6.12*10 <sup>-1</sup>
Val	3.933	2.6%	4.86*10 <sup>-1</sup>	6.183	3.9%	5.06*10 <sup>-1</sup>	1.039	0.7%	8.67*10 <sup>-1</sup>
xLeu	9.916	6.4%	1.56*10 <sup>-1</sup>	9.156	5.7%	3.55*10 <sup>-1</sup>	10.892	7.3%	3.24*10 <sup>-1</sup>
lysoPC a C14:0	-0.047	-1.4%	5.83*10 <sup>-1</sup>	-0.022	-0.6%	8.67*10 <sup>-1</sup>	-0.079	-2.4%	5.04*10 <sup>-1</sup>
lysoPC a C16:0	-1.774	-2.4%	5.84*10 <sup>-1</sup>	0.872	1.3%	8.39*10 <sup>-1</sup>	-5.176	-6.6%	3.34*10 <sup>-1</sup>
lysoPC a C16:1	-0.144	-7.2%	1.58*10 <sup>-1</sup>	-0.061	-3.1%	6.72*10 <sup>-1</sup>	-0.249	-12.3%	1.02*10 <sup>-1</sup>
lysoPC a C17:0	-0.015	-1.3%	7.88*10 <sup>-1</sup>	0.005	0.5%	9.46*10 <sup>-1</sup>	-0.042	-3.4%	6.57*10 <sup>-1</sup>
lysoPC a C18:0	-0.003	0.0%	9.97*10 <sup>-1</sup>	1.077	6.4%	3.22*10 <sup>-1</sup>	-1.392	-6.9%	4.33*10 <sup>-1</sup>
lysoPC a C18:1	-0. 7	-2.9%	5.91*10 <sup>-1</sup>	-0.210	-1.6%	7.91*10 <sup>-1</sup>	-0.728	-4.2%	6.67*10 <sup>-1</sup>
lysoPC a C18:2	0.575	1.9%	8.14*10 <sup>-1</sup>	-1.447	-5.5%	4.00*10 <sup>-1</sup>	3.176	8.9%	5.59*10 <sup>-1</sup>
lysoPC a C20:3	0.008	0.4%	9.43*10 <sup>-1</sup>	0.080	4.0%	5.80*10 <sup>-1</sup>	-0.085	-4.5%	6.19*10 <sup>-1</sup>
lysoPC a C20:4	-0.248	-4.7%	3.27*10 <sup>-1</sup>	-0.302	-6.3%	2.31*10 <sup>-1</sup>	-0.179	-3.1%	7.32*10 <sup>-1</sup>
lysoPC a C24:0	-0.003	-0.5%	9.28*10 <sup>-1</sup>	0.014	3.1%	6.56*10 <sup>-1</sup>	-0.024	-4.7%	6.56*10 <sup>-1</sup>
lysoPC a C26:0	-0.009	-0.9%	9.02*10 <sup>-1</sup>	0.014	1.4%	8.63*10 <sup>-1</sup>	-0.039	-4.1%	7.87*10 <sup>-1</sup>
lysoPC a C26:1	-0.004	-1.1%	8.81*10 <sup>-1</sup>	-0.002	-0.6%	9.42*10 <sup>-1</sup>	-0.007	-1.7%	9.03*10 <sup>-1</sup>
lysoPC a C28:0	0.001	0.1%	9.84*10 <sup>-1</sup>	0.012	1.5%	8.49*10 <sup>-1</sup>	-0.013	-1.7%	8.84*10 <sup>-1</sup>
lysoPC a C28:1	-0.006	-0.6%	9.28*10 <sup>-1</sup>	0.010	1.1%	8.84*10 <sup>-1</sup>	-0.025	-2.6%	8.31*10 <sup>-1</sup>
lysoPC a C6:0	-0.002	-8.4%	4.94*10 <sup>-1</sup>	0.002	10.9%	6.26*10 <sup>-1</sup>	-0.007	-26.5%	1.14*10 <sup>-2</sup>
PC aa C24:0	-0.005	-1.8%	7.93*10 <sup>-1</sup>	-0.005	-1.9%	8.42*10 <sup>-1</sup>	-0.005	-1.8%	8.78*10 <sup>-1</sup>
PC aa C26:0	-0.028	-2.4%	6.22*10 <sup>-1</sup>	-0.040	-3.4%	5.37*10 <sup>-1</sup>	-0.013	-1.1%	9.09*10 <sup>-1</sup>

