

# Genome-Wide Interaction Analysis of Air Pollution Exposure and Childhood Asthma with Functional Follow-up

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## Abstract

**Rationale:** The evidence supporting an association between traffic-related air pollution exposure and incident childhood asthma is inconsistent and may depend on genetic factors.

**Objectives:** To identify gene–environment interaction effects on childhood asthma using genome-wide single-nucleotide polymorphism (SNP) data and air pollution exposure. Identified loci were further analyzed at epigenetic and transcriptomic levels.

**Methods:** We used land use regression models to estimate individual air pollution exposure (represented by outdoor NO<sub>2</sub> levels) at the birth address and performed a genome-wide interaction study for doctors' diagnoses of asthma up to 8 years in three European birth cohorts (n = 1,534) with look-up for interaction in two separate North American cohorts, CHS (Children's Health Study) and CAPPS/SAGE (Canadian Asthma Primary Prevention Study/Study of Asthma, Genetics and Environment) (n = 1,602 and 186 subjects, respectively). We assessed expression quantitative trait locus effects in human lung specimens and blood, as well as associations among air pollution exposure, methylation, and transcriptomic patterns.

**Measurements and Main Results:** In the European cohorts, 186 SNPs had an interaction  $P < 1 \times 10^{-4}$  and a look-up evaluation of these disclosed 8 SNPs in 4 loci, with an interaction  $P < 0.05$  in the large CHS study, but not in CAPPS/SAGE. Three SNPs within adenylate cyclase 2 (*ADCY2*) showed the same direction of the interaction effect and were found to influence *ADCY2* gene expression in peripheral blood ( $P = 4.50 \times 10^{-4}$ ). One other SNP with  $P < 0.05$  for interaction in CHS, rs686237, strongly influenced

UDP-Gal:betaGlcNAc  $\beta$ -1,4-galactosyltransferase, polypeptide 5 (*B4GALT5*) expression in lung tissue ( $P = 1.18 \times 10^{-17}$ ). Air pollution exposure was associated with differential discs, large homolog 2 (*DLG2*) methylation and expression.

**Conclusions:** Our results indicated that gene–environment interactions are important for asthma development and provided supportive evidence for interaction with air pollution for *ADCY2*, *B4GALT5*, and *DLG2*.

**Keywords:** genome-wide interaction study; methylation; gene expression; expression quantitative trait locus; children

## At a Glance Commentary

**Scientific Knowledge on the Subject:** Air pollution exposure early in life has been associated with asthma, but the mechanisms behind this effect are largely unknown. Understanding the biological mechanism that connects air pollutants with asthma and respiratory diseases has the potential to point to new targets for therapeutic intervention and to identify susceptible subgroups in the population.

**What This Study Adds to the Field:** We performed a genome-wide interaction study followed by functional genomics analyses that indicated involvement of several genes at the genomic, epigenomic, and transcriptomic levels for asthma related to air pollution exposure. Our results support the notion that gene–environment interactions are important for asthma development.

Asthma is the most common chronic disease among children (1). Heredity is a well-known risk factor, exemplified by strong associations between chromosome 17q21 variants and childhood asthma (2), but genetic factors cannot solely explain

its increasing prevalence in the past few decades. Exposure to traffic-related air pollution in early childhood (often indicated by the level of nitrogen dioxide [NO<sub>2</sub>]) has been associated with asthma exacerbations (3) and reduced lung function in children

(4–7), but its association with initial asthma development has been less consistent (8–11).

The exact mechanisms by which air pollution may lead to asthma are incompletely understood. Oxidative stress and inflammation represent pathogenic

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pathways involved in asthma development (3). Interactions between air pollution and allele variants in genes related to antioxidative stress systems, inflammation, and innate immunity have been reported in relation to the incidence of asthma (12, 13). Such gene–environment ( $G \times E$ ) interactions may partially explain the inconsistencies between air pollution and the incidence of asthma. A limitation of these previous studies is that they only included candidate genes, and so far, no genome-wide attempt has been made.

We aimed to identify mechanisms of childhood asthma using genome-wide single-nucleotide polymorphism (SNP) data and individual traffic-related air pollution exposure data, expressed as exposure to  $\text{NO}_2$  (see Figure E1 in the online supplement). We present genome-wide interaction data from more than 1,500 children with and without asthma from three European birth cohorts in the discovery phase, followed by look-up in two independent North American cohorts with approximately 1,800 children. For each of the SNPs that was nominally significant for interaction in the largest look-up evaluation cohorts ( $P < 0.05$ ), we evaluated expression quantitative trait locus (eQTL) effects in human lung specimens and traffic-related air pollution–induced gene expression by genotype in peripheral blood cells along with effects of short- and long-term air pollution exposure on peripheral blood DNA methylation patterns. Some of the results of these studies were previously reported in the form of an abstract (14).

## Methods

Additional details are available in the online supplement.

### Study Subjects

In the discovery phase, meta-analysis was performed based on genome-wide interaction study (GWIS) results from three traffic pollution, asthma, genetics consortium (13) cohorts, including 454 children with asthma and 1,080 children without asthma of European ancestry: BAMSE (Children, Allergy, Milieu, Stockholm, Epidemiological Survey) (6), Stockholm, Sweden (children with asthma: 235; children without asthma: 246), GINIplus (German Infant Study on the Influence of Nutrition Intervention Plus Environmental and Genetic Influences on Allergy

Development) (15) and LISAPlus (Influence of Life-Style Factors on the Development of the Immune System and Allergies in East and West Germany Plus the Influence of Traffic Emissions and Genetics) (16), Germany (children with asthma: 64; children without asthma: 661), and PIAMA (Prevention and Incidence of Asthma and Mite Allergy) (17), the Netherlands (children with asthma: 155; children without asthma: 173). Detailed cohort descriptions are provided in the online data supplement and elsewhere (13).

