Research Article

Susceptibility loci of CNOT6 in the general mRNA degradation pathway and lung cancer risk - a re-analysis of eight $GWASs^{\dagger}$

Fei Zhou^{1,2,3,4,5*}, Yanru Wang^{1,2*}, Hongliang Liu^{1,2}, Neal Ready^{1,2}, Younghun Han⁶, Rayjean J.

Hung⁷, Yonathan Brhane⁷, John McLaughlin⁸, Paul Brennan⁹, Heike Bickeböller¹⁰, Albert

Rosenberger¹⁰, Richard S. Houlston¹¹, Neil Caporaso¹², Maria Teresa Landi¹², Irene Brüske¹³,

Angela Risch¹⁴, Yuanqing Ye¹⁵, Xifeng Wu¹⁵, David C. Christiani¹⁶, Gary Goodman^{17,18}, Chu

Chen¹⁷, Transdisciplinary Research in Cancer of the Lung (TRICL) Research Team, Christopher

I. Amos⁶, Wei Qingyi^{1,2**}

¹Duke Cancer Institute, Duke University Medical Center, Durham, NC 27710, USA.

²Department of Medicine, Duke University School of Medicine, Durham, NC 27710, USA.

³Department of Oncology, Shanghai General Hospital, Shanghai Jiaotong University School of Medicine, Shanghai, 200080, China.

⁴Cancer Institute, Collaborative Innovation Center for Cancer Medicine, Fudan University Shanghai Cancer Center, Shanghai, 200032, China.

[†]This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the Version of Record. Please cite this article as doi: [10.1002/mc.22585]

Additional Supporting Information may be found in the online version of this article.

Received 22 June 2016; Revised 28 September 2016; Accepted 28 October 2016

Molecular Carcinogenesis

This article is protected by copyright. All rights reserved

DOI 10.1002/mc.22585

⁵Department of Medical Oncology, Fudan University Shanghai Cancer Center, Department of Oncology, Shanghai Medical College, Fudan University, Shanghai 200032, China.

⁶Community and Family Medicine, Geisel School of Medicine, Dartmouth College, Hanover, NH 03755, USA.

⁷Lunenfeld-Tanenbaum Research Institute of Mount Sinai Hospital, Toronto, Ontario, Canada.

⁸Public Health Ontario, Toronto, Ontario M5T 3L9, Canada.

⁹Genetic Epidemiology Group, International Agency for Research on Cancer (IARC), 69372 Lyon, France.

¹⁰Department of Genetic Epidemiology, University Medical Center, Georg-August-University, Göttingen, 37073 Göttingen, Germany.

¹¹Division of Genetics and Epidemiology, the Institute of Cancer Research, London , SW7 3RP, UK.

¹²Division of Cancer Epidemiology and Genetics, National Cancer Institute, National Institutes of Health, Bethesda, MD 20892, USA.

¹³Helmholtz Centre Munich, German Research Centre for Environmental Health, Institute of Epidemiology I, 85764 Neuherberg, Germany.

¹⁴Department of Molecular Biology, University of Salzburg, 5020 Salzburg, Austria.

¹⁵Department of Epidemiology, The University of Texas MD Anderson Cancer Center, Houston, TX 77030, USA.

¹⁶Massachusetts General Hospital, Boston, MA 02114, USA, Department of Environmental Health, Harvard School of Public Health, Boston, MA 02115, USA.

¹⁷Division of Public Health Sciences, Fred Hutchinson Cancer Research Center, Seattle, WA 98109, USA.

¹⁸Swedish Cancer Institute, Seattle, WA 98104, USA

*Fei Zhou and Yanru Wang contributed equally to this work.

**Correspondence to: Qingyi Wei, M.D., Ph.D., Duke Cancer Institute, Duke University Medical Center, 905 S. LaSalle Street, Durham, NC 27710, USA, Tel.: 1-(919) 660-0562, E-mail: qingyi.wei@duke.edu

Acknowledgments

As Duke Cancer Institute members, QW, KO and NR acknowledge support from the Duke Cancer Institute as part of the P30 Cancer Center Support Grant (Grant ID: NIH CA014236). QW was also supported by the start-up funds from Duke Cancer Institute, Duke University Medical Center.

TRICL

This work was supported by the Transdisciplinary Research in Cancer of the Lung (TRICL) Study and, U19-CA148127 on behalf of the Genetic Associations and Mechanisms in Oncology (GAME-ON) Network. The Toronto study was supported by Canadian Cancer Society Research Institute (020214), Ontario Institute of Cancer and Cancer Care Ontario Chair Award to RH The ICR study was supported by Cancer Research UK (C1298/A8780 andC1298/A8362—Bobby Moore Fund for Cancer Research UK) and NCRN, HEAL and Sanofi-Aventis. Additional funding was obtained from NIH grants (5R01CA055769, 5R01CA127219, 5R01CA133996, and

5R01CA121197). The Liverpool Lung Project (LLP) was supported by The Roy Castle Lung Cancer Foundation, UK. The ICR and LLP studies made use of genotyping data from the Wellcome Trust Case Control Consortium 2 (WTCCC2); a full list of the investigators who contributed to the generation of the data is available from www.wtccc.org.uk. Sample collection for the Heidelberg lung cancer study was in part supported by a grant (70–2919) from the Deutsche Krebshilfe. The work was additionally supported by a Helmholtz-DAAD fellowship (A/07/97379 to MNT) and by the NIH (U19CA148127). The KORA Surveys were financed by the GSF, which is funded by the German Federal Ministry of Education, Science, Research and Technology and the State of Bavaria. The Lung Cancer in the Young study (LUCY) was funded in part by the National Genome Research Network (NGFN), the DFG (BI576/2-1; BI 576/2-2), the Helmholtzgemeinschaft (HGF) and the Federal office for Radiation Protection (BfS: STSch4454). Genotyping was performed in the Genome Analysis Center (GAC) of the Helmholtz Zentrum Muenchen. Support for the Central Europe, HUNT2/Tromsø and CARET genome-wide studies was provided by Institut National du Cancer, France. Support for the HUNT2/Tromsø genome-wide study was also provided by the European Community (Integrated Project DNA repair, LSHG-CT- 2005–512113), the Norwegian Cancer Association and the Functional Genomics Programme of Research Council of Norway. Support for the Central Europe study, Czech Republic, was also provided by the European Regional Development Fund and the State Budget of the Czech Republic (RECAMO, CZ.1.05/2.1.00/03.0101). Support for the CARET genome-wide study was also provided by grants from the US National Cancer Institute, NIH (R01 CA111703 and UO1 CA63673), and by funds from the Fred Hutchinson Cancer Research Center. Additional funding for study coordination, genotyping of replication studies and statistical analysis was provided by the US

