

## REVIEW

# Hohenheim Consensus Workshop: Copper

K Schümann<sup>1</sup>, HG Classen<sup>2</sup>, HH Dieter<sup>3</sup>, J König<sup>4</sup>, G Multhaupt<sup>5</sup>, M Rückgauer<sup>6</sup>, KH Summer<sup>7</sup>, J Bernhardt<sup>2</sup> and HK Biesalski<sup>2\*</sup>

<sup>1</sup>Walther-Straub-Institut für Pharmakologie und Toxikologie der LMU, München, Germany; <sup>2</sup>Institute of Biochemistry and Nutrition, Hohenheim University, Stuttgart, Germany; <sup>3</sup>Umweltbundesamt, WABOLU, Berlin, Germany; <sup>4</sup>Institut für Ernährungswissenschaft der Universität Wien, Vienna, Austria; <sup>5</sup>ZMBH-Center for Molecular Biology, University of Heidelberg, Heidelberg, Germany; <sup>6</sup>Katharinenhospital, Stuttgart, Germany; and <sup>7</sup>GSF, München, Germany

Copper (Cu) is an essential trace element with many physiological functions. Homeostatic mechanisms exist to allow Cu to act as a cofactor in enzymatic processes and to prevent accumulation of Cu to toxic levels. The aim of this commentary is to better understand the role of dietary Cu supply in deficiency and under physiological and pathological conditions. The essentiality of Cu can be attributed to its role as a cofactor in a number of enzymes that are involved in the defence against oxidative stress. Cu, however, has a second face, that of a toxic compound as it is observed with accumulating evidence in hepatic, neurodegenerative and cardiovascular diseases. The destructive potential of Cu can be attributed to inherent physico-chemical properties. The main property is its ability to take part in Fenton-like reactions in which the highly reactive and extremely deleterious hydroxyl radical is formed. Diseases caused by dietary Cu overload could be based on a genetic predisposition. Thus, an assessment of risk-groups, such as infants with impaired mechanisms of Cu homeostasis regarding detoxification, is of special interest, as their Cu intake with resuspended formula milk may be very high. This implies the need for reliable diagnostic markers to determine the Cu status. These topics were introduced at the workshop by the participants followed by extensive group discussion. The consensus statements were agreed on by all members. One of the conclusions is that a re-assessment of published data is necessary and future research is required.

European Journal of Clinical Nutrition (2002) 56, 469–483. doi:10.1038/sj.ejcn.1601315

**Keywords:** copper; physiology; oxidative stress; review

## 1. What is the physiological function of Cu?

‘There are a number of functional Cu-containing enzymes and storage proteins. Some of these have highly specific and some have non-specific functions.’

For more than 80y Cu has been recognised as an essential nutrient in man. The rapid development in the analytical field and in cell biology provided an increasing understanding of the role of Cu in immune defences, connective tissue and skeleton, blood formation, the blood vessels and ner-

vous system. Behind iron and zinc, Cu is the third most common trace element in the human body. The adult organism contains about 4g of iron and 80–100mg Cu (Halliwell & Gutteridge, 1985; Harris, 1991). The average Cu concentration in the cerebrospinal fluid is 70µM and 250µM in the synaptic cleft (Kardos *et al*, 1989; Linder & Hazegh-Azam, 1996).

Cu is an integral part of a number of enzymes. For their function they require either Cu alone or Cu and other metal ions as co-factors. Such enzymes are Cu/Zn-superoxide dismutase (Cu/Zn-SOD), ceruloplasmin, cytochrome oxidase, tyrosinase and lysine oxidase. In addition, Cu-dependent transcription factors also play an important part in gene expression. The enzymes in which Cu is an intrinsic component of the catalytic centre are members of the oxidoreductase family. These enzymes are essential for cellular respiration, defence against free radicals, melanin synthesis, formation of connective tissue and iron metabolism (Table 1). The role of Cu in Cu/Zn-SOD has been intensively studied. SOD catalyses the dismutation of superoxide

\*Correspondence: HK Biesalski, Institute of Biochemistry and Nutrition, Hohenheim University, Fruwirthstrasse 12, D 70593 Stuttgart, Germany. E-mail: biesal@uni-hohenheim.de

Guarantor: S Truswell.

Contributors: HK Biesalski.

Received 8 February 2001; revised 15 July 2001;

accepted 16 July 2001

**Table 1** Some Cu-dependent enzymes and their functions

|                            |                                |
|----------------------------|--------------------------------|
| Cu/Zn-superoxide dismutase | Antioxidative defence          |
| Ceruloplasmin/ferroxidase  | Fe binding to transferrin      |
| Cytochrome oxidase         | Cellular respiration           |
| Tyrosinase                 | Melanin synthesis              |
| Lysin oxidase              | Formation of connective tissue |

anions in the cytoplasm, which is a by-product of cellular respiration. During dismutation, Cu in the active centre of SOD is reduced by the substrate  $O_2^-$ ; the resulting metabolites are  $O_2$  and  $H_2O_2$ . In this reaction Cu cannot be replaced by any other metal. Genetic mutations of SOD1 that alter the enzyme go along with increased apoptosis of neuronal cells, eg in amyotrophic lateral sclerosis (ALS).

Iron and Cu metabolism are interrelated via the ferroxidase activity of ceruloplasmin. In subjects with inherited ceruloplasmin-deficiency the lack of ferroxidase activity decreases the conversion of  $Fe^{2+}$  to  $Fe^{3+}$  and, consequently, iron binding to transferrin (Gittlin, 1998). The consequence is a disturbed iron distribution. Either iron deficiency (Wang *et al*, 1995) or iron excess were observed, leading to neurodegenerative sequelae in certain areas of the brain (Gittlin, 1998). In addition, ceruloplasmin has antioxidative functions as it reduces  $Fe^{2+}$ -dependent oxidative stress (Gutteridge, 1980). In contrast, Cu ions bound to albumin and free amino acids are able to participate in Fenton reactions themselves and convert reactive oxygen species like  $H_2O_2$  and  $O_2^-$  to  $\bullet OH$  radicals (Brigelius *et al*, 1974). Cu binding to viruses, DNA, carbohydrates, enzymes and other proteins, such as amyloid, can cause local damage of these compounds as the result of local  $\bullet OH$  formation (Multhaup *et al*, 1998; Quinlan & Gutteridge, 1988). These observations suggest that reactive oxygen species *in vivo* may only be toxic in the presence of catalysing metal ions. In addition to its role in the catalytic centre of Cu-dependent enzymes, there is increasing evidence that Cu is also engaged in gene expression of various proteins. Cu bound to transcription factors is involved in the modulation of eukaryotic gene expression by activation or repression of genetic transcription.

## 2. How much Cu is absorbed from the diet; where and how is Cu absorbed in the gastrointestinal tract?

'Dietary copper is absorbed in the stomach (small amounts), duodenum and proximal jejunum. About 20–70% of dietary Cu is absorbed. In adults maximum absorption is 50%; in infants it is 77%. Absorption can be strongly affected by ligands and other metals.'

The kind of ligands as well as the size, solubility and stability of the Cu complexes formed in the intestinal lumen, have a strong influence on Cu bioavailability. In contrast to other metals, Cu is absorbed in the stomach. This is of minor

importance, though, because food is incompletely digested at this location. Cu absorption involves active saturable transport mechanisms as well as diffusion (Crampton *et al*, 1965). The absorptive capacity for Cu is comparable in all sections of the small bowel. In contrast to adult animals, the immature Cu absorption process in rat pups is not saturable (Varada *et al*, 1993) and has a greater transportation capacity (Lönnerdal *et al*, 1985). It is predominantly mediated by diffusion and by co-transport with water (solvent drag). Correspondingly, a lack of sodium in the lumen or inhibition of intestinal sodium transport impairs Cu absorption (Wapnir, 1991). The intestinal concentration of metallothionein is lower in rat pups than in adult animals (Varada *et al*, 1993). Therefore, metallothionein is unlikely to be responsible for increased retention of Cu in the pup's intestine. In adult animals, however, intestinal metallothionein appears to reduce absorption when Cu exposure is continuously higher (Hall *et al*, 1979). According to investigations with stable isotopes, Cu absorption decreases with increasing supply levels. Increasing the supply from 0.8 to 7.5 mg Cu/day will no more than double the absorbed Cu quantity. At high supply levels, Cu whole body retention decreases to 12% while its maximum is estimated to be 63–67%. Absorption, thus, seems to be homeostatically controlled with a set point rate of 0.8–1.0 mg Cu/24 hours (Turnlund, 1991). Little is known about intestinal transport mechanisms for Cu and their regulation. Na/K-ATPase is involved in the transfer of Cu from the enterocytes into the blood.

The bioavailability of Cu depends largely upon interactions with other dietary components. Thus, balance studies in full-term breast-fed babies indicate a Cu bioavailability of 75–77% (Dörner *et al*, 1989). In contrast, Cu absorption from fortified infant formulas based on cow's milk amounted to only 23% (Lönnerdal, 1998), ie it was three times lower. In human breast milk 75% of Cu is bound to soluble milk proteins (Fransson & Lönnerdal, 1983), whereas in cow's milk 75–80% of Cu are bound to insoluble casein (Lönnerdal *et al*, 1985), which is not completely hydrolysed by the infant's immature digestive systems. This is why the difference in Cu absorption between cow's milk and breast milk is particularly marked in preterm infants.

Adequate gastric HCl production facilitates peptic digestion in the stomach and, hence, the availability of Cu in the small intestine. Alkaline pH values in the intestine reduce Cu-bioavailability by formation of Cu-hydroxides and Cu-complexes with a low dissociation constant. Phytates do not impair the availability of Cu to the same extent as that of Fe or Zn (Turnlund *et al*, 1985). Dephytinization impairs Cu bioavailability indirectly: Zn and Fe are less well bound in the lumen and may reduce Cu absorption (Morris *et al*, 1988). The impairment of Cu-absorption by Zn is used therapeutically in Wilson's disease.

Glucose polymers increase the co-transport of Cu with water (solvent drag) and, thus, Cu absorption. Correspondingly, substitution of glucose by fructose reduces Cu bioavailability. A reduced energy metabolism may add to this

effect (Wapnir & Devas, 1995). Large concentrations of amino acids and peptides may bind Cu in the lumen and reduce its absorption. In moderate concentrations, however, such ligands may reduce Cu-hydroxide formation and increase Cu absorption. Organic acids such as citrate, lactate or malate increase the solubility and absorption of Cu (Sable-Amplis *et al*, 1987; Table 2).

### 3. How is Cu homeostasis maintained?

'In the blood Cu is predominantly present in the plasma, where it is bound to albumin and ceruloplasmin. Little is known about Cu plasma binding during transport from the intestine to the liver and its regulation. Hepatic Cu uptake seems to vary extensively in response to changes in Cu supply. The form in which Cu is bound when it reaches the peripheral tissues is discussed controversially. Cu uptake into extrahepatic tissues and fetal tissues is incompletely understood. Chaperones are specific transport proteins which are involved in intracellular Cu transport. The liver regulates Cu elimination via biliary excretion. Cu reabsorption after biliary excretion is small.'