PC aa C28:1	0.061	2.7%	3.74*10 <sup>-1</sup>	0.089	3.9%	2.47*10 <sup>-1</sup>	0.026	1.1%	8.47*10 <sup>-1</sup>
PC aa C30:0	0.081	2.7%	6.62*10 <sup>-1</sup>	0.099	3.1%	6.76*10 <sup>-1</sup>	0.058	2.1%	8.60*10 <sup>-1</sup>
PC aa C30:2	0.002	1.2%	9.33*10 <sup>-1</sup>	0.010	7.0%	7.71*10 <sup>-1</sup>	-0.009	-4.3%	8.40*10 <sup>-1</sup>
PC aa C32:0	0.352	4.0%	2.19*10 <sup>-1</sup>	0.490	5.8%	5.12*10 <sup>-2</sup>	0.174	1.9%	7.76*10 <sup>-1</sup>
PC aa C32:1	0.370	3.7%	6.36*10 <sup>-1</sup>	1.063	9.4%	3.68*10 <sup>-1</sup>	-0.521	-6.5%	6.17*10 <sup>-1</sup>
PC aa C32:2	0.020	0.8%	9.12*10 <sup>-1</sup>	-0.038	-1.3%	8.84*10 <sup>-1</sup>	0.095	4.1%	7.38*10 <sup>-1</sup>
PC aa C32:3	0.001	0.2%	9.68*10 <sup>-1</sup>	-0.005	-1.3%	7.93*10 <sup>-1</sup>	0.007	1.9%	7.90*10 <sup>-1</sup>
PC aa C34:1	2.395	1.9%	5.09*10 <sup>-1</sup>	5.489	4.3%	1.84*10 <sup>-1</sup>	-1.583	-1.3%	8.16*10 <sup>-1</sup>
PC aa C34:2	7.304	3.7%	1.04*10 <sup>-1</sup>	6.544	3.3%	1.31*10 <sup>-1</sup>	8.282	4.1%	3.78*10 <sup>-1</sup>
PC aa C34:3	0.353	3.0%	5.32*10 <sup>-1</sup>	0.435	3.5%	5.90*10 <sup>-1</sup>	0.247	2.2%	7.78*10 <sup>-1</sup>
PC aa C34:4	-0.013	-1.2%	8.47*10 <sup>-1</sup>	-0.032	-2.7%	7.55*10 <sup>-1</sup>	0.012	1.3%	8.98*10 <sup>-1</sup>
PC aa C36:0	0.017	0.7%	9.23*10 <sup>-1</sup>	-0.046	-2.1%	8.05*10 <sup>-1</sup>	0.099	3.3%	7.81*10 <sup>-1</sup>
PC aa C36:1	1.673	5.8%	1.72*10 <sup>-1</sup>	2.555	8.7%	1.40*10 <sup>-1</sup>	0.538	1.9%	7.75*10 <sup>-1</sup>
PC aa C36:2	6.029	4.8%	1.46*10 <sup>-1</sup>	6.133	4.9%	1.96*10 <sup>-1</sup>	5.894	4.7%	4.61*10 <sup>-1</sup>
PC aa C36:3	3.808	4.5%	1.64*10 <sup>-1</sup>	6.521	7.5%	5.27*10 <sup>-2</sup>	0.320	0.4%	9.47*10 <sup>-1</sup>
PC aa C36:4	1.441	1.3%	6.29*10 <sup>-1</sup>	1.110	1.0%	7.39*10 <sup>-1</sup>	1.866	1.9%	7.49*10 <sup>-1</sup>
PC aa C36:5	-0.045	-0.3%	9.65*10 <sup>-1</sup>	0.559	4.1%	7.51*10 <sup>-1</sup>	-0.821	-6.8%	3.50*10 <sup>-1</sup>
PC aa C36:6	-0.010	-1.5%	8.53*10 <sup>-1</sup>	0.004	0.5%	9.69*10 <sup>-1</sup>	-0.028	-4.4%	6.29*10 <sup>-1</sup>
PC aa C38:0	0.050	2.0%	6.68*10 <sup>-1</sup>	0.039	1.7%	8.06*10 <sup>-1</sup>	0.065	2.3%	7.41*10 <sup>-1</sup>
PC aa C38:1	0.041	2.3%	6.99*10 <sup>-1</sup>	0.151	9.3%	2.19*10 <sup>-1</sup>	-0.101	-5.0%	5.98*10 <sup>-1</sup>
PC aa C38:3	2.020	7.5%	6.46*10 <sup>-2</sup>	3.788	12.9%	1.68*10 <sup>-2</sup>	-0.254	-1.1%	8.48*10 <sup>-1</sup>
PC aa C38:4	1.113	2.0%	5.25*10 <sup>-1</sup>	1.760	3.0%	4.88*10 <sup>-1</sup>	0.280	0.6%	9.16*10 <sup>-1</sup>
PC aa C38:5	0.060	0.2%	9.55*10 <sup>-1</sup>	0.609	2.3%	7.31*10 <sup>-1</sup>	-0.645	-2.6%	5.62*10 <sup>-1</sup>
PC aa C38:6	-0.622	-1.6%	7.12*10 <sup>-1</sup>	-0.987	-2.3%	7.03*10 <sup>-1</sup>	-0.154	-0.4%	9.47*10 <sup>-1</sup>
PC aa C40:1	0.020	2.5%	6.90*10 <sup>-1</sup>	0.060	8.0%	3.34*10 <sup>-1</sup>	-0.031	-3.5%	7.38*10 <sup>-1</sup>