National Cancer Institute (R01 CA092039). The lung cancer GWAS from Estonia was partly supported by a FP7 grant (REGPOT245536), by the Estonian Government (SF0180142s08), by EU RDF in the frame of Centre of Excellence in Genomics and Estoinian Research Infrastructure's Roadmap and by University of Tartu (SP1GVARENG). The work reported in this paper was partly undertaken during the tenure of a Postdoctoral Fellowship from the IARC (for MNT). The Environment and Genetics in Lung Cancer Etiology (EAGLE), the Alpha-Tocopherol, Beta-Carotene Cancer Prevention Study (ATBC), and the Prostate, Lung, Colon, Ovary Screening Trial (PLCO) studies and the genotyping of ATBC, the Cancer Prevention Study II Nutrition Cohort (CPS-II) and part of PLCO were supported by the Intramural Research Program of NIH, NCI, Division of Cancer Epidemiology and Genetics. ATBC was also supported by US Public Health Service contracts (N01-CN-45165, N01-RC-45035 and N01-RC-37004) from the NCI. PLCO was also supported by individual contracts from the NCI to the University of Colorado Denver (NO1-CN-25514), Georgetown University (NO1-CN-25522), Pacific Health Research Institute (NO1-CN-25515), Henry Ford Health System (NO1-CN-25512), University of Minnesota(NO1-CN-25513), Washington University(NO1-CN-25516), University of Pittsburgh (NO1-CN-25511), University of Utah (NO1-CN-25524), Marshfield Clinic Research Foundation (NO1-CN-25518), University of Alabama at Birmingham (NO1-CN-75022, Westat, Inc. NO1-CN-25476), University of California, Los Angeles (NO1-CN-25404). The Cancer Prevention Study II Nutrition Cohort was supported by the American Cancer Society. The NIH Genes, Environment and Health Initiative (GEI) partly funded DNA extraction and statistical analyses (HG-06-033-NCI-01 and RO1HL091172-01), genotyping at the Johns Hopkins University Center for Inherited Disease Research (U01HG004438 and NIH HHSN268200782096C) and study coordination at the GENEVA Coordination Center (U01

HG004446) for EAGLE and part of PLCO studies. Funding for the MD Anderson Cancer Study was provided by NIH grants (P50 CA70907, R01CA121197, R01CA127219, U19 CA148127, R01 CA55769, and K07CA160753) and CPRIT grant (RP100443). Genotyping services were provided by the Center for Inherited Disease Research (CIDR). CIDR is funded through a federal contract from the NIH to The Johns Hopkins University (HHSN268200782096C). The Harvard Lung Cancer Study was supported by the NIH (National Cancer Institute) grants CA092824, CA090578, and CA074386.

deCODE

The project was funded in part by GENADDICT: LSHMCT-2004-005166), the National Institutes of Health (R01-DA017932)

Abbreviations: AD, Adenocarcinoma; CI, confidence interval; eQTL, expression quantitative trait loci; FDR, false discovery rate; GWAS, genome-wide association study; ILCCO, International Lung Cancer Consortium; LD, linkage disequilibrium; OR, odds ratio; SC, squamous cell carcinoma; SNP, single nucleotide polymorphisms; TCGA, The Cancer Genome Atlas; TRICL, Transdisciplinary Research in Cancer of the Lung.

An abbreviated title: Susceptibility loci in the general mRNA degradation pathway and lung cancer risk

Key Words: lung cancer risk, pathway analysis, molecular epidemiology.

Abstract

Purpose: mRNA degradation is an important regulatory step for controlling gene expression and cell functions. Genetic abnormalities involved in mRNA degradation genes were found to be associated with cancer risk. Therefore, we systematically investigated the roles of genetic variants in the general mRNA degradation pathway in lung cancer risk.

Experimental design: Meta-analyses were conducted using summary data from six lung cancer genome-wide association studies (GWASs) from the Transdisciplinary Research in Cancer of the Lung and additional two GWASs from Harvard University and deCODE in the International Lung Cancer Consortium. Expression quantitative trait loci analysis (eQTL) was used for *in silico* functional validation of the identified significant susceptibility loci.

Results: This pathway-based analysis included 6,816 single nucleotide polymorphisms (SNP) in 68 genes in 14,463 lung cancer cases and 44,188 controls. In the single-locus analysis, we found that 20 SNPs were associated with lung cancer risk with a false discovery rate threshold of <0.05. Among the 11 newly identified SNPs in *CNOT6*, which were in high linkage disequilibrium, the rs2453176 with a RegulomDB score "1f" was chosen as the tagSNP for further analysis. We found that the rs2453176 T allele was significantly associated with lung cancer risk (odds ratio=1.11, 95% confidence interval=1.04-1.18) in the eight GWASs. In the eQTL analysis, we found that levels of *CNOT6* mRNA expression were significantly correlated with the rs2453176 T allele, which provided additional biological basis for the observed positive association.

Conclusion: The *CNOT6* rs2453176 SNP may be a new functional susceptible locus for lung cancer risk. This article is protected by copyright. All rights reserved

Introduction

Lung cancer is one of the most frequently diagnosed cancers with about 1.8 million new lung cancer cases reported in 2012 worldwide, accounting for about 13% of total cancer diagnoses [1]. In the United States, 224,390 new lung cancer cases are estimated to occur in 2016 [2]. In addition to other factors, such as occupational and environmental carcinogens, cigarette smoking is the major risk factor for lung cancer [3,4], but not all smokers develop lung cancer, which suggests that genetic predisposition play an essential role in the lung carcinogenesis [5]. In recent years, some genome-wide association studies (GWASs) of lung cancer have been conducted, and a number of genetic variants, i.e., single nucleotide polymorphisms (SNPs), have been found to be associated with lung cancer risk. For example, the significant susceptibility loci associated with lung cancer risk include 5p15.3 (rs401681, rs4975616 and rs402710 in CLPTM1L and rs2736100 in TERT) [6-11], 6p21.3 (rs3117582 in BAG6 or APOM and rs2395185 in *HLA-DRB5* or *HLA-DRB9*) [6,8,9,11], 6q22.1 (rs9387478 in *RAP1BP3* or *DCBLD1*) [11] and 15q25.1 (rs8034191 in *HYKK* and rs1051730 in *CHRNA3*) [6,8,9,12-15]. Among these SNPs, rs1051730, rs3117582 and rs2736100 were found to be specifically associated with risk of lung adenocarcinoma (AD) [9], whereas rs12296850 (mapped to 12q23.1) in SLC17A8 or NR1H4 was found to be a susceptibility locus for risk of squamous cell carcinoma (SC) [16]. Interestingly, the vast majority of the SNPs identified by GWASs are in introns or intergenic regions, and their functional evidence is limited. In the present study, we employed the pathway-based strategy that dramatically decreases the number of SNPs to be analyzed and thus significantly reduced multiple testing with the aim to identify possible lung cancer risk-associated functional SNPs that may have not been revealed by previous lung cancer GWASs.

The degradation of mRNA is an important regulatory step for controlling gene expression and cell functions [17]. The general cytoplasmic mRNA decay pathway usually begins with the deadenylation, which removes the poly(A) tail Ccr4-Not complex [18], followed by degradation of mRNA proceeding in two directions of 5'-3' or 3'-5'. The 5'-3' mRNA degradation initiates with decapping N⁷-methylguanosine (m⁷G) cap mainly by DCP1/DCP2 proteins and subsequently degraded by the exoribonuclease Xrn1, while the 3'-5' mRNA degradation is mainly catalyzed by 10-12 subunit exosome [19,20].

