

LETTER TO THE EDITOR

Treatable mitochondrial diseases: cofactor metabolism and beyond

Felix Distelmaier,¹ Tobias B. Haack,^{2,3} Saskia B. Wortmann,^{2,3,4} Johannes A. Mayr⁴ and Holger Prokisch^{2,3}

1 Department of General Pediatrics, Neonatology and Pediatric Cardiology, University Children's Hospital, Heinrich-Heine-University Düsseldorf, 40225 Düsseldorf, Germany

2 Institute of Human Genetics, Helmholtz Zentrum München, 85764 Neuherberg, Germany

3 Institute of Human Genetics, Technische Universität München, 81675 München, Germany

4 Department of Paediatrics, Salzburger Landeskliniken, Paracelsus Medical University, Salzburg, Austria

Correspondence to: Felix Distelmaier,

Department of General Pediatrics, Heinrich-Heine-University, Moorenstr. 5, 40225

Düsseldorf, Germany

E-mail: felix.distelmaier@med.uni-duesseldorf.de

Sir,

In the past, numerous articles related to disorders of mitochondrial cofactor metabolism have been published in *Brain* (Ozand *et al.*, 1998; Gempel *et al.*, 2007; Johnson *et al.*, 2012; Foley *et al.*, 2014; Haack *et al.*, 2014; Ortigoza-Escobar *et al.*, 2016). These studies not only facilitated our understanding of the underlying biochemical defects, but also paved the way to specific treatment options. As a consequence our clinical view on mitochondrial diseases has changed substantially during the last years.

The umbrella term 'mitochondrial disease' comprises a large group of inherited metabolic disorders caused by dysfunction of the pyruvate oxidation route. Our common understanding of mitochondrial diseases mainly refers to classical mitochondrial syndromes such as Leigh syndrome or MELAS (mitochondrial encephalomyopathy, lactic acidosis, and stroke-like episodes). However, the spectrum of mitochondrial diseases is much broader and the development of novel genetic tools has undeniably advanced our knowledge about this disease group. During the past 6 years more than 100 novel mitochondrial diseases have been identified via next generation sequencing (NGS) strategies leading to a total number of around 280 known disease genes, affecting diverse mitochondrial pathways. Accordingly, clinical management of affected individuals is challenging and diagnostic strategies are in flux.

Unfortunately, current therapeutic options for the majority of classical mitochondrial syndromes are limited to supportive care. Nevertheless, apart from these prognostically unfavourable diseases, there are several mitochondrial defects that are amendable by specific treatment strategies (Table 1). Among the group of these 'treatable mitochondrial diseases', defects of cofactor metabolism play a major role.

For normal functioning, the mitochondrial oxidative phosphorylation (OXPHOS) system requires a large number of organic and inorganic cofactors that facilitate these enzyme-catalysed reactions. Organic cofactors are mainly derived from vitamins such as thiamine, riboflavin, biotin or niacin but there are also non-vitamin cofactors such as coenzyme Q₁₀ or heme. The group of inorganic cofactors comprises metal ions such as iron, magnesium, copper, zinc, and molybdenum. Most cofactors or its precursors are derived from food and intestinal absorption as well as tissue uptake requires specific transport mechanisms. However, in the cases of coenzyme Q₁₀ or heme, endogenous biosynthesis is the main source for cellular supply.

In conditions with insufficient cofactor availability, mitochondrial energy metabolism might get severely disturbed, leading to tissue damage primarily affecting organ systems with high energy demand such as skeletal muscle and brain. The most common vitamin-related cofactor diseases caused by nutritional deficiency of thiamine (beriberi), niacin (pellagra), or riboflavin (pellagra sine pellagra) were elucidated