Cu homeostasis requires a delicate balance between Cu absorption, distribution, storage, and elimination. Specific transport proteins in small intestinal enterocytes control intestinal Cu absorption, ie its uptake and transfer from the diet into the blood (cf. Section 2). Absorbed Cu reaches the liver via the portal circulation predominantly bound to transcuprein (Weiss & Linder, 1985), histidine and to the high affinity binding sites of albumin (Masuoka *et al*, 1993). The uptake of Cu by hepatocytes is incompletely understood. A specific Cu transport protein, called hCtr1, appears to play a major part in this process (Pena *et al*, 1999). In the hepatocyte, monovalent Cu is bound to low molecular weight proteins in the cytosol, so-called chaperones. Each chaperone has specific tasks regarding the transport of Cu into cell organelles and regarding the incorporation of Cu into enzymes (Lin & Culotta, 1995). Some of the Cu is stored by binding to cytosolic metallothionein. The metallothio-

nein concentration, and hence its binding capacity for Cu, increases with an increasing Cu supply (Bremner, 1987).

Cu export from the hepatocytes is mediated by incorporation of the metal into ceruloplasmin via a Cu-binding ATPase located in the Golgi apparatus (Nagano *et al*, 1998) to mediate its excretion via the bile. At adequate Cu supply levels, a constant amount of Cu is incorporated into hepatic ceruloplasmin. In copper deficiency Cu incorporation into ceruloplasmin is reduced, although its synthesis remains largely unaffected (Nakamura *et al*, 1995). Ceruloplasmin-bound Cu is transported with the plasma to the different tissues. Inhibition of ceruloplasmin synthesis by cyclohexamide has a significant effect on Cu distribution between the organs. However, there is no doubt that organs other than liver and kidney can take up Cu without ceruloplasmin mediation (Lee *et al*, 1993), although to a substantially reduced extent. Though ceruloplasmin excretion controls hepatic Cu export to some extent, it does not affect the amount of Cu in the body. Thus, the role of this protein for Cu homeostasis is presently questioned in favour of its function as ferroxidase (Gittlin, 1998).

Excess Cu is extracted via the bile. This process comprises Cu exocytosis via lysosomes and via a Cu-binding ATPase in the canalicular membrane. Besides, an ATP-independent and GSH-dependent Cu transport process is under discussion for biliary Cu excretion (Dijkstra *et al*, 1996). Only marginal amounts of Cu are reabsorbed after biliary excretion (Linder, 1991).

### 4. Biomarkers, diagnostic parameters of Cu status (plasma concentration, Cu in red blood cells, enzymes, ceruloplasmin), diagnostic costs, when is intensive investigation indicated?

'The best indicator for Cu status is the hepatic Cu concentration (biopsy). SOD concentration in the erythrocyte can be used to assess Cu deficiency. The Cu concentration in the plasma is an inert indicator due to homeostatic regulation. Low plasma concentrations and SOD activity in erythrocytes mark advanced Cu deficiency states. They do not reveal marginal deficiencies. Because of their limited discriminative power, the methods used to characterise Cu status cannot be considered as adequate.'

The classic approach to determine the quality of a biomarker is to monitor its response to dietary depletion and repletion. Studies in dogs, pigs, cattle, sheep and rats showed decreases in serum/plasma Cu concentrations, in the hepatic Cu content, in plasma ceruloplasmin concentrations, in the SOD activity in liver or erythrocytes, or in the activity of cytochrome-C-oxidase in the liver and duodenal mucosa during Cu-deficient feeding (Baker & Ammerman, 1995). A decreasing dietary Cu intake correlated best with the hepatic Cu content, followed by hepatic SOD activity. Increasing dietary

**Table 2** Promotion and inhibition of intestinal Cu absorption

#### Promotion

Adequate gastric digestion  
Moderate luminal aminoacid and peptide concentrations (reduced Cu-hydroxide formation)  
Organic acids (increased Cu solubility)

#### Inhibition

Lack of Na and glucose in the intestinal lumen (reduced solvent drag)  
High intestinal metallothionein levels (presumably only in adults)  
Alkaline pH in intestinal lumen  
Phytates (small impact)  
Large luminal concentrations of amino acids and peptides (high Cu-binding in the lumen)

supply levels correlated best with the hepatic Cu content. However, the relationship between dietary Cu content and the Cu hepatic concentration is strongly species-specific (Baker & Ammerman, 1995).

Depletion and repletion studies in healthy volunteers help to assess the value of the different diagnostic parameters: (1) the concentration of Cu and ceruloplasmin in the plasma remained unaltered, when they were challenged by a daily supply of 0.57 mg Cu for 105 days. The SOD activity in erythrocytes and the cytochrome-C oxidase activity in the platelets, however, was decreased. Administration of 2 mg Cu for 35 days increased the activity of cytochrome-C-oxidase and glutathione peroxidase (Milne & Nielsen, 1996). (2) The plasma concentrations of Cu and ceruloplasmin decreased following a dietary challenge with 0.38 mg Cu for 42 days. A 24 day repletion with 2.5 mg Cu/day led to no significant increase of these parameters (Kelley *et al*, 1995). (3) After dietary Cu depletion (0.79 mg Cu/day for 24 h) and subsequent Cu repletion (7.53 mg Cu/day for 42 and 24 days), no changes in plasma Cu and ceruloplasmin concentrations or in erythrocyte's SOD activity were observed (Turnlund *et al*, 1990). (4) After reduced Cu intake for 77 days (1.03 mg/day 2850 kcal with 20% fructose), the plasma concentrations of Cu and ceruloplasmin were unchanged, while erythrocyte's SOD activity decreased and increased again after repletion (3 mg Cu/day for 21 days, Reiser *et al*, 1985). The serum-Cu concentration is homeostatically regulated. This parameter shows circadian changes with a peak in the morning, which is higher in women than in men, and increases with age. Oestrogen intake, pregnancy, infections, inflammation and stress increase plasma Cu concentration while corticosteroids and corticotrophin administration decrease it (Johnson *et al*, 1992; Milne & Johnson, 1983). Ceruloplasmin in the plasma is regarded as an acute phase protein (Milne, 1994). It is influenced in analogy to plasma Cu concentrations. The activity of Cu/Zn-SOD in erythrocytes, in contrast, is not affected by such influences. It is reduced in Cu depletion and increases again on repletion and, therefore, appears as a useful marker of Cu status (Johnson *et al*, 1992). However, because of the limited sensitivity of all of these parameters, no optimal method is available to assess marginal Cu deficiency.

It is common practice to determine Cu concentration in serum by electrothermal atomic absorption spectrometry. Standard indications are the diagnosis of Wilson's disease, Menkes' syndrome, nutritional Cu deficiency or Cu intoxication. The Cu content in tissues or specific blood cells is not routinely investigated. Such analysis is time-consuming, difficult to standardize and tissue sampling is an invasive process. SOD activity is also not measured routinely. There are guideline values for Cu concentrations in plasma and erythrocyte SOD activity which, however, are not generally accepted reference values.

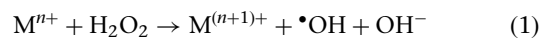
A promising new parameter for the determination of excess Cu intake is an elevated serum diamine oxidase activity (DAO; Kehoe *et al*, 2000). Evans *et al* (1998) sug-

gested determining free plasma Cu concentrations directly, instead of calculating it from other parameters. A suitable method was developed by Gutteridge for other purposes (Gutteridge, 1984). The Cu-specific parameter measured colorimetrically in this case is the extent of oxygen radical formation during Cu-dependent DNA oxidation.

## 5. What is the role of Cu in oxidative stress?

'Cu-overload favours the development of oxidative stress. Cu deficiency has also been proposed as a possible cause of oxidative stress, though this has not been established with certainty. There are, however, individual results to support this suggestion.'

In situations of oxidative stress newly formed oxygen radicals cannot be adequately detoxified by antioxidants and excess free radicals cause cellular damage. Hydroxyl radicals are formed *in vivo* mainly by reaction with  $H_2O_2$ , in accordance with eqn (1):

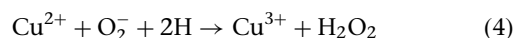
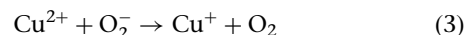


Although the metal ion  $M^{n+}$  can stand for  $Ti^{3+}$ ,  $Fe^{2+}$ ,  $Cu^+$ ,  $Co^{2+}$  as well as for chromium, manganese, vanadium and nickel complexes, only  $Fe^{2+}$  and  $Cu^+$  seem to be capable to catalyse Fenton chemistry *in vivo* (Gutteridge, 1994; Wiedau-Pazos *et al*, 1996). The rate for this second order reaction is as low as  $10^2 M^{-1} s^{-1}$ . If one considers that the reaction partners  $H_2O_2$ ,  $Fe^{2+}$  and  $Cu^+$  are present at a concentration of about  $1 \mu M$  under physiological conditions, the assumed reaction rates for  $Fe^{2+}$  and  $Cu^+$  can be compared. If a cell volume of  $10^{-12} L$  is assumed and a second order reaction constant of  $k = 76 M^{-1} s^{-1}$ , about 46 molecules of  $\bullet OH$  are formed per cell per second (Halliwell & Gutteridge, 1990) when  $Fe^{2+}$  is used as the catalyst (see eqn (2)).

$$N = 7.6 \times 10^{-11} \times 10^{-12} \times 6023 \times 10^{23} \quad (2)$$

In contrast, the turnover rate with  $Cu^+$  as a catalyst is  $4.7 \times 10^3 M^{-1} s^{-1}$ . This is orders of magnitude greater than with  $Fe^{2+}$  as a catalyst and results in the production of 2840  $\bullet OH$  per cell per second. It must be considered here that  $Cu^{3+}$  may be produced in addition to or instead of radicals.

$Cu^+$  and  $Cu^{3+}$  can be produced by the reaction with the superoxide anions (eqns (3) and (4)).



However, it is not certain whether metal ions undergo reactions with superoxide anions *in vivo* and, hence can produce oxidative stress.

Cu deficiency is also discussed to produce oxidative stress, which was studied in different model systems. Assuming that the activity of antioxidative enzymes and the extent of oxidative modification of erythrocyte proteins are reliable indicators for oxidative stress, rats were fed a Cu-deficient

diet. While hepatic Cu concentrations decreased by about 80% during Cu-deficient feeding, the Cu content in nuclei of hepatocytes did not change (Lai *et al*, 1996). The transcription rates of the enzymes Cu,Zn-SOD, glutathione peroxidase and glycerine aldehyde-3-phosphate-dehydrogenase remained unaltered, those of Mn-SOD and of beta-actin increased and the transcription rate of catalase decreased in Cu deficiency (Lai *et al*, 1996). The activity of cytoplasmic Cu,Zn-SOD in erythrocytes decreased as well, while the amount of carbonylated subunits of spectrin, an erythrocyte membrane protein (Sukalski *et al*, 1997) increased significantly. These examples show that experimentally induced Cu deficiency in rats influences the activity of hepatic antioxidant enzymes differentially. The activity of Cu,Zn-SOD is reduced, although the rate of transcription remains unchanged. Thus, presumably, the ratio between apoenzyme to enzyme will increase in situations of inadequate Cu supply. This finding underlines the importance of Cu,Zn-SOD as an indicator for Cu deficiency.

## 6. Which functional deficits are observed in Cu-deficiency?

'Clinical manifestations of Cu deficiency are very rare and are predominantly the results of a dietary Cu deficiency. Among the clinical manifestations of Cu deficiency are anaemia, neutropenia and increased bone fragility. An impact of Cu deficiency on the immune system has been suggested.'

