SUPPLEMENTARY INFORMATION

Supplementary Table 1a | Whole genome sequence cohort information

Supplementary Table 1b | Whole genome sequence sample characteristics

Supplementary Table 2 | Single-variant T2D association analysis descriptions

LRT Likelihood ratio test, GEE Generalized estimating equations.

*Indicator function to account for observed temporal stratification based on sequencing date and center.

Supplementary Table 3a | Imputed cohort information

Supplementary Table 3b | Imputed cohort sample characteristics

Supplementary Table 4a | Exome sequence cohort information

Supplementary Table 4b | Exome sequence sample characteristics

Supplementary Figure 5 | Quality control of 12,940 WES samples. To assess the sequencing quality of each sample, we computed multiple statistics stratified by sample ethnicity. We then identified outlier samples relative to any of the statistical distributions and excluded them from further analysis. Shown are the distributions for nine representative statistics after samples were removed from analysis; note that these metrics are computed prior to any variant quality control and thus measure different statistics than presented in other display items. Number of variants: the number of variants (biallelic or multiallelic SNPs and INDELs) at which the sample exome carries a minor allele. Number of biallelic SNPs: the number of biallelic SNPs at which the sample exome carries a minor allele. Number of biallelic indels: the number of biallelic INDELs at which the sample exome carries a minor allele. Number of singletons: the number of variants carried by the sample alone (e.g., at which all other samples have the reference genotype). Heterozygosity: the heterozygosity of the sample computed across all variant sites. Heterozygosity at low frequency variants: the heterozygosity of the sample computed across low-frequency (MAF < 1%) variant sites. Heterozygous:Homozygous ratio: the ratio of heterozygous non-reference alleles to homozygous non-reference alleles in the sample. Mean allele balance: the fraction of sequence reads containing the non-reference allele, averaged over all heterozygous sites in the sample. Mean allele balance at singleton sites: the fraction of sequence reads containing the nonreference allele, averaged over all singleton heterozygous sites in the sample.

Supplementary Figure 6 | **Quality control of INDELs.** To assess the quality of called INDEL variants, we computed two metrics. (a) The number of INDELs with size equal to x (mod 3), for various values of x. Negative values represent deletions, while positive values represent insertions. As frameshift variants are more likely to disrupt protein sequence than in-frame deletions, spikes at increments of three are expected for INDEL variants in the population. (bc) Principal component analysis of the 12,940 European samples, computed using common (MAF > 1%) (b) SNPs and INDELs and (c) INDELs only. If the majority of common SNPs and INDELs are of high quality, the principal components should be concordant between the two analyses.

 -0.01 $C₁$

 0.00

 0.01

 0.02

 -0.04

 -0.03

 -0.02

 $\mathbf b$

Principal component analysis (SNPs and INDELs)

* Protein truncating

Supplementary Figure 8 | Single variant analyses for exome sequence and combined data sets. QQ plots for each of the three minor allele frequency Supplementary Figure 8 | Single variant analyses for exome sequence and combined data sets. QQ plots for each of the three minor allele frequency N=12,940); (c) all exome array data ("Exome-array", N=79,854); and (d) exome array data combined with exome sequence data ("Combined",N=92,794). N=12,940); (c) all exome array data ("Exome-array", N=79,854); and (d) exome array data combined with exome sequence data ("Combined", N=92,794). categories (common, low-frequency, and rare) for (a) each of the five major ancestry groups included in the exome sequencing study (African American N=2,074; East Asian N=2,165; European N=4,541; Hispanic N=1,943; South Asian N=2,217); (b) the combined exome sequencing results ("13K Meta", categories (common, low-frequency, and rare) for (a) each of the five major ancestry groups included in the exome sequencing study (African American N=2,074; East Asian N=2,165; European N=4,541; Hispanic N=1,943; South Asian N=2,217); (b) the combined exome sequencing results ("13K Meta", The grey region on each plot represents the (analytically estimated) 95% confidence interval. The grey region on each plot represents the (analytically estimated) 95% confidence interval.

