

1 **Novel viral and host targets to cure hepatitis B**

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5 Addresses

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18 **Abstract**

19 Hepatitis B virus (HBV) infection is a global health threat with 240 million chronic carriers at  
20 high risk to develop hepatocellular carcinoma. Current antiviral treatment can efficiently  
21 control viral replication and reduce liver inflammation, but is still quite far from achieving a  
22 cure. Significant progress has been made in understanding the virus life cycle and virus-host  
23 interaction in the past few years. With identification of the HBV receptor, cell-culture infection  
24 systems have become available that allow drug screening and establishing a pipeline of  
25 potential antivirals targeting either viral or host factors. Most of the candidate antivirals  
26 summarized in this review are still in preclinical development, but some have already  
27 entered or are about to enter early clinical trials.

28

## 29 Introduction

30 Currently, an estimated 240 million individuals are chronic HBV carriers, and HBV infection  
31 and its consequences account for 680,000 deaths / year (WHO 2016). Even though HBV  
32 infection can be prevented by a vaccine, there is still no curative treatment for individuals  
33 who are already chronically infected. "Cure" is defined as clearance of circulating hepatitis B  
34 surface antigen (HBsAg) and seroconversion to anti-HBs+, a status in which antibodies  
35 against HBsAg are detectable in the blood. Anti-HBs seroconversion occurs spontaneously  
36 in about 0.2-1% of chronically infected patients per year [1]. Interferon (IFN)-based therapies  
37 achieve this goal in <10% of treated patients [2]. However, even after anti-HBs  
38 seroconversion, low level persistence of HBV is observed for decades [3] indicating on the  
39 one hand a functional rather than a complete cure and on the other hand that the immune  
40 system is able to keep HBV in check.

41

42 Therapy with nucleoside or nucleotide analogs (NAs) controls virus replication and mitigates  
43 inflammatory liver disease but does not result in a "cure" of HBV infection because it does  
44 not directly affect nuclear covalently closed circular HBV DNA (cccDNA), the HBV  
45 persistence form. <1% of HBeAg-negative and 0-10% of HBeAg-positive patients achieve  
46 Anti-HBs seroconversion after 3-5 years of continuous therapy [2], a rate that doesn't  
47 significantly exceed that of spontaneous functional cure [1]. In addition, the risk of  
48 developing hepatocellular carcinoma (HCC) remains significant despite NA treatment  
49 (approx. 7% in 5-7 years) [4,5] and even after anti-HBs seroconversion [1] . A recent study  
50 predicts that only a combination of a better vaccination coverage, innovations in prevention  
51 of mother-to-child transmission, and ambitious population-wide testing and treatment will be  
52 able to eliminate HBV as a major public health threat in the coming decades [6]. Barriers to  
53 implementing this and costs are obviously high, but could be largely reduced if a curative  
54 treatment would exist [6]. Thus, there is a high medical need for development of new  
55 antiviral strategies to achieve at least a functional cure of HBV and reduce HBV-related  
56 disease burden.

57

## 58 The hepatitis B virus life cycle

59

60 HBV is a small, enveloped DNA virus [7]. It attaches to heparan sulfate proteoglycans\_  
61 (HSPGs) [8], and is taken up upon binding to the hepatocyte-specific bile acid transporter  
62 NTCP (sodium taurocholate co-transporting polypeptide) serving as HBV receptor [9,10].  
63 After fusion of the HBV envelope with endosomal membranes, the HBV capsid releases the

64 partially double-stranded relaxed circular DNA genome (rcDNA) into the nucleus, where  
65 rcDNA is converted to cccDNA by cellular enzymes (for details see: [11]). cccDNA serves as  
66 transcription template and is the viral persistence form. Viral RNAs transcribed from cccDNA  
67 in the nucleus are exported to the cytoplasm and translated into viral proteins. One of the  
68 viral transcripts, termed pregenomic RNA (pgRNA) is packaged into a newly formed viral  
69 capsid and therein reverse transcribed into rcDNA. The rcDNA-filled mature capsid are  
70 either enveloped with cellular membranes containing small, medium and large (S, M, L) HBV  
71 envelope proteins and secreted from the infected cell or transported back to the nucleus to  
72 maintain a cccDNA pool (Figure 1).

