

## Primary cell culture systems for human thyroid studies

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### Abstract

**Background:** Cell models are key instruments for *in vitro* studies of the thyroid. Permanent thyroid cell lines that are widely used in laboratory research typically originate from tumors. For many purposes it is desirable to compare tumor cells with cells originating from normal tissue. However, such cultures grow slowly, have a highly limited lifespan and are known to lose their thyroid characteristics. The aim of the present study was to type coding and noncoding thyroid

markers in different culture systems in an attempt to determine the optimal conditions for *in vitro* experimentation.

**Methods:** Human primary thyroid cells were isolated from histologically non-tumorous tissues. Two alternative media (6H and h7H) were used. The morphology and behavior of the ensuing monolayer (2-dimensional) cultures was monitored by microscopy. The expression of key thyroid-related genes (n=9) was monitored by RT-PCR on Days 8, 21 and 43 after initiation. As a pilot study the same markers were studied in a 3-dimensional hanging-drop culture system.

**Results:** In the cultures with 6H or h7H medium, the primary thyroid cells displayed growth in numbers and size. Most cells retained the main morphological characteristics of thyroid cells throughout the first two weeks of culture and fibroblast-like cells appeared around Day 19. By Day 21, most thyroid gene markers were retained but by Day 43 several markers were no longer present. The lncRNA transcripts *PTCSC2* (spliced) and *PTCSC3* were the first to disappear. There were no fundamental differences between the two media in the early period of culture. In the 3-dimensional system most thyroid markers were retained by Day 21.

**Conclusion:** Cultures of thyroid cells retain many thyroid characteristics up to Day 21, thereafter fibroblast-like dedifferentiated cells begin to dominate.

## Introduction

Because of their convenience in long-term culture, thyroid cell lines have been used for decades as preclinical models for research purposes. These cell lines were derived from neoplastic cells and have been selected for strong proliferation *in vitro* (1). Therefore, these cells may have lost

some of their thyroid-specific features right from the beginning. As the cells adapt to the *in vitro* growth conditions they may not be able to maintain important characteristics which play key roles in normal thyroid function and signaling pathways. Moreover, by genetic analysis using short tandem repeats and single nucleotide polymorphisms, many thyroid cell lines were found to be misidentified or cross-contaminated with other cells (2). Consequently the usefulness of these cell lines for research targeting the thyroid cell has important limitations (3, 4). In the past, several laboratories have studied thyroid monolayer primary cultures and assessed their biological behavior. The effects of different culture media on the characteristics of the cells have been assessed using thyroid-specific markers (5-7). More recently novel thyroid specific non-coding genes such as the long noncoding RNA (lncRNA) genes papillary thyroid cancer susceptibility candidate 2 (*PTCSC2*) and papillary thyroid cancer susceptibility candidate 3 (*PTCSC3*) have been detected and their roles in thyroid biology and cancer have begun to be explored (8, 9). Whether these genes might be useful in monitoring the differentiation status of primary thyroid cells *in vitro* is presently not known.

In this study, we evaluated human 2-dimensional (2D) monolayer primary thyroid cell cultures grown in two different culture media, and compared 2D cultures with a 3-dimensional (3D) culture system. We describe cell morphological changes and report gene expression of a panel of 7 thyroid related coding genes and 2 lncRNA genes, *PTCSC2* and *PTCSC3*.

## Materials and Methods

The study was approved by the Institutional Review Board at the Ohio State University (OSU), and all subjects gave written informed consent before participation.

### *Thyroid tissue samples from patients*

A total of 29 fresh thyroid samples were included in the primary culture study. To establish primary cultures, histologically non-tumorous thyroid samples (see below) were obtained freshly from intraoperative processing in the Department of Pathology, The Ohio State University. The fresh tissues were kept in culture medium on ice while being transferred to the laboratory. Upon arrival in the laboratory the first step of the ensuing procedure was immediately initiated.

Three pairs of non-tumorous and tumor thyroid tissue samples were used as controls for RT-PCR and qRT-PCR. These samples were snap-frozen in liquid nitrogen and kept at -80 °C after being obtained from patients with PTC during surgery. All the cases were histologically diagnosed as papillary thyroid carcinoma; clinical information on these samples will be made available on request.

### *Culture media*

The cells were cultured in two previously described alternative media, consisting of modified F-12M medium containing six nutritional factors (6H medium) or alternatively Humanized Seven Homeostatic Additives Medium (h7H medium).

6H medium was made from modified F-12M medium (Coon's modification of Ham's F-12 medium) by adding 5% Heat Inactivated FBS, 2 mM Glutamine, 2.6 g/l NaHCO<sub>3</sub>, 5 ug/ml gentamicin, 1% NEAA and six nutrition factors: 10mU/ml TSH, 10 mU/ml Insulin, 1 nM Hydrocortisone, 2 ng/ml Glycyl-Histidyl-L-Lysine Acetate, 5µg/ml Transferrin and 10 ng/ml Somatostatin (6, 10, 11).

The medium h7H was also made from modified F-12M medium but with 5% Newborn Calf Serum (Life Technologies), 5% FBS (Gibco), 100 IU/ml penicillin–streptomycin, 2.5 ng/ml amphotericin B and other main components as previously described (5): 2.68 g/L Sodium Bicarbonate, 40 mU/L bovine TSH, 25 mU/L human Insulin, 23 nM Hydrocortisone, 0.2 µg/L Growth Hormone, 1 mg/L Apo-Transferrin, 10 µg/L NaI, 75 µg/L Na<sub>2</sub>SeO<sub>3</sub>, 0.2 mg/L L-Glutathione reduced, 0.5 mg/L ±α-tocopherol and 0.5 mg/L DL-α-tocopherol acetate. All the chemical reagents were purchased from Sigma-Aldrich unless specified.