The top discovery SNPs were further evaluated in two independent look-up data sets from North America that included a total of 692 children with asthma and 1,096 children without asthma. The birth cohorts of CAPPS (Canadian Asthma Primary Prevention Study; Vancouver and Winnipeg, Canada) (18) and SAGE (Study of Asthma, Genetics and Environment; Manitoba, Canada) (19), both of which included children of white ancestry, had 49 children with asthma and 137 children without asthma; the larger cohort CHS (Children's Health Study; California) (20), which included children of non-Hispanic white ancestry, had 643 children with asthma and 959 children without asthma. All cohorts obtained ethical approval from their local review boards.

### Exposure and Outcome Assessment

For the European birth cohorts, the annual average of  $\text{NO}_2$  exposure estimates at birth were derived using land use regression modeling (LUR). Site-specific LUR models were developed and validated using the standardized European Study of Cohorts for Air pollution Effects project procedures ([www.escapeproject.eu/manuals](http://www.escapeproject.eu/manuals)), as previously described in detail (21). Using a similar methodology, LUR models of birth exposure were developed for CAPPS and SAGE (22, 23). In CHS,  $\text{NO}_2$  exposure was estimated based on the level in the child's community at baseline (mean age 8.8 yr) obtained from central site monitors placed in each of the study communities (20, 24).  $\text{NO}_2$  was used as a proxy for traffic-related air pollution. Exposure data were entered as a continuous nontransformed variable, and the risk estimates were reported per  $10 \mu\text{g}/\text{m}^3$  increase in  $\text{NO}_2$  (Table E1).

Asthma definitions were based on parental reports of an ever doctor's diagnosis (BAMSE, GINI/LISA, PIAMA

and CHS), clinical examinations by a pediatric allergist (CAPPS), or parental reports with confirmation of diagnoses by a pediatric allergist (SAGE) (Table E2).

### Genotyping and Quality Control

Genotyping, imputation procedure, and quality control steps for each study are described in the online supplement and Table E2.

### SNP $\times$ $\text{NO}_2$ Interaction and Asthma

Logistic regression analyses for estimation of standard SNP  $\times$   $\text{NO}_2$  interaction effects on asthma (multiplicative interaction model using HapMap-imputed, genome-wide association study [GWAS] data) was performed in each cohort separately as the primary model. A genome-wide significant threshold of  $P < 7.2 \times 10^{-8}$  for SNP  $\times$   $\text{NO}_2$  interaction effects was applied (25). Discovery meta-analysis of 2,082,301 overlapping SNPs was conducted using the statistical software METAL with fixed-effect models with default METAL weights (26). In addition to the primary SNP  $\times$   $\text{NO}_2$  GWIS analysis, two other statistical methods to test for genome-wide interaction were used, with the same exposure, outcome, and adjustment factors: a two-step approach, in which in step one, the hypothesis of  $H_0: \beta_{\text{SNP}} = 0$  was tested using  $\text{NO}_2$  as the outcome in a combined set of cases and control subjects; a subset of SNPs that exceeded a given significance threshold ( $P < 0.05$ ) for the test in step one was further analyzed in step two (in our study,  $N_{\text{SNPs}} = 119,521$ , equivalent to a genome-wide significance threshold of meta-analysis  $P < 4 \times 10^{-7}$  after Bonferroni correction of 119,521 tests); and testing the hypothesis that  $H_0: \beta_{\text{SNP} \times \text{NO}_2} = 0$  (analyzing cases and control subjects; regular  $G \times E$  interaction test) (27). The second method was the two degree-of-freedom (2 df) test that jointly tested SNP main and SNP  $\times$   $\text{NO}_2$  interaction effects (28).

To avoid false negative findings, an arbitrary cutoff level for look-up of interacting SNPs was set at  $P < 1 \times 10^{-4}$  (29) for our primary analysis in the discovery data sets (standard interaction model). Thus, SNPs with a combined interaction  $P < 1 \times 10^{-4}$  in the discovery phase were selected for look-up evaluation of standard SNP  $\times$   $\text{NO}_2$  interaction effects on asthma in the CAPPS/SAGE and CHS cohorts. Next, SNPs with  $P < 0.05$  for interaction in the larger CHS cohort (and annotated genes) were included in the functional genomics follow-up described in the following.

## Gene Expression Analysis in Lung Tissue and Peripheral Blood Cells

eQTL analyses were performed to evaluate if the SNPs significant in the look-up ( $P < 0.05$ ) were related to *cis*-acting lung tissue gene expression. Lung tissue from 1,111 human subjects who underwent lung surgery at three academic sites, Laval University, University of British Columbia, and University of Groningen, were previously analyzed (30, 31). Linear regression models were used separately for each cohort, adjusting for age, sex, and smoking status. Meta-analysis was performed using inverse variance weighting. SNPs were considered an eQTL if they survived 5% Benjamini and Hochberg false discovery rate correction for multiple testing of the number of gene probes tested for each SNP. The Genotype-Tissue Expression (GTEx) portal (<http://www.gtexportal.org/home>), which provides tissue-specific global gene expression data from genotyped donors, was used next to evaluate whole blood eQTLs ( $n = 338$  samples), analyzing the same SNPs and genes as in the lung eQTL (32). Furthermore, gene expression analyses (Affy HTA 2.0; Affymetrix Inc., Santa Clara, CA) were performed in peripheral blood cells from 263 16-year-olds in the BAMSE cohort as part of the MeDALL (Mechanisms of the Development of Allergy) project (33, 34). A look-up of GTEx-identified eQTLs was performed in 173 BAMSE samples, with GWAS data available using linear regression, adjusting for age, sex, and peripheral blood cell count. In addition, 250 BAMSE samples with exposure data available were used for linear regression association between  $\text{NO}_2$  at birth and current  $\text{NO}_2$  exposure at 16 years of age and expression levels of the genes annotated to significant look-up SNPs, with further stratification by genotype.