73 Several unique features in HBV infection make the virus difficult to be successfully targeted  
74 by antiviral approaches. Firstly and most importantly, one to five copies of cccDNA hide as  
75 episomal minichromosomes in the nucleus of infected hepatocytes, and can be replenished  
76 via nuclear reimport of rcDNA bypassing the receptor-mediated entry (Figure 1). Secondly,  
77 viral DNA synthesis exclusively occurs inside the capsid and thereby the viral genome and  
78 replicative intermediates are shielded from cytoplasmic pattern recognition receptors during  
79 the entire life cycle. This, together with lack of STING expression in hepatocyte [12], seems  
80 to result in impaired innate immune DNA-sensing. Thirdly, HBV massively sheds non-  
81 infectious subviral particles mainly consisting of S protein (HBsAg) that have no apparent  
82 role in viral replication and are believed to serve as an immunological decoy. Fourthly, HBV  
83 exhibits a strict hepatic tropism and the liver provides a hostile environment for pathogens  
84 under distinct conditions [13]. Last but not least, HBV is divided into 10 genotypes which  
85 differ in clinical course after infection [2] and in their response to IFN treatment [14]. Thus,  
86 different HBV genotypes may also react differently in particular to new immune stimulatory  
87 therapies. Preclinical studies should therefore include the most prevalent HBV genotypes  
88 and clinical studies should not only include HBeAg-positive and –negative patients, but also  
89 monitor if all genotypes are treated with comparable efficacy.

90 Since host-directed therapies are more prone to elicit side effects than antivirals directly  
91 targeting the virus life cycle, a detailed understanding of the HBV life cycle and viral-host  
92 interactions are needed to develop novel and ideally curative treatment strategies. Here we  
93 summarize key viral and host targets for anti-HBV therapy that are currently exploited.

94

## 95 **Viral targets**

### 96 ***The viral transcription template: HBV cccDNA***

97 Persistence of HBV relies on nuclear cccDNA that is refractory to current NA therapeutics  
98 [11]. Once cccDNA is established, it is extraordinarily stable and seems not to be completely  
99 lost via hepatocyte turnover since it can be delivered into daughter cells during cell division  
100 by recycling of viral capsids although it does not contain a nuclear retention signal. Owing to  
101 technical challenges in selective and sensitive cccDNA detection [11] and restriction of the  
102 models used, previous studies gave a wide range of values in terms of the half-life of  
103 cccDNA. We carefully determined by Southern blot analysis that the half-life of cccDNA  
104 following inhibition of viral replication is approximately 30 days in HepG2-NTCP cells, and  
105 cccDNA is not completely lost even after several rounds of cell division (C Ko et al.,  
106 unpublished). Hence, complete loss of cccDNA without direct targeting strategies appears  
107 unlikely as it in average would take more than a lifetime [15].

108 Of note, a recent finding showed that the stable cccDNA pool can be directly targeted for  
109 degradation. We demonstrated that IFN- $\alpha$ , lymphotoxin- $\beta$  receptor agonists, and T cell-  
110 derived cytokines (i.e., IFN- $\gamma$  and TNF- $\alpha$ ) can specifically purge cccDNA from the nucleus of  
111 infected cells in noncytolytic manner [16-18]. Upon treatment, nuclear cytidine deaminases  
112 (APOBEC3A or APOBEC3B) are recruited to the cccDNA, deaminate cytidines and render  
113 the cccDNA molecule sensitive to degradation. Completion of this finding by identifying  
114 downstream cellular glycosylase and nuclease (D. Stadler et al., unpublished), which are  
115 responsible for abasic site formation followed by cccDNA fragmentation, may finally allow  
116 translation of this basic research finding into the clinic.

117 Molecular scissors such as zinc-finger nucleases (ZFNs), transcription activator-like effector  
118 nucleases (TALENs), or the more recently discovered CRISPR/Cas9-based gene editing  
119 approach appears to be promising tools to directly target and reduce the viral cccDNA  
120 reservoir [19,20]. To minimize undesired off-target effects and increase efficacy of cleavage  
121 of the HBV genome, approaches such as a double-nicking strategy by Cas9-nickase and  
122 multiple targeting strategy using two or three guide RNAs in combination have been  
123 performed [21,22]. However, improved strategies are needed for *in vivo* delivery and to steer  
124 the fate of cccDNA upon cleavage (degraded vs. repaired) and target specificity (episomal  
125 cccDNA vs. integrated HBV genome).