#### *Primary cell preparation procedure and establishing 2D cultures*

The starting material was a piece of thyroid excised from a thyroid gland that had been surgically removed because of suspicion of thyroid cancer on preoperative evaluation. To ensure that the specimen did not contain neoplastic cells the piece was preferentially obtained from the lobe contralateral to that harboring the cancer. Alternatively, the specimen was excised as far away from the neoplastic lesion as possible. In either case, hematoxylin-eosin stained tissue sections were analyzed microscopically to verify the non-neoplastic nature of the specimen. The weight of specimens varied between 0.3 g and 1.5 g. The tissue was dissected and minced into fragments as small as possible using a sterile razor blade in a cell culture hood. After one wash in Hanks' Balanced Salt Solution (Life Technologies) the tissue fragments were transferred to 0.25% trypsin solution for an overnight digestion. On Day 2, the fragments were digested with 1% Trypsin (Life Technologies) and 0.35% Collagenase 4 (Worthington Biochemical) solution for 90 min at 37 °C after removal of the 0.25% trypsin from Day 1. The digested material was filtered through a nylon mesh (100 µm, FALCON); the undigested tissue fragments were processed in the same manner two more times. After centrifugation at 1000 g for 5 min, the

supernatant was discarded and 1 ml red blood cell lysing buffer (Sigma) was added for 2 min to eliminate the red blood cells and the reaction was stopped by adding 1 ml culture medium at room temperature. The cells were washed twice with Hanks' solution and centrifuged at 1000 g for 5 min. Finally, the cells were counted using a TC20™ Automated Cell Counter (Bio-rad) and seeded to a density of  $10^5$ ~ $10^6$  cells in 2 ml culture medium per well in six-well plates.

### *Three-dimensional (3D) culture*

Thyroid primary cells were prepared as described above for 2D culture. A total of 8000 to 10,000 cells per well were seeded in a Gravity PLUS™ 96 well plate for the 3D hanging drop culture system (Insphero). The 3D microtissues were formed by seeding cells onto the plate and maturing them for 5 days in hanging drops, and then followed by transferring the single spheroids into the 96-well Gravity TRAP™ assay plates. Commercial 3D InSight™ Cell Line Maintenance Media (Insphero) supplemented with 10 mU/ml TSH (Sigma-Aldrich) was used in the culture system. The medium was changed every 4-5 days in order to keep the 3D microtissue growing in the Gravity TRAP plate.

### *Cell lines and culture condition*

The BCPAP and KTC1 cell lines were incubated in antibiotic-free RPMI 1640 medium supplemented with 10% (vol/vol) FBS (Gibco) at 37 °C in humidified air with 5% (vol/vol) CO<sub>2</sub>.

### *Reverse transcription PCR and quantitative real-time RT-PCR*

Total RNA was extracted from cells using Trizol solution (Life Technologies) followed by a DNA contamination removal step by using DNA-free™ Kit (Ambion) according to the manufacturer's protocol. One microgram of purified RNA was reverse transcribed using High-Capacity cDNA Reverse Transcription Kit (Life Technologies) in the presence of random primers. *GAPDH* was employed as an internal control. RNA samples from fresh frozen thyroid tissue were included in RT-PCR reactions. Quantitative real-time RT-PCR assay was performed in triplicates of three biological replicates on ABI Prism 7900 HT Sequence Detection System (Applied Biosystems) according to the manufacturer's protocol. *PTCSC2*-Spliced and Unspliced isoforms were detected using TaqMan® Fast Universal PCR Master Mix (ThermoFisher SCIENTIFIC), and *PTCSC3*, *TSHR*, *TPO*, *TG*, *E-cadherin* and *FSP1* were detected by Fast SYBR Green Master Mix kit (ThermoFisher SCIENTIFIC). The primer and probe sequences are shown in Supplementary Table 1 and 2. Detailed  $\Delta C_t$  numbers used in qRT-PCR are listed in Supplementary Table 3.

### *Statistical analysis*

The comparison of quantitative real time RT-PCR between two groups was made by applying a *t* test (two tailed). A value of  $P < 0.05$  was considered statistically significant. Data are represented as mean  $\pm$  SD.

## **Results**

### *Cell morphology in thyroid primary cell cultures*

Human primary thyroid cells were obtained from non-tumorous thyroid tissue, disaggregated and digested as described in Material and Methods, then seeded onto Petri dishes in 6H or h7H culture medium. In order to describe the changes occurring in the morphology of the seeded cells, we performed microscopic observation of specific target cell clusters during culture (Figure 1 and Supplementary Figure 1). Comparing the behavior of cells from conventional cell lines such as BCPAP and KTC1 with the primary thyroid cells, we noted a conspicuous early difference. The cell line cells tended to settle evenly on the plate while the primary cells settled as aggregates of two, three or more cells located closely together. Of note, at the time of seeding, the cells formed single cell suspensions without clumping or clustering. During culture, once the primary thyroid cells finished settling down, gradual changes occurred from Day 2 to Day 7. There were increasing cell numbers as a result of cell proliferation and an increase in cell size (Figure 1 and Supplementary Figure 1). Comparing with conventional cell lines originated from tumor tissue (e.g. BCPAP and KTC1 cells), the growth of these primary cells is clearly slower in keeping with their derivation from non-tumorous tissue.

On approximately Day 19 changes in the morphology towards a more fibroblast-like appearance began to be observed (Figure 1). The cells displayed an elongated shape and grew much faster (3~4 days for one generation) than earlier. After approximately Day 27 the fibroblast-like cells constituted the majority of the cells in the culture. This change occurred at about the same time with both 6H medium and h7H medium. The cultures with 6H medium gradually stopped growing and finally reached a complete stop by approximately Day 43 but the cells usually remained attached to the Petri dish for more than 3 months. Notably the cultures in h7H medium kept growing rapidly for more than 11 generations (about 50 to 60 days) after the fibroblast-like cells appeared.



