Appendix for:

Blocking sense strand activity improves potency, safety and specificity of antihepatitis B virus short hairpin RNA

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Appendix Supplementary Discussion

As noted in the main text, we are careful when interpreting complex data on *in vivo* liver gene expression and when assigning dysregulated genes as direct shRNA off-targets, for four reasons. First, roughly half of our genes (even two thirds in (Maczuga *et al*, 2014)) were up- rather than down-regulated, yet only the latter is expected for a direct shRNA (off-)target. Second, although we found seed matches for the shRNA sense strand in over 80% of full transcripts of significantly down-regulated genes, the number was lower when we exclusively analysed the 3'UTR (12.2%; still higher than background or up-regulated genes, Appendix Table S2). Important to note here is that prior to our study, data on *in vivo* specificity of shRNAs were extremely sparse. Therefore, the rules that determine on- versus off-targeting in whole organisms are far from clear, as is the question which algorithms hold the highest predictive value (Birmingham *et al*, 2006; Maczuga *et al*, 2013). Our new data and vectors should help to shed light on the underlying mechanisms and thus foster the implementation of more specific next-generation RNAi therapeutics. Third, as noted, the degree of downregulation amongst all dysregulated genes with shHBV7 sense-strand seed match (irrespective of significance) was small. This was actually expected since we had aimed at pre-minimising toxicity, by using moderate AAV doses and the weak H1 promoter for shRNA expression in all our constructs. The overall mild degree of toxicity with no significant pathohistological changes seen in livers harvested 84 days post-AAV treatment (Fig 5E) was also confirmed in livers from day 15 post-AAV treatment using Hematoxylin/Eosin, ki67 (proliferation marker) or Caspase-3 (apoptosis marker) staining, none of which showed notable abnormalities (Fig EV4). Further noteworthy in

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this context is that shRNA off-targeting likely predominantly occurs via imperfect target binding beyond the seed region, which triggers miRNA-like inhibition on mRNA levels in a range of two-fold or less (congruent with our observations). As these subtle changes might be missed in DNA microarray analyses, future studies should include protein arrays to more comprehensively detect all direct and indirect off-targets.

Fourth and last, it is well possible that most dysregulated genes (especially those that were up-regulated) that we detected 15 days after vector administration were actually secondary or later hits, resulting from earlier perturbance of other genes and compensatory reactions of cellular networks. In addition, a meta-analysis of published RNAi experiments (Khan *et al*, 2009) suggests that shRNA-mediated RISC saturation and miRNA dysregulation can largely and widely alter gene expression profiles, including both up- and down-regulation as noted here. One line of evidence for this more general mechanism could be our pathway analysis in Appendix Table S4 which shows that many of the dysregulated genes are involved in steroid metabolism. The latter is regulated by miRNAs in the liver (Rottiers & Naar, 2012) and is altered upon interference with the predominant liver miRNA miR-122 (Esau *et al*, 2006). We have previously shown that both the expression and function of miR-122 are impaired in cells and mice that express high levels of shRNAs (Grimm *et al*, 2006), which could support this model. Further consistent is that genes involved in steroid metabolism were not dysregulated by the shRNA/TuD, shRNA/Ago2 or the miHBV7 vector, which may support a beneficial effect of all three strategies on RISC/miRNA activity. Finally, many other dysregulated genes that we detected in our mice control essential functions such as cell death/survival, cell morphology and cell cycle. Such hits were also observed by

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Maczuga and co-workers (Maczuga *et al*, 2014) and may generally reflect cellular toxicity rather than representing direct shRNA off-targets.

Appendix Supplementary Methods

Pathohistological analysis

Sections (2 μ m) of livers (fixed in 4% paraformaldehyde and paraffin-embedded) were stained with Hematoxylin/Eosin or anti-ki67 (NeoMarkers/Thermo Scientific, Waltham, Massachusetts, USA; RM-9106-S1; rabbit anti-mouse antibodies; retrieval at 95°C with EDTA for 30 min; 1:200 solution) or anti-Cleaved Caspase 3 (Cell Signaling, Leiden, The Netherlands; #9661; rabbit anti-mouse; retrieval at 100°C for 20 min with EDTA; 1:300 solution) antibodies (Wolf *et al*, 2014). Incubation in Ventana buffer and staining was performed on a NEXES immunohistochemistry robot (Ventana Instruments, Tucson, Arizona, USA) using an IVIEW DAB Detection Kit (Ventana) or on a Bond MAX (Leica, Wetzlar, Germany). For analysis, slides were scanned using a SCN 400 slide scanner (Leica).