Supplementary Table 9A | Distribution of mean age-of-diagnosis by *PAX4* **Arg192His (rs2233580) genotypes in replication studies.**

 $\hat{\beta}$: Regression coefficient estimates. SE: standard error
^a Linear regression *p-value* of age-of- diagnosis with Arg192His

Supplementary Table 9B | Single-variant T2D association analysis of *PAX4* **Arg192His (rs2233580) in each ancestry group from exome-sequence analysis and replication.**

Supplementary Table 9C | Study information

Supplementary Table 9D | Sample characteristics

Supplementary Table 10 | Summary of biological knowledge for genes described in the paper

Supplementary Table 11a | Gene-level mask descriptions. Protein truncating (PTV) and missense variants were further annotated to identify variants predicted deleterious by at least one (NS_{broad}) or each of five (NS_{strict}) prediction algorithms (LRT, Mutation Taster, PolyPhen2-HumDiv, PolyPhen2-HumVar, SIFT). PTV, missense, NS_{strict}, and NS_{broad} classes of variants were combined to generate four masks for gene-level testing (described in first column). The second column lists variant categories (and variant counts) contributing to each mask; the third column indicates the total numbers of variants in each mask; the fourth column indicates the number of genes containing at least one variant meeting mask criteria; the final column indicates the median number (and range) of variants per gene for each mask.

Supplementary Table 11b | Numbers of variants for each mask in 12,940 WES samples.

Supplementary Figure 12 | Manhattan plots for gene-level analysis in 12,940 WES samples

a. Mask 1 - PTV-only

Chromosome

N=84,395). The grey region on each plot represents the (analytically estimated) 95% confidence interval. Across all analyses, there is no compelling N=84,395). The grey region on each plot represents the (analytically estimated) 95% confidence interval. Across all analyses, there is no compelling Supplementary Figure 13 | Aggregate (gene-based) analyses for exome sequence and combined data setQQ plots for each of the four variant **Supplementary Figure 13 | Aggregate (gene-based) analyses for exome sequence and combined data sets.** Q plots for each of the four variant N=4,541; Hispanic N=1,943; South Asian N=2,217); (b) the combined exome sequencing results ("13K Meta", N=12,940); (c) all exome chip data N=4,541; Hispanic N=1,943; South Asian N=2,217); (b) the combined exome sequencing results ("13K Meta", N=12,940); (c) all exome chip data masks for (a) each of the five major ancestry groups in the exome sequencing study (African American N=2,074; East Asian N=2,165; European masks for (a) each of the five major ancestry groups in the exome sequencing study (African American N=2,074; East Asian N=2,165; European ("Exome array", N=79,854); and (d) exome chip data combined with exome sequence data from Europeans only ("Combined European", ("Exome array", N=79,854); and (d) exome chip data combined with exome sequence data from Europeans only ("Combined European", evidence that results depart from the null. evidence that results depart from the null. **Supplementary Figure 14A | Single variant analyses in GWAS regions.** The QQ plots display single variant analyses for all variants (left) and nonsynonymous variants only (right). Analyses of exome sequence (6,504 cases; 6,436 controls) are in the upper panels, and of the combination of exome sequence and exome array data (34,809 cases, 57,985 controls) in the lower. In each panel, variants mapping to established GWAS regions are in pink, and all other variants in blue (only variants with a minor allele count over 9 are included). The plots show enrichment of association signals for coding variants in established GWAS signals resulting from a combination of linkage disequilibrium to known common variant GWAS signals, and secondary signals at a subset of loci (eg *HNF4A, THADA, TSPAN8*).