### *Thyroid marker evidence of dedifferentiation in 6H culture medium*

To examine the expression level of thyroid marker genes in the primary cultures with 6H medium, we performed RT-PCR analysis of the cells. Totally 9 thyroid marker genes were studied including 7 coding genes (*TG*, *TPO*, *TSHR*, *NIS*, *FOXE1*, *NKX2-1* and *PAX8*) and 2 lncRNA genes (*PTCSC2* and *PTCSC3*). These lncRNAs have been implicated in the predisposition to thyroid cancer; they are located close to *FOXE1* and *NKX2-1* on chromosomes 9q22 and 14q13, respectively (8, 9, 12). *PTCSC2* occurs in spliced and unspliced isoforms; both were tested here (Figure 2 A, C and E). On Day 8, most of the coding genes showed slightly reduced expression except for *TSHR* and *NIS* where abundance was much lower than in the two non-tumorous thyroid tissues. While the unspliced isoform of *PTCSC2* did not change much, the two spliced lncRNA genes both showed a strongly reduced abundance *in vitro* (Figure 2A). The expression of most markers did not change by Day 21 comparing to non-tumorous thyroid tissues, but on Day 43, signs of dedifferentiation became strong. (Figure 2C, E). Most marker transcripts including *PTCSC2* spliced isoform, *PTCSC3*, *TSHR*, *TPO* and *NIS* could hardly or not at all be detected. *NKX2-1* showed an extremely low expression, while a few markers such as *TG*, *PAX8* and *PTCSC2* unspliced isoform were still transcribed but at reduced levels.

Taken together, these results showed that the morphology of the primary cells in 2D cultures remained relatively unchanged by Day 8 with concomitant continued expression of most markers. By Day 21 and beyond, fibroblast-like cells increased in numbers and the abundance of the thyroid markers declined. This transformation process was completed by Day 43. The

markers signaling the earliest signs of dedifferentiation were the lncRNAs (*PTCSC2* and *PTCSC3*) and *TSHR*.

#### *Dedifferentiation of thyroid markers in h7H culture medium*

The medium h7H is a recently developed culture medium which may keep thyroid cells proliferating for many generations *in vitro* (5). We performed the same RT-PCR assays as reported above at 3 time points (Day 8, Day 21 and Day 43) on the primary cultures grown in h7H medium (Figure 2 B, D and F). Similar to the results obtained with the 6H medium, most of the markers showed reduced expression already at Day 8, especially *PTCSC2* (spliced), *PTCSC3*, *TSHR* and *NIS*. On Day 43, the *PTCSC2* spliced isoform was almost not transcribed (Figure 2F) and only a few markers (*PAX8*, *FOXE1* and *NKX2-1*) did not show a significant reduction. Compared with non-tumorous thyroid tissue, where all the markers were strongly expressed, the primary cell cultures in h7H medium showed remarkable weakening of most marker transcripts similar to cell cultured in 6H medium. The above results were confirmed by qRT-PCR with 6 selected thyroid markers (*PTCSC2* spliced and unspliced isoforms, *PTCSC3*, *TSHR*, *TPO* and *TG*) (Figure 3).

To further characterize the thyroid primary cultures, we analyzed the expression of 4 genes considered to be either epithelial to mesenchymal transition (EMT) markers and /or fibroblast cell markers. These 4 genes were: cadherin 1 (*CDH1*), also known as E-cadherin; S100 calcium binding protein A4 (*S100A4*), also known as fibroblast-specific protein 1 (*FSP1*); fibronectin 1 (*FNI*) and collagen, type I, alpha 1 (*COL1A1*) (13-17). We observed that *CDH1* and *S100A4* showed decreased expression, similar to most thyroid markers from Day 8 to Day 43 in the

primary cultures. In contrast, *FNI* and *COL1A1* showed increased expression in these cultures (Figure 4).

#### *Primary cells in a novel 3D culture system*

For a variety of laboratory experiments, the cells used should ideally be as close as possible to the tissue *in vivo* (18, 19). The cells grown in the conventional way undergo dedifferentiation, and eventually lose almost all the characteristics of a differentiated thyroid cell. We therefore performed a pilot study using a novel culture method in three dimensions (3D), which has been reported to more faithfully mirror the tissue of origin, although it has never been tested with normal human thyroid cells (20-22). After digestion and the formation of microtissue in a hanging-drop system for around 5 days, a single spheroid made of human thyroid cells could be observed under the microscope. These spheroids displayed a slow growth in size for around 3 weeks (Figure 5A), after which they collapsed into dead cells and cell debris.

In order to compare the 2D and 3D culture systems, the same expression assays were performed with the microtissues after 21 days in culture (Figure 5B). Different from 2D cultures at the same stage, almost all the markers displayed expression levels in the 3D experiment similar to the thyroid tissue control. Notably, the two lncRNAs *PTCSC2* and *PTCSC3* were present but with reduced expression. After 21 days, cell death occurred and cell clusters rapidly disintegrated.

## Discussion

The composition of the culture media is believed to be crucial for the outcome of *in vitro* cultures. Depending on the cell type and purpose, media have been designed by adding different concentrations of a variety of components such as inorganic salts, amino acids, vitamins, hormones and some other chemicals (23). For example, Vitamin C improves the speed and efficiency by which induced pluripotent stem cells are generated in both mouse and man (24). The removal of retinoic acid and triiodothyronine improves the differentiation from embryonic stem cells to pulmonary epithelial cells (25). Heparin has been shown to increase the generation of high yield oligodendrocyte progenitor cell enriched cultures (26).

In the current study, we encountered some, but no dramatic differences in both thyroid marker gene expression and cell morphology between two recommended primary thyroid culture media, 6H and h7H. In our hands, the h7H medium worked better to keep the cultures growing beyond the first few weeks (albeit with fibroblast-like cells), while the 6H medium performed better in maintaining the thyroid marker expression but only at the early stages (before Day 21). The h7H medium supported the growth for several generations (more than 60 days) but increasing dedifferentiation occurred. Some thyroid markers such as *TSHR* and *TPO* almost lost their expression in the h7H medium after 43 days of culture *in vitro*. Consequently, cells that have grown *in vitro* for more than 21 days are not likely to be good substitutes for native thyroid cells in typical laboratory experiments involving e.g. transfection and knock-in or knock-out procedures in either culture medium. This conclusion is not new (7), however the outcome of our experiments may provide a baseline for what should be expected from these cultures. Furthermore, since the loss of E-cadherin is a key marker of EMT, its reduction in cultures with h7H medium indicates that these primary cells can lose their cell-cell adhesion and polarity while

growing (27). The observation of increased expression of *FNI* and *COL1A1* during primary cultures from Day 8 to Day 43 was consistent with the appearance of fibroblast-like cells in late stages of the cultures; however, the *S100A4* gene showed decreased expression. We also noticed that the *PTCSC3* gene was slightly upregulated on Day 43. Future work is needed to understand these observations.