## DNA Methylation in Relation to Long- and Short-Term Air Pollution Exposure

Methylation values for CpG sites within regions  $\pm 50$  kb upstream and downstream of the identified genes were derived from Illumina 450K (Illumina Inc., San Diego, CA) data sets and investigated for association with air pollution exposure. Methylation data from the BAMSE cohort at 8 years ( $n = 460$  with Illumina 450k data available) were investigated for association with long-term  $\text{NO}_2$  exposure at birth using robust linear

regression, adjusting for age, sex, environmental tobacco smoke exposure during the first year of life, municipality at birth, ever doctor's diagnosis of asthma up to 8 years of age, cell type, and batch (bisulfite treatment date) (34). The same CpG sites were also investigated for methylation quantitative trait locus (methQTL) effects to evaluate if the significant SNPs in the  $G \times E$  look-up analyses were associated with methylation changes.

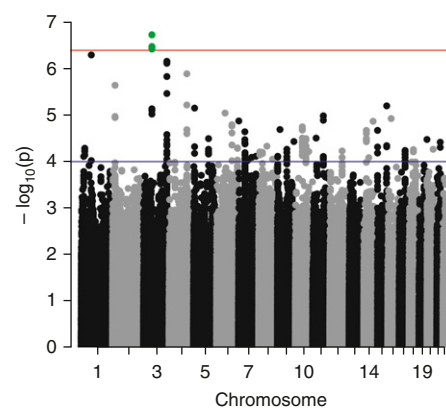
Short-term diesel exhaust exposure (DEP), as a model of particulate air pollution, was next investigated for association with DNA methylation differences in blood samples from 16 19- to 35-year-old nonsmokers with asthma and/or airway hyper-responsiveness using linear mixed effects modeling to compare post-DEP versus pre-DEP, and postfiltered air particles versus prefiltered air particles (35). Adjustment was done using a 5% Benjamini and Hochberg false discovery rate-correction for multiple testing on the set of probes selected for the analysis.

## Results

Tables E1 and E2 in the online supplement present the characteristics of the three European and two North American studies, including  $\text{NO}_2$  exposure assessment, genotyping, and imputation procedures.

### SNP $\times$ $\text{NO}_2$ Interaction and Asthma

In total, 1,534 children of European ancestry aged 7.4 to 11.3 years were included in the primary GWIS meta-analysis (454 children with asthma and 1,080 children without asthma as control subjects). Figure E2 in the online supplement shows the QQ-plot for the SNP  $\times$   $\text{NO}_2$  interaction analysis on asthma ( $\lambda = 1.03$ ). The discovery meta-analysis provided no genome-wide significant hits at the genome-wide significant threshold of  $P < 7.2 \times 10^{-8}$ . The top SNPs for interaction effects (lowest  $P = 1.87 \times 10^{-7}$ ) are located in chromosome 3p14.1, approximately 244 kb downstream of the membrane-associated guanylate kinase, WW, and PDZ domain-containing 1 (*MAGI1*) gene (Figure 1 and see Table E3). Next, we used an alternative two-step analytical approach that was suggested to increase the power to detect  $G \times E$  interactions (27). Four SNPs reached genome-wide significance in this two-step model ( $P < 4 \times 10^{-7}$ ; see Table E4): rs7651862, rs11706125, rs11718057, and rs13066946 close



**Figure 1.** Manhattan plot for the discovery genome-wide interaction meta-analysis of the association between SNP  $\times$   $\text{NO}_2$  and asthma. The horizontal red line indicates the genome-wide significance threshold when using the two-step interaction approach ( $P < 4 \times 10^{-7}$ ). The horizontal blue line indicates the threshold for single-nucleotide polymorphisms selected for look-up ( $P < 1 \times 10^{-4}$ ;  $n = 186$ ). The locus, near *MAGI1* on chromosome 3p14.1, which reached genome-wide significance when using the two-step interaction approach, is marked in green (rs7651862, rs11706125, rs11718057, rs13066946).

to the *MAGI1* gene; these were also identified as top hit SNPs in the primary GWIS meta-analysis. As a third approach, we applied the 2 df test that jointly tested main SNP and SNP  $\times$   $\text{NO}_2$  interaction effects (28). No SNP reached genome-wide significance in this test (lowest  $P$  value was  $1.08 \times 10^{-6}$ ) (see Figures E3 and E4, and Table E5).

### Look-up Evaluation

We selected 186 interaction-effect SNPs with  $P < 1 \times 10^{-4}$  from our primary model, the discovery GWIS meta-analysis for look-up in two different cohorts (Table E3). Of these 186 SNPs, 172 were available for look-up in the larger CHS-imputed, genome-wide SNP data set (643 children with asthma and 959 control subjects), and 8 SNPs showed nominal significant interaction ( $P < 0.05$ ) (Table 1 and see Table E9). The SNP with the lowest  $P$  value for interaction in CHS (rs686237;  $P = 0.0016$ ) is located on chromosome 20q13 in a region located 40 and 59 kb upstream of the genes UDP-Gal:betaGlcNAc  $\beta$ -1,4-galactosyltransferase, polypeptide 5 (*B4GALT5*), and solute carrier family 9, subfamily A (NHE8, cation proton antiporter 8), member 8 (*SLC9A8*), respectively. Three SNPs (rs1057251, rs12455842, and rs12457919) are located



**Table 1.** Single-Nucleotide Polymorphisms from the Genome-Wide Interaction Meta-Analysis of the Association between Single-Nucleotide Polymorphism × NO<sub>2</sub> Interaction and Asthma That Were Statistically Significant in the Look-up Evaluation

