Biomarkers for monitoring clinical efficacy of allergen immunotherapy for allergic rhinoconjunctivitis and allergic asthma: an EAACI Position Paper

M. H. Shamji^{1,2,3}, J. H. Kappen^{1,2,3,4}, M. Akdis²⁰, E. Jensen-Jarolim^{5,6}, E. F. Knol⁷, J. Kleine-Tebbe⁸, B. Bohle⁹, A. M. Chaker^{10,11}, S. J. Till^{12,13}, R. Valenta¹⁴, L. K. Poulsen¹⁵, M. A. Calderon^{1,2,3}, P. Demoly¹⁶, O. Pfaar^{17,18}, L. Jacobsen¹⁹, S. R. Durham^{1,2,3} & C. B. Schmidt-Weber¹⁰

¹Allergy and Clinical Immunology, National Heart and Lung Institute, Imperial College London; ²MRC & Asthma UK Centre in Allergic Mechanisms of Asthma; ³Allergy and Clinical Immunology, Immunomodulation and Tolerance Group, Imperial College London, London, UK; ⁴Department of Pulmonology, STZ Centre of Excellence for Asthma & COPD, Sint Franciscus Vlietland Group, Rotterdam, The Netherlands; ⁵Department of Pathophysiology and Allergy Research, Center of Pathophysiology, Infectiology and Immunology, Medical University Vienna; ⁶The interuniversity Messerli Research Institute, University of Veterinary Medicine Vienna, Medical University Vienna, Vienna, Austria; ⁷Departments Immunology and Dermatology/Allergology, University Medical Center Utrecht, Utrecht, The Netherlands; ⁸Allergy & Asthma Center Westend, Outpatient Clinic and Research Center Hanf, Ackermann & Kleine-Tebbe, Berlin, Germany; ⁹Department of Pathophysiology and Allergy Research, Medical University of Vienna, Vienna, Austria; ¹⁰Center of Allergy and Environment (ZAUM), Technische Universität and Helmholtz Center Munich; ¹¹Department of Otolaryngology, Allergy Section, Klinikum rechts der Isar, Technische Universität, Munich, Germany; ¹²Division of Asthma, Allergy and Lung Biology, King's College London; ¹³Department of Allergy, Guy's and St. Thomas' NHS Foundation Trust, London, UK; ¹⁴Division of Immunopathology, Department of Pathophysiology and Allergy Research, Center for Pathophysiology, Infectiology and Immunology, Medical University of Vienna, Vienna, Austria; ¹⁵Allergy Clinic, Copenhagen University Hospital at Gentofte, Copenhagen, Denmark; ¹⁶Division of Allergy, Department of Pulmonology, Arnaud de Villeneuve Hospital, University Hospital of Montpellier and Sorbonne University, Paris, France; ¹⁷Department of Otorhinolaryngology, Head and Neck Surgery, Universitätsmedizin Mannheim, Medical Faculty Mannheim, Heidelberg University, Mannheim; ¹⁸Center for Rhinology and Allergology, Wiesbaden, Germany; ¹⁹Allergy Learning and Consulting, Copenhagen, Denmark; ²⁰Swiss Institute of Allergy and Asthma Research (SIAF), University of Zürich, Davos, Switzerland

To cite this article: Shamji MH, Kappen JH, Akdis M, Jensen-Jarolim E, Knol EF, Kleine-Tebbe J, Bohle B, Chaker AM, Till SJ, Valenta R, Poulsen LK, Calderon MA, Demoly P, Pfaar O, Jacobsen L, Durham SR, Schmidt-Weber CB. Biomarkers for monitoring clinical efficacy of allergen immunotherapy for allergic rhinoconjunctivitis and allergic asthma: an EAACI position paper. *Allergy* 2017; DOI: 10.1111/all.13138.

Keywords

allergen immunotherapy; basophil activation; biomarkers; IgE-FAB; IgG4.

Correspondence

Mohamed H. Shamji, Immunomodulation and Tolerance Group, Immune Tolerance Network (ITN) Distributed Centre of Excellence for Allergy & Asthma, Allergy & Clinical Immunology Inflammation, Repair and Development National Heart & Lung Institute I Imperial College London 1st Floor, Room 1111 Sir Alexander Fleming Building South Kensington Campus London SW7 2AZ I United Kingdom. Tel: +44 (0) 20 7594 3476 Fax: +44 (0) 20 7594 3476 Mobile: +44 (0) 0771405129 E-mail: m.shamji99@imperial.ac.uk

Accepted for publication 26 January 2017

DOI:10.1111/all.13138

Abstract

Background: Allergen immunotherapy (AIT) is an effective treatment for allergic rhinoconjunctivitis (AR) with or without asthma. It is important to note that due to the complex interaction between patient, allergy triggers, symptomatology and vaccines used for AIT, some patients do not respond optimally to the treatment. Furthermore, there are no validated or generally accepted candidate biomarkers that are predictive of the clinical response to AIT. Clinical management of patients receiving AIT and efficacy in randomised controlled trials for drug development could be enhanced by predictive biomarkers.

Method: The EAACI taskforce reviewed all candidate biomarkers used in clinical trials of AR patients with/without asthma in a literature review. Biomarkers were grouped into seven domains: (i) IgE (total IgE, specific IgE and sIgE/Total IgE ratio), (ii) IgG-subclasses (sIgG1, sIgG4 including SIgE/IgG4 ratio), (iii) Serum inhibitory activity for IgE (IgE-FAB and IgE-BF), (iv) Basophil activation, (v) Cytokines and Chemokines, (vi) Cellular markers (T regulatory cells, B regulatory cells and dendritic cells) and (vii) *In vivo* biomarkers (including provocation tests?).

Results: All biomarkers were reviewed in the light of their potential advantages as well as their respective drawbacks. Unmet needs and specific recommendations on all seven domains were addressed.

Edited by: De Yun Wang

[Corrections added on 17 May 2017 after first online publication: Corrections have been made to the seventeenth and eighteenth affiliations and updates applied to the conflicts of interest in this version.] **Conclusions:** It is recommended to explore the use of allergen-specific IgG4 as a biomarker for compliance. sIgE/tIgE and IgE-FAB are considered as potential surrogate candidate biomarkers. Cytokine/chemokines and cellular reponses provided insight into the mechanisms of AIT. More studies for confirmation and interpretation of the possible association with the clinical response to AIT are needed.

Allergen immunotherapy (AIT) is an effective treatment for allergic rhinoconjunctivitis (AR) with or without asthma (1– 12). Allergen immunotherapy has disease-modifying properties and confers long-term clinical benefit after cessation of treatment (6, 7, 13–17). Allergen immunotherapy is routinely used in daily practice and can be administered either subcutaneously (SCIT) or sublingually (SLIT) (3–12). Although AIT is effective, the degree of remission strongly varies depending on the complex interaction between patient, allergy, symptoms and vaccines used for AIT (3–9). Clinical management of patients receiving AIT and efficacy in randomized controlled trials for drug development could be significantly enhanced if there were means to identify those who are most likely to respond, when to stop treatment, how to predict relapse and

when to perform booster AIT. Furthermore, biomarkers in

AIT can play a central role in personalized medicine (18). Although recommendations for the standardization of clinical outcomes used in AIT trials for AR have recently been defined (1, 19-21), to date there is no consensus on candidate surrogate biomarkers of efficacy or biomarker combinations that would be prognostic, predictive and/or surrogate of the clinical response to AIT (22). In the sense of personalized medicine, biomarkers can be utilized to assist patient selection, identification of responders, target intervention at those who will benefit and to exclude those who are less likely to respond to treatment, thus meeting the criteria of personalized medicine. Additionally, they can be of major importance for the development of novel vaccines and for the optimization of existing therapeutic regimes. According to International Conference on Harmonization (ICH) E15 guidance on 'Definitions for Genomic Biomarkers, Pharmacogenomics, Pharmacogenetics, Genomic Data and Sample Coding Categories', biomarkers are 'indicators of normal biologic processes, pathogenic processes and/or response to therapeutic or other interventions' (22). Biomarkers can be applied in the context of controlled clinical trials for regulatory approval as well as in a clinical practice. Criteria for evaluating and selecting candidate biomarkers are provided by ICH E16 guideline 'Biomarkers Related to Drug or Biotechnology Product Developments: Context, structure and format of Qualification Submissions' (23). Per candidate biomarker, an overview containing the strengths and the limitations, design of the studies supporting its utility should be reported. The 'Guideline on the Clinical Development of Products for Specific Immunotherapy for the Treatment of Allergic Diseases' by the European Medicines Agency (EMA) published in 2008 advises to include immunological changes (e.g. changes in allergen-specific IgG levels, T-cell responses and/or cytokine production) and/or modifications of the end-organ specific response (e.g. provocation tests) in pharmacokinetic and dynamic studies in order to show the effect of AIT on the immune system (24). However, since 2008 several novel immunological markers in AIT have become available and may potentially be used as surrogate/predictive biomarkers for AIT. In this context, laboratory techniques should be reproducible, robust, sensitive and specific following the ICH guidelines for validation of analytical procedures 'Validation of Analytical Procedures: Methodology' (25).

The European Academy of Allergy and Clinical Immunology (EAACI) Immunotherapy Interest Group (IT IG) has conducted a task force (TF) on 'Biomarkers for monitoring the clinical efficacy of allergen Immunotherapy'.

The aim was (i) to collect and evaluate surrogate immunological and clinical biomarker data on the effects of AIT for AR with and without asthma obtained from clinical trials of AIT and (ii) to recommend a consensus position on candidate biomarkers for monitoring AIT and how these biomarkers could be used and implemented in future clinical trials of AIT and daily practice.

Methodology

Taskforce

After two initial meetings (Copenhagen June 2014, London October 2014), the primary objectives of the TF were confirmed: (i) to collect and review surrogate/predictive immunological and clinical biomarker data on the effects of AIT for AR with and without asthma, (ii) to identify surrogate candidate biomarkers that correlate with immunological and clinical effect of AIT, (iii) to identify surrogate/predictive clinical and immunological candidate biomarkers to monitor the effects of AIT in the target organ and systemically during the early and late allergic responses following allergen exposure, (iv) to identify surrogate cellular, humoral and molecular candidate biomarkers to monitor the effects of AIT during and after discontinuation of treatment, (v) to confirm (or reject) the candidate biomarkers for monitoring AIT. Subgroups of the TF drafted these sections on the background, advantages, disadvantages and current critical issues as well as on the unmet needs and recommendations for possible outcomes. During the third TF meeting which was held in London (United Kingdom) in August 2015, individual sections were thoroughly discussed and revised. Following this consensus meeting, the TF committee was responsible for drafting the EAACI TF position paper (PP) in a final draft, which was circulated once again to all TF members for critical review.

The TF report consists of recommendations for seven domains elaborated by the workshop participants (Fig. 1): (i)



Domain v: Cytokines and Chemokines

Figure 1 Seven domains: (i) IgE (total IgE, specific IgE, and sIgE/ Total IgE ratio), (ii) IgG-subtypes (sIgG1, sIgG4 including SIgE/IgG4 ratio), (iii) Serum inhibitory activity for IgE (IgE-FAB and IgE-BF), (iv)

IgE (total IgE, specific IgE and sIgE/total IgE ratio), (ii) IgG subtypes (sIgG1, sIgG4 including SIgE/IgG4 ratio), (iii) serum inhibitory activity for IgE (IgE-FAB and IgE-BF), (iv) basophil activation, (v) cytokines and chemokines, (vi) cellular markers (T regulatory cells, B regulatory cells and dendritic cells) and (vii) *in vivo* biomarkers (including nasal and chamber provocation tests). Health economic outcomes were not considered within the scope of this paper. Representatives from regulatory bodies, pharmaceuticals and biotech companies were invited to join the TF as observers and contribute to the discussion of the PP. EAACI is solely responsible for this PP, which does not represent an official document of any governmental agency such as the Paul-Ehrlich-Institute or the EMA.

Efficacy and biomarkers

The gold standard of efficacy of AIT is the evaluation of clinical symptoms and rescue medications during natural allergen exposure, as defined by the EAACI task force (1) following regulatory guidelines (24). The recommended primary outcome measure is the daily combined symptom and medication scores in AR.

According to the Biomarker Definition Working Group, biomarkers are quantitative measurements that allow clinicians to (i) diagnose, (ii) assess the disease stage, (iii) predict clinical outcomes and (iv) monitor the treatment effects (26).

Table 1 Introduced levels of evidence

Level of evidence	Study type
A	Randomized double-blinded placebo control
D	Nonrandomized double-blinded placebo control
D	Noniandomized open placebo control
С	Untreated control
	Cross-sectional
D	Retrospective
	Responders vs nonresponder

Basophil activation, (v) Cytokines and Chemokines, (vi) Cellular markers (T regulatory cells, B regulatory cells, and dendritic cells), and (vii) *In vivo* biomarkers (including nasal and chamber provocation tests?).

In this report, biomarkers are considered in the context of clinical trials as well as monitoring the response of patients in clinical practice.

Review of literature and level of evidence

Literature was reviewed by a PubMed search using the following MESH terms: immunotherapy, allergic rhinitis, desensitization, biomarkers, allergy. Additional articles were identified by reviewing the reference lists of relevant papers. Limitations were only studies published in English language and no older than 20 years (>1995) and available on PubMed. No limitation was set on the vaccines used. Only studies with a placebo or untreated allergic control group were included. We introduced the following levels of evidence (Table 1): (randomized) double-blinded placebo control (level A), nonrandomized open placebo control (level B), untreated control, cross-sectional (level C), retrospective, responders *vs* nonresponders (level D).