We observed that thyroid cells expand in the traditional 2D culture system soon after the primary cells had attached to the Petri dish. Considering the gene expression findings during this period, the inactivation of the marker genes may be a result of the loss of cell-cell interactions necessary for the cells to work in a functional thyroid follicle. The 3D culture performed better in maintaining the abundance of the thyroid markers, presumably due to better cell-cell interaction in the dense spheroid structure. Although the 3D culture may result in more differentiated cells than the conventional culture, possible disadvantages need to be addressed. The dense 3D structure probably makes it hard to introduce genes and nucleic acids into the cells in the inner part of the spheroid. Such an uneven penetrance was reported from experiments attempting to transfect labeled siRNAs into a cultured kidney rudiment (28). Moreover, the efficiency of electroporation was low in some mesenchymal tissues. Specifically, DNA could only be transfected into a limited number of cell layers (29). Taken together, the 3D culture system might be a future direction for primary cultures in order to establish a real substitution for native thyroid cells but further optimization such as increasing and improving the transfection efficiency is still necessary.

As reported, the two lncRNAs both displayed thyroid tissue-specific expression (8, 12). LncRNA *PTCSC2* has both spliced and unspliced isoforms that are expressed in thyroid tissue (8). *PTCSC2* and *PTCSC3* were discovered through their interaction with SNP markers rs965513

and rs944289 from genome-wide association studies (GWAS) of papillary thyroid cancer (30). These markers are associated with low concentrations of thyroid stimulating hormone (TSH) *in vivo*, and with thyroid cancer susceptibility (31-34). The lncRNAs are thought to perform their function by targeting nearby (*cis*) or far away (*trans*) genes (35, 36). At this point we can only speculate about potential roles of *PTCSC2* and *PTCSC3*, e.g. in maintaining thyroid cell-cell interaction. The mechanisms of action of these lncRNA genes are poorly known. It is plausible that the rapid decrease and disappearance of their transcripts almost immediately after the initiation of culture is related to the disaggregation of the thyrocytes. In this scenario it is the loss of cell-cell interaction that reduces lncRNA transcript abundance, which triggers the downregulation of coding thyroid-specific genes such as the *TSHR*.

In conclusion, we demonstrate that the primary culture of human thyroid cells in the conventional 2D culture produces early proliferation of thyroid-like cells expressing most thyroid markers. Cells harvested from these cultures during the first 3 weeks after seeding should be most useful for *in vitro* experiments. Comparing the commonly used 6H culture medium with the recently developed h7H medium, we observed that the latter helped maintaining long-term proliferation of the cells beyond 6 weeks, but these cells were fibroblast-like. Two lncRNA genes seem to be the first to lose their transcription or abundance when the cells dedifferentiate. A pilot experiment with a recently developed 3D culture method appears to demonstrate that dedifferentiation of the cells is less strong than in conventional 2D cultures for up to about 3 weeks after plating, at which point the cultures die and disintegrate.

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### **Disclosure Statement**

The authors declare that no competing interests exist.

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### **Figure legends**

Thyroid  
Primary cell culture systems for human thyroid studies (doi: 10.1089/thy.2015.0518)  
This article has been peer-reviewed and accepted for publication, but has yet to undergo copyediting and proof correction. The final published version may differ from this proof.