Supplementary Table 14B | *FES* **gene-level association statistics for all ancestry groups (African American N=2,074; East Asian N=2,165; European N=4,541; Hispanic N=1,943; South Asian N=2,217; Total N=12,940) for Mask 4 (PTV + NSbroad***).*

Supplementary Table 15a | Exome array cohort information

Supplementary Table 15b | Exome array sample characteristics

Supplementary Figure 16 | **Overlap of variants detected in 12,940 trans-ethnic exomes and genotyped on exome array in 79,854 Europeans.** Each blue bar indicates the number of coding SNVs, protein-altering SNVs (nonsense, essential splice site, and missense variants), or MAF > 0.5% protein-altering variants observed in 12,940 sequenced samples, broken down by ancestry (African American N=2,074; East Asian N=2,165; European N=4,541; Hispanic N=1,943; South Asian N=2,217). Red bars indicate the numbers of sequence variants that were observed in 79,854 European exome array samples. Exact counts are shown in the table on the right. While a small fraction of all coding variants are represented on exome array, 81.6% of European protein-altering variants with MAF > 0.5% are captured using the array.

Supplementary Figure 17 | **Unconditional regional association plots for coding variants from GoT2D consortium data (N=2,657).** Each plot shows the p-value (on a -log₁₀ scale) as a function of genomic position (NCBI Build 37) covering a 2-Mb window around the novel exome-wide significant coding variant (indicated by the purple symbol). The color-coding of all other SNPs indicates LD with the novel coding SNP estimated from GoT2D data: red, r2 ≥ 0.8; gold, 0.6 ≤ r2 < 0.8; green, 0.4 ≤ r2 < 0.6; cyan, 0.2 ≤ r2 < 0.4; blue, r2 < 0.2; gray, r2 unknown. Recombination rates are estimated from Phase II HapMap, and gene annotations are taken from the UCSC genome browser. Imputation quality was modest for rs60980157 (GSPM1) and rs9379084 (RREB1) (both r²=0.84) and high for all other novel coding SNPs $(r^2>0.99)$.

GCKR (rs1260326)

COBLL1 (rs7607980)

PPARG (rs1801282) 10 100 $\overline{8}$ -log₁₀(p-value) 1282 6 $\overline{4}$ 40 \circ $-$ NUP210 $SYN2 \rightarrow$ $PPARG \rightarrow MKRN2 \rightarrow$ $CAND2 \rightarrow$ $ATG7 \rightarrow$ $- TAMMA1$ $TSEN2 \rightarrow TMEM40$ $+1QSEC1$ \leftarrow VGLL4 $+$ TIMP $-MKRN2OS$ $RPL32$ $+$ BAF1 $+$ SNORAZA 11.5 12 12.5 $\frac{1}{3}$ Position on chr3 (Mb)

WFS1 (rs1801212)

WFS1 (rs734312)

PPIP5K2 (rs36046591)

RREB1 (rs9379084) 10 100 $\overline{\mathbf{8}}$ $-log_{10}(p-value)$ $\ddot{}$ 60 0.2 $\overline{4}$ rs9379 \overline{a} \circ $\frac{RIOK1 \rightarrow DSP \rightarrow}{ENI}$
SNRNP48-> $\begin{array}{ccc}\n\hline\nBMP6 \rightarrow & \leftarrow & BLOC 1S5 \\
\hline\n\end{array}$ $F13A1$ $RREBI \rightarrow$ LY86- $+196-481$ $-$ ssai $+ 7XMOC5$ $+ BLOC 1S5-TXNDC5$ $+$ CAGE1 $PIPSK1PI -EEF1E1-BLOC1S5$ $-EEF1E1$ \leftarrow SCARNA27 6.5 $\overline{7.5}$ $\overline{8}$ Position on chr6 (Mb)

GPSM1 (rs60980157)

ASCC2 (rs28265)

Combined exomes p-values are derived from the meta-analysis of sequence and array datasets, with total sample size up to 92,794 (34,809 cases, 57,985 controls: effective sample size 82,758); smaller sample sizes reflect the fact that many variants were monomorphic in some or all of the non-European sequence cohorts. Exome-array analysis was performed in up to 79,854 samples (28,305 cases, 51,549 controls: effective sample size 69,866). Previously reported p-values from European meta-analysis are taken from Morris et al. (2012) Large-scale association analysis provides insights into the genetic architecture and pathophysiology of type 2 diabetes. *Nat Genet*. 44(9):981-90.