Cu uptake with the diet is essential for man and animals and generally exceeds the minimum requirement of this trace element. Experimental Cu deficiency in animals may cause cardiac and vascular disease, which can partly be explained by a reduced activity of Cu-dependent enzymes. Neurological deficits were observed in Cu-deficient sheep (Williams, 1983), thromboses (Klevay, 1985) and reduced sperm motility and fertility in rats (Battersby & Chandler, 1977). In children Cu deficiency produced hypochromic anaemia and disturbed the maturation of myeloid cells in the bone marrow (Cordano, 1998). The erythrocyte's half-life was reduced, due to altered fluidity and increased sensitivity of the membrane to lipid peroxidation (Rock *et al*, 1995). Accordingly, one of the early clinical signs in Cu deficiency is neutropenia. Neutropenia has been related to an increased frequency of respiratory infections in Cu deficiency. Besides, the number of these cells in the bone marrow is reduced in Cu deficiency, which may in part be attributed to a reduced survival of neutrophilic granulocytes. The skeleton tends to be osteoporotic in Cu deficiency, with an increased risk of fractures. Erosion and reactive spur formation are found in the metaphysis (Cordano, 1998).

Therefore, Cu deficiency is an unlikely cause of teratogenic effects in men, although neural tube defects (Morton *et al*, 1976) and an increased risk of anencephaly (Buamak

*et al*, 1984) have been associated with Cu deficiency. The importance of Cu for foetal development becomes obvious in animal deficiency symptoms. A spastic paralysis of the hind legs in lambs is associated with hypomyelination in the brain in severe Cu deficiency (Hurley & Keen, 1979). This syndrome is called 'sway back disease'. It is thought to be caused by deficiency in Cu-dependent cytochrome-C-oxidase (Mills & Williams, 1962). Deficient elastin incorporation in lung tissue (Abdel Mageed *et al*, 1994) and blood vessels and a defective collagen matrix in dogs and pigs (Hurley & Keen, 1979) appear to be the consequence of a reduced activity of Cu-dependent lysyl-oxidase.

## 7a. Is the recommended dietary Cu intake reached in the average population?

'The daily Cu intake in adults should reach 20 µg/kg body weight; for infants 50 µg Cu/kg body weight should be reached. Regarding the compliance with nutritional advice, Cu has to be counted among the critical elements.'

The DGE recommends an intake of 1.0–1.5 mg Cu/day for adolescents and adults. The corresponding recommendation of the Scientific Committee on Food of the European Commission is 1.1 mg Cu/day for adults (Scientific Committee on Food, 1994). The DGE assessment was based on dietary balance studies, from which intakes of 75 µg Cu/kg in infancy, 50 µg/kg in older infants, 40 µg Cu/kg in children aged 7–10 y of age, and finally 20 µg Cu/kg in adults were derived (Deutsche Gesellschaft für Ernährung, 1991; National Research Council, 1989). In addition to the recommended dietary Cu intake, which is required to maintain normal plasma levels, the WHO derived an upper safe level for Cu in food (12 mg/day for male and 10 mg/day for female adults on the basis of 180 µg/kg body weight and day) which should not be exceeded (WHO, 1996).

Daily Cu intake was studied in Austria using a 7 day weighing protocol (König & Elmadfa, 1993; Elmadfa *et al*, 1999). Data on Cu concentrations in plasma were also available for part of this group. The data show that a Cu intake according to current DGE recommendations was not sufficient in all cases to keep plasma Cu concentrations in the normal range for children, adolescents and elderly (0.8–1.2 mg/l; König & Elmadfa, 1993; Elmadfa *et al*, 1999). According to the 1994–1996 Continuing Survey of Food Intakes by Individuals, Cu intake in individuals over 60 y of age in the US was 1.3 ± 0.7 mg Cu/day for men and 1.0 ± 0.5 mg Cu/day for women which is less than recommended. Main dietary Cu sources were legume, potato and potato products, nuts, seeds and beef (Ma & Betts, 2000). Also, chocolate foods can be a good source of Cu (Joo & Betts, 1996). Correspondingly, the mean Cu intake in various European countries was below the recommended minimum intake of 1.2 mg Cu/day in adult males in about 10% of

subjects under investigation and below the minimum normative population intake in about 25% (WHO, 1996; 1998). In spite of these findings, according to the WHO, Cu deficiency is relatively rare in humans under normal physiological conditions. Clinical signs of deficiency are only observed under exceptional circumstances (Underwood, 1977), eg in children with 'cow's milk anaemia', during rehabilitation after malnutrition or when formula milks are inadequately fortified with Cu, in adults with protein-energy malnutrition, in enteropathies and during total parenteral nutrition as a consequence of inadequate Cu supplementation of infusion solutions (WHO, 1998; see Section 7b).

## 7b. Are there special groups at risk for inadequate Cu supply?

'Deficiency due to increased requirements: disease states, pregnancy (last trimester), premature infants, periods of rapid growth, parenteral nutrition. Deficiency due to inadequate intake: eg maldigestion/malabsorption, reducing diets.'

### Deficiency due to increased requirements

Cu deficiency states in children with massive diarrhoea in developing countries (Castillo-Duran *et al*, 1990; Sachdev *et al*, 1989), especially in association with growth spurts after protein-energy malnutrition seem to be the most common cause of Cu deficiency (Castillo-Duran & Uauy, 1988). The loss of Cu in children with diarrhoea was twice that in controls (Castillo-Duran *et al*, 1988). Consequently, plasma levels of Cu were reduced by half (Rodriguez *et al*, 1985). Also concomitant administration of zinc, fructose or chelators increase the demand and appear to cause Cu deficiency more frequently than inadequate dietary Cu supply (Keen *et al*, 1998). In nephrotic syndrome (Pedraza-Chaveri *et al*, 1994) and in patients with severe burns (Shakespeare, 1982), excessive loss of Cu may occur and the Cu requirement is increased. Other conditions of this type are short bowel syndrome, cystic fibrosis, coeliac disease, tropical and non-tropical sprue, intestinal fistulas and following gastrectomy and ileo-jejunal bypass operations (Beshgetoor & Hambidge, 1998).

During the second half of pregnancy the foetus acquires about 50 µg Cu/kg/day (Widdowson *et al*, 1974). Correspondingly, pregnant women reduce biliary Cu excretion by hormonal adjustment (McArdle, 1995) and serum Cu concentration increases by a factor of about 2 until the final trimester (Keen *et al*, 1998). Therefore, Cu deficiency in pregnancy is rare. Premature babies have an increased risk of Cu deficiency. *In utero* the foetus stores Cu predominantly during the final trimester. A mature infant has acquired Cu stores of approximately, 15–17 mg Cu during pregnancy, 2.5–9.0 mg of which are found in the liver (Widdowson *et al*, 1974). These reserves prevent Cu deficiency in the

first 47 months after full-term birth (Cordano, 1998). If delivery becomes due before the 34th week of pregnancy, plasma Cu concentrations are decreased correspondingly (Sann *et al*, 1980). Cu mobilisation from hepatic stores depends on liver maturation (Mason *et al*, 1981). This is why Cu supply to organs and enzymes does not reach the same level in premature as in full-term babies before the same age after conception is reached (Hillman, 1981).

The question whether abundant Cu stores and ineffective biliary Cu excretion in neonates presents an evolutionary advantage or a toxicological risk (Danks, 1991) (see Section 8), thus, has different answers in premature babies and full-term infants. Due to rapid growth, premature babies have an increased Cu requirement (Shaw, 1992) as their digestive capacity and intestinal Cu absorption is not yet fully matured. Therefore, about a third of very low birth weight infants fed with normal infant formula have a suboptimal Cu status at the age of 6–12 months (Beshgetoor & Hambidge, 1998). The American Academy of Paediatrics recommends to feeding infant formulas with a Cu content of 900 µg Cu/l to premature babies to compensate for these deficits. This recommendation seems to be justified because these children absorb no more than approximately 15% of the Cu supplied with bottled milk due to its high casein content (cf. Section 2), whereas they absorb about 60% from breast milk (Ehrenkranz *et al*, 1989).

### Deficiency due to inadequate intake

Malabsorption in the context of chronic enteropathies may result in hypocupraemia (Cordano & Graham, 1966). Several case reports have described Cu deficiency states in association with coeliac disease (Goyers *et al*, 1985). Total parenteral nutrition without adequate Cu supplementation can lead to overt Cu deficiency not only during rapid growth (Shulman, 1989), but also in adults (Fleming, 1989; Okada, 1994). Accordingly, the American Medical Association has recommended guaranteeing a supply of 0.5–1.5 mg Cu/day during total parenteral nutrition (American Medical Association, 1979). This is based on several observations: A supply of 1.3 mg Cu/day prevented Cu deficiency in adults (Shenkin *et al*, 1987). As little as 0.3 mg Cu/day maintained an adequate Cu status in adults in another study (Shike *et al*, 1981). Cu deficiency was compensated by administration of 1 mg Cu/day over a period of 2 weeks, followed by 0.5 mg Cu/day in adults and 0.2 mg Cu/day in children to maintain the status (Shike *et al*, 1981; Shike, 1984). However in cholestasis, the parenteral Cu supply must be reduced (WHO, 1998).

## 8. How can damage caused by Cu overload be diagnosed? Are there risk groups for oral Cu overload?

'Clinical diagnosis of Cu overload can only be established with certainty by liver biopsy. There is controversy as to

whether Cu poses a special risk for infants and small children that are not breast-fed.'

An infant of 4 kg body weight fed with reconstituted dried milk has an intake of up to 140 ml/kg/day. At a weight of 10–20 kg this is reduced to about 100 ml/kg/day, while an adult of 70 kg body weight consumes about 30 ml of fluid/kg/day. Unlike in bottle-fed babies, adults fluid intake derives from various sources which helps to avoid one-sided exposure. Due to these reasons, the exposure of infants and small children to potentially toxic substances in the drinking water is 3–5 times higher than for adults. This is no problem as long as dose–response relationships are assessed sufficiently conservative (Dieter, 1995). However, our knowledge on the dose-response relationship for hazardous Cu effects is incomplete.

#### Minimum intake vs maximum intake of Cu

A daily dietary intake of 1–2 mg Cu/day seems necessary to provide for Cu-dependent metabolic processes in adults. This corresponds to the most recent lower limit of the 'acceptable range of oral intake' (AROI) for adults of 20 g Cu/kg/day (WHO, 1998). Due to the increased metabolic requirement, the corresponding value for infants is 50 µg Cu/kg/day (=0.2–0.3 mg Cu/day; cf. Section 7a). On the other hand, the WHO (1998) stated that the upper limit of the AROI for adults is not known; it is probably in the order of several, but not many, mg per day per adult. There is no statement regarding the upper limit of AROI in infants or to whether the 'upper limit of the AROI' is a threshold for adverse Cu effects or rather a maximum for a physiologically desirable Cu intake (WHO, 1998). Some calculations may help to differentiate between these two options.