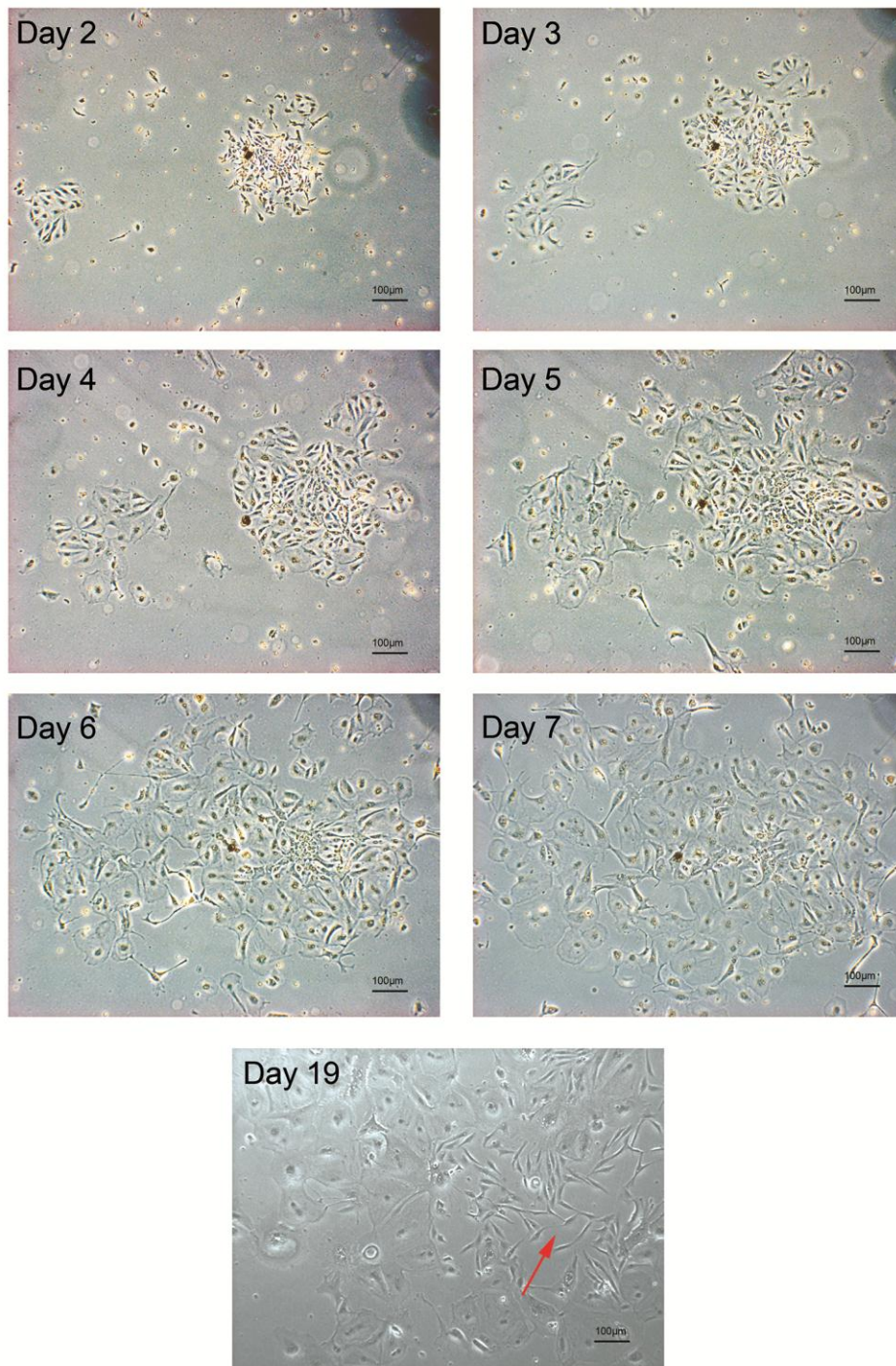


Figure 1. Morphology of 2-dimensional primary thyroid cell cultures after seeding. A red arrow indicates an emerging thyroid fibroblast-like cell. 50X magnification.



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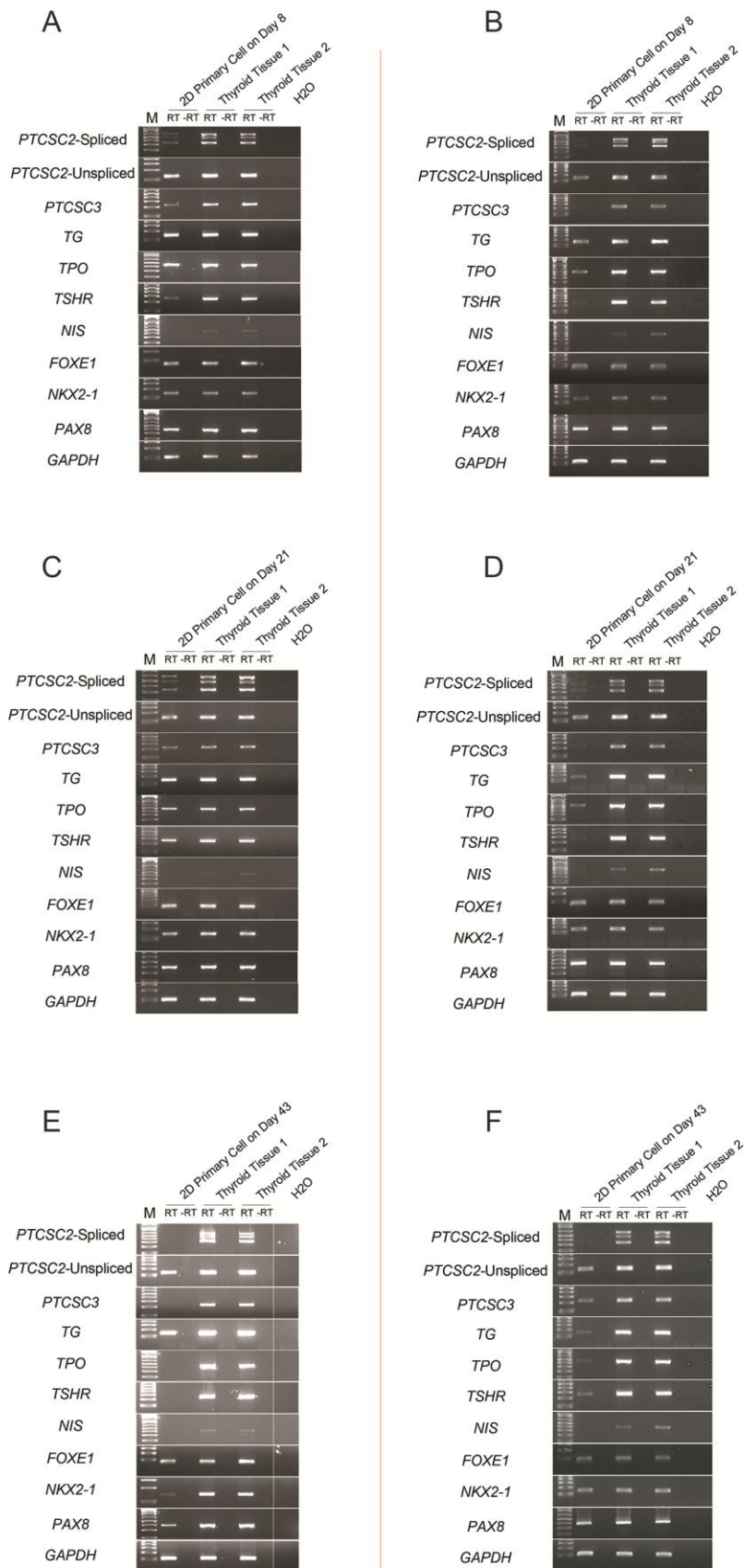


Figure 2. Thyroid marker gene expression profile of 2D primary cells in 6H medium (A, C and E) and h7H medium (B, D and F). A and B, Day 8; C and D, Day 21; E and F, Day 43. *GAPDH* was used as a housekeeping control. RT indicates the presence of reverse transcriptase; -RT indicates reactions without reverse transcriptase.