Recently, new administration forms such as intralymphatic (27, 28) or epidermal (29) routes have been advanced, but the overall clinical efficacy of these treatments is still debated and the treatments are not generally available, so these forms are not included in the present paper.

Results

Literature review

Tables S1 and S2 presents an overview of the results per biomarker and a summary of the included studies (Table S2).

Domain i: IgE

Background and study analyses

Elevated serum specific IgE levels and symptoms on exposure to the sensitizing allergen are currently the sole standard for allergy diagnosis and inclusion criteria for starting AIT (19, 20, 30). Several pollen AIT studies have reported that the levels of specific IgE (sIgE) are transiently increased during treatment (Table S1) and followed by blunting of the seasonal increases. No functional or clinical relevance, for example severe allergic reactions, has been associated with this transient increase in sIgE. In long-term AIT studies, the levels of sIgE have been shown to be decreased over time, for example (31, 32). A heterogeneity in total IgE (tIgE) response during AIT (Table S1) has also been shown. Many studies confirmed no change, while others reported an increase or decrease in the levels of tIgE. These trends seem to depend mainly on the duration of the study or the time of sampling. Like sIgE, an initial increase in tIgE is followed by a decrease (Table S1).

The ratio of sIgE to total IgE (sIgE/tIgE ratio) as a predictive marker has been evaluated in a group of patients who received grass pollen or house dust mite (HDM) AIT for 4 years. The study involved both SCIT and SLIT treatment (33). Clinical outcome was measured using visual analogue scores. A cut-off value of 16.2% of IgE ratio predicted the successful outcome of AIT revealing a sensitivity of 97.2% and specificity of 88.1%. A randomized controlled open-label study of limited size could not replicate these results, while other studies did show a similar correlation between IgE ratio and clinical outcome of AIT (Table S1) (33–37).

Advantages

- Serum-based biomarker and a gold standard for selection of patients for AIT.
- Increases during updosing, maintenance and following AIT reflect immunogenicity and allergen exposure.
- Some studies show that an elevated sIgE/tIgE ratio is a potential positive predictive marker for AIT.

Disadvantages and critical issues

- sIgE shows a clear rise in early phase of SCIT and SLIT (more pronounced in SLIT) without clinical and functional relevance.
- The utility of the sIgE/tIgE ratio has not been properly evaluated and validated in Randomized double-blinded placebo control trial. One Randomized double-blinded placebo control trial of limited size did not replicate the relationship between clinical outcome and IgE ratio.
- A variety of assay platforms could be used to measure sIgE/tIgE ratio; however, equivalence between tIgE units IU/ml (kU/l) and sIgE units (kUa/l) has only been demonstrated for one singleplex IgE assay platform (38).

Unmet needs and recommendations

- More data are needed that assess the relationship between sIgE/tIgE and clinical outcome in responders *vs* nonresponders.
- We recommend that baseline sIgE/tIgE ratio is evaluated as a biomarker in AIT in controlled clinical trials in order to validate cut-off values for (to predict likelihood of response/ nonresponse to treatment) sensitivity and specificity.
- For comparability, there is a need for using standardized assay platforms and established reference ranges and cut-off values.

• The role of locally produced sIgE/tIgE may provide an additional option to capture symptomatic relevance of the biomarker.

Domain ii: IgG subtypes

Background and study analyses

Analysis of the regulation of IgG subtypes following AIT has resulted in specific increases in the range of 10- to 100-fold in the concentrations of IgG1 and particularly of sIgG4, for example (39, 40). A correlation between allergen sIgG4 and clinical outcomes has been reported in some but not all studies (41-44), in a long-term follow-up study until 6 years after termination of AIT no correlation was found (45). In a withdrawal study of AIT, levels of sIgG4 were increased in a time-dependent fashion during treatment followed by a near 90% decline of sIgG4 levels which were still elevated compared to pretreatment levels. sIgG4 is considered to compete with allergen binding of sIgE bound to Fcc receptors of mast cells and basophils, and thus acts as a blocking antibody that prevents the activation and degranulation of effector cells (34, 46). Recent data show that immuno-solid-phase allergy chip (ISAC) can be used to determine the increased blocking of sIgG4 in AIT patients (47, 48). One study demonstrated an association with clinical outcome parameters (49). Immuno-solid-phase allergy chip may also be applicable to monitor the induction of sIgG4 in the updosing phase of SCIT while it shows the application of ISAC in the updosing phase of AIT (50).

In addition, some distinct features of sIgG4 suggest that it may have an anti-inflammatory role. IgG4 antibodies are dynamic molecules that exchange Fab arms by swapping heavy–light chain pairs between IgG4 molecules with different specificities (51). This process results in the production of bispecific antibodies with a substantially decreased capacity for cross-linking, because they are functionally monomeric (52). In addition, serum 'blocking' IgG4 antibodies have the capacity to suppress both allergen-triggered basophil histamine release and the binding of IgE–allergen complexes to B cells. A validated flow cytometry-based assay (IgE-FAB) has been developed as a surrogate for IgE-facilitated antigen presentation and activation of T cells during AIT (53).

It has been known for many years that sIgG levels in allergic individuals are elevated in nasal lavage (54). More recently, it was demonstrated that IgG1 and IgG4 appeared in mucosal fluids after AIT with genetically modified allergens. The increase in IgG4 levels was significantly associated with reduction in nasal sensitivity (40). Furthermore, the ratio of IgE to sIgG4 was shown to be decreased in several SLIT studies and was correlated with a decrease in late-phase skin reaction (55–58). This finding has not consistently been reproduced (59, 60).

Advantages

- Serum-based biomarker.
- Consistent results of elevated serum concentrations of sIgG4 are published in SCIT and SLIT studies.

- sIgG4 indicates allergen exposure and can be informative in combination with functional assay.
- Immuno-solid-phase allergy chip can be used to determine sIgG4-blocking activity.
- Data on local antibody levels are available, and further studies are needed.

Disadvantages and critical issues

• A firm relationship between quantitative levels of sIgG4 antibodies and clinical efficacy such as combined symptoms and rescue medication scores (CSMS) during SCIT and SLIT is missing.

Unmet needs and recommendations

- Low sIgG4 is a potential negative predictive marker.
- Failure in IgG4 induction may also be indicative for inadequate compliance.
- We recommend to use specific IgG4 rather than total IgG as a biomarker for evaluating immunological response to AIT in clinical research and drug development.
- Limited data are available on local antibody levels and activities. More studies, especially comparing local effects to peripheral effects, are needed in order to draw firm conclusions.
- More data are needed to evaluate the role of other IgG subsets, IgD and IgA.

Domain iii: Serum inhibitory activity for IgE

Background and study analyses

In the mid-1930s, Cooke et al. (61) reported on the induction of serum inhibitory antibody activity following AIT; later, this proved to be serum inhibitory activity for IgE. Mainly antibodies in the IgA and IgG fraction of the serum caused this effect (54, 62). Serum inhibitory effect for IgE includes the prevention of allergen binding to IgE (IgE-BF), the binding of IgE–allergen complexes to B cells and the inhibition of basophils. The latter will be discussed in a separate domain (iv) on basophil activation.

IgE-blocking factor (IgE-BF) is the extent to which several factors can hinder IgE from binding to its allergen and thus preventing a pro-allergic response and clinical symptoms (34, 63, 64). To examine this effect, a solid-phase assay is available (65). Several studies confirmed an increase in IgE-BF following AIT (Table S1), associated with clinical outcome in clinical trials. The IgE-BF assay is operated on an Advia Centaur instrument that has limited availability as it is no longer produced, or an alternative reverse-type IgE assay platform.

IgE-FAB is a highly reproducible flow cytometry-based bioassay that was developed to detect the binding of allergen–IgE complexes to B cells that express surface low-affinity IgE receptor FceRII (CD23). This IgE-facilitated allergen presentation via CD23 to B cells has been used as a surrogate rate-limiting step for the subsequent processing of allergen and HLA class II-dependent presentation of allergen peptides by B cells to specific T-cell clones. For example, it has been demonstrated that serum obtained post-birch immunotherapy inhibited allergen–IgE binding to B cells that correlated closely with inhibition of IgE-facilitated presentation to specific T-cell clones (34, 46, 66). Furthermore, serum obtained from patients that received grass pollen AIT could inhibit IgE-facilitated allergen presentation to a grass-specific T-cell clone (67). Specific IgG4 within postimmunotherapy serum appears to play a key role in inhibiting this mechanism (31, 68). IgE-FAB has been shown to decrease after AIT and modestly correlates with clinical response to grass and birch AIT (Table S1) (46, 67). One study showed that increases in serum inhibitory activity for IgE-FAB persisted for 2 years (63). An inverse correlation has been found between symptom scores, rescue medication scores and IgE-FAB (6, 69). To date, there are no data available on the relationship between levels of serum inhibitory activity for IgE-FAB in responders *vs* nonresponders to immunotherapy.

Although the assay is reproducible, it is complex and might be limited to specialized centres or laboratories. Recently, an alternative less complex test has become available (65) – the enzyme-linked immunosorbent-facilitated antigen binding (ELIFAB) assay, a cell-free assay that substitutes EBV-transformed B-cell lines with soluble CD23 monomers bound to a solid surface. The assay follows the basic principles of a standard ELISA protocol using a 96-well plate.

Advantages

- An association between symptom scores, rescue medication scores and IgE-BF has been demonstrated in several studies.
- The IgE-FAB assay is a serum-based assay that is highly reproducible.
- Enzyme-linked immunosorbent-facilitated antigen binding is commercially available, as an alternative test that may be applicable in both research and clinical settings.
- An association between IgE-FAB and symptom and rescue medication scores has been demonstrated in some studies.

Disadvantages and critical issues

- IgE-BF has limited availability, as the Advia Centaur instrument is no longer produced.
- No data on responders vs nonresponders in relation to IgE-FAB have been published.
- There are only limited data exploring the correlation between IgE-FAB and the clinical response to AIT.

Unmet needs and recommendations

- IgE-BF is not a candidate biomarker for clinical use due to the limited availability.
- We recommend strongly that more data be collected on the relationship between IgE-FAB and responders/nonresponders.
- We recommend that IgE-FAB is further evaluated as a surrogate/predictive biomarker for AIT.

Domain iv: Basophil activation

Background and study analyses

Basophils represent 1% of leucocytes in peripheral blood and contain cytoplasmic secretory granules. They are considered as easily accessible cells that share functional characteristics with mast cells and have their own role in systemic allergic responses (70, 71). After allergen cross-linking of specific IgE on basophils, degranulation is induced with release of histamine, leukotrienes and other mediators of the allergic inflammatory response (72, 73). Allergen immunotherapy has been associated with inhibition of basophil activation, and this is achieved via allergen-specific IgG antibodies. Allergen-specific IgG has potential to compete with IgE for allergen binding, thereby preventing allergen-IgE receptor cross-linking on basophils. Alternatively, allergen-IgG complexes may act by triggering basophil surface inhibitory IgG receptors FcyRIIb adjacent to IgE receptors, thereby inhibiting downstream IgE receptor activation (46, 67, 74).

A number of assays to monitor basophil activation are available and are important for allergy diagnosis, particularly in drug hypersensitivity (75, 76). Determination of basophil activation by measuring histamine release or other mediators such as leukotrienes and platelet-activating factor can be complex and time-consuming. Multicolour flow cytometry of basophil surface markers in whole blood enables the evaluation of basophil activation in the presence of potential inhibitory factors including allergen-specific IgGs. CD63 is most commonly used: it detects degranulation of basophils as the epitope is localized within granular membranes and becomes surface exposed upon fusion of the granules with the basophil surface membrane (77). CD203c is an alternative surface marker that not only detects basophil degranulation, but is also a highly selective marker for basophils in peripheral blood. CD203c is located directly underneath the plasma membrane and is induced rapidly on the outside of the plasma membrane after activation (78). Other less frequently used markers for basophil activation are CD13, CD107a and CD164. CD13 and CD164 follow a CD203c pattern, whereas CD107a follows more closely a CD63 pattern of detection by flow cytometry (79). A recently developed reverse staining technique for basophil activation involves measurement by flow cytometry of intracellular phycoerythrin-conjugated diamine oxidase (DAO) that detects intracellular histamine, the natural substrate of DAO. Degranulation of basophils results in a decrease in intracellular DAO corresponding to release of histamine from the cell (80).

The results obtained with basophil activation during AIT in placebo-controlled studies are conflicting. Some authors describe reduction in basophil activation following AIT that is possibly due to serological factors (69, 81–83). Other studies failed to demonstrate suppression of basophil activation in successful trials of AIT (37, 41). These contrasting findings may be partly explained by the route of immunotherapy, with SLIT being possibly less effective in inhibiting basophils than SCIT. Several studies show that basophil activation decreases after AIT, not only at the level of CD63 or CD203c, but also as measured by decreased DAO and increased CD107a (84–88). One study using DAO has shown persistent basophil suppression in four subjects 12–24 months after stopping AIT (84).

Advantages

- *Ex vivo* basophil activation with the sensitizing allergen reflects the FcyRI-mediated *in vivo* response.
- Requires small amount of blood (<2 ml) to perform the test.