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PC aa C40:2	0.078	5.4%	4.46*10	0.149	11.7%	2.16*10	-0.012	-0.7%	9.52*10 -
PC aa C40:3	0.068	4.6%	4.70*10 <sup>-1</sup>	0.136	10.5%	2.55*10 <sup>-1</sup>	-0.020	-1.2%	9.04*10 <sup>-1</sup>
PC aa C40:4	0.154	4.6%	2.35*10 <sup>-1</sup>	0.290	8.9%	1.09*10 <sup>-1</sup>	-0.021	-0.6%	9.16*10 <sup>-1</sup>
PC aa C40:5	0.232	4.1%	3.01*10 <sup>-1</sup>	0.540	9.3%	9.49*10 <sup>-2</sup>	-0.164	-3.0%	5.89*10 <sup>-1</sup>
PC aa C40:6	0.033	0.3%	9.48*10 <sup>-1</sup>	0.413	3.3%	5.51*10 <sup>-1</sup>	-0.454	-4.4%	5.85*10 <sup>-1</sup>
PC aa C42:0	0.028	4.0%	2.80*10 <sup>-1</sup>	0.040	6.2%	2.00*10 <sup>-1</sup>	0.012	1.6%	8.01*10 <sup>-1</sup>
PC aa C42:1	0.009	1.9%	7.04*10 <sup>-1</sup>	0.041	10.0%	1.49*10 <sup>-1</sup>	-0.033	-6.2%	3.74*10 <sup>-1</sup>
PC aa C42:2	0.020	3.1%	6.55*10 <sup>-1</sup>	0.054	9.5%	3.06*10 <sup>-1</sup>	-0.024	-3.1%	7.75*10 <sup>-1</sup>
PC aa C42:4	0.031	5.6%	2.44*10 <sup>-1</sup>	0.053	10.6%	1.88*10 <sup>-1</sup>	0.002	0.4%	9.49*10 <sup>-1</sup>
PC aa C42:5	0.029	5.5%	1.84*10 <sup>-1</sup>	0.047	9.7%	1.34*10 <sup>-1</sup>	0.005	0.8%	8.81*10 <sup>-1</sup>
PC aa C42:6	0.019	3.4%	5.38*10 <sup>-1</sup>	0.049	9.3%	1.60*10 <sup>-1</sup>	-0.020	-3.5%	7.21*10 <sup>-1</sup>
PC ae C30:0	0.002	0.5%	9.20*10 <sup>-1</sup>	0.012	3.5%	6.07*10 <sup>-1</sup>	-0.012	-3.6%	5.95*10 <sup>-1</sup>
PC ae C30:1	-0.003	-1.2%	8.94*10 <sup>-1</sup>	-0.003	-1.2%	9.07*10 <sup>-1</sup>	-0.004	-1.3%	9.43*10 <sup>-1</sup>
PC ae C30:2	0.006	2.5%	6.96*10 <sup>-1</sup>	0.011	5.1%	5.28*10 <sup>-1</sup>	-0.001	-0.3%	9.81*10 <sup>-1</sup>
PC ae C32:1	0.046	2.4%	4.28*10 <sup>-1</sup>	0.035	1.9%	6.30*10 <sup>-1</sup>	0.060	2.9%	5.66*10 <sup>-1</sup>
PC ae C32:2	0.011	2.2%	4.36*10 <sup>-1</sup>	0.017	3.7%	3.50*10 <sup>-1</sup>	0.003	0.6%	8.99*10 <sup>-1</sup>
PC ae C34:0	0.006	0.7%	8.25*10 <sup>-1</sup>	0.015	1.6%	6.90*10 <sup>-1</sup>	-0.005	-0.5%	9.25*10 <sup>-1</sup>
PC ae C34:1	0.159	2.7%	3.75*10 <sup>-1</sup>	0.202	3.3%	4.23*10 <sup>-1</sup>	0.105	1.8%	7.15*10 <sup>-1</sup>
PC ae C34:2	0.274	4.1%	4.06*10 <sup>-1</sup>	0.029	0.4%	9.36*10 <sup>-1</sup>	0.589	8.7%	3.55*10 <sup>-1</sup>
PC ae C34:3	0.213	3.9%	3.04*10 <sup>-1</sup>	0.093	1.8%	7.10*10 <sup>-1</sup>	0.368	6.3%	3.34*10 <sup>-1</sup>
PC ae C36:0	0.048	6.1%	1.66*10 <sup>-1</sup>	0.037	4.6%	3.13*10 <sup>-1</sup>	0.063	8.0%	3.68*10 <sup>-1</sup>
PC ae C36:1	0.832	6.7%	3.11*10 <sup>-1</sup>	0.792	6.9%	3.45*10 <sup>-1</sup>	0.883	6.4%	5.98*10 <sup>-1</sup>
PC ae C36:2	0.308	2.9%	3.51*10 <sup>-1</sup>	0.152	1.4%	7.39*10 <sup>-1</sup>	0.508	4.7%	3.39*10 <sup>-1</sup>
PC ae C36:3	0.143	2.8%	5.30*10 <sup>-1</sup>	0.044	0.9%	8.77*10 <sup>-1</sup>	0.272	5.4%	5.13*10 <sup>-1</sup>
PC ae C36:4	0.257	2.7%	5.65*10 <sup>-1</sup>	-0.112	-1.2%	7.36*10 <sup>-1</sup>	0.732	7.4%	4.57*10 <sup>-1</sup>