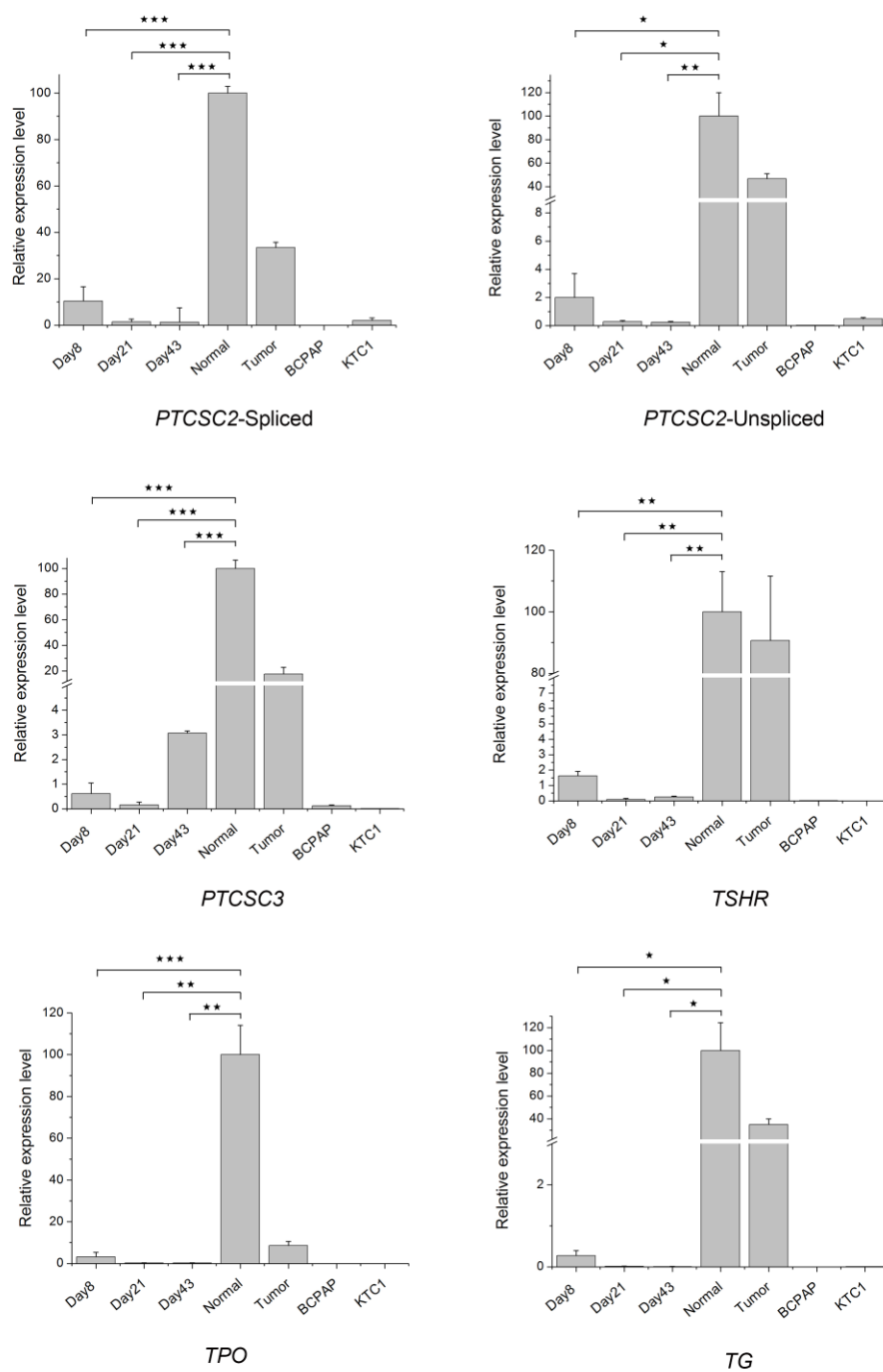


Figure 3. Relative expression of selected thyroid marker genes. Relative expression levels were obtained by the  $2^{-\Delta CT}$  method. *GAPDH* was used as a normalization control. All assays were

performed in three biological replicates and all samples were normalized with the expression level in normal thyroid tissue (set as 100); \* $P < 0.05$ ; \*\*  $P < 0.01$ ; \*\*\*  $P < 0.001$ . (Student  $t$  test). Normal: non-tumorous tissue.

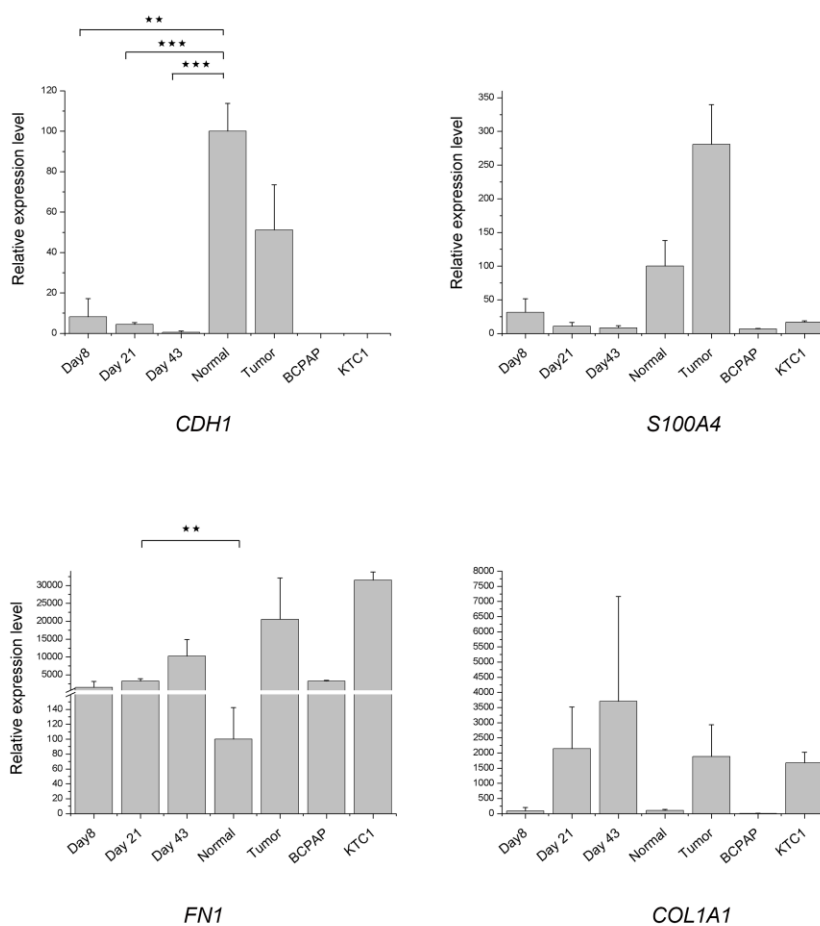


Figure 4. Relative expression of EMT and fibroblast marker genes. Relative expression levels were obtained by the  $2^{-\Delta CT}$  method. *GAPDH* was used as a normalization control. All assays were performed in three biological replicates and all samples were normalized with the expression level in normal thyroid tissue (set as 100); \* $P < 0.05$ ; \*\*  $P < 0.01$ ; \*\*\*  $P < 0.001$ . (Student  $t$  test). Normal: non-tumorous tissue.