Chr	SNP	MAF	Nearest Gene	Discovery GWIS Meta-analysis: BAMSE, GINIplus/LISApplus, PIAMA (n = 1,534)		Look-up			
				Interaction P Value*	Stratification by Genotype <sup>†</sup> : OR (95% CI)	CHS (n = 1,602)		CAPPS/SAGE (n = 186)	
				Interaction P Value*	Stratification by Genotype <sup>†</sup> : OR (95% CI)	Interaction P Value <sup>‡</sup>	Stratification by Genotype <sup>†</sup> : OR (95% CI)	Interaction P Value <sup>‡</sup>	Stratification by Genotype <sup>†</sup> : OR (95% CI)
20	rs686237	0.32	<i>B4GALT5</i> , <i>SLC9A8</i>	$5.43 \times 10^{-5}$	CC: 0.77 (0.48–1.24) AC/AA: 1.69 (1.08–2.64)	0.0016	CC: 1.21 (1.04–1.41) AC/AA: 0.89 (0.78–1.01)	NA	NA
18	rs1057251	0.10	<i>MOCOS</i>	$6.18 \times 10^{-5}$	TT: 1.68 (0.85–3.29) CT/CC: 0.50 (0.22–1.15)	0.0094	TT: 0.95 (0.85–1.06) CT/CC: 1.30 (1.03–1.62)	0.58	TT: 2.59 (1.01–6.66) CT/CC: 2.02 (0.06–66.02)
18	rs12455842	0.10	<i>MOCOS</i>	$6.10 \times 10^{-5}$	TT: 1.70 (0.86–3.39) CT/CC: 0.48 (0.21–1.10)	0.010	TT: 0.95 (0.85–1.06) CT/CC: 1.30 (1.03–1.62)	0.55	TT: 2.59 (1.01–6.66) CT/CC: 2.02 (0.06–66.02)
5	rs4143882	0.33	<i>ADCY2</i>	$4.75 \times 10^{-5}$	GG: 0.81 (0.33–1.99) AG/AA: 1.61 (1.04–2.51)	0.015	GG: 0.88 (0.76–1.02) AG/AA: 1.13 (0.98–1.29)	0.26	GG: 4.90 (1.25–19.24) AG/AA: 1.04 (0.26–4.24)
5	rs727432	0.32	<i>ADCY2</i>	$6.67 \times 10^{-5}$	GG: 0.81 (0.33–1.99) GT/TT: 1.61 (1.04–2.51)	0.016	GG: 0.88 (0.76–1.02) GT/TT: 1.13 (0.98–1.29)	0.27	GG: 4.90 (1.25–19.24) GT/TT: 1.04 (0.26–4.24)
5	rs6886921	0.34	<i>ADCY2</i>	$7.03 \times 10^{-6}$	CC: 0.76 (0.29–1.99) CT/TT: 1.71 (1.11–2.66)	0.016	CC: 0.88 (0.76–1.02) CT/TT: 1.12 (0.98–1.27)	NA	NA
18	rs12457919	0.10	<i>MOCOS</i> , <i>FHOD3</i>	$5.52 \times 10^{-5}$	AA: 1.68 (0.85–3.29) AC/CC: 0.39 (0.09–1.75)	0.012	AA: 0.95 (0.85–1.06) AC/CC: 1.30 (1.03–1.62)	NA	NA
11	rs963146	0.21	<i>DLG2</i>	$8.61 \times 10^{-5}$	AA: 1.56 (1.04–2.33) AG/GG: 0.67 (0.21–2.18)	0.034	AA: 0.93 (0.83–1.06) AG/GG: 1.12 (0.96–1.32)	0.62	AA: 3.02 (0.84–10.87) AG/GG: 2.75 (0.70–10.79)

*Definition of abbreviations:* *ADCY2* = adenylate cyclase 2; *B4GALT5* =  $\beta$ -1,4-galactosyltransferase, polypeptide 5; BAMSE = Children, Allergy, Milieu, Stockholm, Epidemiological Survey; CAPPS = Canadian Asthma Primary Prevention Study; Chr = chromosome; CHS = Children's Health Study; CI = confidence interval; *DLG2* = discs, large homolog 2; *FHOD3* = formin homology 2 domain-containing 3; GINIplus = German Infant Study on the Influence of Nutrition Intervention Plus Environmental and Genetic Influences on Allergy Development; GWIS = genome-wide interaction study; MAF = minor allele frequency, according to BAMSE; *MOCOS* = molybdenum cofactor sulfurase; LISApplus = Influence of Life-Style Factors on the Development of the Immune System and Allergies in East and West Germany Plus the Influence of Traffic Emissions and Genetics; NA = not applicable; OR = odds ratio for asthma associated with exposure to traffic-related NO<sub>2</sub> for different genotypes; PIAMA = Prevention and Incidence of Asthma and Mite Allergy; SAGE = Study of Asthma, Genetics and Environment; *SLC9A8* = solute carrier family 9 member A8; SNP = single-nucleotide polymorphism. SNPs that were nominally significant in CHS ( $P < 0.05$ ), ordered by the CHS interaction  $P$  value. All  $P$  values given are two-sided.

\*Genome-wide significance threshold,  $P < 7.2 \times 10^{-8}$ .

<sup>†</sup>Stratification by genotype using dominant model.

<sup>‡</sup>Significance threshold for look-up evaluation,  $P < 0.05$ .

downstream of, or within the molybdenum cofactor sulfurase (*MOCOS*), on chromosome 18q12 and were in complete linkage disequilibrium ( $r^2 = 1.0$ ). These three SNPs were also among the top SNPs ( $P < 1 \times 10^{-4}$ ) in the two-step interaction approach meta-analysis (Table E4). Three additional SNPs located within adenylate cyclase 2 (*ADCY2*) on chromosome 5p15.3 (rs4143882, rs727432, and rs6886921 with high linkage disequilibrium;  $r^2 = 0.93$ –1.0) and one within discs, large homolog 2 (*DLG2*) on chromosome 11q14.1 (rs963146) reached nominal significance ( $P < 0.05$ ) (Table 1). The four SNPs close to the *MAG11* gene and the eight SNPs with  $P < 0.05$  in CHS were also nominally significant in the 2 df test ( $P$  value range  $5.64 \times 10^{-5}$  to 0.008) (Table E6). In the smaller Canadian CAPPS- and SAGE-imputed, genome-wide SNP data set (49 children with asthma and 137 control subjects), 122 SNPs were available for look-up evaluation. Two of the SNPs (rs3843891 on chromosome 4q31 and rs17265947 on chromosome 8q12.3) reached nominal significance ( $P < 0.05$ ) (Table E7), but none of

the SNPs were significant in CHS. The top SNPs close to *MAG11* identified in the discovery GWIS meta-analysis and the two-step approach were not significant in CHS or CAPPS/SAGE. No overall significant main effects of the 8 SNPs on asthma were observed (Table E8). NO<sub>2</sub> exposure at birth (per 10  $\mu\text{g}/\text{m}^3$  increase) was positively associated with asthma up to 8 years of age, but was not statistically significant (meta-analysis adjusted odds ratio, 1.26; 95% confidence interval, 0.61–2.58).