PC ae C36:5	0.079	1.2%	7.36*10 <sup>-1</sup>	-0.063	-0.9%	8.10*10 <sup>-1</sup>	0.260	3.7%	5.64*10 <sup>-1</sup>
PC ae C38:0	0.054	2.8%	6.14*10 <sup>-1</sup>	0.049	2.4%	7.92*10 <sup>-1</sup>	0.061	3.4%	5.34*10 <sup>-1</sup>
PC ae C38:1	0.263	6.4%	4.50*10 <sup>-1</sup>	0.411	11.8%	3.06*10 <sup>-1</sup>	0.073	1.5%	9.13*10 <sup>-1</sup>
PC ae C38:2	0.414	6.9%	3.07*10 <sup>-1</sup>	0.631	11.3%	1.77*10 <sup>-1</sup>	0.134	2.1%	8.61*10 <sup>-1</sup>
PC ae C38:3	0.776	7.3%	1.96*10 <sup>-1</sup>	0.799	7.6%	2.04*10 <sup>-1</sup>	0.745	6.9%	5.38*10 <sup>-1</sup>
PC ae C38:4	0.189	2.2%	5.31*10 <sup>-1</sup>	0.199	2.3%	6.11*10 <sup>-1</sup>	0.176	2.0%	7.39*10 <sup>-1</sup>
PC ae C38:5	0.132	1.3%	7.42*10 <sup>-1</sup>	-0.130	-1.3%	7.68*10 <sup>-1</sup>	0.470	4.4%	5.50*10 <sup>-1</sup>
PC ae C38:6	0.096	2.5%	5.50*10 <sup>-1</sup>	-0.033	-0.9%	8.70*10 <sup>-1</sup>	0.261	6.8%	3.48*10 <sup>-1</sup>
PC ae C40:0	0.010	0.3%	9.35*10 <sup>-1</sup>	0.044	1.1%	8.20*10 <sup>-1</sup>	-0.032	-0.9%	8.54*10 <sup>-1</sup>
PC ae C40:1	0.040	1.8%	7.94*10 <sup>-1</sup>	0.031	1.5%	8.49*10 <sup>-1</sup>	0.051	2.2%	8.67*10 <sup>-1</sup>
PC ae C40:2	0.144	5.2%	3.60*10 <sup>-1</sup>	0.233	8.5%	2.19*10 <sup>-1</sup>	0.031	1.1%	9.15*10 <sup>-1</sup>
PC ae C40:3	0.349	7.0%	1.55*10 <sup>-1</sup>	0.441	9.1%	1.16*10 <sup>-1</sup>	0.231	4.5%	6.22*10 <sup>-1</sup>
PC ae C40:4	0.223	4.5%	3.41*10 <sup>-1</sup>	0.332	7.1%	2.64*10 <sup>-1</sup>	0.082	1.6%	8.42*10 <sup>-1</sup>