Disadvantages and critical issues

- Basophil responses after AIT are variable with inhibition being shown in some but not all studies.
- Only a limited number of studies of basophil activation are yet available.
- Handling viable basophil cells is technically more challenging than determination of factors in serum.
- Dose–response curves are needed for accurate interpretation of results.
- Five to ten percent of population show no basophil response to IgE cross-linking.

Unmet needs and recommendations

- There is a need to understand the mechanism of allergeninduced basophil hyporesponsiveness during AIT.
- Standardized assays are needed. This applies to markers for accurate selection of basophil selection as well as markers of activation and histamine release.

Domain v: Cytokines and chemokines

Background and study analyses

One postulated mechanism of long-term clinical tolerance following AIT is a shift from a dominant Th2 response towards a Th1 response (13, 15). Hence, from the current state of the art, one would expect down-regulation of Th2 cytokines (IL-4, IL-13, IL-9), of inflammatory cytokines and chemokines such as IL-17, eotaxin or TNF- α , and up-regulation of Th1 (IFN-y, IL-12) and regulatory cytokines (IL-10, TGF- β). In reality, some studies report increases in Th1 cytokines and chemokines, paralleled by an up-regulation of Th1 (IFN-y, IL-12) and regulatory cytokines (IL-10, TGF- β), for example (89–94); others report no changes (95, 96). Furthermore, no clear relationship between serum cytokines and clinical outcome of AIT has been demonstrated. Besides addressing interleukins, numerous studies during AIT investigated chemokines CCR3 (unchanged) and CCR4 (97) (increased) and other original serum markers, like adiponectin (98) (unchanged), apolipoprotein A-IV (99) (increased), beta thromboglobulin (100, 101) (unchanged), complement factors C3a and 5a (102, 103) (decreased), C4a (99) (increased), ECP (104) (unchanged), eotaxin (97, 105, 106) (increased/decreased), soluble HLA molecules (107) (unchanged), leptin (98) (unchanged or increased), signalling lymphocytic activation molecule (108) (increased), thymus and activation-regulated chemokine (TARC) (97) (increased), TRAIL (109) (reduced), transthyretin (110) (increased) to tryptase (105) (unchanged). Importantly, none of these markers showed any correlation with the clinical response. It is thus likely that the changes in serum cytokines and chemokines are immunological paraphenomena of AIT that do not directly correlate with clinical outcome. Local rather than serum levels of cytokines may be more indicative of immunological and clinical effects of AIT, but few studies of local cytokines have been performed (105, 111). For example, a cross-sectional study demonstrated lower concentrations of Th2 cytokines (IL-4, 5, 9 and 13) and chemokines (eotaxin) in local nasal fluid at 2–8 h after nasal allergen provocation following successful AIT compared to untreated controls (105).

Advantages

- These assays explore mechanisms of AIT.
- These assays may be useful for proof of concept at early stages of drug development.

Disadvantages and critical issues

- The low frequency of allergen-specific T cells dilutes the cytokine signal in the pool of cytokines secreted from T cells with other specificities.
- So far, no cytokines or chemokines have been identified that predict the clinical outcome in individual patients before the onset of AIT.
- Results are inconsistent: further studies of local nasal cytokines during AIT are required.

Unmet needs and recommendations

- At this stage, cytokines and chemokines are not applicable as a biomarker. However, nasal cytokines can serve as a marker of the immunological response and be used for proof of concept in drug development.
- Local cytokine production and secretion following allergen challenge may provide increased treatment-associated signals.
- Cytokines secreted from epithelial cells may reflect more closely the condition at the site of inflammation.

Domain vi: Cellular markers

Allergen immunotherapy has been associated with the induction of cellular responses within regulatory T cells (Tregs), regulatory B cells (Bregs) and dendritic cells (DCs). Immunological tolerance induction has been shown to be characterized by the up-regulation of peripheral and local allergenspecific regulatory T (Treg) cells (16, 93, 112-115). Tregs can be grouped into two subsets: (i) Foxp3+ regulatory T cells (nTregs) and (ii) inducible regulatory T cells (iTregs) that produce regulatory cytokines such as IL-10, TGF-B and IL-35 (93, 116, 117). Several studies have reported the immunomodulating properties of both allergen-specific nTregs and iTregs, suggesting that there is an overlap between these subsets of Tregs (118). The early induction of Tregs during AIT has been associated with delayed immune deviation from a Th2-pattern response to a Th1-type response. The association of increased numbers of Tregs in the nasal mucosa after AIT with clinical efficacy and the suppression of seasonal allergic inflammation supports the concept of a role for Tregs in the induction of allergen-specific tolerance (114, 115). A recent study investigating the epigenetic modification of memory Tregs during dual house dust

mite (HDM) and grass pollen SLIT indicated that methylation of the FOXP3 locus might be involved in the mechanism of allergy tolerance after AIT (119).

B cells contribute to immune responses through antigen presentation to T cells, secretion of cytokines and production of antibodies after differentiation to plasma cells. Following receipt of the appropriate signals, plasma cells can reside for many years in the bone marrow and continuously produce antibodies independent of exposure to antigen. Upon activation, IgM+IgD+ naïve B cells undergo class switch recombination (CSR) leading to the expression of IgA, IgG or IgE antibodies. IL-10 suppresses antigen presentation through down-regulation of class II major histocompatibility complex molecules and costimulatory molecules on antigen-presenting cells. Furthermore, IL-10 suppresses the production of proinflammatory chemokines and cytokines. In parallel, IL-10 enhances the survival, proliferation, differentiation and isotype switching of human B cells. IL-10 augments IgG₄ production, and along with IFN- γ , it inhibits IL-4-induced IgE CSR (118). IL-10-mediated immunosuppressive functions of B cells have been described in murine models of autoimmunity, infection and cancer. Bregs expressing IL-10 suppress immune responses and the lack or loss of Bregs leads to exacerbated symptoms in several experimental autoimmune diseases (121). In addition, IL-10-overexpressing B cells produced less IgE and show a general ability to suppress T cells and DCs (122).

Dendritic cells are specialized antigen-presenting cells capable to of integrating a variety of incoming signals and subsequently orchestrating adaptive immune responses. Dendritic cells can either initiate and sustain allergic inflammation, or support tolerance induction. Molecular markers associated with polarized monocyte-derived DCs supporting the differentiation of either effector Th1, Th2, Th17 or regulatory CD4⁺ T cells (termed DC1, DC2, DC17 and DCreg, respectively) have been identified by comparative transcriptomic and proteomic analyses (123, 124). Using such markers, recent AIT studies have documented a significant impact of SLIT on blood DCs. Specifically, 4 months of SLIT has been shown to up-regulate C1Q and stabilin DCreg markers while down-regulating DC2-associated markers such as CD141 in PBMCs from grass pollen-allergic patients (123). Importantly, such molecular alterations were found only in PBMCs from patients exhibiting a significant decrease in rhinoconjunctivitis symptoms, providing further corroboration at the level of the innate immune system for the paradigm that a reorientation of immune responses from a Th2 to a regulatory profile is critical to the success of AIT (123). Interestingly, the C1Q molecule itself, which can be secreted or expressed at the surface of monocyte-derived cells, was shown to be a strong inhibitor of Th2 responses in a murine asthma model. The latter observation suggests that DCregs induced during AIT not only support the differentiation of Tregs, but also mediate a direct anti-inflammatory activity by themselves.

In agreement with these findings, SLIT for 1 year in HDM-allergic children resulted in a decrease in the capacity of monocyte-derived DCs to mature in the presence of

Lipopolysachariden, with a blunted expression of CD86, a low production of IL-12 and an increased IL-10 secretion, consistent with their acquisition of a tolerogenic phenotype (125). Another study reported induction of PD-L1 (programmed cell death ligand 1) and IL-10 in parallel with a reduction in CD80 and CD86 expression on antigen-presenting cells during SLIT in ragweed-allergic patients (126). Also, DCs retrieved from the blood of peanut-allergic patients after 2 years of oral immunotherapy significantly down-regulated Foxp3 CpG methylation when cultured with T lymphocytes, suggesting the induction of DCregs (127).

Collectively, the latter studies confirm a significant and persisting impact of AIT on blood DCs, and suggest that changes in markers associated with DCreg and DC2 cells can be used to detect the early onset of AIT in grass pollen-allergic patients.

Advantages

- Tregs appear to play a key role in the immunological processes of AIT, mainly skewing the Th2 to Th1 immune response.
- It appears that a change in allergen-specific B cells in the direction of Breg cells is one of the major alterations in the course of AIT.
- Markers associated with DC polarization have been identified, which can be monitored in blood by quantitative PCR.
- Changes in such markers could represent an early signature within the innate immune system of the subsequent orientation of adaptive immune responses.
- In contrast with circulating CD4⁺ T cells, which are constantly migrating to tissues, DCs might provide a more persistent signature in the blood of a transition from a Th2 to a regulatory immune response during AIT.

Disadvantages and critical issues

- No specific marker exists for Tregs this means they are difficult to detect without sophisticated experimental approaches (and this cannot be performed routinely).
- There are not enough data to link the appearance or function of Tregs with clinical efficacy.
- Tregs appear very early in the treatment long before it is possible to evaluate clinical efficacy, so it remains difficult to use Tregs as a predictive biomarker in AIT in the absence of analysis of responders *vs* nonresponders.
- The frequency of allergen-specific T and B cells is very low: it is technically challenging and currently impossible to use this in clinical practice.
- Although Bregs can be characterized, this requires sophisticated experimental approaches and cannot be performed routinely.
- Dendritic cell-associated candidate markers of efficacy have been identified in a single short-term SLIT study which requires corroboration before it could be adopted in practice.
- The expression of some of DC-associated markers is shared with other leucocyte subsets (e.g. T or NK cells).

• Changes during AIT have been documented at the level of monocytes and monocyte-derived DCs. No information is available regarding the impact of AIT on myeloid and plasmacytoid DCs.

Unmet needs and recommendations

- At this stage, neither Tregs nor Bregs can serve as biomarkers for monitoring AIT. However, they may be useful in drug development as a marker of immunological response. Future determination of AIT-responsive phenotypes and analysis windows is critical for the practicability of cell-based biomarkers.
- Results remain to be validated in field studies, in the context of natural allergen exposure, in large cohorts of patients allergic to grass, tree pollens or HDM.
- We recommend further study of the impact of AIT on myeloid and plasmacytoid DCs in blood as well as tissues.

Domain vii: In vivo biomarkers

Allergen provocation tests are frequently used in clinical practice to evaluate patients' allergen-specific reactivity in diseaseaffected organs in order to demonstrate the clinical relevance of the underlying IgE-mediated sensitization (4). However, these tests can also be used as *in vivo* methods for stratification of patients for clinical trials as well as for investigating the effects of therapeutic interventions in AIT trials (4).

In the current European Medicinal Agency (EMA) guideline, provocation tests are recommended for proof of concept or phase II dose-finding trials in AIT (24). Therefore, several AIT trials have already included these models as primary objectives (examples in Refs 7, 128-131; reviewed in Refs 1, 132). Allergen provocation tests include skin prick tests, intradermal tests (ID) and direct evaluation of target organ responses with conjunctival provocation tests, nasal provocation tests (NPT) and environmental exposure chambers (EEC). Several protocols have been published using different challenge models (133). As outlined in the EAACI PP on 'Recommendations for the standardization of clinical outcomes used in AIT trials for allergic rhinoconjunctivitis', there is a clear unmet need for thorough harmonization and further validation of the different provocation models (1).

During NPT, subjective nasal symptom scores can be supplemented by objective measurements of peak nasal inspiratory flow (134). Nasal early responses have been shown to be inhibited (135) following intralymphatic cat and epicutaneous grass AIT (136). Nasal provocation test allow evaluation of local cells, mediators and cytokines in nasal fluid. Creticos showed suppression of nasal eosinophil numbers (135, 137, 138) and inflammatory mediators (histamine, TAME esterase during nasal provocation after ragweed immunotherapy). Suppression of the late nasal response by grass pollen (135) and ragweed AIT was associated with decreases in IL-4 mRNA+ cells and increases in IFN- γ mRNA+ cells in nasal biopsies. Recent advances include the use of more precise nasal allergen delivery devices (139) and the availability of synthetic materials for filters, sponges, etc., that enable collection of neat or minimally diluted nasal fluid directly from the nasal mucosa (139, 140). The parallel development of miniaturized assay systems has allowed the reproducible measurement of multiple mediators, cytokines and antibodies in nasal fluid volumes as low as 20-50 µl. Nasal provocation has been shown to result in early increases in local nasal fluid tryptase (at 5-30 min) and later increases in chemokines and Th2 cytokines (eotaxin, IL-4, 5, 9 and 13) and innate lymphoid 2 cells (ILC2s) that parallel the late nasal response. Grass pollen AIT was associated with blunted increases in tryptase, eotaxin and Th2 cytokines (IL-4, 5, 9 and 13) in nasal fluid compared to untreated allergic controls (105). In addition to local suppression of type 2 allergic responses (204), grass pollen AIT has been associated with suppression of systemic basophil activation (84) following nasal provocation, as shown by a decrease in surface CD63 and intracellular DAO, compared to untreated allergic controls.

The EEC represents a recent alternative to NPT that more closely simulates natural exposure. Several recent studies have utilized the EEC to evaluate time-of-onset studies of AIT (141, 142). One study showed a correlation between symptoms provoked in the EEC compared to natural seasonal exposure (143).