Table 1 Inherited mitochondrial diseases with specific treatment options

Affected pathway	Clinical syndrome	Affected gene(s)	Clinical phenotype	Therapeutic substance	Treatment response
Primary disorders of mitochondrial vitamin cofactor metabolism	Brown-Vialetto-Van Laere syndrome / Fazio-Londe disease	SLC52A2, SLC52A3, (SLC52A1) ^a	Sensorineural hearing loss, cranial nerve palsies	Riboflavin (oral: 10–50 mg/kg/day) ^b	Generally good
	Biotin-thiamine-responsive basal ganglia disease	SLC19A3	Episodic encephalopathy, dystonia, seizures	Thiamine (oral: 10–20 mg/kg/day), biotin (oral: 10–15 mg/kg/day) ^c	Generally good
	Biotinidase deficiency	BTBD	Dermatitis, muscular hypotonia, developmental regression	Biotin (oral: 5–10 mg/kg/day) ^d	Generally good
Disorders with indirect response to mitochondrial vitamin cofactor supplementation	Holocarboxylase synthetase deficiency	HLCS	Skin lesions, metabolic acidosis, seizures, developmental delay	Biotin (oral: 10–20 mg/kg/day) ^e	Variable but generally good
	Thiamine pyrophosphokinase deficiency	TPK1	Episodic encephalopathy, dystonia, spasticity	Thiamine (oral: ~20 mg/kg/day) ^f	Variable (< 10 patients treated so far)
	ACAD9 deficiency	ACAD9	Encephalopathy, myopathy, hypertrophic cardiomyopathy	Riboflavin (oral: 10–20 mg/kg/day) ^g	Variable
	Multiple acyl-CoA dehydrogenase deficiency	ETFA, ETFB, ETFDH, SLC25A32, FLAD1	Early childhood multisystem disease or late-onset form with muscle weakness, hepatopathy, etc.	Riboflavin (oral: ~10 mg/kg/day) ^h	Generally good
	Thiamine-responsive pyruvate dehydrogenase deficiency	PDHA1	Neonatal lactic acidosis, seizures, developmental regression, spasticity	Thiamine (oral: 30–40 mg/kg/day) ⁱ	Variable
Disorders of mitochondrial non-vitamin cofactor metabolism	Coenzyme Q ₁₀ deficiency	PDS1, PDS2, COQ2, COQ4, COQ6, COQ7, ADCK3, ADCK4, COQ9	Variable phenotypes, ranging from adult-onset myopathy to fatal neonatal presentations	Coenzyme Q ₁₀ (oral: 10–30 mg/kg/day) ^j	Highly variable depending on the underlying defect
	Cytochrome c oxidase deficiency	SCO2, COA6	Infantile encephalocardiomyopathy	Copper-histidine (dose unclear; subcutaneous injections of up to 500 µg daily were suggested) ^k	Unclear, only one SCO2 patient treated; only <i>in vitro</i> evidence for COA6
'Inhibitors' of mitochondrial metabolism	Molybdenum cofactor deficiency	MOCS1, MOCS2, GPHN	Infantile-onset epileptic encephalopathy, progressive brain damage	Cyclic pyranopterin monophosphate (intravenous: 80–320 µg/kg/day) ^l	Generally good in MoCD type A patients
	3-Hydroxyisobutyryl-CoA hydrolase deficiency	HIBCH	Infantile Leigh-like phenotype	Valine-restricted diet ^m	Unclear, only few patients treated
	Enoyl-CoA hydratase deficiency	ECHS1	Infantile Leigh-like phenotype	Valine-restricted diet ⁿ	Unclear, only few patients treated
	Thioredoxin 2 deficiency	TXN2	Cerebellar atrophy, dystonia, seizures, peripheral neuropathy	Antioxidant treatment (e.g. Idebenone up to 20 mg/kg/day) ^o	Unclear, only few patients treated so far
	Ethylmalonic encephalopathy	ETHE1	Severe, multisystem infantile disorder	Metronidazole, N-acetyl cysteine as glutathione precursor; liver transplantation ^p	Apparently good (only one patient reported)

^aOnly a single case with postulated SLC52A1 haploinsufficiency has been reported so far.^{b–p}The therapeutic regimens are suggestions based on the indicated references: ^bFoley et al., 2014; ^cHaack et al., 2014; ^dJay et al., 2015; ^eDonti et al., 2016; ^fBanka et al., 2014; ^gGerards et al., 2011; ^hOlsen et al., 2007; ⁱvan Dongen et al., 2015; ^jHargreaves, 2014; ^kFraisinger et al., 2004; ^lSchwahn et al., 2015; ^mSoler-Alfonso et al., 2015; ⁿHaack et al., 2015; ^oHolzerova et al., 2016; ^pDionisi-Vici et al., 2016.

more than 70 years ago (Eijkman and Hopkins, 1965; Sebrell and Butler, 1939; Elvehjem *et al.*, 1973). These diseases are characterized by severe clinical phenotypes including progressive neurological deterioration that are highly responsive to supplementation of the lacking vitamin.