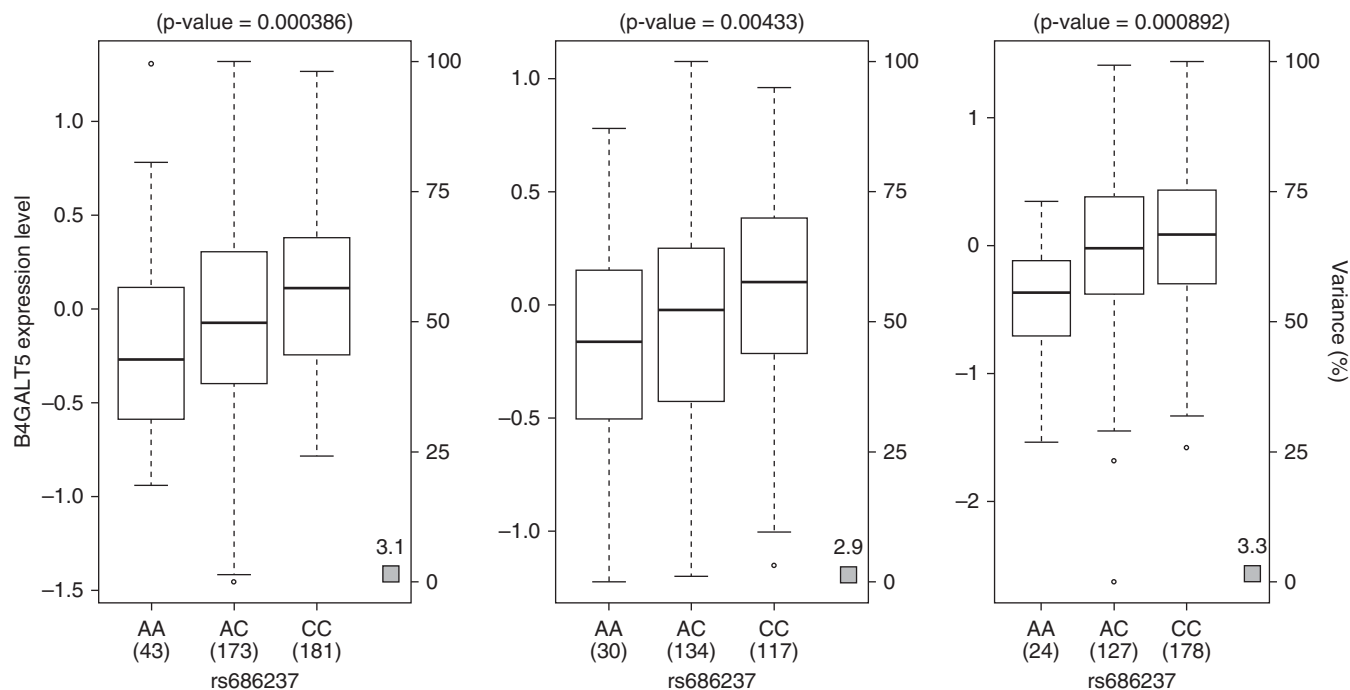
#### Direction of Interaction Effect and Asthma Risk

SNPs that showed nominal significance in the larger CHS sample (all 8 SNPs in Table 1) were investigated for their direction and strength of effect in the association between NO<sub>2</sub> exposure and childhood asthma. Consistent directions of the interaction effect between the discovery meta-analysis and CHS studies were identified for all three SNPs within *ADCY2* (Table 1), with increased risk of asthma associated with NO<sub>2</sub> exposure in carriers of

the minor alleles. The stratified analyses did not show a consistent pattern of asthma risk for the other SNPs, and the odds ratios for asthma across genotypes varied substantially between the data sets. Because the discovery data sets used exposure at birth and the main look-up study (CHS) used exposure at school age, meta-analyses of the interaction effects were not meaningful, because the odds ratios for asthma represented different measures.

#### Gene Expression Analysis in Lung Tissue and Peripheral Blood

We performed eQTL analyses to evaluate if the eight nominally significant SNPs from the look-up showed *cis*-acting eQTL associations in lung tissue ( $n = 1,111$ ). rs686237 was identified as a highly significant *cis*-eQTL of *B4GALT5* (the C allele is associated with increased expression;  $P = 1.18 \times 10^{-17}$ ) (Figure 2 and Table E10). In addition, rs12455842 was a significant *cis*-eQTL of *SLC39A6* ( $P = 0.003$ ) (Table E10) in lung tissue. No



**Figure 2.** Gene expression levels of  $\beta$ -1,4-galactosyltransferase, polypeptide 5 (*B4GALT5*) in lung tissues according to genotyping groups for single-nucleotide polymorphism rs686237 (using an additive model). The *left, middle, and right panels* are results from Laval University ( $n = 397$ ;  $P = 3.86 \times 10^{-4}$ ), University of British Columbia ( $n = 281$ ;  $P = 0.0043$ ), and Groningen University ( $n = 329$ ;  $P = 8.92 \times 10^{-4}$ ), respectively, with a meta-analysis  $P$  value of  $1.18 \times 10^{-17}$ . Expression is presented for probeset 100313047\_TGI\_at. The *left y-axis* represents gene expression levels in the lung. The *x-axis* represents the three genotyping groups for single-nucleotide polymorphism rs686237 (build 37 position 48,370,734) with the number of subjects in parenthesis. The *right y-axis* presents the percent variance in gene expression levels explained by the genotype. *Box boundaries* represent the first and third quartiles, with *whiskers* extending maximum  $\pm 1.5$  times the interquartile range, and the median denoted by the *center mark in the boxplots*. *Circles* represent outliers.