For (pivotal) phase III trials, the EMA guideline (24) highlights that 'Provocation tests in allergen chambers [are] deemed to be a promising tool for the evaluation of efficacy; however, the results of such provocations have to be validated (...)'. An increasing number of EEC have been thoroughly investigated and validated regarding stability and reproducibility of allergen exposure under standardized environmental conditions (144). For AIT, there is a clear unmet need for further validation of treatment effect size as evaluated in EEC challenges to be correlated with effect sizes found under natural exposure in field trials (144). Pending the results, EEC models may become promising candidates in evaluating '*in vivo*' biomarkers in adjunct to natural exposure (1).

Advantages

- Provocation tests (i.e. titrated mucosal challenges) may indicate a change in responsiveness to allergen and/or a change in allergen sensitivity following AIT.
- Provocation tests permit more standardized procedures and the ability to control environmental factors (temperature, humidity) and avoid the variability caused by seasonal variations in pollen exposure.
- Provocation tests have been used as surrogate markers of clinical response to AIT. They are recommended for understanding mechanisms and permit biomarker discovery both at local level and in peripheral blood.
- They permit accurate time-course and dose-response studies, are less expensive to perform, require fewer participants than field studies and are often completed in a single centre, thereby reducing variability of outcome measures.

• European Medicines Agency recommends provocation methods as primary endpoints in proof-of-concept and dose-finding trials of AIT.

Disadvantages and critical issues

- Allergen provocation is not the same as natural exposure: standardization and validation vary for the different challenge protocols.
- Therefore, regulators do not accept replacement of natural allergen exposures by provocation challenges as primary endpoints in pivotal phase III trials.
- Conjunctival provocation test comprises mostly subjective outcomes, and there are no standardized/harmonized scoring methods.
- Different objective methods to measure nasal obstruction after NPT are currently used. Standardization is needed.
- Allergen products for provocation testing require regulatory approval and are not always available internationally for standardization purposes.
- Although environmental chamber studies are attractive, the procedure is expensive and standardization and confirmation of reproducibility within or between sites is required.
- Intradermal tests do not necessarily correlate with improvement of symptoms.

Unmet needs and recommendations

- Comparisons between provocation test results and symptoms evoked under natural exposure should be evaluated (204).
- In AIT, treatment effect sizes as evaluated in provocation tests should be compared with treatment effect sizes as evaluated under natural allergen exposure.
- Meanwhile, provocation tests provide proof of concept for novel approaches and are useful to assess time to onset of effect of AIT. The EMA recognizes their use for allergen dose-finding studies (phase II) before further investigation in (pivotal) phase III AIT trials.
- Provocation tests cannot substitute for assessing symptoms and requirements for rescue medication during natural allergen exposure in phase III trials.
- Pending standardization and clinical validation, EEC are likely to be an optional adjunct to natural exposure studies for phase III trials of AIT.
- Provocation tests should be tightly linked to the local and systemic biomarker assessments described above.

Discussion

Biomarkers, in the context of AIT, are defined as quantitative measurements that can predict clinical and immunological responses during treatment and could assist in patient selection, identification of responders, target intervention of those who will benefit and to exclude those who are less likely to respond to AIT as well as efficacy monitoring during intervention. Biomarkers for AIT would thus facilitate the introduction of personalized medicine in allergy. Furthermore, they could be of assistance in clinical trials for the development of treatment modalities. An overview and recommendations for the standardization of clinical outcomes used in AIT are available (1), but to date there is no consensus on candidate biomarkers that are predictive of the clinical response to AIT (22). Although several biomarkers such as sIgG4, IgE-BF or IgE ratio have been included as secondary measurements in AIT studies, there are only very limited data on the relationship between biomarkers and clinical response vs nonresponse. This EAACI Task Force PP presents an overview of biomarkers tested in AIT trials in relation to clinical outcome. It emphasizes the pros and cons of different biomarkers and, finally, gives recommendations on the use of biomarkers in future research and AIT trials. It is important to note that currently available biomarkers are experimental and are confined to research and AIT trials. There is no evidence they may predict responses in individual patients in a clinical setting, but this is the ultimate goal of biomarker development.

We applied the definition of biomarkers provided by ICH E15 guidance on 'Definitions for Genomic Biomarkers, Pharmacogenomics, Pharmacogenetics, Genomic Data and Sample Coding Categories', which states that a biomarker is 'an indicator of normal biologic processes, pathogenic processes and/or response to therapeutic or other interventions' (22). European Medicines Agency advises in the 2008 guide-lines to include immunological changes (e.g. changes in allergen-specific IgG levels, T-cell responses and/or cytokine production) and/or modifications of the end-organ specific response (e.g. provocation tests) in pharmacokinetic and dynamic studies (24). No advice is provided on the use of immunological changes as a predictive biomarker.

Humoral changes are included in many AIT clinical trials as secondary outcome measures (e.g. IgE, IgG₄, IgA). Several pollen AIT studies have reported transient increase in the levels of specific IgE but no functional relevance or severe allergic reactions have been associated with this transient increase in sIgE (31, 32). The ratio of sIgE to total IgE (sIgE/tIgE ratio) is more promising as a predictive marker (33). With a cut-off value of 16.2%, the sIgE/ tIgE ratio predicted the successful outcome of AIT revealing a sensitivity of 97.2% and specificity of 88.1%. A limited number of studies showed a similar correlation between sIgE/tIgE ratio and clinical outcome of AIT; one open-label study could not replicate these data (34-36). We therefore recommend that more studies, including prospective cohort studies, should include the sIgE/tIgE ratio and correlate with responders and nonresponders. For sIgG₄, a small number of studies demonstrate correlation between allergen sIgG₄ and clinical outcomes (41-44). Most studies, however, have shown an increase in IgG₄ soon after the initiation of AIT which is followed by a decrease after cessation of treatment. The increase in the IgG₄ levels after AIT is likely to indicate an immunological response following allergen exposure during treatment and could potentially be used as a marker of therapy compliance reflecting the standardized preparation of the vaccine and effective administration.

The serum inhibitory activity of IgE following AIT has been known for many years and includes IgE-BF and IgE-FAB (46, 61). Although several studies have confirmed that IgE-BF increases after AIT, the limited availability of this assay means that this surrogate biomarker cannot be widely used in clinical practice or clinical trials. Two tests are available for IgE-FAB, a flow cytometry-based bioassay and ELIFAB. Enzyme-linked immunosorbent-facilitated antigen binding can easily be used in clinical settings, whereas the IgE-FAB assay may be better for clinical trials. IgE-FAB has been shown to decrease after AIT and remains decreased even after discontinuation of AIT (46, 63, 67). We recommend that IgE-FAB is explored further as a biomarker in clinical trials and in prospective cohort studies.

Cellular responses following AIT, including cytokines and chemokines, have been reported in several studies. Investigating cellular responses requires highly technically skilled personnel to perform the assays. To date, no cellular marker or cell-derived serum marker has been identified that would be useful in clinical practice. However, studies on cellular responses are of utmost importance in our quest to understand the underlying mechanisms of AIT and identify novel biomarkers. We recommend that the use of cellular markers is limited to clinical trials and mechanistic studies of AIT.

Environmental chambers and provocation tests are included here as they can be combined with the evaluation of local cells, mediators or cytokines (4). The current EMA guidelines already recommend provocation tests as a proof of concept in phase II dose-finding trials (24). This resulted in several AIT trials that included provocation testing in analysing their primary objectives (7, 128-131). There is, however, a clear unmet need for thorough harmonization and further validation of different provocation models (1, 133). Some data are available on local cytokines in NPTs showing that Th2 cytokine responses are blunted after AIT (204). Data so far are very limited; we recommend that protocols of provocation tests are harmonized and that studies that include provocation tests also include serological biomarkers. For example, provocation tests could be combined with sIgE/tIgE ratios or IgE-FAB.

So far, EMA guidelines advise inclusion of immunological changes in pharmacokinetic and dynamic studies only in order to show the effect of AIT on the immune system and they recommend provocation tests for assessing proof of principle (24). There is an urgent need for standardizing the use of potential biomarkers that are related to clinical outcome and reflect the immunogenicity of vaccines in inducing clinical and immunological tolerance. We propose that measurements of $sIgG_4$, IgE-FAB, sIgE/tIgE ratio and local cellular responses are investigated and implemented in future guidelines for registration of novel vaccines.

Conclusions

To date, there are no validated and generally accepted candidate biomarkers that are predictive or indicative of the clinical response to AIT. Although several studies include biomarkers as secondary outcomes, current guidelines do not include biomarkers in the recommendations for clinical trials or clinical response. Therefore, this PP on biomarkers in AIT, as proposed by the EAACI Immunotherapy Interest Group, has reviewed all of the candidate biomarkers used in clinical trials of AR patients with or without asthma and grouped them into seven related domains. All biomarkers have been reviewed in the light of their potential advantages as well as their respective drawbacks. Furthermore, unmet needs and specific recommendations in all seven domains have been addressed. In order to raise the evidence level for candidate biomarkers from each domain, it is critical to conduct biomarker studies with a novel approach in design (i.e. responders vs nonresponders) and determine their clinical relevance as surrogate or predictive markers of the efficacy of AIT. In the light of the evidence above, this EAACI PP recommends exploration of the use of allergen-specific sIgG4 as a biomarker for compliance. Candidate biomarkers for clinical outcome are sIgE/tIgE ratio and IgE-FAB: more studies are needed to confirm and to interpret their association with the clinical response to immunotherapy and how they relate to persistence of clinical benefit after discontinuation of immunotherapy.

Acknowledgments

The authors would like to thank PS Andersen (ALK), P Moingeon (Stallergenes), M Kramer and T Higgenbottam (Allergy Therapeutics), A Nandy and C Willer (Allergopharma) for their input. The TF was financed by EAACI. The authors would like to thank EAACI for their financial support in the development of this TF report.

Conflicts of interest

Dr. Shamji reports grants from Immune Tolerance Network, NIAID, during the conduct of the study; grants from Regeneron, USA, grants from Biotech Tools, personal fees from ALK, Horsholm, Denmark, personal fees from ASIT Biotech inc., outside the submitted work; Dr. Kappen reports personal fees from ALK, personal fees from Chiesi, personal fees from GSK, outside the submitted work; Drs Kappen, Akdis, Jensen-Jarolim and Knol have nothing to disclose. Dr Jörg Kleine-Tebbe reports grants from Circassia, UK; Leti, Germany; Stallergenes. He also reports receiving lecture fees from Allergopharma, ALK-Abelló, Bencard, HAL Allergy, LETI, Lofarma, Novartis, Stallergenes. He has board membership for ALK-Abelló Advisory Board, Novartis Advisory Board, Leti Advisory Board, Bencard Advisory Board. He has received consultancy fees from MERCK, US and Circassia, UK. Dr. Bohle reports grants from Austrian Science Funds, grants from Christian Doppler Society, during the conduct of the study. Dr. Chaker reports consultancy and speaker arrangements via TUM with ALK-Abello, grants and other from Allergopharma, consultancy arrangements via TUM from Lofarma and HAL Allergy, clinical

trial arrangements via TUM with Bencard/Allergy Therapeutics and ASIT Biotech, grants and other from Novartis, grants from German Federal Environmental Agency, grants from Zeller AG, grants and other from LETI, outside the submitted work. Dr. Till reports personal fees from ALK-Abello, outside the submitted work. Dr Valenta has received research grants from Biomay AG, Vienna, Austria, Thermo-Fisher, Uppsala, Sweden and Fresenius Medical Care, Bad Homburg, Germany, and serves as a consultant for these companies. Dr. Poulsen reports grants and personal fees from ALK, grants from Anergis, grants from EU Commission, outside the submitted work. Dr. Calderon has received consultancy fees from ALK-Abelló and HAL Allergy and has received lecturing fees from ALK-Abelló, Merck, and Stallergenes Greer. Dr. Demoly reports personal fees from ALK, personal fees from Stallergénes, personal fees from BTT, personal fees from Chiesi, personal fees from ThermoFisherScientific, personal fees from Meda, personal fees from Yslab, personal fees from AstraZeneca, personal fees from Ménarini, outside the submitted work; Dr. Pfaar reports grants and personal fees from ALK-Abelló, grants and personal fees from Allergopharma, grants and personal fees from Stallergenes Greer, grants and personal fees from HAL Allergy Holding B.V./HAL Allergie GmbH, grants and personal fees from Bencard Allergie GmbH/Allergy Therapeutics, grants and personal fees from Lofarma, grants from Biomay, grants from Nuvo, grants from Circassia, grants and personal fees from Biotech Tools S.A., grants and personal fees from Laboratorios LETI/LETI Pharma, personal fees from Novartis Pharma, personal fees from MEDA Pharma, grants and personal fees from Anergis S.A., personal fees from Sanofi US Services, personal fees from Mobile Chamber Experts (a GA²LEN Partner), personal fees from Pohl Boskamp, outside the submitted work. Dr. Jacobsen reports and Secretary EAACI immunotherapy interest group Speaker for ALK, ThermoFischer. Dr. Durham reports grants from Immune Tolerance Network, NIAID, nonfinancial support from ALK, Horsholm Denmark, during the conduct of the study; grants from Regeneron, USA, grants from Biotech Tools, grants from ALK, Horsholm, Denmark, personal fees from Anergis, Switzerland, personal fees from Circassia, UK, personal fees from Biomay, Austria, personal fees from Merck, personal fees from Allergy Therapeutics, UK, personal fees from ALK, Horsholm, Denmark, personal fees from med Update GmbH, Germany, outside the submitted work. Dr. Schmidt-Weber C Schmidt-Weber reports grants from Allergopharma, Leti, Regeneron; is member of the scientific advisory board of Leti; consultant for PLS-Design, Allergopharma, Leti; shares of PLS-Design.