PC ae C40:5	0.278	4.4%	3.21*10 <sup>-1</sup>	0.241	3.8%	5.41*10 <sup>-1</sup>	0.325	5.2%	4.65*10 <sup>-1</sup>
PC ae C40:6	0.051	1.8%	6.17*10 <sup>-1</sup>	0.057	2.0%	7.26*10 <sup>-1</sup>	0.044	1.5%	7.36*10 <sup>-1</sup>
PC ae C42:0	0.028	4.0%	2.93*10 <sup>-1</sup>	0.049	7.1%	1.08*10 <sup>-1</sup>	0.000	0.1%	9.94*10 <sup>-1</sup>
PC ae C42:1	0.044	4.5%	4.52*10 <sup>-1</sup>	0.075	8.4%	2.37*10 <sup>-1</sup>	0.004	0.3%	9.75*10 <sup>-1</sup>
PC ae C42:2	0.023	2.3%	6.72*10 <sup>-1</sup>	0.057	6.1%	4.38*10 <sup>-1</sup>	-0.020	-1.8%	8.26*10 <sup>-1</sup>
PC ae C42:3	0.047	3.4%	5.37*10 <sup>-1</sup>	0.090	7.0%	3.21*10 <sup>-1</sup>	-0.007	-0.5%	9.59*10 <sup>-1</sup>
PC ae C42:4	0.060	4.2%	2.59*10 <sup>-1</sup>	0.120	8.7%	1.16*10 <sup>-1</sup>	-0.017	-1.1%	8.27*10 <sup>-1</sup>
PC ae C42:5	0.119	4.3%	1.96*10 <sup>-1</sup>	0.149	5.4%	2.06*10 <sup>-1</sup>	0.081	2.9%	6.20*10 <sup>-1</sup>
PC ae C44:3	0.011	3.0%	6.00*10 <sup>-1</sup>	0.022	6.8%	4.11*10 <sup>-1</sup>	-0.003	-0.8%	9.26*10 <sup>-1</sup>
PC ae C44:4	0.016	3.3%	2.93*10 <sup>-1</sup>	0.040	8.1%	5.12*10 <sup>-2</sup>	-0.014	-2.7%	5.62*10 <sup>-1</sup>
PC ae C44:5	0.041	2.8%	3.86*10 <sup>-1</sup>	0.058	3.9%	3.78 <sup>*</sup> 10 <sup>-1</sup>	0.018	1.2%	8.06 <sup>*</sup> 10 <sup>-1</sup>
PC ae C44:6	0.021	1.9%	6.18*10 <sup>-1</sup>	0.060	5.7%	2.69*10 <sup>-1</sup>	-0.031	-2.7%	6.51*10 <sup>-1</sup>