Some studies suggest that genetic abnormalities of genes involved in the general mRNA degradation pathway may be associated with lung cancer. For example, various genetic variants in *LSM2-LSM8*, which encode cofactors for mRNA decapping, were recently found in lung cancer cell lines [21]. Therefore, we hypothesize that genetic variants of the general mRNA degradation pathway are associated with lung cancer risk. To test the hypothesis, we conducted the comprehensive meta-analysis of the eight published lung cancer GWASs from the ILCCO (International Lung Cancer Consortium)-TRICL (Transdisciplinary Research in Cancer of the Lung) consortia, focusing on the SNPs of the genes in the general mRNA degradation pathway.

Materials and Methods

Study populations

The first part of the study populations came from the TRICL consortium, which included 12,160 lung cancer cases and 16,838 controls (all Europeans) of six previously published GWASs from: the MD Anderson Cancer Center (MDACC), the Institute of Cancer Research (ICR), the National Cancer Institute (NCI), the International Agency for Research on Cancer (IARC), Toronto study from Samuel Lunenfeld Research Institute study (Toronto), and the German Lung Cancer Study (GLC) [22]. The second part of the study populations included GWASs of

European ancestry from Harvard Lung Cancer Study (984 cases and 970 controls) [23] and Icelandic Lung Cancer Study (deCODE) (1,319 cases and 26,380 controls) [15] of the ILCCO. Written informed consents were achieved for all participants, and the present study was approved by each institutional review board of the participating institutions.

GWAS genotyping and imputation

Genotyping in the eight GWASs was performed by Illumina HumanHap 317, 317+240S, 370Duo, 550, 610 or 1M arrays. The imputation was conducted by IMPUTE2 v2.1.1 or MaCH v1.0 software using the reference panel from the 1000 Genomes Project (phase I integrated release 3, March 2012). Standard quality control on samples was performed on all scans in the analysis, excluding any participants with low call rate (< 90%), extremely high or low heterozygosity ($P < 1.0 \times 10^{-4}$), non-European (with the HapMap phase II CEU, JPT/CHB and YRI populations as a reference) and imputed SNPs with an information score < 0.40 in IMPUTE2 or $r^2 < 0.30$ in MaCH.

Gene and SNP selection

We first identified genes in the general mRNA degradation pathway from the Molecular Signatures Database [24] and the literature [18]. Overall, 75 genes located on autosomal chromosomes were selected, of which seven genes were pseudogenes or duplicates or withdrawn from updated NCBI. As a result, we then extracted genotype data of 68 genes (detailed in **Table 1**), including 2-kb of the flanking regions of each gene, from the GWAS datasets that also included those SNPs generated by imputation. The final meta-analysis contained 6,816 SNPs and covariates provided by the TRICL consortium in the summary data with the following standards:

genotyping rate \geq 90%, minor allele frequency \geq 1%, and Hardy Weinberg Equilibrium exact *P* value \geq 10⁻⁵. The overall workflow is shown in **Figure 1**.

In silico functional validation

Two *in silico* tools, SNPinfo (http://snpinfo.niehs.nih.gov/snpinfo/snpfunc.htm) [25], RegulomeDB (http://regulomedb.org/) [26], were used to predict potential functions. Expression quantitative trait loci (eQTL) analysis was performed by using the expression data of lymphoblastoid cell lines from 373 Europeans available in the 1000 Genomes Project (http://www.1000genomes.org/category/frequently-asked-questions/gene-expression) [27] and The Cancer Genome Atlas (TCGA) (https://tcga-data.nci.nih.gov/tcga/) [28]. In this TCGA dataset, 107 subjects had adjacent normal lung cancer samples used for the different expression testing, which were matched by 105 adjacent normal cancer tissue samples from the same individuals with the genotype data.

Statistical analysis

Logistic regression model was used to calculate the odds ratios (ORs) and their 95% confidence intervals (CIs) in an additive genetic model with PLINK (v1.06) software. A meta-analysis with the inverse variance method was employed on the 6,816 SNPs with Stata software (v12, State College, Texas, US). Cochran's Q statistic was applied to test for heterogeneity and the I^2 statistic for the proportion of the total variation in the meta-analysis [29]. The fixed-effects model was used when there was no heterogeneity among GWASs (Q-test P > 0.100 and $I^2 < 50\%$); otherwise, the random-effects model was used. Multiple testing correction was conducted with false discovery rate (FDR) with a threshold < 0.050 [30]. A linear regression model was also performed to evaluate the correlation between SNPs and mRNA expression levels of the

corresponding genes. A paired t-test was used to compare the mRNA expression levels of genes in the lung cancer and normal adjacent tissue from the TCGA database. LocusZoom (http://locuszoom.sph.umich.edu/locuszoom/) was applied to construct regional association plots using Europeans from the 1000 Genomes Project as the reference (phase I integrated release 3, March 2012) [31]. Haploview v4.2 was used to generate the Manhattan plot and LD plots [32]. All analyses were conducted with SAS (version 9.4; SAS Institute, Cary, NC, USA) except for those specified otherwise.

Results

Associations of the SNPs with lung cancer risk

We first performed a meta-analysis in the TRICL database consisted of six previously published GWAS datasets with 12,160 cases and 16,838 controls. The basic information of these six studies is presented in **Supplemental Table S1**. A total of 6,816 SNPs in the pathway were extracted, of which 466 SNPs were associated with lung cancer risk at P < 0.05 in the additive model and 20 SNPs on LSM2, SKIV2L and CNOT6 remained significantly associated with lung cancer risk with FDR < 0.05 after multiple testing corrections (**Figure 2A and Table 2**). Among these SNPs, we excluded those of LSM2 and SKIV2L, because they were mapped to and in high LD with previously GWAS-reported locus at 6p21.33 [6,8]. As a result, 11 SNPs of CNOT6 located at 5q35.3 were left for further analysis. In the LD analysis, these 11 SNPs shared moderate to high LD ($r^2 \ge 0.60$, **Figure 2B and 2C**). We finally chose rs2453176 as the tag SNP, because it was significantly associated with lung cancer risk (OR = 1.13, 95% CI = 1.06-1.19, $P = 4.33 \times 10^{-5}$) (**Table 2**) and potentially functional according to function prediction and its imputation quality was the best among the 11 SNPs (**Table 3**). We used the forest plot to illustrate the association between rs2453176 and lung cancer risk in the six GWASs (**Figure 3**),

and the rs2453176 T allele was associated with an increased lung cancer risk in five GWASs, except for the GLC GWAS.

We expanded our analysis to include additional two independent lung cancer GWASs (Supplemental Table S1). The deCODE GWAS validated our result of the *CNOT6* rs2453176 tag SNP (OR = 1.14, 95% CI = 1.01-1.28, P = 0.032), while the GWAS from Harvard University displayed the same trend as the GLC GWAS (OR = 0.85, 95% CI = 0.68-1.05, P = 0.133) (Figure 3 and Table 4).

As we combined the above results from the eight GWASs, the functional CNOT6 rs2453176 tag SNP was found to be significantly associated with an increased risk of lung cancer (OR = 1.11, 95% CI = 1.04-1.18, P = 0.001) after the FDR correction (**Figure 3 and Table 4**).