126

### 127 ***HBV transcripts***

128 Another approach is not to directly target the nuclear cccDNA, but to target HBV RNA  
129 transcripts. This may be achieved by cytokine-induced pathways with type I IFN destabilizing

130 viral RNA or IL-6 inhibiting HBV transcription [23]. Alternatively, one can block translation  
131 using an antisense RNA (asRNA) approach [24] or to induce degradation of the RNAs by  
132 employing RNA interference in a sequence-specific fashion. The latter can be achieved  
133 using either short interfering RNA (siRNA) that is brought into infected hepatocytes or short  
134 hairpin RNA (shRNA) that is expressed from a transgene which was delivered into the  
135 infected cell by a viral gene therapy vector [25]. Targeting HBV RNAs is particularly  
136 attractive as all HBV open reading frames use the same polyadenylation site downstream of  
137 the X ORF and therefore all HBV transcripts share a common 3' terminus. This allows  
138 simultaneous suppression of all viral transcripts (including that of the HBV pregenome) and  
139 antigens with one single RNA-targeting molecule. Immune stimulatory and siRNA function  
140 may even be combined in a single molecule [26]. As data accumulate revealing  
141 immunomodulatory as well as carcinogenic properties of the secreted HBV antigens (HBsAg  
142 and HBeAg) as well as HBx, this approach is very appealing. Anti-HBV siRNAs are already  
143 in early clinical trials, developed by Arbutus Biopharma (phase II) and Alnylam  
144 Pharmaceuticals (phase I). A clinical trial by Arrowhead Pharmaceuticals was halted  
145 (company decision; potential toxicity of the delivery agent used in this trial in primates at very  
146 high doses). Although siRNA therapeutics are well tolerated in general, a drawback of the  
147 approach is that these strategies targeting HBV transcripts do not affect cccDNA persistence  
148 or turn-over and thus need to be applied repeatedly. However, one may hope that  
149 suppressing viral proteins does not only suppress viral replication, but allows to restore  
150 antiviral immunity and enhance spontaneous clearance rates - or at least improve efficacy of  
151 co-administered immune stimulatory drugs.

152

### 153 ***HBV polymerase: an enzyme with multiple activities***

154 Since HBV polymerase (HBV Pol) is the only viral protein with enzymatic activities [DNA  
155 polymerase and ribonuclease H (RNase H) activity], it has been a primary target for drug  
156 development [27]. Consequently, five FDA-approved NAs that all target the reverse  
157 transcriptase activity of HBV Pol are in the mainstay of current therapies [28]. In addition,  
158 next-generation prodrugs of NAs (e.g., Tenofovir alafenamide, CMX157, AGX-1009,  
159 Besifovir, and Lagociclovir valactate) with the goal of enhancing antiviral efficacy, oral  
160 bioavailability, and long-term tolerability are currently undergoing evaluation in clinical trials.

161 Enzymatic RNase H activity of HBV Pol is also essential for viral DNA synthesis since it  
162 removes the pgRNA template during HBV minus-strand DNA synthesis. Recently, it was  
163 reported that chemical compounds being active against HIV RNase H are able to inhibit HBV

164 replication by blocking HBV RNase H activity [29,30]. Nonetheless, specific RNase H  
165 inhibitors have not received much attention for several decades probably due to early  
166 introduction of potent NAs into hepatitis B therapy and the interdependence between DNA  
167 polymerase and RNase H activity.

168 Targeting HBV Pol may also allow to target encapsidation of the HBV pregenome. Viral DNA  
169 synthesis begins with specific recognition of a stem-loop structure at the 5' end of pgRNA by  
170 HBV Pol, followed by incorporation of both into an assembling capsid. Subsequently, HBV  
171 Pol, which is covalently attached to a short DNA oligonucleotide (5'-GAA-3'), acts as a  
172 protein primer initiating reverse transcription. Attempts to develop specific inhibitors blocking  
173 those two largely unexplored steps are undergoing [31,32].

174

### 175 ***Capsid-forming core protein***

176 Core protein (Cp) that assembles into the viral capsid is involved in many aspects of the  
177 HBV life cycle (Figure 1), suggesting that Cp represents a valid target for antivirals.  
178 Targeting Cp may lead to simultaneous inhibition of multiple steps. Interference with the  
179 capsid-forming activity of core protein has been thoroughly studied due to the importance for  
180 subsequent viral DNA synthesis. As a result, two groups of Cp allosteric modulators (CpAM)  
181 which misdirect core assembly and thereby induce either empty capsid (e.g., AT130) or  
182 aberrant capsid formation (e.g., HAPs) were discovered [33]. Much effort to increase antiviral  
183 efficacy of known CpAM by chemical optimization as well as the search for new classes of  
184 CpAM are ongoing and the first compounds have entered early clinical trials (e.g., NVR 3-  
185 778 and GLS-4).