#### Physiologically desirable Cu concentrations in the drinking water for bottle-fed babies

The Cu content in 75 different commercial milk formulas from 10 manufacturers varies between 400 and 700 µg/l (mean *ca* 500 µg/l), when the milk was rehydrated with Cu-free drinking water (Blasco *et al*, 1999). The corresponding Cu intake of bottle-fed babies is up to 75 µg/kg/day, which is 1.5 times the minimum intake recommended by the WHO (1998). Additional Cu intake from the water, thus is not needed from a physiological point of view. The upper limit of the excretory capacity in infants is estimated to be 50 µg Cu/kg/day (Aggett, 1999). Taking this value and assuming 50% absorption for Cu in the gastrointestinal tract, no more than 0.17 mg Cu/l in the milk are needed to satisfy physiological requirements. However, the preliminary WHO drinking water standard (2 mg Cu/l, WHO, 1993) is more than 10 times higher. This value derives from data on the acute gastrointestinal side effects of Cu that were characterised as not very solid. It was noted that studies are underway to quantify Cu-induced effects in man more exactly, especially in sensitive subgroups (WHO, 1993).

Due to this situation it was discussed that exposure to milk containing 2 mg Cu/l, as approved by the WHO (1993) and possibly also to lower Cu concentrations might be hazardous for infants regarding the development of liver cirrhosis (Hadorn, 1999). On the other hand, a single case of liver cirrhosis was proven to be associated with public water supply, although millions of German homes are equipped with Cu plumbing (Vergara *et al*, 2000). Still, the scarcity of positive observations does not rule out completely that Cu in the drinking water may increase the hazard for hepatic damage in infants (Dieter *et al*, 1999).

#### How much Cu have infants with 'idiopathic copper toxicosis (ICT)' ingested with resuspended formula milk?

The idea that Cu concentrations between 0.2 and 2.0 mg/l may be harmful to the health of bottle-fed babies seems unlikely on the basis of plausible pharmacokinetic considerations (Tanner, 1998): assumed that the liver of a 1-y-old child suffering from ICT weighs 900 g (for comparison, a normal, healthy liver would weigh about 300–400 g) and has the average pathological content of *ca* 400 µg Cu/g wet weight, the total liver Cu content would be 360 mg Cu. Up to 9 mg of Cu are acquired by the liver during pregnancy (see Section 7b) and are likely to be excreted or redistributed during the first year of life. If the child is assumed to consume 1 l of formula milk containing 3 mg Cu/l over a year, which is a high estimate, and if absorption amounted to 50%, total Cu absorption corresponds to 550 mg Cu. Thus, the liver would store over 70% of the absorbed Cu quantity without providing for Cu excretion. However, a healthy child excretes *ca* 30 µg Cu/kg/day, corresponding to 100–200 µg/day, which sums up to at least 50 mg during the first year of life (Aggett, 1999).

During the perinatal period, the liver contains 50–60% of the body Cu content at most. From the 6th month on this fraction decreases to 10–20%, unless a genetic disorder of Cu metabolism were present. Taking these figures, the daily absorption of 1.5 mg Cu/day from the diet would correspond to a hepatic Cu content of 60–70 mg Cu at the end of the first year of life. This corresponds to a Cu concentration of about 200 µg/g wet weight in a normal-weight liver (300–400 g) and to just 70 µg/g wet weight in an enlarged liver of 900 g. However, hepatic Cu concentrations determined in ICT children are 20–30 times higher than that! Assuming an intake of 3 mg Cu/day with reconstituted milk, such high hepatic Cu concentrations can accumulate only if biliary Cu excretion were far below the normal rate (30 µg/kg/day) and if much less than 80% of the body Cu content were distributed to other organs after the 6th month of life, ie when Cu metabolism were markedly disturbed.

#### Remaining uncertainties

Ceruloplasmin synthesis and, thus, Cu distribution to other tissues is underdeveloped during the first 3–6 months of life.

Between the first and fourth month the mean plasma concentration of ceruloplasmin increases from barely 1 to 2  $\mu\text{mol/l}$ , and reaches adult levels (2–3  $\mu\text{mol/l}$ ) by the 12th month of life. This development was observed in breast-fed infants as well as after intake of formula milk with low Cu and high Cu content (85 vs 500  $\mu\text{g Cu/l Cu}$ ). The Cu content in the supplementary diet was not determined in this study (Salmenperä *et al*, 1989).

Biliary Cu excretion increased in parallel to the increase in ceruloplasmin synthesis, reaching adult levels during the 3rd to 6th months. Thus, biliary excretion is as ineffective as ceruloplasmin synthesis in early infancy. In addition, the frail bile ducts in infants are particularly susceptible to infection. 'Neonatal hepatitis' may impair or stop the bile flow and lead to cholestatic Cu retention (Epstein, 1983; Danks, 1991). For these reasons, normal hepatic Cu concentrations found in fetuses and during the first 6 months of life (200–400  $\mu\text{g/g}$  dry weight) are about 10 times higher than in adults (Aggett, 1999). The assumption that such high Cu levels are essential stores for physiological requirement in full-term neonates is difficult to maintain (Danks, 1991), as the Cu amount supplied with the breast milk under normal conditions (0.1–0.3  $\text{mg/l}$ ) is easily adequate for its essential requirements (Aggett, 1999; Salmenperä *et al*, 1989).

It cannot be decided at present whether the high hepatic Cu levels in infants indicate an increased sensitivity for Cu exposure. If one compares the example of an adult who developed severe cirrhosis after 3 y exposure to 0.5–1.0  $\text{mg Cu/kg/day}$  (O'Donohue *et al*, 1993, to that physiological Cu intake during the first 6 months of life (0.4–0.4  $\text{mg Cu/kg/day}$ ) one feels tempted to speculate that a healthy infants might be less sensitive to high Cu exposure than the adult liver, in spite of its underdeveloped Cu excretion capacity. Correspondingly, a prospective exposure study involving 128 infants than 3 months (Olivares *et al*, 1998) and the analysis of copper exposure in 1178 children older than 9 months of age (Pettersson & Rasmussen, 1999) did not indicate any particular sensitivity to Cu in those children. Despite daily administration of 2  $\text{mg Cu/L}$  (Olivares *et al*, 1998) or an estimated load of up to 3.2  $\text{mg Cu/day}$  with the drinking water (Pettersson & Rasmussen, 1999) no signs of liver damage were observed in the exposed children. However, none of these studies included the critical age group of infants younger than 3 months, who might be particularly sensitive to Cu.

## 9. Is there a relationship between Cu metabolism and (a) neurodegenerative diseases and (b) cardiovascular diseases?

'Findings in the field of molecular biology suggest an association between Cu metabolism and neurodegenerative diseases. Also, there are mechanisms that suggest a

relationship between Cu intake and coronary heart disease (CHD).'

### Neurodegenerative diseases

Neurodegenerative diseases appear to be directly related to Cu metabolism. Menkes' syndrome and Wilson's disease are inherited disorders with a functional disturbance of two membrane-located ATPases for the transport of Cu ions (Bull *et al*, 1993; Chelly & Monaco, 1993). In Wilson's disease Cu excretion via the bile ducts is disturbed. Consequently, Cu accumulates predominantly in the liver. In Menkes' syndrome, on the other hand, Cu-dependent enzymes, such as cytochrome-C oxidase and lysyloxidase, are not adequately supplied with Cu.

In Menkes' syndrome, clinical and pathological features are already present at birth, implying that Cu-dependent enzymes are of importance already in the prenatal phases of CNS growth and development. Myelination disorders of nerve cells, diffuse atrophy of the brain, focal disintegration of the grey matter with destruction of axons in the white matter and extensive necrosis of Purkinje cells in the cerebellum are observed in Menkes' syndrome. These are sequels of inadequate Cu supply to Cu-containing enzymes and lead to mental retardation of the patients.

Hepatic Cu deposits in Wilson's disease damage the liver. In adolescents and young adults an increased number of neurological deficits show up. Cu deposition in the basal ganglia leads to gliosis and to neurological deficits with symptoms of Parkinson's disease. They go along with changes in personality, depression and schizophrenia, although the molecular mechanisms of these events are not known.

Amyotrophic lateral sclerosis (ALS) is a neurodegenerative disease with a sporadic and an inherited form. A loss of motor neurones in the spinal cord and in the brain causes progressive muscular weakness. Observations in siblings have shown that mutations in Cu, Zn-superoxide dismutase (SOD1) may be responsible for the disease (Deng *et al*, 1993; Rosen *et al*, 1993). Animal experiments showed that the disease is not due to decreased enzyme activity. The mutant SOD1 enzyme can also act as a peroxidase as a result of a change in protein conformation (Matzuk *et al*, 1998; Reaume *et al*, 1996). In the presence of Cu the peroxidase activity increases free radical activity, which can be demonstrated by indirect methods in transgenic animals and in patients (Liu *et al*, 1998; Ferrante *et al*, 1997). As cofactor for the mutant SOD-1 molecules, Cu is likely to be directly responsible for the toxic effects. The radicals seem to oxidise aggregate proteins. These changes may retard axonal transport and damage the neurofilaments of motor neurones (Brujin *et al*, 1998; Williamson & Cleveland, 1999).

Alzheimer's disease is a progressive neurodegenerative disease. It is regarded the most common type of dementia. Its particular characteristic is a deposition of amyloid-A $\delta$ -proteins which are observed in all sporadic and familial

forms of the disease (Selkoe, 1998). There is direct evidence that oxidative stress is involved in Alzheimer pathogenesis. Thus, increase oxidation of proteins and DNA is seen in those brain regions that show amyloid deposits first (Markesbery & Carney, 1999). Cu ions can stimulate the aggregation of synthetic A $\beta$ -protein *in vitro*. The concentration is increased 6–8-fold in the characteristic protein deposits in affected patients (Atwood *et al*, 1998; Lovell *et al*, 1998). This fits in with the finding that amyloid-A $\delta$  deposits can be redissolved in the presence of zinc- or Cu-specific chelators in the brain of Alzheimer patients post mortem (Cherny *et al*, 1999). A probable source for the Cu that concentrates in the deposits is the amyloid precursor protein (APP). It contains a Cu-binding domain from which amyloid-A $\delta$  is naturally formed by proteolytic activity (Selkoe, 1998; Multhaup *et al*, 1997).

Numerous findings support the hypothesis that APP is involved in the transport of Cu into the cell. APP is synthesised as a membrane-located and secreted protein. It binds Cu and can reduce Cu<sup>2+</sup> to Cu<sup>+</sup> (Hesse *et al*, 1994; Multhaup, 1997; Multhaup *et al*, 1996). The protein is transported from the cell to the axonal membrane and to the dendritic plasma membrane and could, thus, transport Cu<sup>+</sup> along this way (Simons *et al*, 1995). Cu<sup>+</sup> bound to this protein is rapidly oxidized by reactive oxygen species (Multhaup *et al*, 1999). This reaction splits APP. Some fragments contain the entire A $\delta$  domain and are regarded as neurotoxic derivatives of APP within the cell (Simons *et al*, 1996). By virtue of their Cu-binding domains, APP–Cu complexes themselves can cause oxidative stress in neuronal cell culture systems (White *et al*, 1999). The most recent evidence for a role of APP in cellular Cu homeostasis comes from cell and animal models used in Alzheimer's research. In mice that do not produce any APP, Cu concentrations in the liver and cerebral cortex are increased by 80% and 40%, respectively (White *et al*, 1999). Besides, additional Cu in the medium also affects the physiological processing of APP, presumably by regulation of APP transport via a conformational change in the protein (Borchardt *et al*, 1999). Thus, an involvement of Cu in Alzheimer pathogenesis is conceivable, eg when the transport of APP–Cu complexes were disturbed or when APP molecules would aggregate in axons and thereby initiate the amyloid cascade.