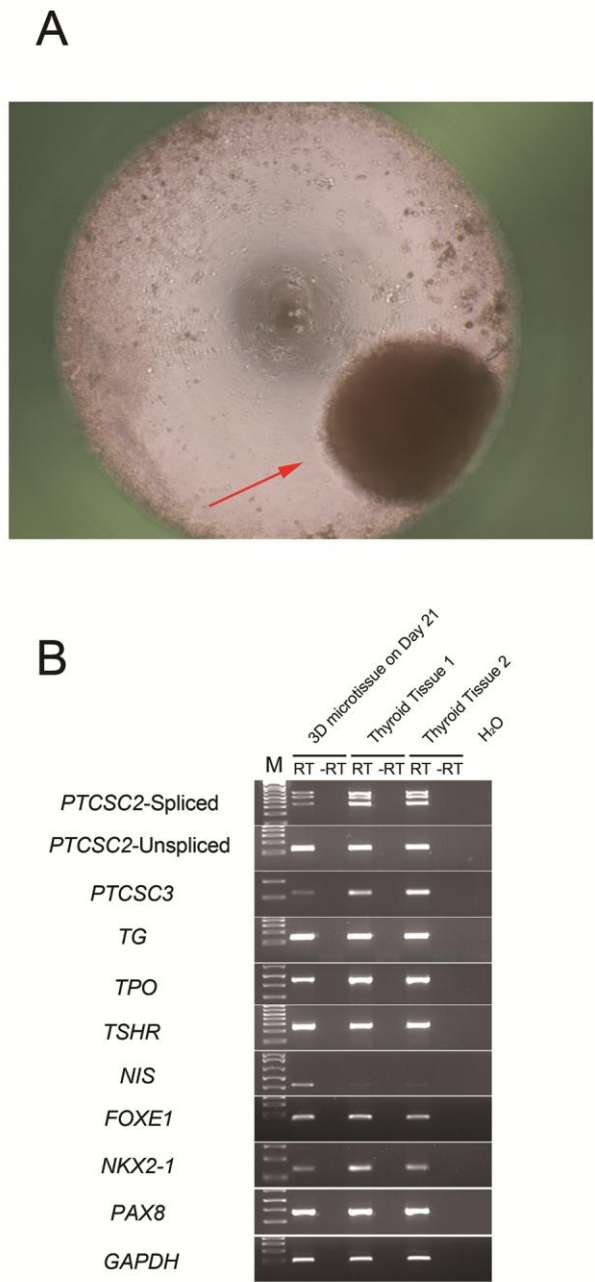
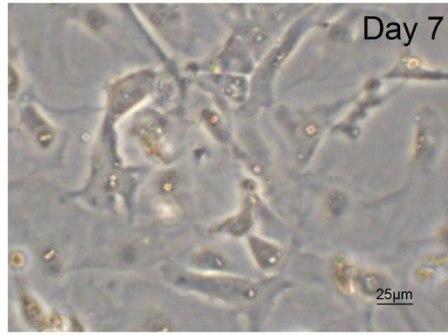
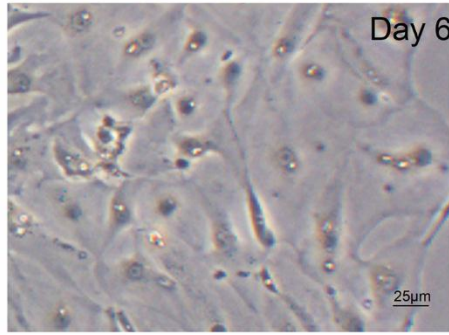
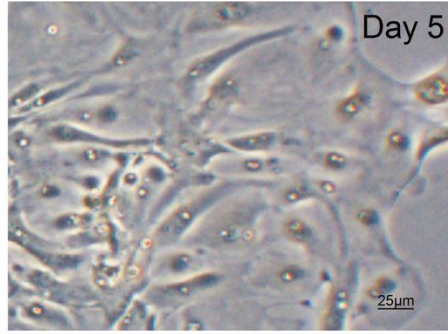
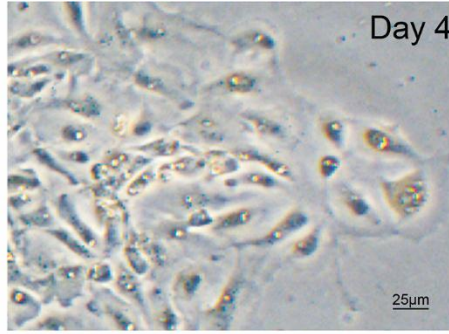
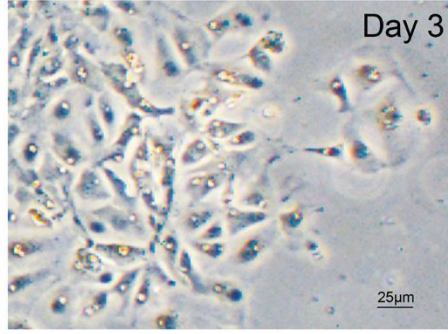
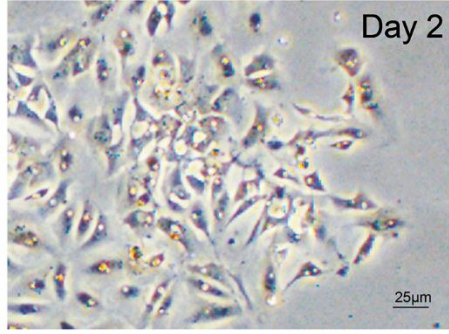


Figure 5. 3D microtissue derived from primary thyroid cells. A, the 3D spheroid formed by hanging-drop system and pictured on Day 21. Red arrow shows the 3D microtissue under a microscope with 50X magnification. B, Thyroid marker gene expression profile of the 3D

microtissue. *GAPDH* was used as a housekeeping control. RT indicates a reverse transcription reaction with reverse transcriptase; -RT indicates a reaction without reverse transcriptase.