Stratified analyses by lung cancer histology

Since lung cancer has different histological types that could have distinct biological behaviors, we performed AD and SC subgroup analysis and found that the rs2453176 T allele was associated with a borderlinely increased risk in AD (OR = 1.13, 95% CI = 1.00-1.27, P = 0.050, **Table 4**), but it was significantly associated with SC risk (OR = 1.12, 95% CI = 1.03-1.22, P = 0.006, **Table 4**). Because smoking is a major risk factor for lung cancer, we further stratified the data into smokers and non-smokers and found that that the rs2453176 T allele was associated with a significantly increased risk in smokers (OR = 1.09, 95% CI = 1.02-1.17, P = 0.011, **Table 5**), while the allele was not statistically significant in non-smokers (OR = 1.10, 95% CI = 0.89-1.36, P = 0.363, **Table 5**). Homogeneity tests suggested that there was no heterogeneity between strata either in subgroups of histologic types or smoking status (**Table 4 and Table 5**, all P > 0.05).

Functional validation by eQTL analysis

Because the *CNOT6* rs2453176 SNP was predicted with a score of "1f", suggesting the most confident functional annotation by regulomeDB [26], we further explored the underlying molecular mechanism by performing the eQTL analysis. With mRNA expression data of lymphoblastoid cell lines from 373 Europeans available from the 1000 Genomes Project, We found that expected mRNA expression levels of *CNOT6* were significantly decreased with an increased number of the rs2453176 T allele in both the additive (P = 0.008) (**Figure 4A**) and dominant (P = 0.007) (**Figure 4B**) models but not the recessive model (**Figure 4C**). However, only 105 subjects had both DNA and RNA samples tested in this dataset. We also used the 105 normal adjacent tissue samples in the TCGA to further explore the correlation between the rs2453176 genotypes and their corresponding mRNA expression levels, but we did not observe a statistical significance (P > 0.05) (**Supplemental Figure S1A-S1C**). We also compared the mRNA expression level of *CNOT6* in the 107 paired samples and did not find a statistically significant difference (P > 0.05) (**Supplemental Figure S1D**).

Discussion

In the present study, we found that a novel potentially functional susceptibility locus rs2453176 C>T of *CNOT6* in the general mRNA degradation pathway was associated with an increased lung cancer risk in 14,463 cases and 44,188 controls. This association was further supported by a significant correlation between a decreased mRNA expression level and an increasing number of the A allele in the eQTL analysis.

Gene expression disorder is one of cancer hallmarks, and instability of mRNA may result in altered transcript/protein levels of oncogenes and tumor repressor genes [33]. The degradation of mRNA is a key step in controlling the expression of genes related to cell proliferation. For

example, the CCR4-Not complex consists of highly conserved exoribonucleases and adaptor proteins that hydrolyze and shorten the poly(A) tail, which starts the initial and the rate-limiting step of mRNA degradation [18,34-36]. Located at 5q35.3, *CNOT6* encodes a protein that has a 3'-5' RNase activity and acts as a catalytic subunit of the CCR4-Not deadenylation complex [37]. Although it remains unclear how the catalytic subunit works during the deadenylation process, some studies reported that its expression level was associated with carcinogenesis or prognosis. For example, one study of lung cancer found that the CNOT6 overexpression in lung SC predicted a significantly less metastasis [33]. Another study of acute leukemia discovered that CNOT6 had a significantly lower expression in patients than in controls [38]. These two studies suggest that high expression levels of CNOT6 may promote the degradation of mRNA of some oncogenes and the suppression of cell proliferation in carcinogenesis.

In the present study, we identified that the *CNOT6* rs2453176 T allele was associated with an increased risk of lung cancer, which was supported by the association of *CNOT6* rs2453176 T allele with a decreased mRNA expression level in lymphoblastoid cell lines from 373 Europeans. This finding is consistent with the role of CNOT6 in lung cancer prognosis as previously described [33]. The ENCODE project data from University of California Santa Cruz show that the *CNOT6* rs2453176 locus is located at the DNase I hypersensitive region (**Supplemental Figure S2**). Usually such an area has a loose chromatin structure and renders it a region with a high affinity for transcription factors (TFs). As a result, some TFs, including MAFK and MAFF, bond to this region in many cell types (**Supplemental Figure S2**). For example, MAFK and MAFF were found to form heterodimers with a series of TFs and suppressed gene transcriptions [39,40]. Based on these, we speculate that the rs2453176 T allele may have a relatively high affinity with MAFK or MAFF and thus leads to the decreased mRNA expression of *CNOT6*. It is

likely that a reduced quantity of CNOT6 may not be optimal in the mRNA degradation of some aberrant genes, which may in turn increases lung cancer risk, but these speculations need to be further investigated.

In the stratification analysis, rs2453176 was associated with lung cancer risk in both AD and SC subtypes, but it was significantly associated with cancer risk in the smokers but not in the non-smokers. Genetic susceptibility to smoking-related lung cancer risk may determine smoking behavior and tobacco metabolism [41]. Indeed, we found that the rs2453176 T allele was associated with a higher risk of lung cancer in smokers than in non-smokers. One study reported that smoking would enhance the activity of the GATA family [42], and another study reported that nicotine would increase the expression of EP300 and promote the lung cancer growth [43]. From the **Supplemental Figure S2**, GATA1, GATA2 and EP300 are the TFs that bind to the rs2453176 locus, possibly explaining why carriers of the rs2453176 T allele may have an increased risk of lung cancer in smokers than non-smokers.

There are some limitations in the present study. First, we employed the Molecular Signatures

Database [24] to define the general mRNA degradation pathway to be investigated, but we may
have missed some newly discovered genes in the pathway. However, we searched the literatures
and added genes as many as possible. Second, due to the data limitation, we had no access to
family history and others factors that may have an impact on lung cancer risk. Third, we used the
eQTL analyses from lymphoblastoid cell lines and normal adjacent tissue in TCGA database to
validate the risk association. Although the results from the cell lines support our identified
association, they may only reflect the baseline or genetically determined expression levels
without exposure to smoking. The gene expressions in the normal adjacent lung tissues may be
in some degree different from the normal lung tissue and did not support the association.

Overall, the present study of eight published GWASs identified a novel *CNOT6* rs2453176 SNP in the general mRNA degradation pathway to be significantly associated with lung cancer risk in European populations, and the risk was more evident in smokers than in non-smokers. Although we used the publically available gene expression database from blood to confirm the biological significance of the variant, further functional evaluations in normal lung tissue are warranted to validate our findings.

Conflict of interest:

The authors disclose no potential conflicts of interest.

U

References

- 1. Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. CA: a cancer journal for clinicians 2015;65(2):87-108.
- 2. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2016. CA: a cancer journal for clinicians 2016;66(1):7-30.
- 3. Field RW, Withers BL. Occupational and environmental causes of lung cancer. Clinics in chest medicine 2012;33(4):681-703.
- 4. Schottenfeld D, Fraumeni JF. Cancer epidemiology and prevention. Oxford; New York: Oxford University Press; 2006. xviii, 1392 p. p.
- 5. Sun S, Schiller JH, Gazdar AF. Lung cancer in never smokers--a different disease. Nature reviews Cancer 2007;7(10):778-790.
- 6. Wang Y, Broderick P, Webb E et al. Common 5p15.33 and 6p21.33 variants influence lung cancer risk. Nature genetics 2008;40(12):1407-1409.
- 7. McKay JD, Hung RJ, Gaborieau V et al. Lung cancer susceptibility locus at 5p15.33.