Supporting Information

Additional Supporting Information may be found in the online version of this article:

Table S1. Summary of study results.

Table S2. Study overview.

References

- Pfaar O, Demoly P, Gerth van Wijk R, Bonini S, Bousquet J, Canonica GW et al. Recommendations for the standardization of clinical outcomes used in allergen immunotherapy trials for allergic rhinoconjunctivitis: an EAACI position paper. *Allergy* 2014;69:854-867.
- Jutel M, Agache I, Bonini S, Burks AW, Calderon M, Canonica W et al. International consensus on allergy immunotherapy. J Allergy Clin Immunol 2015;136:556-568.
- Bousquet J, Van Cauwenberge P, Khaltaev N, Aria Workshop G, World Health O. Allergic rhinitis and its impact on asthma. *J Allergy Clin Immunol* 2001;108(5 Suppl.):S147-S334.
- Bousquet J, Khaltaev N, Cruz AA, Denburg J, Fokkens WJ, Togias A et al. Allergic rhinitis and its impact on asthma (ARIA) 2008 update (in collaboration with the World Health Organization, GA(2)LEN and Aller-Gen). *Allergy* 2008;63(Suppl. 86):8-160.
- Bousquet J, Lockey R, Malling HJ. Allergen immunotherapy: therapeutic vaccines for allergic diseases. A WHO position paper. J Allergy Clin Immunol 1998;102: 558-562.
- Shamji MH, Ljorring C, Francis JN, Calderon MA, Larche M, Kimber I et al. Functional rather than immunoreactive levels of IgG4 correlate closely with clinical response to grass pollen immunotherapy. *Allergy* 2012;67:217-226.
- Durham SR, Walker SM, Varga EM, Jacobson MR, O'Brien F, Noble W et al. Long-term clinical efficacy of grass-pollen immunotherapy. *N Engl J Med* 1999;**341**:468-475.
- Calderon MA, Alves B, Jacobson M, Hurwitz B, Sheikh A, Durham S. Allergen injection immunotherapy for seasonal allergic rhinitis. *Cochrane Database Syst Rev* 2007;CD001936.
- Radulovic S, Calderon MA, Wilson D, Durham S. Sublingual immunotherapy for allergic rhinitis. *Cochrane Database Syst Rev* 2010:CD002893.
- Radulovic S, Wilson D, Calderon M, Durham S. Systematic reviews of sublingual immunotherapy (SLIT). *Allergy* 2011;66:740-752.
- Calderon MA, Penagos M, Sheikh A, Canonica GW, Durham S. Sublingual immunotherapy for treating allergic conjunctivitis. *Cochrane Database Syst Rev* 2011;CD007685.
- Compalati E, Braido F, Canonica GW. An update on allergen immunotherapy and asthma. *Curr Opin Pulm Med* 2014;20:109-117.

- Shamji MH, Durham SR. Mechanisms of immunotherapy to aeroallergens. *Clin Exp Allergy* 2011;41:1235-1246.
- Matsuoka T, Shamji MH, Durham SR. Allergen immunotherapy and tolerance. *Allergol Int* 2013;62:403-413.
- Akdis CA, Akdis M. Mechanisms of allergen-specific immunotherapy and immune tolerance to allergens. *World Allergy Organ* J 2015;8:17.
- Akdis M, Akdis CA. Mechanisms of allergen-specific immunotherapy: multiple suppressor factors at work in immune tolerance to allergens. J Allergy Clin Immunol 2014;133:621-631.
- Calderon MA, Penagos M, Sheikh A, Canonica GW, Durham SR. Sublingual immunotherapy for allergic conjunctivitis: Cochrane systematic review and meta-analysis. *Clin Exp Allergy* 2011;41:1263-1272.
- Galli SJ. Toward precision medicine and health: Opportunities and challenges in allergic diseases. J Allergy Clin Immunol 2016;137:1289-1300.
- Burks AW, Calderon MA, Casale T, Cox L, Demoly P, Jutel M et al. Update on allergy immunotherapy: American Academy of Allergy, Asthma & Immunology/ European Academy of Allergy and Clinical Immunology/PRACTALL consensus report. J Allergy Clin Immunol 2013;131:1288-1296.
- Calderon MA, Casale T, Cox L, Akdis CA, Burks AW, Nelson HS et al. Allergen immunotherapy: a new semantic framework from the European Academy of Allergy and Clinical Immunology/American Academy of Allergy, Asthma and Immunology/ PRACTALL consensus report. *Allergy* 2013;68:825-828.
- Calderon MA, Bernstein DI, Blaiss M, Andersen JS, Nolte H. A comparative analysis of symptom and medication scoring methods used in clinical trials of sublingual immunotherapy for seasonal allergic rhinitis. *Clin Exp Allergy* 2014;44:1228-1239.
- Food, Drug Administration HHS. International Conference on harmonisation; guidance on E15 pharmacogenomics definitions and sample coding; availability. Notice. *Fed Regist* 2008;73:19074-19076.
- Food, Drug Administration HHS. International conference on harmonisation; guidance on E16 biomarkers related to drug or biotechnology product development: context, structure, and format of qualification submissions; availability. Notice. *Fed Regist* 2011;**76**:49773-49774.
- 24. CHMP EMA Guideline on the Clinical Development of Products for Specific

Immunotherapy for the Treatment of Allergic Diseases. 2008. No separate publisher

- 25. International Conference on Harmonisation of Technical Requirements for registration of pharmaceuticals for human use (ICH) adopts consolidated guideline on good clinical practice in the conduct of clinical trials on medicinal products for human use. *Int Dig Health Legis* 1997;48:231-234.
- Atkinson AJ, Colburn WA, DeGruttola VG, DeMets DL, Downing GJ, Hoth DF et al. Biomarkers and surrogate endpoints: Preferred definitions and conceptual framework. *Clin Pharmacol Ther* 2001;69:89-95.
- 27. Senti G, Prinz Vavricka BM, Erdmann I, Diaz MI, Markus R, McCormack SJ et al. Intralymphatic allergen administration renders specific immunotherapy faster and safer: a randomized controlled trial. *Proc Natl Acad Sci U S A* 2008;105:17908-17912.
- Witten M, Malling HJ, Blom L, Poulsen BC, Poulsen LK. Is intralymphatic immunotherapy ready for clinical use in patients with grass pollen allergy? J Allergy Clin Immunol 2013;132:1248-1252.
- Senti G, Graf N, Haug S, Ruedi N, von Moos S, Sonderegger T et al. Epicutaneous allergen administration as a novel method of allergen-specific immunotherapy. J Allergy Clin Immunol 2009;124:997-1002.
- Cox L, Nelson H, Lockey R, Calabria C, Chacko T, Finegold I et al. Allergen immunotherapy: a practice parameter third update. J Allergy Clin Immunol 2011;127(1 Suppl.):S1-S55.
- Nouri-Aria KT, Wachholz PA, Francis JN, Jacobson MR, Walker SM, Wilcock LK et al. Grass pollen immunotherapy induces mucosal and peripheral IL-10 responses and blocking IgG activity. *J Immunol* 2004:172:3252-3259.
- 32. Pilette C, Nouri-Aria KT, Jacobson MR, Wilcock LK, Detry B, Walker SM et al. Grass pollen immunotherapy induces an allergen-specific IgA2 antibody response associated with mucosal TGF-beta expression. J Immunol 2007;178:4658-4666.
- 33. Di Lorenzo G, Mansueto P, Pacor ML, Rizzo M, Castello F, Martinelli N et al. Evaluation of serum s-IgE/total IgE ratio in predicting clinical response to allergenspecific immunotherapy. J Allergy Clin Immunol 2009;123:1103-1110.
- 34. Wurtzen PA, Lund G, Lund K, Arvidsson M, Rak S, Ipsen H. A double-blind placebo-controlled birch allergy vaccination study II: correlation between inhibition of IgE binding, histamine release and facilitated allergen presentation. *Clin Exp Allergy* 2008;**38**:1290-1301.

- Li Q, Li M, Yue W, Zhou J, Li R, Lin J et al. Predictive factors for clinical response to allergy immunotherapy in children with asthma and rhinitis. *Int Arch Allergy Immu*nol 2014:164:210-217.
- 36. Eifan AO, Akkoc T, Yildiz A, Keles S, Ozdemir C, Bahceciler NN et al. Clinical efficacy and immunological mechanisms of sublingual and subcutaneous immunotherapy in asthmatic/rhinitis children sensitized to house dust mite: an open randomized controlled trial. *Clin Exp Allergy* 2010;40:922-932.
- 37. Van Overtvelt L, Baron-Bodo V, Horiot S, Moussu H, Ricarte C, Horak F et al. Changes in basophil activation during grass-pollen sublingual immunotherapy do not correlate with clinical efficacy. *Allergy* 2011;66:1530-1537.
- Kober H, Perborn H. Quantitation of mouse-human chimeric allergen-specific IgE antibodies with ImmunoCAP technology. J Allergy Clin Immunol 2006;117:Abstract 845.
- Jutel M, Jaeger L, Suck R, Meyer H, Fiebig H, Cromwell O. Allergen-specific immunotherapy with recombinant grass pollen allergens. *J Allergy Clin Immunol* 2005;**116**:608-613.
- 40. Reisinger J, Horak F, Pauli G, van Hage M, Cromwell O, Konig F et al. Allergenspecific nasal IgG antibodies induced by vaccination with genetically modified allergens are associated with reduced nasal allergen sensitivity. *J Allergy Clin Immunol* 2005;**116**:347-354.
- Gomez E, Fernandez TD, Dona I, Rondon C, Campo P, Gomez F et al. Initial immunological changes as predictors for House Dust Mite immunotherapy response. *Clin Exp Allergy* 2015;45:1542-1553.
- Gehlhar K, Schlaak M, Becker WM, Bufe A. Monitoring allergen immunotherapy of pollen-allergic patients: the ratio of allergen-specific IgG4 to IgG1 correlates with clinical outcome. *Clin Exp Allergy* 1999;**29**:497-506.
- 43. Moverare R, Elfman L, Vesterinen E, Metso T, Haahtela T. Development of new IgE specificities to allergenic components in birch pollen extract during specific immunotherapy studied with immunoblotting and Pharmacia CAP System. *Allergy* 2002;57:423-430.
- Nelson HS, Nolte H, Creticos P, Maloney J, Wu JM, Bernstein DI. Efficacy and safety of timothy grass allergy immunotherapy tablet treatment in North American adults. J Allergy Clin Immunol 2011;127:72-80.
- Jarolim E, Poulsen LK, Stadler BM, Mosbech H, Oesterballe O, Kraft D et al. A long-term follow-up study of

hyposensitization with immunoblotting. J Allergy Clin Immunol 1990;**85**:996-1004.

- 46. van Neerven RJ, Wikborg T, Lund G, Jacobsen B, Brinch-Nielsen A, Arnved J et al. Blocking antibodies induced by specific allergy vaccination prevent the activation of CD4+ T cells by inhibiting serum-IgE-facilitated allergen presentation. J Immunol 1999;163:2944-2952.
- Lupinek C, Wollman E, Valenta R. Monitoring allergen immunotherapy effects by microarray. *Curr Treat Options Allergy* 2016;3:189-203.
- Lupinek C, Wollmann E, Baar A, Banerjee S, Breiteneder H, Broecker BM et al. Advances in allergen-microarray technology for diagnosis and monitoring of allergy: the MeDALL allergen-chip. *Methods* 2014;66:106-119.
- Wollmann E, Lupinek C, Kundi M, Selb R, Niederberger V, Valenta R. Reduction in allergen-specific IgE binding as measured by microarray: A possible surrogate marker for effects of specific immunotherapy. J Allergy Clin Immunol 2015;136:806-809.
- Schmid JM, Wurtzen PA, Dahl R, Hoffmann HJ. Pretreatment IgE sensitization patterns determine the molecular profile of the IgG4 response during updosing of subcutaneous immunotherapy with timothy grass pollen extract. J Allergy Clin Immunol 2016;137:562-570.
- Rispens T, Ooijevaar-de Heer P, Bende O, Aalberse RC. Mechanism of immunoglobulin G4 Fab-arm exchange. J Am Chem Soc 2011;133:10302-10311.
- 52. Aalberse RC, Schuurman J. IgG4 breaking the rules. *Immunology* 2002;**105**:9-19.
- 53. Shamji MH, Wilcock LK, Wachholz PA, Dearman RJ, Kimber I, Wurtzen PA et al. The IgE-facilitated allergen binding (FAB) assay: validation of a novel flow-cytometric based method for the detection of inhibitory antibody responses. *J Immunol Methods* 2006;**317**:71-79.
- 54. Platts-Mills TA, von Maur RK, Ishizaka K, Norman PS, Lichtenstein LM. IgA and IgG anti-ragweed antibodies in nasal secretions. Quantitative measurements of antibodies and correlation with inhibition of histamine release. J Clin Invest 1976;57:1041-1050.
- 55. La Rosa M, Ranno C, Andre C, Carat F, Tosca MA, Canonica GW. Double-blind placebo-controlled evaluation of sublingualswallow immunotherapy with standardized *Parietaria judaica* extract in children with allergic rhinoconjunctivitis. J Allergy Clin Immunol 1999;104:425-432.
- Troise C, Voltolini S, Canessa A, Pecora S, Negrini AC. Sublingual immunotherapy in *Parietaria* pollen-induced rhinitis: a double-

blind study. J Investig Allergol Clin Immunol 1995;5:25-30.