SM (OH) C14:1	0.146	2.6%	2.81*10 <sup>-1</sup>	0.213	4.0%	1.67*10 <sup>-1</sup>	0.059	1.0%	8.19*10 <sup>-1</sup>
SM (OH) C16:1	0.096	3.7%	1.60*10 <sup>-1</sup>	0.084	3.3%	3.39*10 <sup>-1</sup>	0.111	4.2%	3.54*10 <sup>-1</sup>
SM (OH) C22:1	0.271	2.4%	5.34*10 <sup>-1</sup>	0.730	6.9%	1.15*10 <sup>-1</sup>	-0.320	-2.7%	7.02*10 <sup>-1</sup>
SM (OH) C22:2	-0.036	-0.4%	8.93*10 <sup>-1</sup>	0.084	1.0%	7.57*10 <sup>-1</sup>	-0.191	-2.0%	7.29*10 <sup>-1</sup>
SM (OH) C24:1	0.021	1.8%	7.53*10 <sup>-1</sup>	0.075	6.5%	9.51*10 <sup>-2</sup>	-0.049	-4.0%	7.43*10 <sup>-1</sup>
SM C16:0	3.468	3.9%	1.56*10 <sup>-1</sup>	4.587	5.4%	8.66*10 <sup>-2</sup>	2.030	2.1%	6.71*10 <sup>-1</sup>
SM C16:1	0.585	4.0%	1.77*10 <sup>-1</sup>	0.568	4.3%	1.42*10 <sup>-1</sup>	0.606	3.8%	5.14*10 <sup>-1</sup>
SM C18:0	0.859	5.0%	9.13*10 <sup>-2</sup>	0.375	2.1%	5.47*10 <sup>-1</sup>	1.482	8.9%	9.81*10 <sup>-2</sup>
SM C18:1	0.265	3.2%	3.15*10 <sup>-1</sup>	0.014	0.2%	9.65*10 <sup>-1</sup>	0.588	6.8%	2.07*10 <sup>-1</sup>
SM C20:2	0.024	5.5%	3.80*10 <sup>-1</sup>	0.010	2.3%	7.28*10 <sup>-1</sup>	0.042	9.9%	4.41*10 <sup>-1</sup>
SM C22:3	-0.017	-5.8%	7.54*10 <sup>-1</sup>	-0.021	-7.4%	8.15*10 <sup>-1</sup>	-0.061	-19.9%	2.00*10 <sup>-1</sup>
SM C24:0	1.132	5.0%	1.61*10 <sup>-1</sup>	2.201	9.9%	3.84*10 <sup>-2</sup>	-0.242	-1.0%	8.48*10 <sup>-1</sup>
SM C24:1	2.222	4.7%	6.72*10 <sup>-2</sup>	2.991	6.6%	3.74*10 <sup>-2</sup>	1.234	2.5%	5.82*10 <sup>-1</sup>
SM C26:0	0.032	51.0%	1.31*10 <sup>-1</sup>	0.023	24.4%	3.09*10 <sup>-1</sup>	0.050	139.3%	9.71*10 <sup>-2</sup>
SM C26:1	0.017	5.1%	4.07*10 <sup>-1</sup>	-0.008	-2.5%	7.53*10 <sup>-1</sup>	0.048	14.8%	1.48*10 <sup>-1</sup>
H1	419.618	7.8%	3.58*10 <sup>-1</sup>	126.200	2.8%	3.61*10 <sup>-1</sup>	796.870	12.3%	4.65*10 <sup>-1</sup>