 Nature genetics 2008;40(12):1404-1406.
- 8. Broderick P, Wang Y, Vijayakrishnan J et al. Deciphering the impact of common genetic variation on lung cancer risk: a genome-wide association study. Cancer research 2009;69(16):6633-6641.
- 9. Landi MT, Chatterjee N, Yu K et al. A genome-wide association study of lung cancer identifies a region of chromosome 5p15 associated with risk for adenocarcinoma.

 American journal of human genetics 2009;85(5):679-691.

- 0
 - Hu Z, Wu C, Shi Y et al. A genome-wide association study identifies two new lung cancer susceptibility loci at 13q12.12 and 22q12.2 in Han Chinese. Nature genetics 2011;43(8):792-796.
 - 11. Lan Q, Hsiung CA, Matsuo K et al. Genome-wide association analysis identifies new lung cancer susceptibility loci in never-smoking women in Asia. Nature genetics 2012;44(12):1330-1335.
 - 12. Amos CI, Wu X, Broderick P et al. Genome-wide association scan of tag SNPs identifies a susceptibility locus for lung cancer at 15q25.1. Nature genetics 2008;40(5):616-622.
 - 13. Liu P, Vikis HG, Wang D et al. Familial aggregation of common sequence variants on 15q24-25.1 in lung cancer. Journal of the National Cancer Institute 2008;100(18):1326-1330.
 - Hung RJ, McKay JD, Gaborieau V et al. A susceptibility locus for lung cancer maps to nicotinic acetylcholine receptor subunit genes on 15q25. Nature 2008;452(7187):633-637.
 - Thorgeirsson TE, Geller F, Sulem P et al. A variant associated with nicotine dependence, lung cancer and peripheral arterial disease. Nature 2008;452(7187):638-642.
 - Dong J, Jin G, Wu C et al. Genome-wide association study identifies a novel susceptibility locus at 12q23.1 for lung squamous cell carcinoma in han chinese. PLoS genetics 2013;9(1):e1003190.
 - 17. Parker R, Song H. The enzymes and control of eukaryotic mRNA turnover. Nat Struct Mol Biol 2004;11(2):121-127.
 - 18. Balagopal V, Fluch L, Nissan T. Ways and means of eukaryotic mRNA decay.

 Biochimica et biophysica acta 2012;1819(6):593-603.

- 19. Siwaszek A, Ukleja M, Dziembowski A. Proteins involved in the degradation of cytoplasmic mRNA in the major eukaryotic model systems. RNA Biol 2014;11(9):1122-
- 20. Houseley J, Tollervey D. The many pathways of RNA degradation. Cell 2009;136(4):763-776.

1136.

- Young JH, Peyton M, Kim HS et al. Computational Discovery of Pathway-Level Genetic Vulnerabilities in Non-Small-Cell Lung Cancer. Bioinformatics 2016.
- Wang Y, McKay JD, Rafnar T et al. Rare variants of large effect in BRCA2 and CHEK2 affect risk of lung cancer. Nature genetics 2014;46(7):736-741.
- 23. Su L, Zhou W, Asomaning K et al. Genotypes and haplotypes of matrix metalloproteinase 1, 3 and 12 genes and the risk of lung cancer. Carcinogenesis 2006;27(5):1024-1029.
- 24. Liberzon A, Birger C, Thorvaldsdottir H, Ghandi M, Mesirov JP, Tamayo P. The Molecular Signatures Database (MSigDB) hallmark gene set collection. Cell systems 2015;1(6):417-425.
- 25. Xu ZL, Taylor JA. SNPinfo: integrating GWAS and candidate gene information into functional SNP selection for genetic association studies. Nucleic Acids Res 2009;37:W600-W605.
- Boyle AP, Hong EL, Hariharan M et al. Annotation of functional variation in personal genomes using RegulomeDB. Genome research 2012;22(9):1790-1797.
- 27. Lappalainen T, Sammeth M, Friedlander MR et al. Transcriptome and genome sequencing uncovers functional variation in humans. Nature 2013;501(7468):506-511.

- 28. Rodgers K, Network CGAR. Comprehensive molecular profiling of lung adenocarcinoma (vol 511, pg 543, 2014). Nature 2014;514(7521).
- 29. Higgins JP, Thompson SG, Deeks JJ, Altman DG. Measuring inconsistency in metaanalyses. BMJ 2003;327(7414):557-560.
- Benjamini Y, Hochberg Y. Controlling the False Discovery Rate a Practical and Powerful Approach to Multiple Testing. J Roy Stat Soc B Met 1995;57(1):289-300.
- 31. Pruim RJ, Welch RP, Sanna S et al. LocusZoom: regional visualization of genome-wide association scan results. Bioinformatics 2010;26(18):2336-2337.
- Barrett JC, Fry B, Maller J, Daly MJ. Haploview: analysis and visualization of LD and haplotype maps. Bioinformatics 2005;21(2):263-265.
- Maragozidis P, Papanastasi E, Scutelnic D et al. Poly(A)-specific ribonuclease and Nocturnin in squamous cell lung cancer: prognostic value and impact on gene expression.

 Molecular cancer 2015;14(1):187.
- 34. Collart MA, Panasenko OO. The Ccr4--not complex. Gene 2012;492(1):42-53.
- 35. Goldstrohm AC, Wickens M. Multifunctional deadenylase complexes diversify mRNA control. Nat Rev Mol Cell Biol 2008;9(4):337-344.
- Wahle E, Winkler GS. RNA decay machines: deadenylation by the Ccr4-not and Pan2-Pan3 complexes. Biochimica et biophysica acta 2013;1829(6-7):561-570.
- 37. Mittal S, Aslam A, Doidge R, Medica R, Winkler GS. The Ccr4a (CNOT6) and Ccr4b (CNOT6L) deadenylase subunits of the human Ccr4-Not complex contribute to the prevention of cell death and senescence. Molecular biology of the cell 2011;22(6):748-758.

38. Maragozidis P, Karangeli M, Labrou M et al. Alterations of deadenylase expression in acute leukemias: evidence for poly(a)-specific ribonuclease as a potential biomarker.

Acta haematologica 2012;128(1):39-46.

- 39. Kannan MB, Solovieva V, Blank V. The small MAF transcription factors MAFF, MAFG and MAFK: current knowledge and perspectives. Biochimica et biophysica acta 2012;1823(10):1841-1846.
- 40. Katsuoka F, Yamamoto M. Small Maf proteins (MafF, MafG, MafK): History, structure and function. Gene 2016.
- Shields PG. Molecular epidemiology of smoking and lung cancer. Oncogene 2002;21(45):6870-6876.
- 42. Zhao J, Harper R, Barchowsky A, Di YP. Identification of multiple MAPK-mediated transcription factors regulated by tobacco smoke in airway epithelial cells. American journal of physiology Lung cellular and molecular physiology 2007;293(2):L480-490.
- Dasgupta P, Rizwani W, Pillai S et al. ARRB1-mediated regulation of E2F target genes in nicotine-induced growth of lung tumors. Journal of the National Cancer Institute 2011;103(4):317-333.