Supplementary Table 19 | Association summary statistics for T2D and fasting glucose levels from exome-array and exome sequence for selected *RREB1* **coding variants.**

N_{eff}: effective sample size. EAF: effect allele frequency. OR: odds-ratio. CI: confidence interval. I²: heterogeneity measure in %. p_het: p-value for Cochran's Q statistic. N: number of individuals analysed. $\hat{\beta}$: regression coefficient estimates. SE: standard error.

Summary statistics of just the two coding variants showing significant association signals for either T2D or fasting glucose have been summarized.

For Supplementary Table 20 see Excel File "20Supp20 - T2D loci and genes.xlsx"

Supplementary Table 21 | Characterization of role of coding variants within genes in established common variant GWAS regions through reciprocal conditional analysis.

Combined exomes p-values are derived from the meta-analysis of sequence and array datasets, with total sample size up to 92,794 (34,809 cases, 57,985 controls: effective sample size 82,758). Conditional analysis was perfor only on the exome-array component (28,305 cases, 51,549 controls: effective sample size 69,866). However, the previously reported non-coding GWAS SNP at the THADA locus (rs10203174) is not available on the exome array, pvalues reported here come from approximate conditional analyses undertaken in GCTA in a genome-wide imputed meta-analysis from the GoT2D Consortium (**METHODS**). We also examined genome-wide sequence and imputed data sets from the GoT2D consortium (N=2,657; **METHODS**) to determine whether these causal inferences were robust to more comprehensive coverage of regional variation**.**

Supplementary Table 22 | List of monogenic analysis categories and genes

Supplementary Figure 23 | Age of diagnosis of variant carriers. To assess whether individuals carrying variants in genes for monogenic forms of diabetes were enriched for patients with undiagnosed monogenic diseases, we examined the ages of diagnosis for carriers and compared them to those of non-carriers. As some diseases typically manifest at an earlier age than does T2D (e.g. MODY), a lower age of diagnosis for carriers might suggest that the monogenic phenotype, rather than late-onset T2D, is responsible for the diabetes phenotype in carriers. Shown is the mean age of diagnosis for carriers of variants in (a) the Monogenic All gene set, (b) the Monogenic Primary gene set, and (c) the Monogenic OMIM gene set. In each case, the mean ages are computed for carriers of variants in each of the five variant masks discussed in the text. Error bars indicate one standard deviation. Numbers of carriers and non-carriers for each mask are listed in parentheses at the bottom of the plot.

Monogenic All

Monogenic OMIM

Non − carriers (3190)

Non-carriers (3190)

 $PTV + NS_{Strict}$

PTV - only

H G M D

Supplementary Figure 24 | Accumulation of ultra-rare deleterious alleles amongst genes contributing to pre-specified "biologically-driven" gene-sets. a, Using the SMP approach, we confirmed association in the 'Monogenic All" gene set (81 genes: p= 0.006, OR=1.35 for singletons; p=0.04, OR=1.07 for ultra-rare alleles) and the "Monogenic OMIM" gene set (13 genes: p=0.0088, OR=2.4 for singletons; p=0.02, OR=1.82 for ultra-rare alleles). We also detected a separate "burden" signal for increased T2D-risk attributable to singleton alleles within the MTOR pathway (p=0.012, OR=3.61). **b**, Individual gene-ranking of composite set genes (set genes with *p*< .05 are shown). Genes are ranked by their case burden of rare PTVs, from top to bottom, for the Mtor and the monogenic all gene sets (labeled MTOR and MODY ALL, respectively). The squares along the bottom indicate to which sets each gene belongs. The red and blue triangles represent case and control counts for each gene. The lines represent the statistical significance of the best test for this set: that is, the significance of the top K genes, evaluated by permutation. For example, the drivers of the MTOR pathway signal include three case-only PTV singletons in both MNK1 and MTOR.