other SNP showed significant *cis*-eQTL association in lung tissue after 5% FDR correction for multiple testing. GTEx eQTL analyses in whole blood confirmed that rs686237 was a significant *cis*-eQTL of *B4GALT5* ( $P = 4.00 \times 10^{-4}$ ) (Figure E5), but with an opposite effect, in that the C allele was associated with decreased expression. rs6886921, rs727432, and rs4143882 were significant *cis*-eQTLs for *ADCY2* (lowest  $P = 4.50 \times 10^{-4}$  for rs6886921, with the T allele associated with decreased expression) (Figure E6 and Table E11). However, these blood eQTLs were not statistically significant in the smaller BAMSE data set ( $n = 173$ ).

Next, we explored  $\text{NO}_2$  exposure association with gene expression in BAMSE ( $n = 250$ ).  $\text{NO}_2$  exposure at birth significantly influenced *ADCY2*, *DLG2* and *MOCOS* expression, with increased expression levels in peripheral blood cells in relation to  $\text{NO}_2$  (Table 2; similar associations were also seen for exposure at 16 years; Table E12). For the top lung eQTL SNP, rs686237, an interacting

SNP  $\times \text{NO}_2$  effect was detected for *B4GALT5* expression (interaction  $P = 0.001$ , also FDR significant; Table 2), in which the effect of  $\text{NO}_2$  exposure at birth on gene expression differed depending on genotype status.

#### DNA Methylation and Air Pollution Exposure

Because air pollution exposure has been associated with differential DNA methylation patterns in peripheral blood cells (36), we explored potential links between  $\text{NO}_2$  exposure and methylation at the 278 CpG sites identified in a region  $\pm 50$  kb of the identified genes.

In the BAMSE cohort ( $n = 460$ ),  $\text{NO}_2$  exposure at birth was significantly associated with 2.7% decreased methylation in CpG site cg02275784 within *DLG2* (per  $10 \mu\text{g}/\text{m}^3 \text{NO}_2$  increase;  $P = 1.21 \times 10^{-4}$ ). Methylation in other CpG sites was not associated with  $\text{NO}_2$  exposure after 5% FDR correction for multiple testing (data not shown). Minor effects of DNA methylation changes were detected in

the methQTL analysis with a level of methylation change up to 1% per allele (nominal  $P < 0.05$ ). None of the associations remained significant at the 5% FDR level (Table E13).

As a marker for short-term, traffic-related air pollution exposure, 16 adult nonsmokers with asthma were exposed to 2 hours of DEP, at an average concentration of  $300 \mu\text{g}/\text{m}^3$ , containing high levels of  $\text{NO}_2$  (0.22 ppm) (35, 37). The difference in DNA methylation level was tested in blood samples pre-exposure versus postexposure. A total of 13 CpG sites were differentially methylated after 5% FDR correction for multiple testing (Table 3). Decreased methylation at eight CpG sites at the *DLG2* locus was detected (lowest  $P = 4.64 \times 10^{-5}$  for a 2% difference; cg26449294), and increased methylation was detected at two CpG sites close to transcription start sites (lowest  $P = 1.07 \times 10^{-4}$  for a 4% difference; cg20275558) (Table 3). Decreased methylation was also identified at one *ADCY2* CpG site, and increased methylation was seen at one *MOCOS* CpG site.

**Table 2.** Association between NO<sub>2</sub> Exposure Levels at Birth and Peripheral Blood Gene Expression Levels at 16 Years of Age in BAMSE (n = 250\*)

Chr	Gene	Probe	Associated SNP	Genotype	Coef	P Value	Interaction P Value
5	ADCY2	TC05000054.hg.1	rs6886921	All (n = 250)	0.03	<b>0.05</b>	0.85
				CC (n = 72)	0.04	0.17	
				TC (n = 83)	0.04	0.17	
5	ADCY2	TC05000055.hg.1	rs6886921	All (n = 250)	0.04	0.09	0.17
				CC (n = 72)	0.08	0.07	
				TC (n = 83)	-0.05	0.53	
11	DLG2	TC11002159.hg.1	rs963146	All (n = 250)	0.04	<b>0.008</b>	0.35
				AA (n = 104)	0.02	0.34	
				AG (n = 64)	0.05	0.08	
18	MOCOS	TC18000149.hg.1	rs1057251	All (n = 250)	0.03	<b>0.046</b>	0.59
				TT (n = 147)	0.04	0.09	
				CC (n = 22)	0.08	0.13	
20	B4GALT5	TC20000928.hg.1	rs686237	All (n = 250)	0.01	0.73	<b>0.001</b>
				CC (n = 88)	-0.11	<b>0.03</b>	
				AC (n = 66)	0.17	<b>0.02</b>	
20	SLC9A8	TC20000391.hg.1	rs686237	All (n = 250)	-0.01	0.53	0.18
				CC (n = 88)	-0.06	0.09	
				AC (n = 66)	0.08	0.08	
				AA (n = 19)	-0.03	0.66	

*Definition of abbreviation:* ADCY2 = adenylyl cyclase 2; B4GALT5 =  $\beta$ -1,4-galactosyltransferase, polypeptide 5; BAMSE = Children, Allergy, Milieu, Stockholm, Epidemiological Survey; Chr = chromosome; Coef = coefficient; DLG2 = discs, large homolog 2; MOCOS = molybdenum cofactor sulfurase; SLC9A8 = solute carrier family 9 member A8; SNP = single-nucleotide polymorphism.