Figure Legend

Figure 1. Study workflow SNP: single nucleotide polymorphism; FDR: false discovery rate; TRICL: Transdisciplinary Research in Cancer of the Lung; GWAS: genome-wide association study; eQTL: expression quantitative trait loci.

Figure 2. Screening of SNPs in the general mRNA degradation pathway. A, Manhattan Plot of genome-wide association results from the general mRNA degradation pathway in TRICL. The x-axis shows SNPs' positions on each chromosome. The y-axis shows the association P values with lung cancer risk (as $-\log 10 P$ values). The FDR threshold of 0.05 was shown by a horizontal blue line. The P value of 0.05 was shown by a horizontal red line. B, Regional association plot for SNP rs2453176 in 500 kb up- and downstream region. The left-hand y-axis shows P values of the SNPs, which are transformed as $-\log 10 (P)$ against chromosomal base pair positions. The right-hand y-axis shows the recombination rate estimated from HapMap Data Rel 22/phase II European population; C, The linkage disequilibrium plots of 11 SNPs in CNOT6. The value within each diamond represents the pairwise correlation between SNPs (measured as r^2) defined by the upper left and the upper right sides of the diamond.

Figure 3. Forest plots for associations between CNOT6 rs2453176 and lung cancer risk for all participants (P = 0.0013).

Figure 4. The eQTL analysis of *CNOT6* mRNA expression for rs2453176 with lymphoblastoid cell data of 373 Europeans from 1000 Genomes Project. A. additive model, P = 0.008; B. dominant model, P = 0.007; C. recessive model, P = 0.634.



Table 1 The mRNA degradation pathway gene sets

Dataset	Name of pathway	Gene number	Gene name					
KEGG*	KEGG_RNA_DEGRADATIONE	59	C1D, C1DP2**, C1DP3**, CNOT1, CNOT10, CNOT2, CNOT3,					
			CNOT4, CNOT6, CNOT6L, CNOT7, CNOT8, DCP1A, DCP1B, DCP2,					
			DCPS, DDX6, DIS3, EDC3, EDC4, ENO1, ENO2, ENO3, EXOSC1,					
			EXOSC10, EXOSC2, EXOSC3, EXOSC4, EXOSC5, EXOSC6, EXOSC7,					
			EXOSC8, EXOSC9, HSPA9, HSPD1, LSM1, LSM2, LSM3, LSM4, LSM5, LSM6, LSM7, MPHOSPH6, NAA38, PAPD7, PAPOLA,					
			PAPOLB, PAPOLG, PARN, PATL1, PNPT1, RQCD1, SKIV2L,					
			SKIV2L2, TTC37, WDR61, XRN1, XRN2, ZCCHC7.					
Reactome	REACTOME_DEADENYLATION_DEPENDENT_MRNA_DECAY	48	C2orf29**, CNOT10, CNOT2, CNOT3, CNOT4, CNOT6, CNOT7,					
			CNOT8, DCP1A, DCP1B, DCP2, DCPS, DDX6, DIS3, EDC3, EDC4,					
			EIF4A1, EIF4A2, EIF4A3, EIF4B, EIF4E, EIF4G1, EXOSC1,					
			EXOSC2, EXOSC3, EXOSC4, EXOSC5, EXOSC6, EXOSC7, EXOSC8,					
			EXOSC9, LOC645139**, LOC645947**, LOC651789**,					
			LOC652607**, LSM1, LSM2, LSM3, LSM4, LSM5, LSM6, PABPC1,					
	DEL CTOLE DEL DENNI L'EION DE MONT	22	PAIP1, PARN, PATL1, RQCD1, TNKS1BP1, XRN1.					
Reactome	REACTOME_DEADENYLATION_OF_MRNA	22	C2orf29**, CNOT10, CNOT2, CNOT3, CNOT4, CNOT6, CNOT7,					
			CNOT8, EIF4A1, EIF4A2, EIF4A3, EIF4B, EIF4E, EIF4G1, LOC645139**, LOC651789**, LOC652607**, PABPC1, PAIP1,					
			PARN, RQCD1, TNKS1BP1.					
Reactome	REACTOME MRNA DECAY BY 3 TO 5 EXORIBONUCLEASE	11	DCPS, DIS3, EXOSC1, EXOSC2, EXOSC3, EXOSC4, EXOSC5,					
			EXOSC6, EXOSC7, EXOSC8, EXOSC9,					
Reactome	REACTOME_MRNA_DECAY_BY_5_TO_3_EXORIBONUCLEASE	15	DCP1A, DCP1B, DCP2, DDX6, EDC3, EDC4, LOC645947, LSM1,					
			LSM2, LSM3, LSM4, LSM5, LSM6, PATL1, XRN1.					
PID*	NO DATA	0						
GO*	NO DATA	0						
BioCarta	NO DATA	0						
Literature		2	PAN2, PAN3					
Total		68***						

^{*}KEGG, Kyoto encyclopedia of genes and genomes; GO, gene ontology; PID, pathway interaction database;

Search keyword: mRNA degradation; Search Filters: Collection, canonical pathways + GO gene sets; Organism, Homo sapiens; Contributor, all contributors.

^{**}Pseudo gene: C1DP2, C1DP3, LOC645139; same gene with different name: C2orf29; withdrawn by updated NCBI: LOC645947, LOC651789, LOC652607.

^{***}After removing the duplicate genes and those genes mentioned in **;

Table 2 Associations between SNPs in the general mRNA degradation pathway and lung cancer risk with FDR < 0.050 in TRICL GWASs