<sup>1</sup> Difference in metabolite levels ( $\mu$ M) as measured by Biocrates/ $DQ^{TM}$  p150 kit between short and normal sleep duration. Positive mean difference indicates an increase after short sleep duration. Negative mean difference indicates a decrease after short sleep duration.

<sup>2</sup>Change (%) represents percentage of change in metabolite level in short compared to normal sleep (metabolite level (short sleep) – metabolite level (normal sleep)) / metabolite level (normal sleep).

DM1 = individuals with type 1 diabetes. P-values are based on paired Students t-tests. Abbreviations of all metabolites are shown in Supplemental Table S3. N= 16 (healthy: n=9, DM1: n=7).

**Table S5:** Acylcarnitine levels after normal sleep duration.

	Healthy		DM1	Healthy vs. DM1	
Metabolite	Mean	SD	Mean	SD	P-value
C0	31.9	5.8	26.6	4.8	0.073
C2	4.24	0.57	3.58	1.06	0.133
С3	0.273	0.043	0.192	0.042	0.002
C3:1	0.010	0.002	0.012	0.003	0.139
C3-DC (C4-OH)	0.074	0.062	0.049	0.011	0.304
СЗ-ОН	0.020	0.003	0.024	0.005	0.020
C4	0.105	0.019	0.105	0.057	0.979
C4:1	0.022	0.003	0.031	0.007	0.005
C5	0.120	0.032	0.097	0.027	0.153
C5:1	0.022	0.003	0.024	0.004	0.148
C5:1-DC	0.020	0.004	0.019	0.002	0.628
C5-DC (C6-OH)	0.017	0.003	0.019	0.005	0.273
C5-M-DC	0.035	0.003	0.039	0.004	0.047
С5-ОН (С3-DC-М)	0.023	0.003	0.025	0.004	0.405
C6 (C4:1-DC)	0.066	0.022	0.058	0.014	0.385
C6:1	0.027	0.003	0.026	0.002	0.828
C7-DC	0.025	0.008	0.024	0.008	0.975
C8	0.172	0.126	0.121	0.066	0.356
C8:1	0.065	0.009	0.088	0.043	0.151
С9	0.025	0.010	0.024	0.005	0.817
C10	0.274	0.228	0.181	0.073	0.321
C10:1	0.124	0.073	0.093	0.039	0.324
C10:2	0.027	0.006	0.028	0.004	0.593
C12	0.084	0.051	0.055	0.015	0.179
C12:1	0.080	0.035	0.059	0.018	0.163
C12-DC	0.087	0.005	0.101	0.005	0.000#
C14	0.029	0.008	0.022	0.004	0.044
C14:1	0.071	0.028	0.049	0.015	0.080
С14:1-ОН	0.009	0.002	0.009	0.001	0.684
C14:2	0.027	0.016	0.020	0.008	0.300
C14:2-OH	0.007	0.001	0.007	0.001	0.277
C16	0.072	0.018	0.060	0.010	0.136
C16:1	0.059	0.004	0.056	0.006	0.231
C16:1-OH	0.006	0.001	0.005	0.001	0.174
C16:2	0.007	0.002	0.006	0.001	0.164
С16:2-ОН	0.010	0.001	0.010	0.002	0.684

C16-OH	0.006	0.001	0.007	0.003	0.714
C18	0.025	0.007	0.024	0.007	0.798
C18:1	0.073	0.009	0.074	0.019	0.949
C18:1-OH	0.009	0.001	0.009	0.001	0.668
C18:2	0.025	0.006	0.028	0.006	0.467

Mean = mean plasma metabolite level ( $\mu$ M). DM1 = individuals with type 1 diabetes. <sup>#</sup>P<0.001 (0.05/41). P-values are based on independent Students t-tests. Abbreviations of all metabolites are shown in Supplemental Table S3. Healthy individuals n=9, DM1 n=7.