The first inborn errors of mitochondrial cofactor metabolism were already described in the 19th century (Brown, 1894; Londe, 1894). However, it took decades until advances in genetic diagnostic tools allowed the identification of the underlying inherited defects affecting the tissue-specific transport and metabolism of these cofactors. Classical examples of inherited vitamin-related disorders of cofactor metabolism are Brown-Vialetto-Van Laere syndrome (BVVLS1 and 2; OMIM #614707, #211530), biotin-thiamine-responsive basal ganglia disease (OMIM #607483) or biotinidase deficiency (OMIM #253260; Gompertz *et al.*, 1973; Brown, 1894; Ozand *et al.*, 1998). These disorders are autosomal recessive defects of specific transporter proteins, causing severe neurometabolic diseases with onset in childhood. In the case of BVVL, mutations in *SLC52A2* or *SLC52A3* impair cellular supply with riboflavin (Green *et al.*, 2010; Johnson *et al.*, 2012). This leads to sensorineural hearing loss and cranial nerve palsies. The progression of these neurological problems can be slowed down or even stopped by supplementation of high-dose riboflavin (Foley *et al.*, 2014). Apart from classical BVVLS, Fazio-Londe disease (OMIM #211500; Londe, 1894) caused by *SLC52A3* mutations is a recognized riboflavin-responsive clinical entity similar to BVVLS but without sensorineural deafness. Biotin-thiamine-responsive basal ganglia disease is characterized by episodes of encephalopathy, dystonia and seizures. The disease is caused by mutations in *SLC19A3*, leading to impaired thiamine transport into the CNS with subsequent mitochondrial dysfunction (Zeng *et al.*, 2005). Thiamine is beneficial in affected patients and prevents further neurological deterioration (Haack *et al.*, 2014). In addition, biotin supplementation is helpful via increasing the gene expression of *SLC19A3* (Debs *et al.*, 2010). During the past years, the spectrum of vitamin-related disorders of cofactor metabolism has broadened substantially. Treatable genetic defects include holocarboxylase synthetase deficiency (OMIM #253270), thiamine pyrophosphokinase deficiency (OMIM #614458) or inherited riboflavin deficiency (OMIM #615026). The recent reports of Schiff *et al.* (2016) and Olsen *et al.* (2016) add to this rapidly growing list of potentially treatable diseases affecting mitochondrial cofactor metabolism (Table 1). They illustrate that in some individuals, the clinical course can be alleviated with simple measures such as vitamin supplementation.

Importantly, beyond the field of such ‘classical’ cofactor-related mitochondrial diseases, there is also a growing group of treatable disorders that affect mitochondrial metabolic pathways via inhibitory/toxic mechanism. This involves, for example, 3-hydroxyisobutyryl-CoA hydrolase (HIBCH) deficiency (OMIM #250620) and enoyl-CoA hydratase (ECHS1) deficiency (OMIM #616277), both

defects of the valine catabolic pathway. The disorders induce secondary OXPHOS deficiency, which is most likely caused by the accumulation of the toxic metabolites methacrylyl-CoA and acryloyl-CoA (Ferdinandusse *et al.*, 2013; Peters *et al.*, 2014). The diseases present with a severe infantile Leigh-like phenotype. Preliminary clinical evidence suggests that affected children might be responsive to a valine-restricted diet (Haack *et al.*, 2015; Soler-Alfonso *et al.*, 2015). Another example is the recently described disorder thioredoxin 2 deficiency (OMIM #616811), which affects an essential mitochondrial pathway that is protective against oxidative stress. The clinical phenotype is characterized by early-onset neurodegeneration, dystonia, and seizures. *In vitro* and *in vivo* studies indicated a benefit of high-dose antioxidant treatment (Holzerova *et al.*, 2016).

The studies mentioned above illustrate how the application of NGS technologies facilitates our understanding of the pathophysiology of mitochondrial diseases in general and potentially treatable disease subgroups in particular. Importantly, many of the above-mentioned treatable defects show clinical and neuroradiological similarities to classical mitochondrial disorders such as Leigh syndrome (e.g. *SLC19A3*, *ECHS1* or *HIBCH* mutations). Therefore, it is crucial to recognize that a subset of patients with a clinical diagnosis of ‘mitochondrial disease’ may suffer from a vitamin/cofactor-responsive condition. These disorders need to be urgently explored in patients with mitochondrial diseases. In our view, early empirical therapy in combination with rapid genetic diagnostic strategies should be applied to maximize treatment benefits and to avoid any delay in appropriate treatment. This especially holds true in critically ill patients. For example in neonates with severe lactic acidosis or children with acute neurological deterioration, a therapy regimen including thiamine (20 mg/kg/day), biotin (5 mg/kg/day), riboflavin (20 mg/kg/day) and coenzyme Q₁₀ (15 mg/kg/day) might be considered. This approach covers the most important disorders summarized in Table 1. Importantly, empirical application of these cofactors is not expected to cause any serious side effects. In case of biotin, potential interaction with streptavidin-biotin laboratory methods should be kept in mind (Kummer *et al.*, 2016).