Supplementary Figure 1. Morphology of 2-dimensional primary thyroid cell cultures after seeding with high magnification (200X). A red arrow indicates an emerging thyroid fibroblast-like cell.

Supplementary Table 1. Forward (sense) and reverse (antisense) primers for RT-PCR test.

Gene	Direction	Sequence 5'-3'
<i>PTCSC2-Spliced</i>	Forward	ccaggatgatggcctgtg
	Reverse	aatcccctgcagttcctgatt
<i>PTCSC2-unspliced</i>	Forward	tggtgatggtatggtcatgg
	Reverse	cccaggctcaggttatgtct
<i>PTCSC3</i>	Forward	tcaaactccagggtgaac
	Reverse	attacggctgggtctacct
<i>TSHR</i>	Forward	cccagtagcagactctgaa
	Reverse	ccagtgtgaaaatgccaag
<i>FOXE1</i>	Forward	tgacctgggctggtttcc
	Reverse	tgctgacgcacagttcaactc
<i>PAX8</i>	Forward	cctttgtgaatggcagacct
	Reverse	tagggagggtgaatggtgc
<i>NKX2-1</i>	Forward	ccatgaggaacagcgctc
	Reverse	ctcacgtccccagcga
<i>TPO</i>	Forward	cgggtcatctgtgacaacac
	Reverse	cggagtctacgcaggttctc
<i>NIS</i>	Forward	ctccctgctaacgactccag

<i>TG</i>	Reverse	gaccacatcatgtccaaca
	Forward	ccaagaaaaccatctcaa
<i>GAPDH</i>	Reverse	attgtccaccaaccacctc
	Forward	caccaactgcttagcacc
	Reverse	tcttctgggtggcagtgatg

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Supplementary Table 2. qRT-PCR primers and probes in the SyberGreen and Taqman methods.

Gene	Detection method	Direction	Sequence 5'-3'
<i>PTCSC3</i>	SyberGreen	Forward	tcaaactccagggttgaac
		Reverse	attacggctgggtctacct
<i>TSHR</i>	SyberGreen	Forward	cccagtacgcagactctgaa
		Reverse	ccagtgtgaaaatccaag
<i>TPO</i>	SyberGreen	Forward	acagcatcactggcatgaac
		Reverse	gtgcacaaagtccccattct
<i>TG</i>	SyberGreen	Forward	ctggtgtgtcatggacagcggagaa
		Reverse	cccagattgtctcacacaggat
<i>CDH1</i>	SyberGreen	Forward	gtcactgacaccaacgataatcct
		Reverse	ttcagtgtggtgattacgacgtta
<i>S100A4</i>	SyberGreen	Forward	gagctgccagcttcttg
		Reverse	tgcaggacaggaagacacag
<i>FNI</i>	SyberGreen	Forward	aaacttgcactctggaggcaaaccc
		Reverse	agctctgatcagcatggaccact
<i>COL1A1</i>	SyberGreen	Forward	tgttcagcttgtggacctccg
		Reverse	cgcagggtgattggggatgtct
<i>PTCSC2-spliced</i>	Taqman	Forward	atggtccttgagctgacat
		Reverse	ttcgaactggatataggaca
		Probe	tcctctctttaggcattcccact
<i>PTCSC2_unspliced</i>	Taqman	Forward	tggatgatggatggatcatgg
		Reverse	cccaggctcaggttatgtct
		Probe	aggtaatgagtggctggaatggaaca

Supplementary Table 3.  $\Delta C_t$  values for all the 10 markers tested in qRT-PCR.

Gene name	Day 8	Day 21	Day 43	Normal tissue	Tumor tissue	BCPAP	KTC1
<i>PTCSC2</i> -Spliced	13.51 ± 0.92	16.57 ± 1.14	16.43 ± 0.32	10.10 ± 0.04	11.69 ± 0.10	Undetectable	15.95 ± 1.01
<i>PTCSC2</i> -Unspliced	8.36 ± 1.44	10.79 ± 0.36	11.00 ± 0.36	2.29 ± 0.27	3.38 ± 0.14	15.39 ± 0.67	10.05 ± 0.29
<i>PTCSC3</i>	13.62 ± 1.36	15.60 ± 1.47	10.96 ± 0.04	5.93 ± 0.10	8.48 ± 0.43	15.59 ± 0.33	20.09 ± 0.99
<i>TSHR</i>	8.45 ± 0.27	13.31 ± 1.48	11.13 ± 0.25	2.49 ± 0.18	2.64 ± 0.32	17.18 ± 0.31	Undetectable
<i>TPO</i>	6.76 ± 1.14	11.13 ± 2.36	9.94 ± 0.17	1.56 ± 0.20	5.12 ± 0.32	Undetectable	19.38 ± 2.85
<i>TG</i>	6.97 ± 0.76	11.56 ± 0.94	12.13 ± 0.57	-1.65 ± 0.33	-0.15 ± 0.21	19.70 ± 0.27	13.19 ± 0.29
<i>CDH1</i>	4.36 ± 1.63	4.69 ± 0.34	8.79 ± 2.07	-0.18 ± 0.55	0.98 ± 0.61	18.46 ± 0.44	19.80 ± 0.94
<i>S100A4</i>	5.04 ± 0.84	6.54 ± 0.79	6.83 ± 0.51	2.72 ± 0.95	1.34 ± 0.72	7.11 ± 0.21	5.79 ± 0.20
<i>FN1</i>	2.75 ± 1.49	1.15 ± 0.33	-0.41 ± 0.77	4.85 ± 2.63	-1.68 ± 1.45	1.12 ± 0.09	-2.15 ± 0.11
<i>COL1A1</i>	5.05 ± 2.15	-0.33 ± 1.34	-1.05 ± 1.28	3.34 ± 0.90	-1.20 ± 1.54	7.30 ± 0.25	-0.28 ± 0.32

Data were shown as Mean ± Standard variation. And all the measurements were made from 3 - 5 repeated experiments.

