1 Supplemental Material:

Virion encapsidated HIV-1 Vpr induces NFAT to prime non-activated T cells for
 productive infection.

Kristin Höhne^{1,2}, Ramona Businger⁴, Anouk van Nuffel³, Sebastian Bolduan¹,
Herwig Koppensteiner¹, Ann Baeyens³, Jolien Vermeire³, Eva Malatinkova⁵, Bruno
Verhasselt^{3,#}, Michael Schindler^{1,2,4,#}

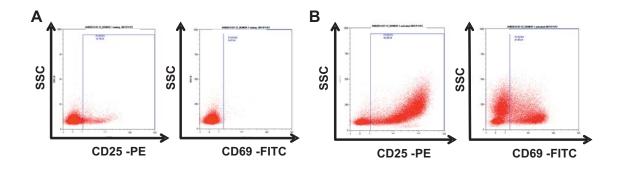
- 7
- 8 ¹Institute of Virology, Helmholtz Zentrum München, German Research Center for
- 9 Environmental Health, Neuherberg, Germany
- ²Heinrich Pette Institute, Leibniz Institute for Experimental Virology, Hamburg, Germany
- ³Department of Clinical Chemistry, Microbiology and Immunology, Ghent University,
- 12 Ghent, Belgium
- 13 ⁴Institute of Medical Virology and Epidemiology of Viral Diseases, University Hospital
- 14 Tübingen, Tübingen, Germany
- 15 ⁵HIV Translational Research Unit, Department of Internal Medicine, Ghent University,
- 16 Ghent, Belgium
- 17
- 18 # Corresponding authors:
- 19 Michael Schindler, <u>michael.schindler@med.uni-tuebingen.de</u>
- 20 Bruno Verhasselt, bruno.verhasselt@ugent.be

21

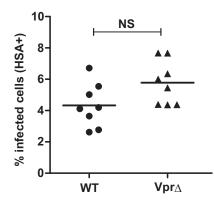
22 Supplemental Figures S1-S5

23

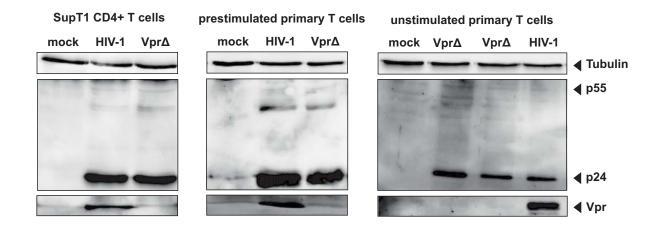
24



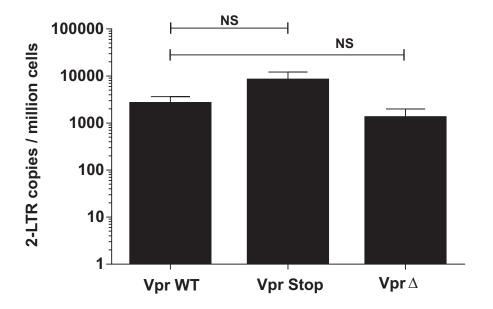
Supplemental Figure 1: Analyses of T cell activation markers in cultured CD4+ T cells. Presence of activation markers CD25 and CD69 were assessed by cell surface antibody FACS staining after 3 days of CD4+ T cell culture in IL-2 alone (A) or in IL-2 and PHA (B).



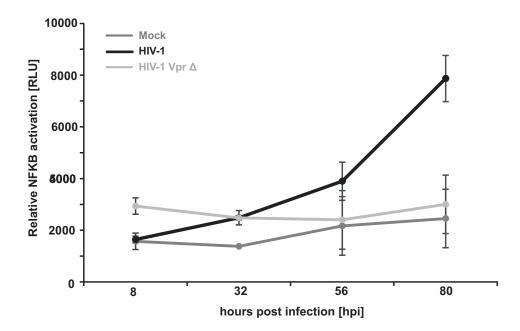
Supplemental Figure 2: Productive WT and Vpr Δ HIV-1 infection of primary prestimulated CD4+ T cells. Primary CD4+ T cells were stimulated with PHA/IL-2 for 3 days and subsequently infected with equal p24 amounts of HIV-1 WT or the Vpr Δ variant. Another 3 days later the amount of infected cells was determined by HSA staining and flow cytometry. Results show the mean of at least 8 independent experiments with CD4+ T cells from different donors and independent virus stocks.



Supplemental Figure 3: Vpr does not affect the ability of HIV-1 to enter T cells as measured by capsid delivery. Western blots show staining for tubulin, p24 (Gag) and Vpr as indicated, on the cell line SupT1, peripheral blood CD4+ T cells (PHA/IL2 stimulated for 3 days; prestimulated primary T cells) or left untreated (unstimulated primary T cells), infected as described in methods. Per cell type, all were infected with equal amounts of virus, measured by p24 antigen concentration. Mock: uninfected, HIV: pBR NL4-3; Δ Vpr: pBR NL4-3 Δ Vpr. For unstimulated primary T cells left Δ Vpr lane: pBR NL4-3 Δ Vpr and right Δ Vpr lane: NL4-3-IRES-HSA Δ Vpr. 24 hours after infection, cells were washed and lysates were prepared for WB analysis. Representative experiments are shown.



Supplemental Figure 4: Vpr's effect on productive infection in primary CD4+ T cells is not the result of defective nuclear import. Primary CD4+ T cells were infected as before (compare Fig. 1A). 3 days post-infection cells were collected and DNA was extracted. 2-LTR circles were quantified by qPCR. Results show the mean of 4 independent experiments carried out on 4 individual donors. No statistical difference was found (Kruskal-Wallis test with Dunn's correction).



Supplemental Figure 5: Vpr stimulates NFKB at a late time point in the HIV-1 replication cycle. 293 cells expressing luciferase under control of the NFKB promoter were infected with HIV-1 similar to the experiment described in Fig. 2A and stimulated with 10 ng TNF α eight hours before each measurement. The experiment shows the mean and SD of three independent infections.