However, despite the scientific progress discussed above, it is important to realize that not all of the disorders summarized in Table 1 have an ideal treatment prognosis and caution should be emphasized against an overly enthusiastic approach. Moreover, clinical evidence for the suggested treatment options in some of the disorders remains limited, owing largely to the fact that these are rare and recently discovered genetic defects. This underlines the need of further clinical and genetic data collection, for example via international mitochondrial disease registries, to achieve generalizable recommendations regarding optimal treatment and long-term prognosis for these patients.

Funding

This work was supported by the BMBF funded German Network for Mitochondrial Disorders (mitoNET #01GM1113C) and by the E-Rare project GENOMIT (01GM1603). T.B.H. was supported by the BMBF through the Juniorverbund in der Systemmedizin ‘mitOmics’ (FKZ 01ZX1405C). F.D. was supported by a grant of the Forschungskommission of the Medical Faculty of the Heinrich-Heine-University Düsseldorf.

References

- Banka S, de Goede C, Yue WW, Morris AA, von Bremen B, Chandler KE, et al. Expanding the clinical and molecular spectrum of thiamine pyrophosphokinase deficiency: a treatable neurological disorder caused by TPK1 mutations. *Mol Genet Metab* 2014; 113: 301–6.
- Brown C. Infantile amyotrophic lateral sclerosis of the family type. *J Nerv Ment Dis* 1894; 19: 707–16.
- Debs R, Depienne C, Rastetter A, Bellanger A, Degos B, Galanaud D, et al. Biotin-responsive basal ganglia disease in ethnic Europeans with novel SLC19A3 mutations. *Arch Neurol* 2010; 67: 126–30.
- Dionisi-Vici C, Diodato D, Torre G, Picca S, Pariente R, Giuseppe Picardo S, et al. Liver transplant in ethylmalonic encephalopathy: a new treatment for an otherwise fatal disease. *Brain* 2016; 139: 1045–51.
- Donti TR, Blackburn PR, Atwal PS. Holocarboxylase synthetase deficiency pre and post newborn screening. *Mol Genet Metab Rep* 2016; 7: 40–4.
- Eijkman C, Hopkins F. Antineuritic vitamin and beriberi: Nobel lecture. Nobel lectures, physiology or medicine 1922–1941. Amsterdam: Elsevier Publishing Company; 1965.
- Elvehjem CA, Madden RJ, Strong FM, Woolley DW. Relation of nicotinic acid and nicotinic acid amide to canine black tongue. *J Am Chem Soc* 1973; 95: 1767–8.
- Ferdinandusse S, Waterham HR, Heales SJ, Brown GK, Hargreaves IP, Taanman JW, et al. HIBCH mutations can cause Leigh-like disease with combined deficiency of multiple mitochondrial respiratory chain enzymes and pyruvate dehydrogenase. *Orphanet J Rare Dis* 2013; 8: 188.
- Foley AR, Menezes MP, Pandraud A, Gonzalez MA, Al-Odaib A, Abrams AJ, et al. Treatable childhood neuronopathy caused by mutations in riboflavin transporter RFVT2. *Brain* 2014; 137: 44–56.
- Freisinger P, Horvath R, Macmillan C, Peters J, Jaksch M. Reversion of hypertrophic cardiomyopathy in a patient with deficiency of the mitochondrial copper binding protein Sco2: is there a potential effect of copper? *J Inher Metab Dis* 2004; 27: 67–79.
- Gempel K, Topaloglu H, Talim B, Schneiderat P, Schoser BG, Hans VH, et al. The myopathic form of coenzyme Q10 deficiency is caused by mutations in the electron-transferring-flavoprotein dehydrogenase (ETFDH) gene. *Brain* 2007; 130: 2037–44.
- Gerards M, van den Bosch BJ, Danhauser K, Serre V, van Weeghel M, Wanders RJ, et al. Riboflavin-responsive oxidative phosphorylation complex I deficiency caused by defective ACAD9: new function for an old gene. *Brain* 2011; 134: 210–19.
- Gompertz D, Goodey PA, Bartlett K. Evidence for the enzymic defect in beta-methylcrotonylglycinuria. *FEBS Lett* 1973; 32: 13–14.