- 57. Lima MT, Wilson D, Pitkin L, Roberts A, Nouri-Aria K, Jacobson M et al. Grass pollen sublingual immunotherapy for seasonal rhinoconjunctivitis: a randomized controlled trial. *Clin Exp Allergy* 2002;**32**:507-514.
- Bahceciler NN, Arikan C, Taylor A, Akdis M, Blaser K, Barlan IB et al. Impact of sublingual immunotherapy on specific antibody levels in asthmatic children allergic to house dust mites. *Int Arch Allergy Immunol* 2005;**136**:287-294.
- Rolinck-Werninghaus C, Kopp M, Liebke C, Lange J, Wahn U, Niggemann B. Lack of detectable alterations in immune responses during sublingual immunotherapy in children with seasonal allergic rhinoconjunctivitis to grass pollen. *Int Arch Allergy Immunol* 2005;**136**:134-141.
- Baron-Bodo V, Horiot S, Lautrette A, Chabre H, Drucbert AS, Danze PM et al. Heterogeneity of antibody responses among clinical responders during grass pollen sublingual immunotherapy. *Clin Exp Allergy* 2013;43:1362-1373.
- Cooke RA, Barnard JH, Hebald S, Stull A. Serological evidence of immunity with coexisting sensitization in a type of human allergy (hay fever). *J Exp Med* 1935;62:733-750.
- Lichtenstein LM, Norman PS, Winkenwerder WL. Antibody response following immunotherapy in ragweed hay fever: Allpyral vs. whole ragweed extract. *J Allergy* 1968;41:49-57.
- 63. Durham SR, Emminger W, Kapp A, de Monchy JG, Rak S, Scadding GK et al. SQ-standardized sublingual grass immunotherapy: confirmation of disease modification 2 years after 3 years of treatment in a randomized trial. J Allergy Clin Immunol 2012;129:717-725.
- 64. Petersen AB, Gudmann P, Milvang-Gronager P, Morkeberg R, Bogestrand S, Linneberg A et al. Performance evaluation of a specific IgE assay developed for the ADVIA centaur immunoassay system. *Clin Biochem* 2004;37:882-892.
- Shamji MH, Francis JN, Wurtzen PA, Lund K, Durham SR, Till SJ. Cell-free detection of allergen-IgE cross-linking with immobilized phase CD23: inhibition by blocking antibody responses after immunotherapy. J Allergy Clin Immunol 2013:132:1003-1005.
- 66. Subbarayal B, Schiller D, Mobs C, de Jong NW, Ebner C, Reider N et al. Kinetics, cross-reactivity, and specificity of Bet v 1specific IgG4 antibodies induced by immunotherapy with birch pollen. *Allergy* 2013;68:1377-1386.

- Wachholz PA, Soni NK, Till SJ, Durham SR. Inhibition of allergen-IgE binding to B cells by IgG antibodies after grass pollen immunotherapy. J Allergy Clin Immunol 2003:112:915-922.
- James LK, Shamji MH, Walker SM, Wilson DR, Wachholz PA, Francis JN et al. Long-term tolerance after allergen immunotherapy is accompanied by selective persistence of blocking antibodies. J Allergy Clin Immunol 2011;127:509-516.
- Schmid JM, Wurtzen PA, Dahl R, Hoffmann HJ. Early improvement in basophil sensitivity predicts symptom relief with grass pollen immunotherapy. J Allergy Clin Immunol 2014;134:741-744.
- Ishizaka T, De Bernardo R, Tomioka H, Lichtenstein LM, Ishizaka K. Identification of basophil granulocytes as a site of allergic histamine release. *J Immunol* 1972:108:1000-1008.
- Falcone FH, Knol EF, Gibbs BF. The role of basophils in the pathogenesis of allergic disease. *Clin Exp Allergy* 2011;41:939-947.
- Schroeder JT, Kagey-Sobotka A, Lichtenstein LM. The role of the basophil in allergic inflammation. *Allergy* 1995;50:463-472.
- MacGlashan D Jr. Expression of CD203c and CD63 in human basophils: relationship to differential regulation of piecemeal and anaphylactic degranulation processes. *Clin Exp Allergy* 2010;**40**:1365-1377.
- Kepley CL, Cambier JC, Morel PA, Lujan D, Ortega E, Wilson BS et al. Negative regulation of FcepsilonRI signaling by FcgammaRII costimulation in human blood basophils. J Allergy Clin Immunol 2000;106:337-348.
- Hoffmann HJ, Santos AF, Mayorga C, Nopp A, Eberlein B, Ferrer M et al. The clinical utility of basophil activation testing in diagnosis and monitoring of allergic disease. *Allergy* 2015;**70**:1393-1405.
- Hoffmann HJ, Santos AF, Mayorga C, Nopp A, Eberlein B, Ferrer M et al. The clinical utility of basophil activation testing in diagnosis and monitoring of allergic disease. *Allergy* 2015;**70**:1393-1405.
- Knol EF, Mul FP, Jansen H, Calafat J, Roos D. Monitoring human basophil activation via CD63 monoclonal antibody 435. *J Allergy Clin Immunol* 1991;88:328-338.
- Buhring HJ, Streble A, Valent P. The basophil-specific ectoenzyme E-NPP3 (CD203c) as a marker for cell activation and allergy diagnosis. *Int Arch Allergy Immunol* 2004;133:317-329.
- Hennersdorf F, Florian S, Jakob A, Baumgartner K, Sonneck K, Nordheim A et al. Identification of CD13, CD107a, and CD164 as novel basophil-activation markers and dissection of two response patterns

in time kinetics of IgE-dependent upregulation. *Cell Res* 2005;**15**:325-335.

- Ebo DG, Bridts CH, Mertens CH, Hagendorens MM, Stevens WJ, De Clerck LS. Analyzing histamine release by flow cytometry (HistaFlow): a novel instrument to study the degranulation patterns of basophils. *J Immunol Methods* 2012;375:30-38.
- Ozdemira SK, Sin BA, Gulogu D, Ikinciogullari A, Gencturk Z, Misirligil Z. Short-term preseasonal immunotherapy: is early clinical efficacy related to the basophil response? *Int Arch Allergy Immunol* 2014;164:237-245.
- 82. Aasbjerg K, Backer V, Lund G, Holm J, Nielsen NC, Holse M et al. Immunological comparison of allergen immunotherapy tablet treatment and subcutaneous immunotherapy against grass allergy. *Clin Exp Allergy* 2014;**44**:417-428.
- Ceuppens JL, Bullens D, Kleinjans H, van der Werf J, Group PBES. Immunotherapy with a modified birch pollen extract in allergic rhinoconjunctivitis: clinical and immunological effects. *Clin Exp Allergy* 2009;**39**:1903-1909.
- 84. Shamji MH, Layhadi JA, Scadding GW, Cheung DK, Calderon MA, Turka LA et al. Basophil expression of diamine oxidase: a novel biomarker of allergen immunotherapy response. J Allergy Clin Immunol 2015;135:913-921.
- Zidarn M, Kosnik M, Silar M, Bajrovic N, Korosec P. Sustained effect of grass pollen subcutaneous immunotherapy on suppression of allergen-specific basophil response; a real-life, nonrandomized controlled study. *Allergy* 2015;**70**:547-555.
- 86. Gokmen NM, Ersoy R, Gulbahar O, Ardeniz O, Sin A, Unsel M et al. Desensitization effect of preseasonal seven-injection allergoid immunotherapy with olive pollen on basophil activation: the efficacy of olive pollen-specific preseasonal allergoid immunotherapy on basophils. *Int Arch Allergy Immunol* 2012;**159**:75-82.
- Lalek N, Kosnik M, Silar M, Korosee P. Immunoglobulin G-dependent changes in basophil allergen threshold sensitivity during birch pollen immunotherapy. *Clin Exp Allergy* 2010;40:1186-1193.
- Nopp A, Cardell LO, Johansson SG, Oman H. CD-sens: a biological measure of immunological changes stimulated by ASIT. *Allergy* 2009;64:811-814.
- Jutel M, Akdis M, Budak F, Aebischer-Casaulta C, Wrzyszcz M, Blaser K et al. IL-10 and TGF-beta cooperate in the regulatory T cell response to mucosal allergens in normal immunity and specific immunotherapy. *Eur J Immunol* 2003;33:1205-1214.

- Faith A, Richards DF, Verhoef A, Lamb JR, Lee TH, Hawrylowicz CM. Impaired secretion of interleukin-4 and interleukin-13 by allergen-specific T cells correlates with defective nuclear expression of NF-AT2 and jun B: relevance to immunotherapy. *Clin Exp Allergy* 2003;33:1209-1215.
- 91. Ebner C, Siemann U, Bohle B, Willheim M, Wiedermann U, Schenk S et al. Immunological changes during specific immunotherapy of grass pollen allergy: reduced lymphoproliferative responses to allergen and shift from TH2 to TH1 in T-cell clones specific for Phl p 1, a major grass pollen allergen. *Clin Exp Allergy* 1997;27:1007-1015.
- 92. Fanta C, Bohle B, Hirt W, Siemann U, Horak F, Kraft D et al. Systemic immunological changes induced by administration of grass pollen allergens via the oral mucosa during sublingual immunotherapy. *Int Arch Allergy Immunol* 1999;**120**:218-224.
- Bohle B, Kinaciyan T, Gerstmayr M, Radakovics A, Jahn-Schmid B, Ebner C. Sublingual immunotherapy induces IL-10producing T regulatory cells, allergen-specific T-cell tolerance, and immune deviation. J Allergy Clin Immunol 2007; 120:707-713.
- 94. Cosmi L, Santarlasci V, Angeli R, Liotta F, Maggi L, Frosali F et al. Sublingual immunotherapy with *Dermatophagoides* monomeric allergoid down-regulates allergen-specific immunoglobulin E and increases both interferon-gamma- and interleukin-10-production. *Clin Exp Allergy* 2006;**36**:261-272.
- 95. Wachholz PA, Nouri-Aria KT, Wilson DR, Walker SM, Verhoef A, Till SJ et al. Grass pollen immunotherapy for hayfever is associated with increases in local nasal but not peripheral Th1:Th2 cytokine ratios. *Immunology* 2002;**105**:56-62.
- Francis JN, Till SJ, Durham SR. Induction of IL-10+CD4+CD25+ T cells by grass pollen immunotherapy. J Allergy Clin Immunol 2003;111:1255-1261.
- Plewako H, Holmberg K, Oancea I, Gotlib T, Samolinski B, Rak S. A follow-up study of immunotherapy-treated birch-allergic patients: effect on the expression of chemokines in the nasal mucosa. *Clin Exp Allergy* 2008;**38**:1124-1131.
- Ciprandi G, De Amici M, Murdaca G, Filaci G, Fenoglio D, Marseglia GL. Adipokines and sublingual immunotherapy: preliminary report. *Hum Immunol* 2009;**70**:73-78.
- Makino Y, Noguchi E, Takahashi N, Matsumoto Y, Kubo S, Yamada T et al. Apolipoprotein A-IV is a candidate target molecule for the treatment of seasonal

allergic rhinitis. J Allergy Clin Immunol 2010;**126**:1163-1169.

- Kasperska-Zajac A, Brzoza Z, Rogala B. Effect of allergen-specific immunotherapy on platelet secretory activity in patients with grass-pollen allergy. *Vaccine* 2006;24:6990-6993.
- 101. Kasperska-Zajac A, Brzoza Z, Rogala B. Effect of allergen-specific immunotherapy on plasma level of platelet factor 4 (PF-4) and beta-thromboglobulin (beta-TG), platelet activation markers in patients with house dust mite allergy. *Vaccine* 2007:25:3595-3598.
- Li H, Xu E, He M. Cytokine responses to specific immunotherapy in house dust miteinduced allergic rhinitis patients. *Inflammation* 2015;**38**:2216-2223.
- 103. Sakashita M, Yamada T, Imoto Y, Hirota T, Tamari M, Ito Y et al. Long-term sublingual immunotherapy for Japanese cedar pollinosis and the levels of IL-17A and complement components 3a and 5a. *Cytokine* 2015;**75**:181-185.
- 104. Ohashi Y, Nakai Y, Kakinoki Y, Ohno Y, Okamoto H, Sakamoto H et al. The effect of immunotherapy on the serum levels of eosinophil cationic protein in seasonal allergic rhinitis. *Clin Otolaryngol Allied Sci* 1997;**22**:100-105.
- 105. Scadding GW, Eifan AO, Lao-Araya M, Penagos M, Poon SY, Steveling E et al. Effect of grass pollen immunotherapy on clinical and local immune response to nasal allergen challenge. *Allergy* 2015;**70**:689-696.
- 106. Jahnz-Rozyk K, Targowski T, Glodzinska-Wyszogrodzka E, Plusa T. Cc-chemokine eotaxin as a marker of efficacy of specific immunotherapy in patients with intermittent IgE-mediated allergic rhinoconjunctivitis. *Allergy* 2003;**58**:595-601.
- 107. Ciprandi G, Continia P, Fenoglio D, Sormani MP, Negrini S, Puppo F et al. Relationship between soluble HLA-G and HLA-A,-B,-C serum levels, and interferongamma production after sublingual immunotherapy in patients with allergic rhinitis. *Hum Immunol* 2008;69: 409-413.
- 108. Nieminen K, Laaksonen K, Savolainen J. Three-year follow-up study of allergeninduced in vitro cytokine and signalling lymphocytic activation molecule mRNA responses in peripheral blood mononuclear cells of allergic rhinitis patients undergoing specific immunotherapy. *Int Arch Allergy Immunol* 2009;**150**:370-376.
- 109. Yalcin AD, Gumuslu S, Parlak GE, Bisgin A. Soluble trail as a marker of efficacy of allergen-specific immunotherapy in patients with allergic rhinoconjunctivitis. *Med Sci Monit* 2012;18:CR617-CR621.