Prion diseases generally occur as sporadic, dominantly inherited or infective diseases (Aguzzi & Brandner, 1999). Creutzfeldt–Jacob disease (CJD) in man, for example, is characterised by a rapidly progressing dementia and cerebellar ataxia. Spongiform changes are found in the tissue with gliosis of the astrocytes. In the course of pathogenesis, the cellular prion protein PrP<sup>C</sup> is transformed into an infectious prion protein PrP<sup>Sc</sup> and aggregates in the form of amyloid-like plaques in affected brains (Merz *et al*, 1981).

There are reasons to believe that cellular PrP is involved in CNS Cu homeostasis. Thus, the Cu chelating agent cuprizone induces similar neurological alterations in mice as were observed in prion diseases (Kimberlin *et al*, 1974; Pattison *et al*, 1971). In contrast to APP-deficient mice (see above), the

brain cells of mice in which the PrP gene is turned off are more sensitive to Cu salts (Bueler *et al*, 1992). They show a reduced SOD1 activity and a lower Cu content in the cell membranes (Brown *et al*, 1997; 1998). Since Cu ions stimulate PrP uptake into cells (Pauly & Harris, 1998), it is assumed that PrP might be involved in cellular uptake.

Cu binds to the N-terminus of PrP (Hornshaw *et al*, 1995; Muira *et al*, 1996) and changes the conformation of the protein into a  $\delta$ -folding structure (Stöckel *et al*, 1998). *In vitro*, Cu binding causes the prion protein to regain its infective potential and its resistance to proteolytic destruction (McKenzie *et al*, 1998). PrP<sup>Sc</sup> molecules are found in the affected tissues as protein/metal ion complexes containing zinc and Cu. Obviously, Cu binding causes conformational changes in PrP<sup>Sc</sup> which could also explain the diversity of prion strains (Wadsworth *et al*, 1999).

The physiological significance of Cu binding to PrP is not yet elucidated. However, on the basis of the findings described above one can assume that PrP co-regulates cellular Cu homeostasis. It is involved in cellular Cu uptake and subsequently, as Cu chaperone, it delivers Cu to corresponding target proteins.

### Coronary heart disease (CHD)

Disturbed Cu uptake has been proposed as a risk factor for coronary heart disease (Klevay, 1975; 1980). HMG-CoA-reductase is a key enzyme in endogenous cholesterol synthesis and its activity is increased in Cu deficiency (Yount *et al*, 1990). In parallel, the sensitivity of VLDL and LDL to oxidation increases in Cu deficiency (Raysiguier *et al*, 1993). LDL oxidation is a significant pathogenetic factor in the development of arteriosclerosis and hence of CHD. Correspondingly, there is evidence that an inadequate Cu intake favours the development of hypercholesterolaemia. An increase in total cholesterol, LDL-cholesterol and decrease in HDL-cholesterol were observed in Cu deficiency (Klevay *et al*, 1984). The effect of Cu depletion in animal vessels and the changes found in patients with CHD show apparent similarities (elastin destruction, fibrosis, intramural haemorrhages, increase in mucopolysaccharides, necrosis and the proliferation of smooth muscle cells; (Klevay, 1993). The Yi People Study (He *et al*, 1992) found an inverse correlation between serum Cu and the risk of CHD. However, Cu intake was not monitored in this study, and the problems related to the use of serum copper concentrations as the only indicator of Cu status were commented on earlier. In contrast to these observations, some epidemiological studies found a positive correlation between Cu uptake and the risk of CHD (Manthey *et al*, 1981; Kok *et al*, 1988).

Macroangiopathy in Cu-deficient animals is primarily attributable to a decreased activity of lysyloxidase. This Cu-dependent enzyme cross-links collagen and elastin (Reiser *et al*, 1992), and a decreased activity enzyme impairs the integrity of the arterial wall (Schuschke, 1997). Cu deficiency might also impair the effect on the elasticity of the arterial

wall via a reduction in NO-induced vasodilatation (Saari, 1992; Schuschke *et al*, 1992). This effect is supposed to be mediated via increased NO degradation by superoxide anions as a consequence of decreased SOD activity in Cu deficiency (Dubick *et al*, 1988; Belch *et al*, 1989). These points are reviewed in greater detail by Strain (1994).

## 10. What damage can be caused by Cu overload?

'Acute: gastrointestinal symptoms, death. Chronic: damage to target organs show individual variation.'

### Acute toxicity

After acute oral intake Cu is relatively non-toxic for man. Acute intoxications have primarily been observed after accidental or suicidal ingestion, and due to inhalation of Cu dusts and fumes at work. Milligram amounts of soluble Cu salts can be ingested with fruit juices that were stored in Cu containers or with water from Cu plumbing. Such intake may cause vomiting and diarrhoea. In severe cases, soluble Cu compounds in gram quantities lead to haemolysis, liver and kidney damage, coma and death. The lowest acutely fatal dose in man is about 10 g Cu. Inhalation of Cu fumes irritates the upper airways, and may lead to malaise, stomach pain and diarrhoea.

### Chronic toxicity

Chronic Cu intoxication from contaminated food is rare and mainly affects the gastrointestinal tract and the liver. In patients with Wilson's disease, haemolysis, neurological symptoms and renal damage are observed in addition. A family in Vermont experienced recurrent gastrointestinal irritation as a result of Cu-contaminated water consumption (2.8–7.8 mg Cu/l; Spitalny *et al*, 1984). Excess Cu concentrations are also found in infant milk stored in Cu containers or resuspended in Cu contaminated water (Tanner, 1998; Sethi *et al*, 1993; see Section 8). A 26-y-old man, who had taken 30 mg Cu/day as Cu gluconate for 2 y and then 60 mg Cu for a further year developed cirrhosis of the liver and acute liver failure (O'Donohue *et al*, 1993).

### Mechanisms of toxicity

Cu toxicity is linked to its unique electron configuration. Thus, Cu<sup>+</sup> ions easily be polarised and bind predominantly to ligands rich in nitrogen and sulphur. Cu<sup>2+</sup> ions tend to form complexes with ligands containing nitrogen and sulphur. Therefore, Cu is comparably reactive in biological matrices and can form strong bonds with a wide variety of structures that are rich in electrons. The replacement of other essential metals in enzymes by Cu or Cu-binding to macromolecules may have deleterious effects on their function and structure, resulting, for example, in enzyme inhibi-

tion and alterations of DNA and membrane structures (Alt *et al*, 1990). In addition, Cu is involved in redox reactions. Cu ions are able to catalyse the formation of hydroxyl radicals via Fenton chemistry (Goldstein & Czapsky, 1986; see Section 5). These redox-reactions can increase cytoplasmatic calcium concentrations, cause ATP depletion and excess oxidation of thiols, DNA and membranes, which may finally lead to a loss of cellular integrity (Bremner, 1998).

Apart from whole body Cu content, the subcellular distribution and binding behaviour is of crucial importance for Cu toxicity. Thus, Cu transfer from the cytoplasm into lysosomes appears to be essential for the development of Cu-induced liver damage (Goldfischer & Sternlieb, 1968; Klein *et al*, 1998). Chaperones mediate intracellular Cu distribution to the sites of functional or toxic action (Pena *et al*, 1999). The low-molecular protein metallothionein is crucial for intracellular Cu storage and detoxification. Thus, Cu administration induces metallothionein synthesis *in vivo* (Bremner, 1987) and high metallothionein concentrations reduce Cu toxicity (Freedman *et al*, 1989). Metallothionein is also assumed to have a direct antioxidant effect by reacting readily with hydroxyl radicals (Thornalley & Vasak, 1985). Correspondingly, mouse cells that can't synthesise metallothionein proved to be particularly sensitive to oxidative stress (Liu *et al*, 1995).

## 11. Future research requirements

'Markers for the determination of Cu status and Cu requirement (SOD); mechanisms of Cu homeostasis at the molecular level; Cu and gene expression; polymorphisms, the regulation of Cu transport at the blood–tissue barrier, molecular mechanisms of age-dependent Cu transport across the blood–brain barrier and of the distribution in blood (fractions); molecular mechanisms of chronic toxicity in relation to the cellular and subcellular Cu distribution.'

The significance of Cu for human nutrition has not been adequately elucidated. In particular, reliable biomarkers are lacking to permit reliable determination and interpretation of the individual Cu status. Research preferences in this field should be on the clinical applicability. The lack of reliable markers for Cu status makes it difficult to assess the benefit of increased Cu intake for risk groups or in the prevention of disease and to determine threshold values for Cu toxicity. From a toxicological point of view, Cu metabolism in the neonate needs to be investigated in detail, particularly absorption and hepatic retention and excretion of Cu. Therefore, a better understanding of the mechanisms of Cu homeostasis would be helpful, eg the dependence of Cu transport across the blood–brain barrier on concentration and age. Using molecular methods it should be possible to study the effects of marginal Cu intake on the gene expression of Cu-

dependent transcription factors. A particular challenge for the future is the demonstration of genetic polymorphisms in the expression of Cu-dependent proteins and their functional consequences. With such data it will be possible to define risk groups for excessive or inadequate Cu intake and prophylactic treatment.

## Note

The 15th Hohenheim Consensus Workshop took place in March 1999 at the University of Stuttgart-Hohenheim. The subject of this conference was copper (Cu). Seven German and Austrian scientists who had published and reviewed scientific and regulatory papers on Cu were invited, among them basic researchers, toxicologists, clinical chemists, nutritionists and scientific regulators from the Umweltbundesamt. The participants were presented with 11 questions, which were discussed and answered at the workshop, with the aim of summarising the current state of knowledge on Cu. The explicatory text accompanying the short answers was produced and agreed on after the conference and was backed up by corresponding references. Because of the lack of space, it was not possible to include all references provided by the authors.