Supplementary Table 25 | All pathway enrichment signals with uncorrected FDR <= 10% from gene-set enrichment analyses conducted on the ancestry-specific and combined exome sequence data. Ancestries are denoted as European (EUR), East Asian (EA), South Asian (SA), Hispanic (HS) and African American (AA). For hand-curated gene-sets see **Supplementary Table 32**. We detected no study-wide significant signals (defined FDR < 5% after correction for multiple testing on four masks and five gene set collections). However, we detect nominal associations (uncorrected FDR <= 10%) in a subset of analyses, as listed below.

Supplementary Figure 26 | PPI analyses. A, QQ-plot of Fisher aggregated empirical *p*values ("PTV+NS_{strict}" mask) from the 2418 clusters generated by clusterONE based on 100,000 iterations. Cluster 630, consisting of ASB (ankyrin repeat and SOC box protein) protein family members interacting with RNF7 and CUL5, showed the strongest enrichment ("PTV+NS_{strict}" mask, empirical p-value P=5x10⁻⁵); **B**, Membership of the cluster 630 subnetwork highlighted by the clusterONE analyses. ASB6 is adipocyte-specific and interacts with APS to enable recruitment of elongins to the insulin receptor-signaling complex; **C**, PPI sub-network constructed using the top 15 modules generated with dmGWAS from genebased association *p*-values derived using the "PTV+NS_{strict}" variant mask. The sub-network includes the cluster of ASB proteins found in the clusterONE method as significant (cluster 630, shaded area), a cluster of mitochondrial-activity related genes, and the *PAM* gene; **D,** PPI sub-network built using the 15 top modules generated with dmGWAS from gene-based association *p*-values derived using the "PTV+NS_{broad}" mask. The sub-network includes *PAM* and *FES*, both of which contain exome-wide significant coding variants associated with T2D. The darkness of the node in the sub-networks is proportional to its *p*-value (lighter color indicates lower *p*-values) and the thickness of the edge is proportional to confidence score for the interaction between each pair of proteins.

Supplementary Figure 27 | Use of permutations to evaluate synthetic association hypothesis at 10 T2D GWAS loci.

Number of low frequency variants

Number of low frequency variants

Supplementary Table 28 | Properties of credible sets constructed at all T2D GWAS loci. Loci are sorted by the size of the final 99% credible set (from smallest to largest; column 5 below). Up to 2 credible sets were constructed for independent signals (r^2 <0.1) at all previously known autosomal T2D GWAS loci. Only loci where the index SNV had MAF>1% in the GoT2D sequencing data were included; *RBM43* and *SGCG* were excluded due to low index SNV MAF. *CILP2* was excluded due to poor sequencing quality across this region in the GoT2D experiment. At loci where the two independent signals have opposite directions of effect at the minor allele (risk, protective), the credible sets are labeled as such ("risk", "prot"); at loci where both signals are in the same direction, they are labeled "sig1" and "sig2".

Supplementary Figure 29 | **Trans-ethnic principal component analysis for exome-sequence samples.** African American studies (N=2,074): Jackson Heart Study (AJ) and Wake Forest School of Medicine Study (AW); East Asian studies (N=2,165): Korea Association Research Project (EK) and Singapore Diabetes Cohort Study and Singapore Prospective Study Program (ES); Hispanic studies (N=1,943): San Antonio Family Heart Study (HA) and Starr County (HS); South Asian studies (N=2,217): London Life Sciences Population Study (SL) and Singapore Indian Eye study (SS); and European studies (N=4,541): Ashkenazi (UA), Metabolic Syndrome in Men Study (UM), and GoT2D study (GO). A total of 10,348 independent QC passed, autosomal variants (trans-ethnic r^2 <0.05) with MAF>1% in all ancestry groups were considered for constructing axes of genetic variation through principal components analysis implemented in EIGENSTRAT to identify ethnic outliers.

Supplementary Table 30 | **Summary of samples and adjustments for EMMAX and WALD singlevariant association analysis for 12,940 exome-sequence samples**. Genomic control inflation factors (λ) were calculated on the basis of independent autosomal variants with MAF>1% within each ancestry group.

N: total number of samples. PC: principal component.

Supplementary Figure 31 | Global ancestry estimates for 1,943 Hispanic samples.

Supplementary Table 32 | Premium gene sets

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