- 110. Lou W, Wang C, Wang Y, Han D, Zhang L. Enhancement of the frequency and function of IL-10-secreting type I T regulatory cells after 1 year of cluster allergen-specific immunotherapy. *Int Arch Allergy Immunol* 2012;**159**:391-398.
- 111. Kirmaz C, Ozenturk Kirgiz O, Bayrak P, Yilmaz O, Vatansever S, Ozbilgin K et al. Effects of allergen-specific immunotherapy on functions of helper and regulatory T cells in patients with seasonal allergic rhinitis. *Eur Cytokine Netw* 2011;**22**:15-23.
- 112. O'Hehir RE, Gardner LM, de Leon MP, Hales BJ, Biondo M, Douglass JA et al. House dust mite sublingual immunotherapy: the role for transforming growth factor-beta and functional regulatory T cells. *Am J Respir Crit Care Med* 2009;**180**:936-947.
- 113. Rolland JM, Gardner LM, O'Hehir RE. Functional regulatory T cells and allergen immunotherapy. *Curr Opin Allergy Clin Immunol* 2010;**10**:559-566.
- 114. Radulovic S, Jacobson MR, Durham SR, Nouri-Aria KT. Grass pollen immunotherapy induces Foxp3-expressing CD4+ CD25+ cells in the nasal mucosa. J Allergy Clin Immunol 2008;121:1467-1472.
- 115. Scadding GW, Shamji MH, Jacobson MR, Lee DI, Wilson D, Lima MT et al. Sublingual grass pollen immunotherapy is associated with increases in sublingual Foxp3expressing cells and elevated allergen-specific immunoglobulin G4, immunoglobulin A and serum inhibitory activity for immunoglobulin E-facilitated allergen binding to B cells. *Clin Exp Allergy* 2010:40:598-606.
- 116. Shamji M, Achkova D, Layhadi J, Perera-Webb A, Scadding G, Khan S et al. editors. IL-35 producing regulatory T cells modulate grass pollen-specific Th2 responses and are induced following sublingual grass pollen immunotherapy. *Clin Exp Allergy* 2012;**42**:1816-1817.
- 117. Shamji M, Layhadi J, Perera-Web A, Scadding G, Durham SR. IL-35+ regulatory T cells suppress grass pollen-driven Th2 responses and are induced following grass pollen-specific sublingual immunotherapy. J Allergy Clin Immunol 2013;2:AB146.
- Akdis CA, Akdis M. Advances in allergen immunotherapy: aiming for complete tolerance to allergens. *Sci Transl Med* 2015;7:280.
- 119. Mitra M, Kandalam M, Harilal A, Verma RS, Krishnan UM, Swaminathan S et al. EpCAM is a putative stem marker in retinoblastoma and an effective target for T-cell-mediated immunotherapy. *Mol Vis* 2012;18:290-308.
- van der Neut Kolfschoten M, Schuurman J, Losen M, Bleeker WK, Martinez-Martinez P, Vermeulen E et al. Anti-

inflammatory activity of human IgG4 antibodies by dynamic Fab arm exchange. *Science* 2007;**317**:1554-1557.

- Mauri C, Bosma A. Immune regulatory function of B cells. *Annu Rev Immunol* 2012;30:221-241.
- 122. Stanic B, van de Veen W, Wirz OF, Ruckert B, Morita H, Sollner S et al. IL-10overexpressing B cells regulate innate and adaptive immune responses. J Allergy Clin Immunol 2015;135:771-780.
- 123. Zimmer A, Bouley J, Le Mignon M, Pliquet E, Horiot S, Turfkruyer M et al. A regulatory dendritic cell signature correlates with the clinical efficacy of allergen-specific sublingual immunotherapy. J Allergy Clin Immunol 2012;129:1020-1030.
- 124. Gueguen C, Bouley J, Moussu H, Luce S, Duchateau M, Chamot-Rooke J et al. Changes in markers associated with dendritic cells driving the differentiation of either TH2 cells or regulatory T cells correlate with clinical benefit during allergen immunotherapy. J Allergy Clin Immunol 2016;137:545-558.
- 125. Angelini F, Pacciani V, Corrente S, Silenzi R, Di Pede A, Polito A et al. Dendritic cells modification during sublingual immunotherapy in children with allergic symptoms to house dust mites. *World J Pediatr* 2011;7:24-30.
- 126. Piconi S, Trabattoni D, Rainone V, Borgonovo L, Passerini S, Rizzardini G et al. Immunological effects of sublingual immunotherapy: clinical efficacy is associated with modulation of programmed cell death ligand 1, IL-10, and IgG4. *J Immunol* 2010;**185**:7723-7730.
- 127. Syed A, Garcia MA, Lyu SC, Bucayu R, Kohli A, Ishida S et al. Peanut oral immunotherapy results in increased antigen-induced regulatory T-cell function and hypomethylation of forkhead box protein 3 (FOXP3). J Allergy Clin Immunol 2014;133:500-510.
- 128. Passalacqua G, Albano M, Fregonese L, Riccio A, Pronzato C, Mela GS et al. Randomised controlled trial of local allergoid immunotherapy on allergic inflammation in mite-induced rhinoconjunctivitis. *Lancet* 1998;**351**:629-632.
- 129. Patel D, Couroux P, Hickey P, Salapatek AM, Laidler P, Larche M et al. Fel d 1derived peptide antigen desensitization shows a persistent treatment effect 1 year after the start of dosing: a randomized, placebo-controlled study. J Allergy Clin Immunol 2013;131:103-109.
- 130. Meyer W, Narkus A, Salapatek AM, Hafner D. Double-blind, placebo-controlled, dose-ranging study of new recombinant hypoallergenic Bet v 1 in an environmental exposure chamber. *Allergy* 2013;68:724-731.

- 131. Pfaar O, van Twuijver E, Boot JD, Opstelten DJ, Klimek L, van Ree R et al. A randomized DBPC trial to determine the optimal effective and safe dose of a SLITbirch pollen extract for the treatment of allergic rhinitis: results of a phase II study. *Allerev*, 2016**71**-99-107
- Makatsori M, Pfaar O, Calderon MA. Allergen immunotherapy: clinical outcomes assessment. J Allergy Clin Immunol Pract 2014;2:123-129.
- Agache I, Bilo M, Braunstahl GJ, Delgado L, Demoly P, Eigenmann P et al. In vivo diagnosis of allergic diseases–allergen provocation tests. *Allergy* 2015;**70**:355-365.
- 134. Scadding G, Hellings P, Alobid I, Bachert C, Fokkens W, van Wijk RG et al. Diagnostic tools in Rhinology EAACI position paper. *Clin Transl Allergy* 2011;1:2.
- 135. Durham SR, Ying S, Varney VA, Jacobson MR, Sudderick RM, Mackay IS et al. Grass pollen immunotherapy inhibits allergen-induced infiltration of CD4+ T lymphocytes and eosinophils in the nasal mucosa and increases the number of cells expressing messenger RNA for interferongamma. J Allergy Clin Immunol 1996;97:1356-1365.
- Senti G, Crameri R, Kuster D, Johansen P, Martinez-Gomez JM, Graf N et al. Intralymphatic immunotherapy for cat allergy induces tolerance after only 3 injections. J Allergy Clin Immunol 2012;129:1290-1296.
- 137. Creticos PS, Marsh DG, Proud D, Kagey-Sobotka A, Adkinson NF Jr, Friedhoff L et al. Responses to ragweed-pollen nasal challenge before and after immunotherapy. J Allergy Clin Immunol 1989;84:197-205.
- 138. Furin MJ, Norman PS, Creticos PS, Proud D, Kagey-Sobotka A, Lichtenstein LM et al. Immunotherapy decreases antigeninduced eosinophil cell migration into the nasal cavity. *J Allergy Clin Immunol* 1991;88:27-32.
- 139. Scadding GW, Calderon MA, Bellido V, Koed GK, Nielsen NC, Lund K et al. Optimisation of grass pollen nasal allergen challenge for assessment of clinical and immunological outcomes. J Immunol Methods 2012;384:25-32.
- Scadding GW, Eifan A, Penagos M, Dumitru A, Switzer A, McMahon O et al. Local and systemic effects of cat allergen nasal provocation. *Clin Exp Allergy* 2015;45:613-623.
- 141. Horak F, Zieglmayer P, Zieglmayer R, Lemell P, Devillier P, Montagut A et al. Early onset of action of a 5-grass-pollen 300-IR sublingual immunotherapy tablet evaluated in an allergen challenge chamber. J Allergy Clin Immunol 2009;**124**:471-477.
- 142. Nolte H, Maloney J, Nelson HS, Bernstein DI, Lu S, Li Z et al. Onset and dose-

related efficacy of house dust mite sublingual immunotherapy tablets in an environmental exposure chamber. *J Allergy Clin Immunol* 2015;**135**:1494-1501.

- 143. Jacobs RL, Harper N, He W, Andrews CP, Rather CG, Ramirez DA et al. Responses to ragweed pollen in a pollen challenge chamber versus seasonal exposure identify allergic rhinoconjunctivitis endotypes. J Allergy Clin Immunol 2012;130:122-127.
- 144. Rosner-Friese K, Kaul S, Vieths S, Pfaar O. Environmental exposure chambers in allergen immunotherapy trials: Current status and clinical validation needs. J Allergy Clin Immunol 2015;135:636-643.
- 145. Didier A, Malling HJ, Worm M, Horak F, Jager S, Montagut A et al. Optimal dose, efficacy, and safety of once-daily sublingual immunotherapy with a 5-grass pollen tablet for seasonal allergic rhinitis. J Allergy Clin Immunol 2007;120:1338-1345.
- 146. Keskin O, Tuncer A, Adalioglu G, Sekerel BE, Sackesen C, Kalayci O. The effects of grass pollen allergoid immunotherapy on clinical and immunological parameters in children with allergic rhinitis. *Pediatr Allergy Immunol* 2006;**17**:396-407.
- 147. Durham SR, Yang WH, Pedersen MR, Johansen N, Rak S. Sublingual immunotherapy with once-daily grass allergen tablets: a randomized controlled trial in seasonal allergic rhinoconjunctivitis. J Allergy Clin Immunol 2006;117:802-809.
- 148. Lue KH, Lin YH, Sun HL, Lu KH, Hsieh JC, Chou MC. Clinical and immunologic effects of sublingual immunotherapy in asthmatic children sensitized to mites: a double-blind, randomized, placebo-controlled study. *Pediatr Allergy Immunol* 2006;17:408-415.
- 149. Ozdemir C, Yazi D, Gocmen I, Yesil O, Aydogan M, Semic-Jusufagic A et al. Efficacy of long-term sublingual immunotherapy as an adjunct to pharmacotherapy in house dust mite-allergic children with asthma. *Pediatr Allergy Immunol* 2007;18:508-515.
- 150. Moverare R, Vesterinen E, Metso T, Sorva R, Elfman L, Haahtela T. Pollen-specific rush immunotherapy: clinical efficacy and effects on antibody concentrations. *Ann Allergy Asthma Immunol* 2001;**86**:337-342.
- 151. Chakraborty P, Roy I, Chatterjee S, Chanda S, Gupta-Bharracharya S. Phoenix sylvestris Roxb pollen allergy: a 2-year randomized controlled trial and follow-up study of immunotherapy in patients with seasonal allergy in an agricultural area of West Bengal, India. J Investig Allergol Clin Immunol 2006;16:377-384.
- 152. Klimek L, Schendzielorz P, Pinol R, Pfaar O. Specific subcutaneous immunotherapy with recombinant grass pollen allergens:

first randomized dose-ranging safety study. *Clin Exp Allergy* 2012;**42**:936-945.