## References

- Abdel Mageed AB, Welti R, Oehme FW & Pickrell RA (1994): Perinatal hypocuprosis affects synthesis and composition of neonatal lung collagen, elastin, and surfactant. *Am. J. Physiol.* **267**, L679–685.
- Aggett PJ (1999): An overview of the metabolism of copper. *Eur. J. Med. Res.* **4**, 214–216.
- Aguzzi A & Brandner S (1999): The genetics of prions—a contradiction in terms? *Lancet* **354**(Suppl 1), 122–125.
- Alt ER, Sternlieb I & Goldfischer S (1990): The cytopathology of metal overload. *Int. Rev. Exp. Pathol.* **31**, 165–188.
- American Medical Association (1979): Guidelines for essential trace element preparations for parenteral use. *J.A.M.A.* **241**, 2051–2054.
- Atwood CS, Moir RD, Huang X, Scarpa RC, Bacarra NM, Romano DM, Hartshorn MA, Tanzi RE & Bush AI (1998): Dramatic aggregation of Alzheimer abeta by Cu(II) is induced by conditions representing physiological acidosis. *J. Biol. Chem.* **273**, 12817–12826.
- Baker DH & Ammerman CB (1995): Copper bioavailability. In *Bioavailability of Nutrients For Animals*, ed. DH Baker, CB Ammerman & Lewis AJ, pp 127–156. San Diego: Academic Press.
- Battersby S & Chandler JA (1977): Correlation between elemental composition and motility of human spermatozoa. *Fertil. Steril.* **28**, 557–561.
- Belch JFF, Chopra M, Hutchinson S, Stannick RD, Forbes CD & WE Smith (1989): Free radical pathology in chronic arterial disease. *Free Rad. Biol. Med.* **6**, 375–378.
- Beshgetoor D & Hambidge M (1998): Clinical conditions altering copper metabolism in humans. *Am. J. Clin. Nutr.* **67**(Suppl), 1017S–1021S.
- Blasco N, Galindo A & Pérez M (1999): Estimated daily intakes and concentrations of essential trace elements in infant formulas. Abstract on behalf of TEMA 10, Evian/France, 2–7 May.
- Borchardt T, Camakaris J, Cappai R, Masters CL, Beyreuther K & Multhaup G (1999): Copper inhibits  $\beta$ -amyloid production and stimulates the non-amyloidogenic pathway of amyloid precursor protein (APP) secretion. *Biochem. J.* **344**, 461–467.
- Bremner I (1987): Involvement of metallothionein in the hepatic metabolism of copper. *J. Nutr.* **117**, 19–29.
- Bremner I (1998): Manifestations of copper excess. *Am. J. Clin. Nutr.* **67**, 1069S–1073S.
- Brigelius R, Spottl R, Bors W, Lengfelder E, Saran M & Weser U (1974): Superoxide dismutase activity of low molecular weight Cu<sup>2+</sup> chelates studies by pulse radiolysis. *F.E.B.S. Lett.* **47**, 72–74.
- Brown DR, Quin K, Herms JW, Madlung A, Manson J, Strome R, Fraser PE, Kruck T, von Bohlen A, Schulz-Schaeffer W, Giese A, Westaway D & Kretschmar H (1997): The cellular prion protein binds copper in vivo. *Nature* **390**, 684–687.
- Brown DR, Schmidt B & Kretschmar HA (1998): Effects of copper on survival of prion protein knockout neurons and glia. *J. Neurochem.* **70**, 1686–1693.
- Brujin LI, Houseweart MK, Kato S, Anderson KL, Anderson SD, Ohama E, Reaume AG, Scott RW & Cleveland DW (1998): Aggregation and motor neuron toxicity of an ALS-linked SOD1 mutant independent from wild-type SOD1. *Science* **281**, 1851–1854.
- Buamak PH, Russell M, Milford-Ward A, Taylor P & Roberts DF (1984): Serum copper concentration significantly less in abnormal pregnancies. *Clin. Chem.* **30**, 1667–1670.
- Bueler H, Fischer M, Lang Y, Bluethmann H, Lipp HP, DeArmond SJ, Prusiner SB, Aguet M & Weissmann C (1992): Normal development and behaviour of mice lacking the neuronal cell-surface PrP protein. *Nature* **356**, 577–582.
- Bull PC, Thomas GR, Rommenes JM, Forbes JR & Cox DW (1993): The Wilson disease gene is a putative copper transporting P-type ATPase similar to the Menkes gene. *Nature Genet.* **5**, 327–337.
- Castillo-Duran C & Uauy R (1988): Copper deficiency impairing growth of infants recovering from malnutrition. *Am. J. Clin. Nutr.* **47**, 710–714.
- Castillo-Duran C, Vial P & Uauy R (1988): Trace element balance during acute diarrhea in infants. *J. Pediatr.* **113**, 452–457.
- Castillo-Duran C, Vial P & Uauy R (1990): Oral copper supplementation: effect on copper and zinc balance during acute gastroenteritis in infants. *Am. J. Clin. Nutr.* **51**, 1088–1092.
- Chelly J & Monaco AP (1993): Cloning the Wilson disease gene. *Nature Genet.* **5**, 317–318.
- Cherny RA, Legg JT, McLean CA, Fairlie DP, Huang X, Atwood CS, Beyreuther K, Tanzi RE, Masters CL & Bush AI (1999): Aqueous dissolution of Alzheimer's disease abeta amyloid deposits by biometal depletion. *J. Biol. Chem.* **274**, 23223–23228.
- Cordano A (1998): Clinical manifestation of nutritional copper deficiency in infants and children. *Am. J. Clin. Nutr.* **67**(Suppl), 1012S–1016S.
- Cordano A & Graham GG (1966): Copper deficiency complicating severe chronic intestinal malabsorption. *Pediatrics* **38**, 506–604.
- Crampton RF, Matthews DM & Poisner R (1965): Observations on the mechanism of absorption of copper by small intestine. *J. Physiol.* **178**, 111–126.
- Danks DM (1991): Copper and liver disease. *Eur. J. Pediatr.* **50**, 142–148.
- Deng HX, Hentati A, Tainer JA, Iqbal Z, Cayabyab A, Hung WY, Getzoff ED, Hu P, Herzfeldt B, Roos RP, Warner C, Deng G, Soriano E, Smyth C, Parge HE, Ahmed A, Roses AD, Hallewell RA, Pericak-Vance MA & Siddique T (1993): Amyotrophic lateral sclerosis and structural defects in Cu, Zn superoxide dismutase. *Science* **261**, 1047–1051.
- Deutsche Gesellschaft für Ernährung (DGE) (2000): *Referenzwerte für die Nährstoffzufuhr*. Frankfurt: Umschau.
- Dieter HH (1995): Risikoquantifizierung: Abschätzungen, Unsicherheiten und Gefahrenbezug. *Bundesgesundh. bl.* **38**, 250–257.
- Dieter HH, Schimmelpfennig W, Meyer E & Tabert M (1999): Early Childhood Cirrhosis (ECC) in Germany between 1982 and 1994 with Special Consideration of Copper Etiology. *Eur. J. Med. Res.* **4**, 233–242.
- Dijkstra M, Vonk RJ & Kuipers F (1996): How does copper get into bile? New insights into the mechanism(s) of hepatobiliary copper transport. *J. Hepatol.* **24**, 109–120.
- Dörner K, Dziadzka S & Hohn A (1989): Longitudinal manganese and copper balances in young infants and preterm infants fed on breastmilk and adapted cow's milk formulas. *Br. J. Nutr.* **61**, 559–572.

- Dubick MA, Zidenberg-Cherr S, Rucker RB & Keen CL (1988): Super-oxide dismutase activity in lung from copper and manganese-deficient mice exposed to ozone. *Toxicol. Lett.* **42**, 149–154.
- Ehrenkranz RA, Gettner PA & Nelli CM (1989): Zinc and copper nutritional studies in very low birth weight infants: comparison of stable isotopic extrinsic tag and chemical balance methods. *Pediatr. Res.* **26**, 298–307.
- Elmadfa I, Burger P, Derndorfer E, Kiefer I, Kunze M, König J, Leimüller G, Manafi M, Mecl M, Papathanasiou V & Rust P (1999): *Österreichischer Ernährungsbericht 1998*. Wien: Bundesministerium für Gesundheit, Arbeit und Soziales und Bundesministerium für Frauenangelegenheiten und Verbraucherschutz.
- Epstein O (1983): Liver copper in health and disease. *Postgrad. Med. J.* **59**(Suppl), 88–94.
- Evans PJ, Bomford A & Halliwell B (1998): Non-coeruloplasmin copper in human serum, does it exist? Abstract on behalf of the 5th Meeting of the European Copper Research Group, Kinsale, Ireland, 28–29 April
- Ferrante RJ, Browne SE, Shinobu LA, Bowling AC, Baik MJ, MacGarvey U, Kowall NW, Brown RH Jr & Beal MF (1997): Evidence of increased oxidative damage in both sporadic and familial amyotrophic lateral sclerosis. *J. Neurochem.* **69**, 2064–2074.
- Fleming CR (1989): Trace element metabolism in adult patients requiring total parenteral nutrition. *Am. J. Clin. Nutr.* **49**, 573–579.
- Fransson G-B & Lönnerdal B (1983): Distribution of trace elements and minerals in human and cow's milk. *Pediatr. Res.* **17**, 912–915.
- Freedman JH, Ciriolo MR & Peisach J (1989): The role of glutathione in copper metabolism and toxicity. *J. Biol. Chem.* **264**, 5598–5605.
- Gittlin JD (1998): Review: aceruloplasminemia. *Pediatr. Res.* **44**, 271–276.
- Goldstein S & Czapsky G (1986): The role and mechanism of metal ions and their complexes in enhancing damage in biological systems or in protecting these systems from the toxicity of O<sub>2</sub>. *J. Free Radic. Biol. Med.* **2**, 3–11.
- Goldfischer S & Sternlieb I (1968): Changes in the distribution of hepatic copper in relation to the progression of Wilson's disease (hepatolenticular degeneration). *Am. J. Pathol.* **53**, 883–901.
- Goyers P, Brasseur D & Cadranel S (1985): Copper deficiency in infants with active celiac disease. *J. Pediatr. Gastroenterol. Nutr.* **4**, 677–680.
- Gutteridge JM (1980): Iron–oxygen reactions and their use in clinical chemistry. *Med. Lab. Sci.* **37**, 267–273.
- Gutteridge JM (1994): Hydroxyl radicals, iron, oxidative stress, and neurodegeneration. *Ann. N.Y. Acad. Sci.* **738**, 201–213.
- Gutteridge JMC (1984): Copper-phenantroline induced site specific oxygen radical damage to DNA. Detection of loosely bound trace copper in biological fluids. *Biochem. J.* **218**, 983–985.
- Hadorn HB (1999): Copper-associated liver diseases in children. Proceedings of the 4th Emma-Thaler-Symposium (Introduction). *Eur. J. Med. Res.* **4**, 212–213.
- Hall AC, Young BW & Bremner I (1979): Intestinal metallothionein and the mutual antagonism between copper and zinc in the rat. *J. Inorg. Biochem.* **11**, 57–66.
- Halliwell B & Gutteridge JM (1985): The importance of free radicals and catalytic metal ions in human diseases. *Mol. Asp. Med.* **8**, 89–193.
- Halliwell B & Gutteridge JM (1990): Role of free radicals and catalytic metal ions in human disease: an overview. *Meth. Enzymol.* **186**, 1–85.
- Harris ED (1991): Copper transport: an overview. *Proc. Soc. Exp. Biol. Med.* **196**, 130–140.
- He J, Tell GS, Tang Y-C, Mo P-S & He G-Q (1992): Relation of serum zinc and copper to lipids and lipoproteins: the Yi people study. *J. Am. Coll. Nutr.* **11**, 74–78.
- Hesse L, De Beher, CL Masters & G Multhaup (1994): The beta A4 amyloid precursor protein binding to copper. *F.E.B.S. Lett.* **349**, 109–116.
- Hillman LS (1981): Serial serum copper concentrations in premature and SGA infants during the first three month of life. *J. Pediatr.* **98**, 305–308.
- Hornshaw MP, McDermott JR, Candy JM & Lakey JH (1995): Copper binding to the N-terminal tandem repeat region of mammalian and avian prion protein: structural studies using synthetic peptides. *Biochem. Biophys. Res. Commun.* **214**, 993–999.
- Hurley LS & Keen CL (1979): Teratogenic effects of copper. In *Copper in the Environment. Part II: Health Effects*, ed. JO Nriagu, pp 33–56. New York: John Wiley.
- Johnson PE, Milne DB & Lykken GI (1992): Effects of age and sex on copper absorption, biological half-life, and status in humans. *Am. J. Clin. Nutr.* **56**, 917–925.
- Joo S-J & Betts NM (1996): Copper intakes and consumption patterns of chocolate foods as sources of copper for individuals in the 1987–88 nationwide food consumption survey. *Nutr. Res.* **16**, 41–52.
- Kardos J, Kovacs I, Hajos F, Kalman M & Simonyi M (1989): Nerve endings from rat brain tissue release copper upon depolarization. A possible role in regulation neuronal excitability. *Neurosci. Lett.* **103**, 139–144.
- Keen CL, Uriu-Hare JY, Hawk SN, Jankowski MA, Daston GP, Kwik-Uribe CL & Rucker RB (1998): Effect of copper deficiency on prenatal development and pregnancy outcome. *Am. J. Clin. Nutr.* **67**(Suppl), 1003S–1011S.
- Kehoe CA, Turley E, Bonham MP, O'Connor JM, McKeown A, Faughnan MS, Coulter JS, Gilmore WS, Howard AN & Strain JJ (2000): Response of putative indices of copper status to copper supplementation in human subjects. *Br. J. Nutr.* **84**, 151–156.
- Kelley DS, Daudu PA, Taylor PC, Mackey BE & Turnlund JR (1995): Effects of low-copper diets on human immune response. *Am. J. Clin. Nutr.* **62**, 412–416.
- Kimberlin RH, GC Millson, L Bountiff & SC Collis (1974): A comparison of the biochemical changes induced in mouse brain cuprizone toxicity and by scrapie infection. *J. Comp. Pathol.* **84**, 263–270.
- Klein D, Lichtmannegger J, Heinzmann U, Müller-Höcker J, Michaelson S & Summer KH (1998): Association of copper to metallothionein in hepatic lysosomes of Long-Evans cinnamon (LEC) rats during the development of hepatitis. *Eur. J. Clin. Invest.* **28**, 302–310.
- Klevay LM (1975): Coronary heart disease: the zinc/copper hypothesis. *Am. J. Clin. Nutr.* **28**, 764–774.
- Klevay LM (1980): Interactions of copper and zinc in cardiovascular disease. *Ann. N.Y. Acad. Sci.* **355**, 140–151.
- Klevay LM (1985): Atrial thrombosis, abnormal electrocardiograms and sudden death in mice due to copper deficiency. *Atherosclerosis* **54**, 213–224.
- Klevay LM (1993): Ischaemic heart disease: nutrition or pharmacotherapy. *J. Trace Elem. Electr. Health Dis.* **7**, 63–69.
- Klevay LM, Inman I, Johnson LK, Lawler M, Milne DB, Lukaski HC, Bolonchuk W & Sandstead HH (1984): Increased cholesterol in plasma in a young man during experimental copper depletion. *Metabolism* **33**, 1112–1118.
- Kok FJ, Duijn CMV, Hofman A, Voet GBVD, Wolff FAD, Paays CHC & Valkenburg HA (1988): Serum copper and zinc and the risk of death from cancer and cardiovascular disease. *Am. J. Epidemiol.* **128**, 352–359.
- König JS & Elmadfa I (1993): Copper status in Austrian school and Kindergarten children at the age of 4–19 years. In *Proceedings of the Eighth International Symposium on Trace Elements in Man and Animals—TEMA-8*, ed. M Anke, pp 896–897. Dresden.
- Lai CC, Huang WH, Klevay L, Gunning WT III & Chiu TH (1996): Antioxidant enzyme gene transcription in copper-deficient rat liver. *Free Rad. Biol. Med.* **21**, 233–240.
- Lee SH, Lancey R, Montaser A, Madani N & Linder MC (1993): Ceruloplasmin and copper transport during the latter part of gestation in the rat. *Proc. Soc. Exp. Biol. Med.* **203**, 428–439.