186 Although CpAMs and clinically approved NAs suppress HBV DNA accumulation, it is  
187 tempting to speculate that CpAMs may have additional antiviral activities e.g. via targeting  
188 the incoming capsid that establishes cccDNA in a newly infected cell or activation of innate  
189 immunity by abnormal Cp aggregates. HBV rcDNA contained in incoming capsid must be  
190 delivered to and released into the nucleus; otherwise, no cccDNA formation is expected. In  
191 fact, we observed that a novel CpAM significantly reduced cccDNA levels when it was  
192 administered prior to cccDNA establishment in a *de novo* HBV infection (C Ko et al.,  
193 unpublished). Given that the HBV capsid has a certain degree of flexibility and needs to  
194 dissociate at the nuclear pore to release the rcDNA genome [33,34], it is conceivable that  
195 CpAMs have an effect on establishing HBV infection through interference with capsid  
196 dissociation. In addition, Cp has been described to be associated with cccDNA [35],

197 although it is not clear if Cp regulates viral transcription or exerts particular function by  
198 altering the structure of the cccDNA minichromosome which may also be influenced by  
199 CpAMs.

200

### 201 ***Regulatory HBx protein***

202 HBx is a small regulatory protein that is essential for viral gene expression and has been  
203 implicated in the pathogenesis of HCC [36-38]. It also seems to play a role in a number of  
204 cellular processes influencing the HBV life cycle including epigenetic control of the cccDNA  
205 minichromosome. Similar to cellular chromatin, cccDNA is organized as a viral  
206 minichromosome wrapped around histone and non-histone proteins in the nucleus of HBV-  
207 infected hepatocytes [39]. The cccDNA minichromosome can be in a transcriptionally open  
208 conformation carrying high levels of active chromatin markers [40]. HBx, as a viral protein  
209 attached to the cccDNA minichromosome, may play a role in the establishment of an  
210 accessible chromatin by regulating the recruitment of chromatin modifying enzymes [41-44].

211 HBx, however, seems to exert its main function by interacting with distinct cellular proteins  
212 and the Cul4-DDB1 E3 ubiquitin ligase machinery [45]. Thus, HBx targets the Smc5/6  
213 complex, which assembles around the cccDNA and prevents transcription of HBV genes  
214 [46,47]. This finding was sufficient to bring all the interest in HBx back to the stage and to  
215 consider it an antiviral target. It may be worth tracking if additional restriction factors exist  
216 that are targeted by HBx. One drawback is that HBx seems to be an intrinsically unfolded  
217 protein [48] with a short half-life. Another drawback is that targeting HBx will have to be  
218 continuous as experiments have shown that cccDNA persists in the absence of HBx and is  
219 reactivated as soon as HBx becomes available again [36]. The fact that cccDNA persists  
220 even in patients with resolved chronic hepatitis B (CHB) and can be reactivated under  
221 immunosuppression, strongly supports the notion that influencing epigenetic control of HBV  
222 transcription does maximally lead to functional control of HBV and needs to be persistent.

223

### 224 ***HBV envelope proteins***

225 The HBV envelope proteins are primarily responsible for virus entry. Thus, targeting S or L  
226 protein (e.g., by neutralizing antibodies) allows to prevent HBV infection. This is exploited  
227 using polyclonal antibodies from human serum as a passive vaccination in neonates or after  
228 accidental exposure to HBV. Alternatively, a number of monoclonal antibodies have been



229 developed that neutralize also escape-variant HBV and can serve this purpose [49].

230 Since HBsAg that is released in large amounts from infected cells serves as an antibody  
231 decoy and probably also skews T cell responses, blocking its release is also an interesting  
232 strategy which may be achieved by administration of nucleic acid polymers (NAPs). A recent  
233 report revealed that NAP monotherapy in CHB patients led to reduction of serum HBV DNA  
234 and concurrent appearance of anti-HBs antibodies even after treatment cessation,  
235 suggesting that NAPs may have a clinical benefit by restoring host immune response to  
236 some extent [50].

237 In addition, three independent studies recently showed that tetherin/BST-2, an IFN-inducible  
238 antiviral protein targeting various enveloped viruses, inhibits HBV virion egress [51-53]. In  
239 turn, HBsAg or HBx were proposed to counteract tetherin-mediated antiviral activity [52,53],  
240 suggesting that targeting these viral antagonists might be one therapeutic option to trap the  
241 infectious particle and thereby prevent virus spread.