PCR

For analysis of HBV RNA from liver lysate, RNA was extracted from 30 mg liver tissue with the RNeasy mini kit (Qiagen) and cDNA synthesized with the Superscript III kit (Thermo Fisher Scientific). HBV transcripts were amplified with primers specific for only the 3.5 kb transcripts, or with primers binding to the common 3´ end of all HBV transcripts (Yan *et al*, 2012). Results were normalised to two housekeeping genes (Hypoxanthine-guanine phosphoribosyltransferase (HPRT) and cyclophylin). To analyse

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viremia in HBV-transgenic mice, DNA was extracted from 20 µl serum with the High

Pure Viral Nucleic Acid Kit (Roche Diagnostics) and amplified with primers HBV 1844

and HBV 1745. All PCRs were performed on a LightCycler 480 (Roche Diagnostics)

using the primers and PCR conditions shown in Appendix Table S5.

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Appendix Figure S1 - Location and conservation of selected shHBV7 binding site.

- A Schematic HBV genome. Grey arrows in the center indicate open reading frames. Blue lines denote the partially doublestranded DNA genome. Thin outer lines symbolise transcripts, with arrows showing transcriptional start sites. The white box represents the RNA encapsidation signal.
- B Alignment of all HBV transcripts and indication of shHBV7 target region.
- C Alignment of shHBV7 with all HBV genotypes. The shRNA seed region, *i.e.*, nt 1 to 8 from the 5' end, is highlighted in capital letters. Note how this seed region, whose binding to the target site is most important for shRNA specificity and activity, is 100% conserved across all HBV genotypes. The bar on the bottom left shows the p-distance.

Appendix Figure S2 - **Structure of the dual AAV vector (center).**

Shown on top is the U6 promoter-driven TuD with two perfect binding sites (blue) for the shHBV7 sense strand. Depicted at the bottom is the H1 promoter-driven anti-HBV shHBV7 (sense strand in orange, antisense strand in blue).

Appendix Table S1 - Number of biological replicates (n) and exact p values. vs., versus.

Appendix Table S2 - **Frequency of shHBV7 2-7 nt seed matches in whole transcripts or 3'UTRs of dysregulated genes, and background frequency.**

Shown are data for all genes represented on the Affymetrix Mouse Gene 2.0 ST Array. Dysregulated genes in shHBV7-treated animals (compared to mock; adjusted p<0.25) were identified as explained in Methods. Absolute numbers of genes with at least one 2-7 nt seed match for the shHBV7 sense or antisense strand are shown with frequencies across all genes that could be analysed in brackets.

Appendix Table S3 - **List of dysregulated genes in shHBV7-treated animals.**

Shown are significantly dysregulated genes in the respective treatment group compared to mock-treated controls. p-values and adjusted p-values (false discovery rate) were controlled by Benjamini-Hochberg and defined as described in Methods. The four columns on the right show the number of shHBV7 sense- or antisense-strand seed matches in the 3'UTR or the whole transcript of the respective gene. NA, non-applicable (because the RNA is non-coding or the sequence could not be retrieved).

Appendix Table S4 - **List of dysregulated pathways in shHBV7-treated animals.**

Pathway enrichment of genes significantly dysregulated in shHBV7-treated animals compared to the empty control vector was obtained by querying pathway annotations using the R-package RDAVIDWebService as described in Methods.

Appendix Table S5 - **List of oligonucleotides including PCR conditions used in this study.** examplies Table SE . List of oliganual satisfaction including PCP conditions used in this study.

BHQ1, Black Hole Quencher 1; FAM, Carboxyfluorescein; Fwd/Fw, forward; rev/Rev, reverse; B2M, Beta-2-microglobulin; mHPRT, murine Hypoxanthine-guanine phosphoribosyltransferase.