- Lin SJ & Culotta VC (1995): The ATX1 gene of *Saccharomyces cerevisiae* encodes a small homeostatic factor that protects cells against reactive oxygen toxicity. *Proc. Natl. Acad. Sci. USA* **92**, 3784–3788.
- Linder MC (1991): *The Biochemistry of Copper*. New York: Plenum Press.
- Linder MC & Hazegh-Azam M (1996): Copper biochemistry and molecular biology. *Am. J. Clin. Nutr.* **63**, 797S–811S.
- Liu R, Althaus JS, Ellerbrock BR, Becker DA & Gurney ME (1998): Enhanced oxygen radical production in a transgenic mouse model of familial amyotrophic lateral sclerosis. *Ann. Neurol.* **44**, 763–770.
- Liu Y, Liu J, Iszard MB, Andrews GK, Palmiter RD & Klaassen CD (1995): Transgenic mice that over-express metallothionein-1 are protected from cadmium lethality and toxicity. *Toxicol. Appl. Pharmac.* **135**, 222–228.
- Lönnerdal B (1998): Copper nutrition during infancy and childhood. *Am. J. Clin. Nutr.* **67**(Suppl), 1046S–1053S.
- Lönnerdal B, Bell JG & Keen CL (1985): Copper absorption from human milk, cow's milk and infant formulas using a suckling rat model. *Am. J. Clin. Nutr.* **42**, 836–844.
- Lovell MA, Robertson JD, Teesdale WJ, Campbell JL & Markesbery WR (1998): Copper, iron and zinc in Alzheimer's disease senile plaques. *J. Neurol. Sci.* **158**, 47–52.
- Ma J & Betts NM (2000): Zinc and copper intakes and their food sources for older adults in the 1994–96 continuing survey of food intakes by individuals (CSFII). *J. Nutr.* **130**, 2838–2843.
- McArdle HJ (1995): The metabolism of copper during pregnancy—a review. *Food Chem.* **54**, 79–84.
- McKenzie D, Bartz J, Mirwald J, Olander D, Marsh R & Aiken J (1998): Reversibility of scrapie inactivation is enhanced by copper. *J. Biol. Chem.* **273**, 25545–25547.
- Manthey J, Stoeppler M, Morgenstern W, Nussel E, Opherk D, Weintraut A, Wesch H & Kubler W (1981): Magnesium and trace metals: risk factors for coronary heart disease? Associations between blood levels and angiographic findings. *Circulation* **64**, 722–729.
- Markesbery WR & JM Carney (1999): Oxidative alterations in Alzheimer's disease. *Brain Pathol.* **9**, 133–146.
- Mason R, Bakka A, Samarawickrama GP & Webb M (1981): Metabolism of zinc and copper in the neonate: accumulation and function of (Zn-Cu)-metallothionein in the liver of the newborn rat. *Br. J. Nutr.* **45**, 375–389.
- Masuoka J, Hegenauer J, Van Dyke BR & Saltman P (1993): Intrinsic stoichiometric equilibrium constants for the binding of zinc (II) and copper (II) to the high affinity site of serum albumin. *J. Biol. Chem.* **268**, 21533–21537.
- Matzuk MM, Dionne L, Guo Q, Kumar TR & Lebovitz RM (1998): Ovarian function in superoxide dismutase 1 and 2 knockout mice. *Endocrinology* **139**, 4008–4011.
- Merz PA, Somerville RA, Wisniewski HM & Iqbal K (1981): Abnormal fibrils from scrapie-infected brain. *Acta Neuropathol.* **54**, 63–74.
- Mills CF & Williams RB (1962): Copper concentrations and cytochrome-oxidase and ribonuclease activities in the brains of copper deficient lambs. *Biochem. J.* **85**, 629–632.
- Milne DB (1994): Assessment of copper nutritional status. *Clin. Chem.* **40**, 1479–1484.
- Milne DB & Johnson PE (1993): Assessment of copper status: effect of age and gender on reference ranges in healthy adults. *Clin. Chem.* **39**, 883–887.
- Milne DB & Nielsen FH (1996): Effects of a diet low in copper on copper-status indicators in postmenopausal women. *Am. J. Clin. Nutr.* **63**, 358–364.
- Miura T, Horii A & Takeuchi H (1996): Metal-dependent alpha-helix formation promoted by the glycine-rich octapeptide region of prion protein. *F.E.B.S. Lett.* **396**, 248–252.
- Morris ER, Ellis R, Steele P & Moser PB (1988): Mineral balance of adult men consuming whole or dephytinized wheat bran. *Nutr. Res.* **8**, 445–458.
- Morton MS, Elwood PC & Abernethy M (1976): Trace elements in water and congenital malformation of the central nervous system in South Wales. *Br. J. Prev. Soc. Med.* **30**, 36–39.
- Multhaup G (1997): Amyloid precursor protein, copper and Alzheimer's disease. *Biomed. Pharmacother.* **51**, 105–111.
- Multhaup G, Schlicksupp A, Hesse L, Behr D, Ruppert T, Masters CL & Beyreuther K (1996): The amyloid precursor protein of Alzheimer's disease in the reduction of copper (II) to copper (I). *Science* **271**, 1406–1409.
- Multhaup G, Ruppert T, Schlicksupp A, Hesse L, Behr D, Masters CL & Beyreuther K (1997): Reactive oxygen species and Alzheimer's disease. *Biochem. Pharmac.* **54**, 533–539.
- Multhaup G, Ruppert T, Schlicksupp A, Hesse L, Bill E, Pipkorn R, Masters CL & Beyreuther K (1998): Copper-binding amyloid precursor protein undergoes a site-specific fragmentation in the reduction of hydrogen peroxide. *Biochemistry* **37**, 7224–7230.
- Multhaup G, Hesse L, Borchardt T, Ruppert T, Cappai R, Masters CL & Beyreuther K (1999): Autooxidation of amyloid precursor protein and formation of reactive oxygen species. *Adv. Exp. Med. Biol.* **448**, 183–192.
- Nagano K, Nakamura K, Urakami KL, Uneyama H, Koiwai K, Hattori S, Yamamoto T, Matsuda I & Endo F (1998): Intracellular distribution of the Wilson's disease gene product (ATPase 7B) after in vitro and in vivo exogenous expression in hepatocytes from the LEC rat: an animal model of Wilson's disease. *Hepatology* **27**, 799–807.
- Nakamura K, Endo F, Ueno T, Awata H, Tanoue A & Matsuda I (1995): Excess copper and ceruloplasmin biosynthesis in long-term cultured hepatocytes from Long-Evans cinnamon (LEC) rat, a model of Wilson disease. *J. Biol. Chem.* **270**, 7656–7660.
- National Research Council (NRC) (1989): *Recommended Dietary Allowances*. Washington, DC: National Academy of Sciences.
- O'Donohue JW, Reid MA, Varghese A, Portmann B & Williams R (1993): Case report: Micronodular cirrhosis and acute liver failure due to chronic copper self-intoxication. *Eur. J. Gastroenterol. Hepatol.* **5**, 561–562.
- Okada A (1994): Copper deficiency with pancytopenia during total parenteral nutrition. *J. Parenter. Enteral Nutr.* **18**, 190–192.
- Olivares M, Pizarro F, Speisky H, Lönnerdal B & Uauy R (1998): Copper in infant nutrition: Safety of World Health Organization Provisional Guideline Value for copper content of drinking water. *J. Pediatr. Gastroenterol. Nutr.* **26**, 251–257.
- Pattison IH, Clarke MC, Haig DA & Jebbitt JN (1971): Brain cell cultures from mice affected with scrapie or fed with cuprizone. *Res. Vet. Sci.* **12**, 478–480.
- Pauly PC & Harris DA (1998): Copper stimulates endocytosis of the prion protein. *J. Biol. Chem.* **273**, 33107–33110.
- Pedraza-Chaveri J, Torres-Rodriguez GA & Cruz C (1994): Copper and zinc metabolism in amino nucleoside-induced nephrotic syndrome. *Nephron* **92**, 87–92.
- Pena MM, Lee OJ & Thiele DJ (1999): A delicate balance: homeostatic control of copper uptake and distribution. *J. Nutr.* **129**, 1251–1260.
- Pettersson R & Rasmussen F (1999): Daily intake of copper from drinking water among young children in Sweden. *Environ. Health Persp.* **107**, 441–446.
- Quinlan GJ & Gutteridge JM (1988): Oxidative damage to DNA and deoxyribose by beta-lactam antibiotics in the presence of iron and copper salts. *Free Rad. Res. Commun.* **5**, 149–158.
- Raysiguer Y, Gueux E, Bussiere L & Mazur A (1993): Copper deficiency increases the susceptibility of lipoproteins and tissues to peroxidation in rats. *J. Nutr.* **123**, 1343–1348.
- Reaume AG, Elliott JL, Hoffmann EK, Kowall NW, Ferrante RJ, Siwek DF, Wilcox HM, Flood DG, Beal MF, Brown RH Jr, Scott RW & Snider WD (1996): Motor neurons in Cu/Zn superoxide dismutase-deficient mice develop normally but exhibit enhanced cell death after axonal injury. *Nature Genet.* **13**, 43–47.
- Reiser S, Smith JC Jr, Mertz W, Holbrook JT, Powell AS, Canfield WK & Canary JJ (1985): Indices of copper status in humans consuming a typical American diet containing either fructose or starch. *Am. J. Clin. Nutr.* **42**, 242–251.