- 153. Reich K, Gessner C, Kroker A, Schwab JA, Pohl W, Villesen H et al. Immunologic effects and tolerability profile of in-season initiation of a standardized-quality grass allergy immunotherapy tablet: a phase III, multicenter, randomized, double-blind, placebo-controlled trial in adults with grass pollen-induced rhinoconjunctivitis. *Clin Ther* 2011;**33**:828-840.
- 154. TePas EC, Hoyte EG, McIntire JJ, Umetsu DT. Clinical efficacy of microencapsulated timothy grass pollen extract in grass-allergic individuals. Ann Allergy Asthma Immunol 2004;92:25-31.
- 155. Ariano R, Kroon AM, Augeri G, Canonica GW, Passalacqua G. Long-term treatment with allergoid immunotherapy with Parietaria. Clinical and immunologic effects in a randomized, controlled trial. *Allergy* 1999;**54**:313-319.
- 156. Wahn U, Klimek L, Ploszczuk A, Adelt T, Sandner B, Trebas-Pietras E et al. Highdose sublingual immunotherapy with single-dose aqueous grass pollen extract in children is effective and safe: a doubleblind, placebo-controlled study. J Allergy Clin Immunol 2012;130:886-893.
- 157. Cox LS, Casale TB, Nayak AS, Bernstein DI, Creticos PS, Ambroisine L et al. Clinical efficacy of 300IR 5-grass pollen sublingual tablet in a US study: the importance of allergen-specific serum IgE. J Allergy Clin Immunol 2012;130:1327-1334.
- 158. Ott H, Sieber J, Brehler R, Folster-Holst R, Kapp A, Klimek L et al. Efficacy of grass pollen sublingual immunotherapy for three consecutive seasons and after cessation of treatment: the ECRIT study. *Allergy* 2009;64:1394-1401.
- 159. Mosges R, Bruning H, Hessler HJ, Gotz G, Knaussmann HG. Sublingual immunotherapy in pollen-induced seasonal rhinitis and conjunctivitis: a randomized controlled trial. Acta Dermatovenerol Alp Pannonica Adriat 2007;16:143-148.
- 160. Smith H, White P, Annila I, Poole J, Andre C, Frew A. Randomized controlled trial of high-dose sublingual immunotherapy to treat seasonal allergic rhinitis. J Allergy Clin Immunol 2004;114:831-837.
- 161. Pfaar O, Biedermann T, Klimek L, Sager A, Robinson DS. Depigmented-polymerized mixed grass/birch pollen extract immunotherapy is effective in polysensitized patients. *Allergy* 2013;68:1306-1313.
- 162. Bousquet J, Scheinmann P, Guinnepain MT, Perrin-Fayolle M, Sauvaget J, Tonnel AB et al. Sublingual-swallow immunotherapy (SLIT) in patients with asthma due to house-dust mites: a double-blind, placebocontrolled study. *Allergy* 1999;54:249-260.

- 163. Corzo JL, Carrillo T, Pedemonte C, Martin AMP, Hurtado SM, Dige E et al. Tolerability during double-blind randomized phase I trials with the house dust mite allergy immunotherapy tablet in adults and children. J Investig Allergol Clin Immunol 2014:24:154-161.
- 164. Yukselen A, Kendirli SG, Yilmaz M, Altintas DU, Karakoc GB. Effect of one-year subcutaneous and sublingual immunotherapy on clinical and laboratory parameters in children with rhinitis and asthma: a randomized, placebo-controlled, double-blind, double-dummy study. *Int Arch Allergy Immunol* 2012;157:288-298.
- 165. Keles S, Karakoc-Aydiner E, Ozen A, Izgi AG, Tevetoglu A, Akkoc T et al. A novel approach in allergen-specific immunotherapy: combination of sublingual and subcutaneous routes. J Allergy Clin Immunol 2011;128:808-815.
- 166. Bush RK, Swenson C, Fahlberg B, Evans MD, Esch R, Morris M et al. House dust mite sublingual immunotherapy: results of a US trial. *J Allergy Clin Immunol* 2011;**127**:974-981.
- 167. Zielen S, Kardos P, Madonini E. Steroidsparing effects with allergen-specific immunotherapy in children with asthma: a randomized controlled trial. *J Allergy Clin Immunol* 2010;**126**:942-949.
- 168. Pham-Thi N, Scheinmann P, Fadel R, Combebias A, Andre C. Assessment of sublingual immunotherapy efficacy in children with house dust mite-induced allergic asthma optimally controlled by pharmacologic treatment and mite-avoidance measures. *Pediatr Allergy Immunol* 2007;18:47-57.
- 169. Wang H, Lin X, Hao C, Zhang C, Sun B, Zheng J et al. A double-blind, placebo-controlled study of house dust mite immunotherapy in Chinese asthmatic patients. *Allergy* 2006;**61**:191-197.
- 170. Pajno GB, Morabito L, Barberio G, Parmiani S. Clinical and immunologic effects of long-term sublingual immunotherapy in asthmatic children sensitized to mites: a double-blind, placebo-controlled study. *Allergy* 2000;55:842-849.
- 171. Wang DH, Chen L, Cheng L, Li KN, Yuan H, Lu JH et al. Fast onset of action of sublingual immunotherapy in house dust mite-induced allergic rhinitis: a multicenter, randomized, double-blind, placebo-controlled trial. *Laryngoscope* 2013;**123**:1334-1340.
- 172. Hoiby AS, Strand V, Robinson DS, Sager A, Rak S. Efficacy, safety, and immunological effects of a 2-year immunotherapy with Depigoid birch pollen extract: a randomized, double-blind, placebo-controlled study. *Clin Exp Allergy* 2010;**40**:1062-1070.

- 173. Pfaar O, Robinson DS, Sager A, Emuzyte R. Immunotherapy with depigmented-polymerized mixed tree pollen extract: a clinical trial and responder analysis. *Allergy* 2010;65:1614-1621.
- 174. Horiguchi S, Okamoto Y, Yonekura S, Okawa T, Yamamoto H, Kunii N et al. A randomized controlled trial of sublingual immunotherapy for Japanese cedar pollinosis. *Int Arch Allergy Immunol* 2008;**146**:76-84.
- 175. Cortellini G, Spadolini I, Patella V, Fabbri E, Santucci A, Severino M et al. Sublingual immunotherapy for *Alternaria*-induced allergic rhinitis: a randomized placebo-controlled trial. *Ann Allergy Asthma Immunol* 2010;**105**:382-386.
- 176. Srivastava D, Gaur SN, Arora N, Singh BP. Clinico-immunological changes postimmunotherapy with *Periplaneta americana*. *Eur J Clin Invest* 2011;41:879-888.
- 177. Dahl R, Kapp A, Colombo G, de Monchy JG, Rak S, Emminger W et al. Sublingual grass allergen tablet immunotherapy provides sustained clinical benefit with progressive immunologic changes over 2 years. J Allergy Clin Immunol 2008;121:512-518.
- 178. Durham SR, Emminger W, Kapp A, Colombo G, de Monchy JG, Rak S et al. Long-term clinical efficacy in grass polleninduced rhinoconjunctivitis after treatment with SQ-standardized grass allergy immunotherapy tablet. J Allergy Clin Immunol 2010;125:131-138.
- 179. Saleem N, Chaker A, Zissler U, Schmidt-Weber CB, Durham SR, Shamji MH. Local nasal 'protective' immunoglobulin G4 (IgG4) responses in nasal fluid following grass pollen sublingual immunotherapy. J Allergy Clin Immunol 2013;131:AB202.
- 180. Blaiss M, Maloney J, Nolte H, Gawchik S, Yao R, Skoner DP. Efficacy and safety of timothy grass allergy immunotherapy tablets in North American children and adolescents. J Allergy Clin Immunol 2011;127:64-71.
- 181. Purohit A, Niederberger V, Kronqvist M, Horak F, Gronneberg R, Suck R et al. Clinical effects of immunotherapy with genetically modified recombinant birch pollen Bet v 1 derivatives. *Clin Exp Allergy* 2008;**38**:1514-1525.
- 182. Amar SM, Harbeck RJ, Sills M, Silveira LJ, O'Brien H, Nelson HS. Response to sublingual immunotherapy with grass pollen extract: monotherapy versus combination in a multiallergen extract. J Allergy Clin Immunol 2009;124:150-156.
- 183. Pauli G, Larsen TH, Rak S, Horak F, Pastorello E, Valenta R et al. Efficacy of recombinant birch pollen vaccine for the treatment of birch-allergic

rhinoconjunctivitis. *J Allergy Clin Immunol* 2008;**122**:951-960.

- 184. Ohashi Y, Tanaka A, Kakinoki Y, Ohno Y, Sakamoto H, Kato A et al. Serum level of soluble interleukin-2 receptor in patients with seasonal allergic rhinitis. *Scand J Immunol* 1997;45:315-321.
- 185. Barberi S, Villa MP, Pajno GB, La Penna F, Barreto M, Cardelli P et al. Immune response to sublingual immunotherapy in children allergic to mites. *J Biol Regul Homeost Agents* 2011;25:627-634.
- 186. Schulten V, Tripple V, Sidney J, Greenbaum J, Frazier A, Alam R et al. Association between specific timothy grass antigens and changes in TH1- and TH2-cell responses following specific immunotherapy. J Allergy Clin Immunol 2014;134:1076-1083.
- 187. Bonvalet M, Moussu H, Wambre E, Ricarte C, Horiot S, Rimaniol AC et al. Allergen-specific CD4+ T cell responses in peripheral blood do not predict the early onset of clinical efficacy during grass pollen sublingual immunotherapy. *Clin Exp Allergy* 2012;**42**:1745-1755.
- Campbell JD, Buchmann P, Kesting S, Cunningham CR, Coffman RL, Hessel EM. Allergen-specific T cell responses to immunotherapy monitored by CD154 and intracellular cytokine expression. *Clin Exp Allergy* 2010;**40**:1025-1035.
- Nomura T, Tsuge I, Inuo C, Nakajima Y, Tanaka K, Naruse N et al. Effect of Japanese cedar specific immunotherapy on allergen-specific T(H)2 cells in peripheral blood. *Ann Allergy Asthma Immunol* 2013;110:380-385.
- 190. Lent AM, Harbeck R, Strand M, Sills M, Schmidt K, Efaw B et al. Immunologic response to administration of standardized dog allergen extract at differing doses. J Allergy Clin Immunol 2006;118:1249-1256.
- 191. Potter PC, Baker S, Fenemore B, Nurse B. Clinical and cytokine responses to house dust mite sublingual immunotherapy. Ann Allergy Asthma Immunol 2015;114:327-334.
- 192. Francis JN, James LK, Paraskevopoulos G, Wong C, Calderon MA, Durham SR et al. Grass pollen immunotherapy: IL-10 induction and suppression of late responses precedes IgG4 inhibitory antibody activity. J Allergy Clin Immunol 2008;121:1120-1125.
- 193. Francis JN, Lloyd CM, Sabroe I, Durham SR, Till SJ. T lymphocytes expressing CCR3 are increased in allergic rhinitis compared with non-allergic controls and following allergen immunotherapy. *Allergy* 2007;62:59-65.
- 194. Moed H, Gerth van Wijk R, Hendriks RW, van der Wouden JC. Evaluation of clinical and immunological responses: a 2-year follow-up study in children with

allergic rhinitis due to house dust mite. *Mediators Inflamm* 2013;**2013**: 345217.

- 195. Savolainen J, Jacobsen L, Valovirta E. Sublingual immunotherapy in children modulates allergen-induced in vitro expression of cytokine mRNA in PBMC. *Allergy* 2006;61:1184-1190.
- 196. Ciprandi G, De Amici M, Marseglia GL. Serum IL-9 levels and sublingual immunotherapy: preliminary report. J Biol Regul Homeost Agents 2011;25:295-297.
- 197. Yukselen A, Kendirli SG, Yilmaz M, Altintas DU, Karakoc GB. Two year follow-up of clinical and inflammation parameters in children monosensitized to mites undergoing subcutaneous and sublingual immunotherapy. *Asian Pac J Allergy Immunol* 2013;**31**:233-241.
- 198. Ciprandi G, Cirillo I, Fenoglio D, Marseglia G, Tosca MA. Sublingual immunotherapy induces spirometric improvement associated with IL-10 production: prelimi-

nary reports. Int Immunopharmacol 2006;6:1370-1373.

- 199. Ciprandi G, Cirillo I, Tosca MA, Marseglia G, Fenoglio D. Sublingual immunotherapyinduced IL-10 production is associated with changed response to the decongestion test: preliminary results. *Allergy Asthma Proc* 2007:28:574-577.
- 200. Savolainen J, Laaksonen K, Rantio-Lehtimaki A, Terho EO. Increased expression of allergen-induced in vitro interleukin-10 and interleukin-18 mRNA in peripheral blood mononuclear cells of allergic rhinitis patients after specific immunotherapy. *Clin Exp Allergy* 2004;**34**:413-419.
- 201. Nieminen K, Valovirta E, Savolainen J. Clinical outcome and IL-17, IL-23, IL-27 and FOXP3 expression in peripheral blood mononuclear cells of pollen-allergic children during sublingual immunotherapy. *Pediatr Allergy Immunol* 2010;**21**:e174-e184.
- 202. Murakami D, Kubo K, Sawatsubashi M, Kikkawa S, Ejima M, Saito A et al. Phase

I/II study of oral immunotherapy with Cry j1-galactomannan conjugate for Japanese cedar pollinosis. *Auris Nasus Larynx* 2014;**41**:350-358.

- 203. Arvidsson MB, Lowhagen O, Rak S. Effect of 2-year placebo-controlled immunotherapy on airway symptoms and medication in patients with birch pollen allergy. J Allergy Clin Immunol 2002;109:777-783.
- 204. Scadding GW, Calderon MA, Shamji MH, Eifan AO, Penagos M, Dumitru F, Sever ML, Bahnson HT, Lawson K, Harris KM, Plough AG, Panza JL, Qin T, Lim N, Tchao NK, Togias A, Durham SR. Effect of 2 Years of Treatment With Sublingual Grass Pollen Immunotherapy on Nasal Response to Allergen Challenge at 3 Years Among Patients With Moderate to Severe Seasonal Allergic Rhinitis: The GRASS Randomized Clinical Trial. JAMA. 2017;**317**:615-625.

References 145–203 are cited in Supporting information.