SNP	Gene	Chr.	Position (hg19)	Allele ^a	EAF	Q^{b}	I^2	Effect ^c	OR (95% CI)	P	FDR
rs115834633	LSM2	6	31765984	G/A	0.11	0.200	30.79	+++++	1.20 (1.14-1.27)	3.92E-11	<.0001
rs114312980	LSM2	6	31768799	A/C	0.11	0.230	26.77	+++++	1.20 (1.14-1.27)	4.39E-11	<.0001
rs115801685	LSM2	6	31772093	C/A	0.11	0.220	27.36	+++++	1.20 (1.14-1.27)	4.54E-11	<.0001
rs115489726	LSM2	6	31766660	C/T	0.11	0.240	25.69	+++++	1.20 (1.14-1.27)	8.76E-11	<.0001
rs114637560	LSM2	6	31765864	T/A	0.15	0.260	22.42	+-++++	1.14 (1.08-1.20)	3.04E-07	0.0003
rs114984862	SKIV2L	6	31936668	C/T	0.27	0.290	18.64	+++++	1.09 (1.05-1.13)	1.87E-05	0.013
rs9800264	CNOT6	5	179940091	G/A	0.10	0.750	0.00	++++-	1.13 (1.07-1.19)	3.12E-05	0.013
rs2387281	CNOT6	5	179988283	T/C	0.10	0.743	0.00	++++-	1.13 (1.07-1.19)	3.28E-05	0.013
rs6877400	CNOT6	5	179996111	T/C	0.10	0.747	0.00	++++-	1.13 (1.07-1.19)	3.41E-05	0.013
rs116188106	SKIV2L	6	31927342	G/A	0.27	0.298	17.82	+++++	1.09 (1.04-1.13)	3.70E-05	0.013
rs114011334	SKIV2L	6	31928799	C/T	0.27	0.297	17.92	+++++	1.09 (1.04-1.13)	3.77E-05	0.013
rs115002281	SKIV2L	6	31929014	C/A	0.27	0.297	17.95	+++++	1.08 (1.04-1.13)	3.79E-05	0.013
rs10052782	CNOT6	5	179975104	C/T	0.10	0.723	0.00	++++-	1.13 (1.06-1.19)	4.09E-05	0.013
rs6422334	CNOT6	5	179982151	C/T	0.10	0.734	0.00	++++-	1.13 (1.06-1.19)	4.14E-05	0.013
rs2453176	CNOT6	5	179975792	C/T	0.10	0.723	0.00	++++-	1.13 (1.06-1.19)	4.33E-05	0.013
rs2387285	CNOT6	5	179982278	A/G	0.10	0.700	0.00	++++-	1.12 (1.06-1.19)	4.92E-05	0.014
rs2447734	CNOT6	5	179968674	G/C	0.10	0.720	0.00	++++-	1.12 (1.06-1.19)	4.97E-05	0.014
rs76820754	CNOT6	5	179936737	G/A	0.10	0.735	0.00	++++-	1.12 (1.06-1.19)	8.06E-05	0.021
rs780126	CNOT6	5	179963034	C/T	0.13	0.758	0.00	++++-	1.12 (1.06-1.18)	8.67E-05	0.021
rs812956	CNOT6	5	179953048	G/C	0.10	0.653	0.00	++++-	1.12 (1.06-1.18)	1.18E-04	0.027

SNP: single nucleotide polymorphism; FDR: false discovery rate; TRICL: Transdisciplinary Research in Cancer of the Lung; GWAS: genome-wide association study; Chr.: chromosome; EAF: effect allele frequency; OR: odds ratio; CI: confidence interval;

a Reference allele/effect allele;

^bFixed effect models were used when no heterogeneity was found between studies (Q-test P > 0.100 and $I^2 < 50.0\%$); otherwise, random effect models were used;

[&]quot;c"+" means a positive association, and "-" means a negative association.

Table 3 Linkage disequilibrium between the 11 SNPs of *CNOT6* in European populations included in the 1000 Genomes Project and imputation quality scores

SNP	Position		2	Function	prediction		Imputation quality				
	(hg19)	D'	r²	SNPinfo ^a	Regulome DB ^b	Info ICR	Rsq MDACC	Rsq IARC	Info NCI	Info Toronto	Rsq GLC
rs2453176	179975792				1f	1.000	0.999	1.000	1.000	1.000	1.000
rs780126	179963034	1.00	0.71			0.874	0.751	0.703	0.859	0.857	0.784
rs2387281	179988283	1.00	0.97			0.998	0.972	0.969	0.996	0.990	0.967
rs6877400	179996111	1.00	0.97	Splicing site	5	0.998	0.964	0.965	0.996	0.990	0.953
rs2387285	179982278	1.00	0.97		4	0.990	0.966	0.923	0.988	0.981	0.964
rs812956	179953048	1.00	0.97		6	0.991	0.961	0.962	0.988	0.978	0.976
rs9800264	179940091	1.00	0.99			0.999	0.970	0.977	0.998	0.993	1.000
rs6422334	179982151	1.00	0.99		5	0.999	0.982	0.976	0.997	0.994	0.979
rs10052782	179975104	1.00	1.00		6	1.000	0.998	1.000	1.000	1.000	1.000
rs76820754	179936737	1.00	1.00		6	1.000	0.970	0.971	0.999	0.998	0.999
rs2447734	179968674	1.00	1.00			1.000	0.994	0.999	0.999	0.997	0.999

SNP: single nucleotide polymorphism;

Imputation quality: Rsq: MaCH r-squared; Info: IMPUTE2 information score;

ICR: the Institute of Cancer Research Genome-wide Association Study, UK;

MDACC: the MD Anderson Cancer Center Genome-wide Association Study, US;

IARC: the International Agency for Research on Cancer Genome-wide Association Study, France;

NCI: the National Cancer Institute Genome-wide Association Study, US;

Toronto: the Samuel Lunenfeld Research Institute Genome-wide Association Study, Toronto, Canada;

GLC: German Lung Cancer Study, Germany;

ahttps://snpinfo.niehs.nih.gov/snpinfo/snpfunc.htm;

bhttp://regulomedb.org/.

Table 4 Associations between of CNOT6 rs2453176 (C >T) and lung cancer risk stratified by histologic types in all eight lung cancer GWASs from ILCCO-TRICL

	Overall				AD					SC			
Study	Case	Control	OR (95% CI)	P	Case	Control	OR (95% CI)	P	Case	Control	OR (95% CI)	P	
ICR	1952	5200	1.15 (1.02-1.28)	0.020	465	5200	1.38 (1.12-1.70)	0.002	611	5200	1.14 (0.95-1.38)	0.158	0.181
MDACC	1150	1134	1.25 (1.03-1.52)	0.027	619	1134	1.06 (0.92-1.47)	0.206	306	1134	1.45 (1.08-1.94)	0.013	0.102
IARC	2533	3791	1.14 (1.00-1.29)	0.053	517	2824	1.13 (0.90-1.42)	0.301	911	2968	1.18 (0.98-1.41)	0.081	0.771
NCI	5713	5736	1.10 (1.01-1.21)	0.025	1841	5736	1.17 (1.03-1.33)	0.016	1447	5736	1.04 (0.91-1.20)	0.543	0.220
Toronto	331	499	1.12 (0.76-1.63)	0.057	90	499	1.48 (0.83-2.64)	0.186	50	499	1.00 (0.44-2.25)	0.998	0.442
GLC	481	478	0.93 (0.69-1.26)	0.064	186	478	1.25 (0.86-1.83)	0.240	97	478	0.90 (0.52-1.54)	0.695	0.330
Discovery combined	12160	16838	1.13 (1.06-1.19)	4.33E-05	3818	15871	1.21 (1.11-1.32)	2.04E-05	3424	16015	1.13 (1.03-1.23)	0.009	0.268
Harvard	984	970	0.85 (0.68-1.05)	0.133	597	970	0.79 (0.62-1.01)	0.130	216	970	1.08 (0.75-1.56)	0.678	0.164
deCODE	1319	26380	1.14 (1.01-1.28)	0.032	547	26380	1.02 (0.85-1.21)	0.858	259	26380	1.11 (0.86-1.43)	0.436	0.592
Replication combined	2303	27350	1.00 (0.75-1.32)	0.098	1144	27350	0.91 (0.71-1.16)	0.449	475	27350	1.10 (0.89-1.36)	0.381	0.252
Overall	14463	44188	1.11 (1.04-1.18)	0.001	4862	43221	1.13 (1.00-1.27)	0.050	3897	43365	1.12 (1.03-1.22)	0.006	0.905

GWAS: genome-wide association study; ILCCO: International Lung Cancer Consortium; TRICL: Transdisciplinary Research in Cancer of the Lung; AD, adenocarcinoma; SC, squamous cell carcinoma; OR: odds ratio; CI: confidence interval.