Analyses were adjusted for age, sex, and cell count. Coef is the log-fold change in gene expression per 10  $\mu\text{g}/\text{m}^3$  increase in NO<sub>2</sub> exposure. P value is the P value for association between NO<sub>2</sub> exposure and gene expression. Interaction P value is the P value for association between SNP  $\times$  NO<sub>2</sub> and gene expression using additive effect of SNP. P values in bold have a value  $\leq 0.05$ .

\*n = 250 for the NO<sub>2</sub> to gene expression association analyses and n = 173 for the SNP  $\times$  NO<sub>2</sub> to gene expression analyses.

## Discussion

We presented a comprehensive GWIS with functional follow-up integrating genomics and environmental data that identified novel and previously identified loci for childhood asthma in relation to traffic-related air pollution exposure. Identified loci from the genome-wide SNP by NO<sub>2</sub> interaction approach, with significant look-up in 1,602 independent samples, were investigated for effects at genomic, epigenomic, and transcriptomic levels. We provided supportive evidence for interaction with air pollution for the novel loci B4GALT5 and the previously lung disease associated loci ADCY2 (38, 39) and DLG2 (40).

The GWIS was used as screening to detect genomic regions with a potential link to traffic-related air pollution exposure and childhood asthma. The SNP with the lowest P value in the look-up evaluation, rs686237 on chromosome 20, was found to be a strong eQTL for expression of B4GALT5 in the lung and was also identified as an eQTL

for B4GALT5 in whole blood. These results suggested a potential SNP-mediated effect of the association between NO<sub>2</sub> and childhood asthma with a functional consequence as indicated by differential B4GALT5 expression in blood depending on genotype.

The enzyme B4galt5 is involved in the biosynthesis of lactosylceramide, which is a common precursor of glycosphingolipids (41). Previous GWASs have identified a locus on chromosome 17q21, encompassing ORMDL sphingolipid biosynthesis regulator 3 (ORMDL3) and gasdermin B (GSDMB), to be strongly associated with childhood asthma (2). Interestingly, the endoplasmic reticulum transmembrane protein ORMDL3 is involved in the regulation of eosinophil trafficking (42).

ADCY2 encodes a member of the family of adenylyl cyclases, which are membrane-associated enzymes involved in G-protein-coupled receptor signaling. Three SNPs in ADCY2 showed statistical significance in the look-up evaluation of SNP  $\times$  NO<sub>2</sub> interaction effects on asthma,

and they had all similar direction of effect between the discovery cohorts and the main look-up study, CHS. The three SNPs were also identified as eQTLs for ADCY2 in whole blood. For the ADCY2 eQTL rs6886921, the minor allele T was associated with decreased expression in blood, and CT/TT carriers had the highest risk of asthma associated with NO<sub>2</sub> exposure in both the discovery and CHS data sets. ADCY2 was also differentially expressed in relation to air pollution exposure, and decreased methylation levels were found in relation to short-term air pollution exposure. ADCY2 SNPs were previously associated with pulmonary function and chronic obstructive pulmonary disease (COPD) (38, 39, 43).

Using the complementary alternative two-step statistical approach for the genome-wide interaction analysis, genome-wide significance was reached in the discovery data set for four SNPs located near the MAG11 locus. MAG11 acts as a scaffolding protein, stabilizing and

**Table 3.** Significant Associations of Short-Term Diesel Exhaust Exposure and CpG Site Methylation Difference (Postexposure – Preexposure) in Adults with Asthma (n = 16)

Chr	GWIS Locus	Probe	Probe Position (Build 37)	CpG Site Location	$\Delta$ F <sup>A</sup> * (Post – Pre)	$\Delta$ E <sup>†</sup> (Post – Pre)	DE P Value	DE Adjusted P Value <sup>‡</sup>
5	ADCY2	cg04119977	7,826,972	ADCY2 (Body)	–0.015	–0.019	0.0011	<b>0.041</b>
5	ADCY2	cg10995381	7,877,198	MTRR (Body)	–0.013	–0.032	0.0011	<b>0.041</b>
11	DLG2	cg26449294	83,169,193	DLG2 (3' UTR)	–0.010	–0.021	$4.64 \times 10^{-5}$	<b>0.017</b>
11	DLG2	cg09080874	83,284,905	DLG2 (Body)	–0.015	–0.027	$3.01 \times 10^{-4}$	<b>0.029</b>
11	DLG2	cg27373604	83,372,714	DLG2 (5' UTR;Body)	–0.021	–0.026	0.0021	<b>0.041</b>
11	DLG2	cg08432013	83,393,570	DLG2 (Body;TSS200)	–0.010	–0.025	$5.85 \times 10^{-4}$	<b>0.031</b>
11	DLG2	cg02675969	83,526,604	DLG2 (Body)	–0.010	–0.017	0.0018	<b>0.041</b>
11	DLG2	cg05405389	84,386,472	DLG2 (Body)	–0.017	–0.035	0.0016	<b>0.041</b>
11	DLG2	cg18023263	84,403,466	DLG2 (Body)	–0.020	–0.022	0.0013	<b>0.041</b>
11	DLG2	cg14716968	84,635,906	DLG2 (TSS1500;Body)	–0.003	–0.037	$4.28 \times 10^{-4}$	<b>0.029</b>
11	DLG2	cg20275558	85,338,473	TMEM126B; DLG2 (TSS1500;TSS200)	0.011	0.042	$1.07 \times 10^{-4}$	<b>0.020</b>
11	DLG2	cg06698742	85,359,218	TMEM126A (5' UTR)	0.0009	0.0041	0.0014	<b>0.041</b>
18	MOCOS	cg19453250	33,710,783	SLC39A6; ELP2 (TSS1500;Body)	0.0076	0.024	0.0019	<b>0.041</b>

*Definition of abbreviations:* ADCY2 = adenylate cyclase 2; Chr = chromosome; DE = diesel exhaust exposure; DLG2 = discs, large homolog 2; ELP2 = elongator acetyltransferase complex subunit 2; FA = filtered air; GWIS = genome-wide interaction study; MOCOS = molybdenum cofactor sulfurase; MTRR = 5-methyltetrahydrofolate-homocysteine methyltransferase reductase; SLC39A6 = solute carrier family 39 member A6; TMEM126A = transmembrane protein 126A; TMEM126B = transmembrane protein 126B; UTR = untranslated region.