- Reiser K, McCormick RJ & RB Rucker (1992): Enzymatic and none-enzymatic cross-linking of collagen and elastin. *F.A.S.E.B. J.* **6**, 2439–2449.
- Rock E, Gueux E, Mazur A, Motta C & Rayssiguier Y (1995): Anemia in copper-deficient rats: role of alterations in erythrocyte membrane fluidity and oxidative damage. *Am. J. Physiol.* **269**, C1245–C1249.
- Rodriguez A, Soto G, Torres S, Venegas G & Castillo-Duran C (1985): Zinc and copper in hair and plasma of children with chronic diarrhea. *Acta Paediatr. Scand.* **74**, 770–774.
- Rosen DR, Siddique T, Patterson D, Figlewicz DA, Sapp P, Hentati A, Donaldson D, Goto J, O Regan JP, Deng HX, Rahmani Z, Krizus A, McKenna-Yasek D, Cayabyab A, Gaston SM, Berger R, Tanzi RE, Halperin JJ, Herzfeldt B, Van den Bergh R, Hung WY, Bird T, Deng G, Mulder DW, Smyth C, Laing NG, Soriano E, Pericak-Vance MA, Haines J, Rouleau GA, Gusella JS, Horovitz HR & Brown RH Jr (1993): Mutation in Cu/Zn superoxide dismutase gene are associated with familial amyotrophic lateral sclerosis. *Nature* **362**, 59–62.
- Saari JT (1992): Dietary copper deficiency and endothelium-dependent relaxation of rat aorta. *Proc. Soc. Exp. Biol. Med.* **200**, 19–24.
- Sable-Amplis R, Sicart R & Reynier B (1987): Apparent retention of copper, zinc and iron in hamster. Influence of a fruit-enriched diet. *Nutr. Rep. Int.* **35**, 811–818.
- Sachdev HPS, Mittal NK & Yadan HS (1989): Serum and rectal mucosal copper status in acute and chronic diarrhea. *J. Pediatr. Gastroenterol. Nutr.* **8**, 212–216.
- Salmenperä L, Siimes MA, Nantö V & Perheentupa J (1989): Copper supplementation: failure to increase plasma copper and coeruleo-plasmin concentrations in healthy infants. *Am. J. Clin. Nutr.* **50**, 843–847.
- Sann L, Rigal D, Galy G, Benvenu F & Bourgeois J (1980): Serum copper and zinc concentrations in premature and small for date infants. *Pediatr. Res.* **14**, 1041–1046.
- Schuschke DA (1997): Dietary copper in the physiology of the microcirculation. *J. Nutr.* **127**, 2274–2281.
- Schuschke DA, Reed MWR, Saari JT & Miller FN (1992): Copper deficiency alters vasodilation in white rat cremaster muscle microcirculation. *J. Nutr.* **122**, 1547–1552.
- Scientific Committee on Food (SCF) (1994): *Nährstoff- und Energiezufuhr in der Europäischen Gemeinschaft* (Stellungnahme vom 11. Dezember 1992). Ber. wiss. Lebensmittelausschusses, ed. Europäische Kommission, 31. Folge. Amt. f. amlt. Veröff. Eur. Gem., Luxemburg.
- Selkoe DJ (1998): The cell biology of beta-amyloid precursor protein and presenilin in Alzheimer's disease. *Trends Cell Biol.* **8**, 447–453.
- Sethi S, Grover S & Khodaskar MB (1993): Role of copper in Indian childhood cirrhosis. *Ann. Trop. Pediatr.* **13**, 3–6.
- Shakespeare PG (1982): Studies on the serum levels of iron, copper and zinc and urinary excretion of zinc after burn injury. *Burns Incl. Therm. Inj.* **8**, 358–364.
- Shaw JCL (1992): Copper deficiency in term and preterm infants. In *Nutritional Anemias*, ed. SJ Fomon & S. Zlotkin. pp 105–109. New York: Raven Press Nestle Nutrition Workshop Series, Vol 30.
- Shenkin A, Fraser WD, McLalland JD, Fell GS & Garden OJ (1987): Maintenance of vitamin and trace element status in intravenous nutrition using a complete nutritive mixture. *J. Parenter. Enter. Nutr.* **11**, 238–242.
- Shike M (1984): Copper in parenteral nutrition. *Bull. N.Y. Acad. Med.* **60**, 132–143.
- Shike M, Roulet M, Kurian R & Jeejeebhoy K (1981): Copper metabolism and requirements in total parenteral nutrition. *Gastroenterology* **81**, 290–297.
- Shulman RJ (1989): Zinc and copper balance studies in infants requiring total parenteral nutrition. *Am. J. Clin. Nutr.* **49**, 879–883.
- Simons M, Ikonen E, Tienari PJ, Cidarregui A, Monning U, Beyreuther K & Dotti CG (1995): Intracellular routing of human amyloid protein precursor: axonal delivery followed by transport to the dendrites. *J. Neurosci. Res.* **41**, 121–128.
- Simons M, de Strooper B, Multhaup G, Tienari PJ, Dotti CG & Beyreuther K (1996): Amyloidogenic processing of the human precursor protein in primary cultures of rat hippocampal neurons. *J. Neurosci.* **16**, 899–908.
- Spitalny KC, Brondum J, Vogt RL, Sargent HE & Kappel S (1984): Drinking-water-induced copper intoxication in a Vermont family. *Pediatrics* **74**, 1103–1106.
- Stöckel J, Safar J, Wallace AC, Cohen FE & Prusiner SB (1998): Prion protein selectively binds copper(II) ions. *Biochemistry* **37**, 7185–7193.
- Strain JJ (1994): Newer aspects of micronutrients in chronic disease: copper. *Proc. Nutr. Soc.* **53**, 583–598.
- Sukalski KA, LaBerge TP & Johnson WT (1997): In vivo oxidative modification of erythrocyte membrane proteins in copper deficiency. *Free Rad. Biol. Med.* **22**, 835–842.
- Tanner MS (1998): Role of copper in Indian childhood cirrhosis. *Am. J. Clin. Nutr.* **67**(Suppl), 1074S–1081S.
- Thornalley PJ & Vasak M (1985): Possible role for metallothionein in protection against radiation-induced oxidative stress. Kinetics and mechanism of its reaction with superoxide and hydroxyl radicals. *Biochim. Biophys. Acta* **827**, 36–44.
- Turnlund JR (1991): Bioavailability of dietary minerals to humans: the stable isotope approach. *Crit. Rev. Food Sci. Nutr.* **30**, 387–396.
- Turnlund JR, King JC, Gong B, Keyes WR & Michel MC (1985): A stable isotope study of copper absorption in young men: effect of phytate and cellulose. *Am. J. Clin. Nutr.* **42**, 18–23.
- Turnlund JR, Keen CL & Smith RG (1990): Copper status and urinary and salivary copper in young men at three levels of dietary copper. *Am. J. Clin. Nutr.* **51**, 658–664.
- Underwood EJ (1977): *Trace Elements in Human and Animal Nutrition*, 4th edn, pp 56–108. New York: Academic Press.
- Varada KR, Harper RG & Wapnir RA (1993): Development of copper intestinal absorption in the rat. *Biochem. Med. Metab. Biol.* **50**, 277–283.
- Vergara JD, Zietz B & Dunkelberg H (2000): Gesundheitliche Gefährdung ungestillter Säuglinge durch Kupfer in Haushalten mit kupfernen Trinkwasserleitungen. Erste Ergebnisse einer prospektiven Studie. *Bundesgesundheitsbl. Gesundheitsforsch. Gesundheitsschutz* **43**, 272–278.
- Wadsworth JD, Jackson GS, Hill AF & Collinge J (1999): Molecular biology of prion propagation. *Curr. Opin. Genet. Dev.* **9**, 338–345.
- Wang X, Manganaro F & Schipper HM (1995): A cellular stress model for the sequestration of redox-active glial iron in the ageing and degenerating nervous system. *J. Neurochem.* **64**, 1868–1877.
- Wapnir RA (1991): Copper-sodium linkage during intestinal absorption: inhibition by amiloride. *Proc. Soc. Exp. Biol. Med.* **196**, 410–414.
- Wapnir RA & Devas G (1995): Copper deficiency: interaction with high fructose and high fat diets in rats. *Am. J. Clin. Nutr.* **61**, 105–110.
- Weiss KC & Linder MC (1985): Copper transport in rats involving a new plasma protein. *Am. J. Physiol.* **249**, E77–E88.
- White AR, Reyes R, Mercer JFB, Camakaris J, Zheng H, Bush AI, Multhaup G, Beyreuther K, Masters CL & Cappai R (1999): Copper levels are increased in the cerebral cortex and liver of APP and APLP2 knockout mice. *Brain Res.* **842**, 439–444.
- WHO (1993): *WHO Guidelines for Drinking Water Quality*, 2nd edn, pp 6–7, Addendum to Vol 1 (Recommendations). Geneva: WHO.
- WHO (1996): *Trace Elements in Human Nutrition and Health*. Geneva: World Health Organization.
- WHO (1998): *Copper. Environmental Health Criteria 2000*. Geneva: World Health Organization.
- Widdowson EM, Dancy J & Shaw JLC (1974): Trace elements in foetal and early postnatal development. *Proc. Nutr. Soc.* **33**, 275–284.
- Wiedau-Pazos M, Goto JJ, Rabizadeh S, Gralla EB, Roe JA, Lee MK, Valentine JS & Bredesen DE (1996): Altered reactivity of superoxide dismutase in familial amyotrophic lateral sclerosis. *Science* **271**, 515–518.

Williams DR (1983): Historical outline of the biological importance of trace metals. *J. Inherit. Metab. Dis.* 6(Suppl 1), 1–4.  
Williamson TL & Cleveland DW (1999): Slowing of axonal transport is a very early event in the toxicity of ALS-linked SOD1 mutants to motor neurons. *Nature Neurosci.* 2, 50–56.

Yount NY, Mc Namara DJ, Al-Othman AA & Lei KY (1990): The effect of copper deficiency on rat hepatic 3-hydroxy-3-methylglutaryl co-enzyme A reductase activity. *J. Nutr. Biochem.* 1, 27–33.