*Homogeneity tests suggest that there is no heterogeneity between the subgroups of AD and SC in each GWAS and overall result (P > 0.05).

ICR: the Institute of Cancer Research Genome-wide Association Study, UK;

MDACC: the MD Anderson Cancer Center Genome-wide Association Study, US;

IARC: the International Agency for Research on Cancer Genome-wide Association Study, France;

NCI: the National Cancer Institute Genome-wide Association Study, US;

Toronto: the Samuel Lunenfeld Research Institute Genome-wide Association Study, Toronto, Canada;

GLC: German Lung Cancer Study, Germany;

Harvard: Harvard Lung Cancer Study;

DeCODE: Icelandic Lung Cancer Study.



Table 5 Associations between of *CNOT6* rs2453176 (C > T) and lung cancer risk stratified by smoking status in six lung cancer GWASs from ILCCO-TRICL Consortia

Study -			Smoker			P^*			
	Case	Control	OR (95% CI)	P	Case	Control	OR (95% CI)	P	•
MDACC	1150	1134	1.25 (1.03-1.52)	0.027					
IARC	2367	2508	1.12 (0.97-1.29)	0.131	159	1253	1.40 (0.94-2.09)	0.096	0.303
NCI	5342	4336	1.10 (1.00-1.22)	0.058	350	1379	0.99 (0.72-1.37)	0.972	0.540
Toronto	236	272	1.14 (0.70-1.86)	0.606	95	217	1.13 (0.61-2.11)	0.702	0.983
GLC	433	258	1.00 (0.67-1.49)	0.995	35	220	1.64 (0.71-3.82)	0.250	0.298
Harvard	892	809	0.87 (0.70-1.09)	0.221	92	161	0.71 (0.38-1.33)	0.288	0.549
Overall	10420	9317	1.09 (1.02-1.17)	0.011	731	3230	1.10 (0.89-1.36)	0.363	0.936

GWAS: genome-wide association study; ILCCO: International Lung Cancer Consortium; TRICL: Transdisciplinary Research in Cancer of the Lung; OR: odds ratio; CI: confidence interval.

MDACC: the MD Anderson Cancer Center Genome-wide Association Study, US;

IARC: the International Agency for Research on Cancer Genome-wide Association Study, France;

NCI: the National Cancer Institute Genome-wide Association Study, US;

Toronto: the Samuel Lunenfeld Research Institute Genome-wide Association Study, Toronto, Canada;

GLC: German Lung Cancer Study, Germany;

Harvard: Harvard Lung Cancer Study;

*Homogeneity tests suggest there is no heterogeneity between the subgroups of smoker and non-smoker in each GWAS and overall result (P > 0.05).



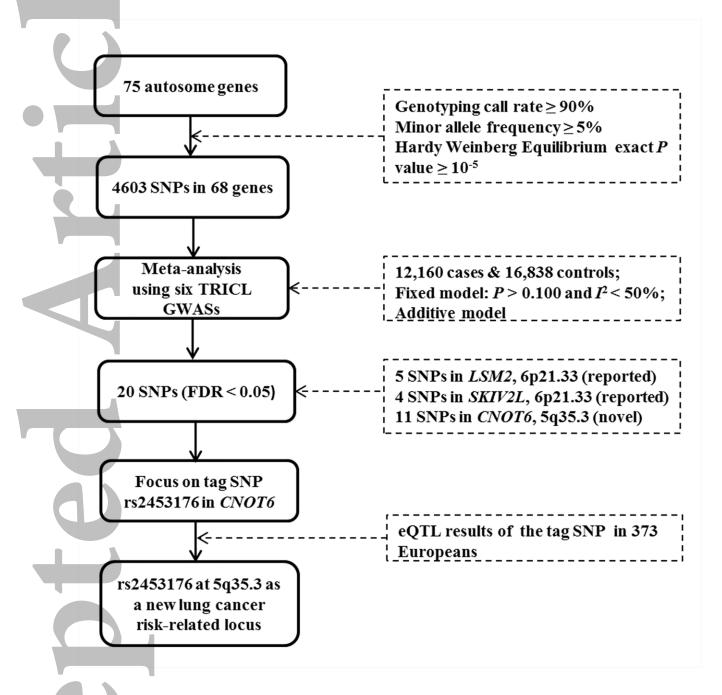
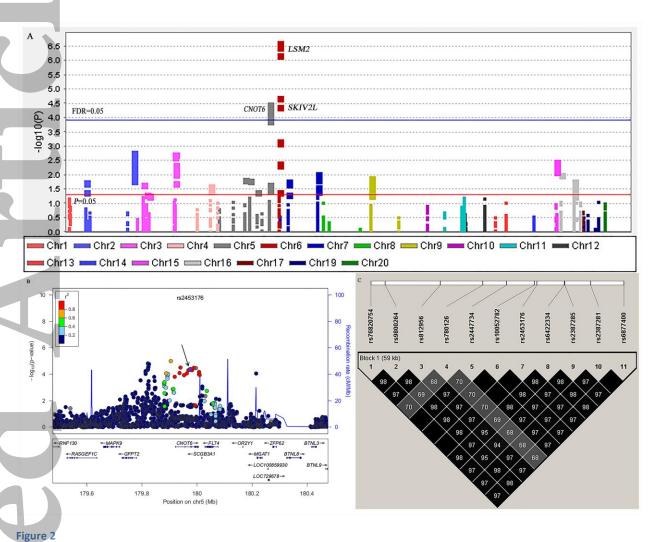


Figure 1







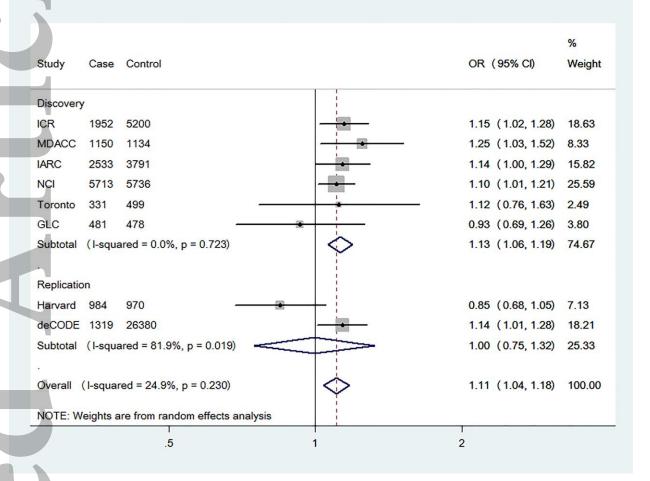
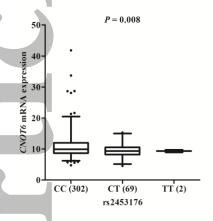
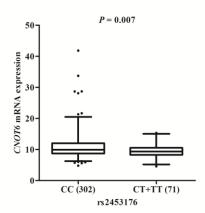


Figure 3







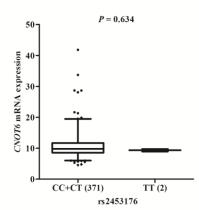


Figure 4