P values in bold have a value  $\leq 0.05$ .

\* $\Delta$ F<sup>A</sup>: relative methylation change postexposure versus preexposure of filtered air.

<sup>†</sup> $\Delta$ E: relative methylation change postexposure versus preexposure of diesel exhaust.

<sup>‡</sup>Adjusted P values using the false discovery rate method for multiple testing at a 5% level.

recruiting various molecules to the cell–cell contacts, and it is widely distributed at tight junctions in epithelial cells (44). Involvement of the airway epithelium is of importance in asthma pathogenesis because disruption of barrier functions could potentially lead to air pollution–related adverse effects. However, the genome-wide significant SNP  $\times$  NO<sub>2</sub> interaction results for *MAG11* did not show statistical significance for interaction in the look-up evaluation, and functional analyses were therefore not pursued. The 2 df test that jointly tested for main genetic and interaction effects is an attractive method in genome-wide interaction studies (28). It was primarily developed to detect main effects while fully taking the environmental exposure into account, and has been successfully used in large-scale lung function studies (45). However, our analyses revealed no statistically significant hits at the genome-wide level, and limited power might have contributed to these results. The choice of method to detect interactions depends on study aims and availability of data, and from our study, it was difficult to draw conclusions about any preferred model. The main focus in our study was to perform functional interaction follow-up analyses on promising hits identified in the GWIS

analyses, which we believe, is of crucial importance.

In the diesel exposure study on adults, methylation changes were most notable for CpG sites in the *DLG2* gene, with reduced methylation levels at most sites. Analyses of long-term NO<sub>2</sub> exposure and DNA methylation profiles also indicated an association between air pollution exposure and *DLG2* methylation changes. We did not identify any significant association between the top *DLG2* SNP rs963146 and *DLG2* methylation, which indicated that the difference in *DLG2* methylation levels associated with air pollution was not SNP-mediated. NO<sub>2</sub> exposure was associated with higher expression levels of *DLG2* in blood cells (Table 2), which provided further evidence that exposure might induce functional changes related to this gene. *DLG2* (and *MAG11*) belong to the membrane-associated guanylate kinase (MAGUK) family (46). Disruption of *Drosophila melanogaster* DLG results in acute disorganization of epithelial structure, with disruption of intercellular junction formation (46). *DLG2* has recently been associated with COPD (38).

Three *MOCOS* SNPs were nominally significant in the CHS study, but we did not find convincing data in our functional analyses to support G  $\times$  E interactions of importance.

FANTOM5 (Functional Annotation of the Mammalian Genome 5) (47) and the Human Protein Atlas (HPA) (48) results showed that the identified genes were expressed at mRNA and protein levels in tissues relevant for asthma, although B4GALT5 could not be evaluated for protein expression in HPA (*see* the online supplement for additional details).

This study included all available data sets that we are aware of, with the required childhood phenotype, exposure, and genetic data needed for interaction analyses. Nevertheless, it would have been preferable to have larger sample sizes for G  $\times$  E analyses and functional analyses to decrease the likelihood of both type I and type II errors, and inclusion of non-white populations would have increased the generalizability of our results. We acknowledge that none of the identified SNPs were actually genome-wide significant in the discovery data set, and at the same time, were significant in the look-up data sets. Low statistical power is common in studies using GWIS data, and previous GWIS efforts to detect G  $\times$  E interaction effects for asthma and lung function indicate that new loci are challenging to discover (45, 49, 50).

For all cohorts, exposures were based on modeled outdoor concentrations of NO<sub>2</sub> (a surrogate for traffic-related pollution)



at the home and school addresses, but personal exposure to different pollution components, including indoor exposures, were not considered. NO<sub>2</sub> level is a good indicator for local air pollution, mainly from motor vehicles, and is highly correlated with other components of motor vehicle emissions, such as exhaust particles (21). However, we observed quite heterogeneous interaction effects in the discovery and look-up data sets, and only *ADCY2* SNPs showed similar directions of effect in the discovery and main look-up study. Differences in the levels or constituents of air pollutants, co-exposures, and unmeasured confounding factors between the North American and European cohorts could possibly explain the observed results. We also acknowledge that we used a rather liberal and unspecific definition of asthma [similar to the GABRIEL (A Multidisciplinary Study to Identify the Genetic and Environmental Causes of Asthma in the European Community) GWAS (2)], and the maximum age of asthma definition in CHS was up to 3 years older compared with the other cohorts, which might have contributed to heterogeneous effects (51). Because of these differences, we did not perform meta-analysis of the interaction  $\beta$ s, but we did present interaction  $\beta$ s and *P* values for each data set.

Previous  $G \times E$  interaction analyses using candidate gene approaches suggested that genes related to antioxidative stress systems, inflammation, and innate immunity (e.g., *GSTP1*, *TNF* and *TLR2/4*) are important effect modifiers (12, 13). These genes were not among the top hits in our GWIS, but this did not exclude true interaction effects for key SNPs as previously reported.

A key strength of our study was the extensive functional follow-up, and we

provided data for asthma that indicated the involvement of identified genes at the genomic, epigenomic, and transcriptomic levels in both lung tissue and peripheral blood cells in relation to air pollution exposure. These results were unlikely to be biased due to ethnic differences in our study populations, because all data were based on an ethnically homogenous (European or non-Hispanic white ancestry) population. In all steps of our study, we corrected for multiple testing to minimize false positive findings.

Our  $G \times E$  analysis using genome-wide data and multiple functional DNA methylation and gene expression analyses provided promising results for further understanding of the pathogenesis of childhood asthma. Our results supported the notion that  $G \times E$  interactions are important for asthma development, and that functional genomics analyses in conjunction with detailed environmental exposures provide valuable insight about pathophysiologic mechanisms. ■

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