Table S6: Acylcarnitine levels after short sleep duration.

	Healthy		DM1	Healthy vs. DM1	
Metabolite	Mean	SD	Mean	SD	P-value
C0	32.3	7.0	26.6	6.4	0.113
C2	0.287	0.212	0.208	0.078	0.434
С3	0.120	0.057	0.115	0.037	0.839
C3:1	0.027	0.005	0.028	0.005	0.680
C3-DC (C4-OH)	0.094	0.056	0.070	0.018	0.309
СЗ-ОН	0.094	0.034	0.080	0.023	0.356
C4	0.089	0.006	0.104	0.011	0.004
C4:1	0.032	0.008	0.028	0.006	0.300
C5	0.087	0.032	0.073	0.026	0.336
C5:1	0.009	0.001	0.010	0.003	0.261
C5:1-DC	0.032	0.016	0.028	0.008	0.595
C5-DC (C6-OH)	0.007	0.001	0.008	0.002	0.447
C5-M-DC	0.077	0.016	0.070	0.014	0.398
С5-ОН (С3-DC-М)	0.063	0.006	0.061	0.011	0.577
C6 (C4:1-DC)	0.007	0.002	0.007	0.002	0.610
C6:1	0.008	0.002	0.008	0.003	0.715
C7-DC	0.010	0.001	0.010	0.001	0.937
C8	0.006	0.001	0.007	0.002	0.213
C8:1	0.027	0.009	0.027	0.006	0.932
C9	0.088	0.014	0.092	0.024	0.664
C10	0.010	0.002	0.011	0.002	0.225
C10:1	0.031	0.006	0.037	0.009	0.103
C10:2	4.421	1.169	4.859	1.478	0.517
C12	0.271	0.065	0.180	0.039	0.006
C12:1	0.010	0.002	0.010	0.003	0.810
C12-DC	0.080	0.047	0.057	0.015	0.241
C14	0.021	0.003	0.024	0.005	0.255
C14:1	0.113	0.028	0.120	0.072	0.793

C14:1-OH	0.024	0.004	0.032	0.007	0.012
C14:2	0.132	0.038	0.106	0.036	0.188
C14:2-OH	0.021	0.005	0.026	0.006	0.121
C16	0.019	0.004	0.021	0.004	0.209
C16:1	0.015	0.003	0.019	0.003	0.028
С16:1-ОН	0.034	0.005	0.040	0.007	0.086
C16:2	0.023	0.003	0.025	0.005	0.222
C16:2-OH	0.068	0.016	0.069	0.025	0.919
С16-ОН	0.027	0.003	0.027	0.004	0.637
C18	0.027	0.008	0.033	0.008	0.159
C18:1	0.166	0.109	0.135	0.063	0.507
C18:1-OH	0.073	0.028	0.104	0.048	0.128
C18:2	0.024	0.007	0.025	0.005	0.645

Mean = mean plasma metabolite level ( $\mu$ M). DM1 = individuals with type 1 diabetes. P-values are based on independent Students t-tests. Abbreviations of all metabolites are shown in Supplemental Table S3. Healthy individuals n=9, DM1 n=7.

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- One night of short sleep acutely increases plasma acylcarnitine levels
- Short sleep increases acylcarnitine in healthy individuals and patients with DM1
- Acylcarnitines reflect fatty acid oxidation and have pro-inflammatory properties
- Acylcarnitines may mediate the relation between short sleep and insulin resistance