- Green P, Wiseman M, Crow YJ, Houlden H, Riphagen S, Lin JP, et al. Brown-Vialetto-Van Laere syndrome, a ponto-bulbar palsy with deafness, is caused by mutations in C20orf54. *Am J Hum Genet* 2010; 86: 485–9.
- Haack TB, Jackson CB, Murayama K, Kremer LS, Schaller A, Kotzaeridou U, et al. Deficiency of ECHS1 causes mitochondrial encephalopathy with cardiac involvement. *Ann Clin Transl Neurol* 2015; 2: 492–509.
- Haack TB, Klee D, Strom TM, Mayatepek E, Meitinger T, Prokisch H, et al. Infantile Leigh-like syndrome caused by SLC19A3 mutations is a treatable disease. *Brain* 2014; 137: e295.
- Hargreaves IP. Coenzyme Q10 as a therapy for mitochondrial disease. *Int J Biochem Cell Biol* 2014; 49: 105–11.
- Holzerova E, Danhauser K, Haack TB, Kremer LS, Melcher M, Ingold I, et al. Human thioredoxin 2 deficiency impairs mitochondrial redox homeostasis and causes early-onset neurodegeneration. *Brain* 2016; 139: 346–54.
- Jay AM, Conway RL, Feldman GL, Nahhas F, Spencer L, Wolf B. Outcomes of individuals with profound and partial biotinidase deficiency ascertained by newborn screening in Michigan over 25 years. *Genet Med* 2015; 17: 205–9.
- Johnson JO, Gibbs JR, Megarbane A, Urtizberea J A, Hernandez DG, Foley AR, et al. Exome sequencing reveals riboflavin transporter mutations as a cause of motor neuron disease. *Brain* 2012; 135: 2875–82.
- Kummer S, Hermesen D, Distelmaier F. Biotin treatment Mimicking Graves’ disease. *N Engl J Med* 2016; 375: 704–6.
- Londe P. Paralysie bulbaire progressive, infantile et familiale. *Rev Med* 1894; 14: 212–54.
- Olsen RK, Koňářková E, Giancaspero TA, Mosegaard S, Boczonadi V, Mataković L, et al. Riboflavin-responsive and -non-responsive mutations in FAD synthase cause multiple Acyl-CoA dehydrogenase and combined respiratory-chain deficiency. *Am J Hum Genet* 2016; 98: 1130–45.
- Olsen RK, Olpin SE, Andresen BS, Miedzybrodzka ZH, Pourfarzam M, Merinero B, et al. ETFDH mutations as a major cause of riboflavin-responsive multiple acyl-CoA dehydrogenation deficiency. *Brain* 2007; 130: 2045–54.
- Ortigoza-Escobar JD, Molero-Luis M, Arias A, Oyarzabal A, Darin N, Serrano M, et al. Free-thiamine is a potential biomarker of thiamine transporter-2 deficiency: a treatable cause of Leigh syndrome. *Brain* 2016; 139: 31–8.
- Ozand PT, Gascon GG, Al Essa M, Joshi S, Al Jishi E, Bakheet S, et al. Biotin-responsive basal ganglia disease: a novel entity. *Brain* 1998; 121: 1267–79.
- Peters H, Buck N, Wanders R, Ruiters J, Waterham H, Koster J, et al. ECHS1 mutations in Leigh disease: a new inborn error of metabolism affecting valine metabolism. *Brain* 2014; 137: 2903–8.
- Schiff M, Veauville-Merllié A, Su CH, Tzagoloff A, Rak M, Ogier de Baulny H, et al. SLC25A32 mutations and riboflavin-responsive exercise intolerance. *N Engl J Med* 2016; 374: 795–7.
- Schwahn BC, Van Spronsen FJ, Belaidi AA, Bowhay S, Christodoulou J, Derks TG, et al. Efficacy and safety of cyclic pyranopterin monophosphate substitution in severe molybdenum cofactor deficiency type A: a prospective cohort study. *Lancet* 2015; 386: 1955–63.
- Sebrell WH, Butler RE. “Riboflavin deficiency in man (Ariboflavinosis)”. *Public Health Rep* 1939; 54: 2121–31.
- Soler-Alfonso C, Enns GM, Koenig MK, Saavedra H, Bonfante-Mejia E, Northrup H. Identification of HIBCH gene mutations causing autosomal recessive Leigh syndrome: a gene involved in valine metabolism. *Pediatr Neurol* 2015; 52: 361–5.
- van Dongen S, Brown RM, Brown GK, Thorburn DR, Boneh A. Thiamine-responsive and non-responsive patients with PDHC-E1 deficiency: a retrospective assessment. *JIMD Rep* 2015; 15: 13–27.
- Zeng WQ, Al-Yamani E, Acierio JS Jr, Slaugenhaupt S, Gillis T, MacDonald ME, et al. Biotin-responsive basal ganglia disease maps to 2q36.3 and is due to mutations in SLC19A3. *Am J Hum Genet* 2005; 77: 16–26.