242 Clinical studies showed that serum HBV DNA in chronic patients declines 3-4 log<sub>10</sub> scales  
243 after loss of HBeAg, whereas HBsAg reduction was significantly lower by only 1 log<sub>10</sub> scale  
244 [54]. This intriguing finding suggests that HBV DNA that becomes integrated into the host  
245 genome already early during HBV infection [55] is a significant source of HBsAg. This  
246 renders HBsAg an interesting “marker” of cells either infected with HBV or carrying an  
247 integrated HBV genome and thus bearing a risk for HCC development. Targeting HBsAg on  
248 the surface of these cells using single chain antibodies allows the development of T-cell  
249 retargeting strategies to eliminate the cells. This can either be achieved using chimeric  
250 antigen receptors for T cell grafting [56,57] or by applying bispecific antibodies to redirect T  
251 cells (U. Protzer et al., unpublished). This immunotherapeutic approach to induce killing of  
252 hepatocyte expressing HBsAg through specific recognition by redirect T cells is a potentially  
253 promising, alternative antiviral strategy.

254

## 255 **Host targets**

256 Achieving HBV cure means targeting cccDNA persistence. Besides directly targeting the  
257 cccDNA molecule as discussed above, the only alternatives are to either prevent  
258 establishment or at least replenishment of a cccDNA pool. At the level of the host, this may  
259 be achieved by preventing HBV entry into cells or by preventing the formation of cccDNA.

260

## 261 ***Viral entry***

262 Viral entry into host cells is mediated by a number of host factors in a highly orchestrated  
263 manner. Viral protein motifs mediating entry are highly conserved. Targeting the host factors  
264 involved in viral entry thus is a reasonable approach to prevent both initial viral infection and  
265 virus propagation. HBV entry initiates by adsorption of viral particles to HSPGs [8], followed  
266 by a specific engagement of viral particles with NTCP via motif aa 9-NPLGF(F/L)P-15 that is  
267 located in the N-terminal preS1 domain of the L protein [9,10] and highly conserved among  
268 different human and primate HBV genotypes (Figure 1, summarized in: [58]). Specific  
269 receptor recognition drives the internalization of virus particles, most likely via receptor-  
270 mediated endocytosis, but the underlying endocytic pathway as well as the detailed fusion  
271 events remain elusive [59,60]. The cellular factors involved in endocytosis and endosomal  
272 fusion may serve as therapeutic targets.

273

274 A synthetic peptide spanning this motif and neighboring regions (aa 2-48) in the preS1  
275 domain of L has been identified that inhibits HBV entry by competitively occupying its liver-  
276 specific receptor NTCP. This peptide inhibitor, called Myrcludex B, has proven its efficacy at  
277 low picomolar concentrations in preclinical models, seems safe in first-in-men trials and is  
278 currently in phase II clinical trials [58,61]. Alternatively, compounds targeting NTCP function  
279 (e.g., cyclosporine A, ezetimibe, and irbesartan) have been shown to inhibit HBV infection  
280 [62-65]. Taken together, antivirals targeting HBV entry may serve as an alternative to  
281 neutralizing antibodies to prevent virus transmission (e.g., after liver transplantation or from  
282 mother to child) if safety proves equivalent, but also in combination with other antivirals or  
283 immune therapies to prevent virus spread or re-establishment of infection.

284

## 285 ***Conversion of rcDNA to cccDNA***

286 The molecular mechanism by which rcDNA is converted to cccDNA is still enigmatic. Recent  
287 studies proposed tyrosyl-DNA-phosphodiesterase (TDP2) as the enzyme responsible for  
288 removal of covalently attached HBV Pol from rcDNA *in vitro* [66,67], and the cellular DNA  
289 polymerases  $\kappa$  being crucial to fill the plus-stand DNA gaps [68]. Further studies are needed  
290 to identify additional cellular factors, for instance a cellular exonuclease responsible for  
291 removing a short pgRNA remnant which serves as primer for the plus-stand DNA synthesis,  
292 or a cellular DNA ligase responsible for ligating the end of each stand. In this regard, a novel  
293 screening strategy aiming at identifying drug candidates that prevent the conversion of  
294 rcDNA to cccDNA would draw much attention [69,70]. However, targeting host molecules

295 involved in this process risks side effects.

296

## 297 **Conclusion**

298 Currently available NAs are well-tolerated and can control HBV replication. Although a  
299 variety of viral and host targets are currently exploited for the development of new antivirals,  
300 the holy grail of HBV cure, viral persistence via nuclear cccDNA, is only targeted with very  
301 few of the approaches summarized above. To achieve the final goal, HBV cure, approaches  
302 not directly targeting cccDNA would either require the host immune system to spontaneously  
303 “recover” and control the virus or would have to be combined with a potent trigger increasing  
304 HBV-specific immunity.

305

## 306 **Conflict of interest**

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309

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315

316

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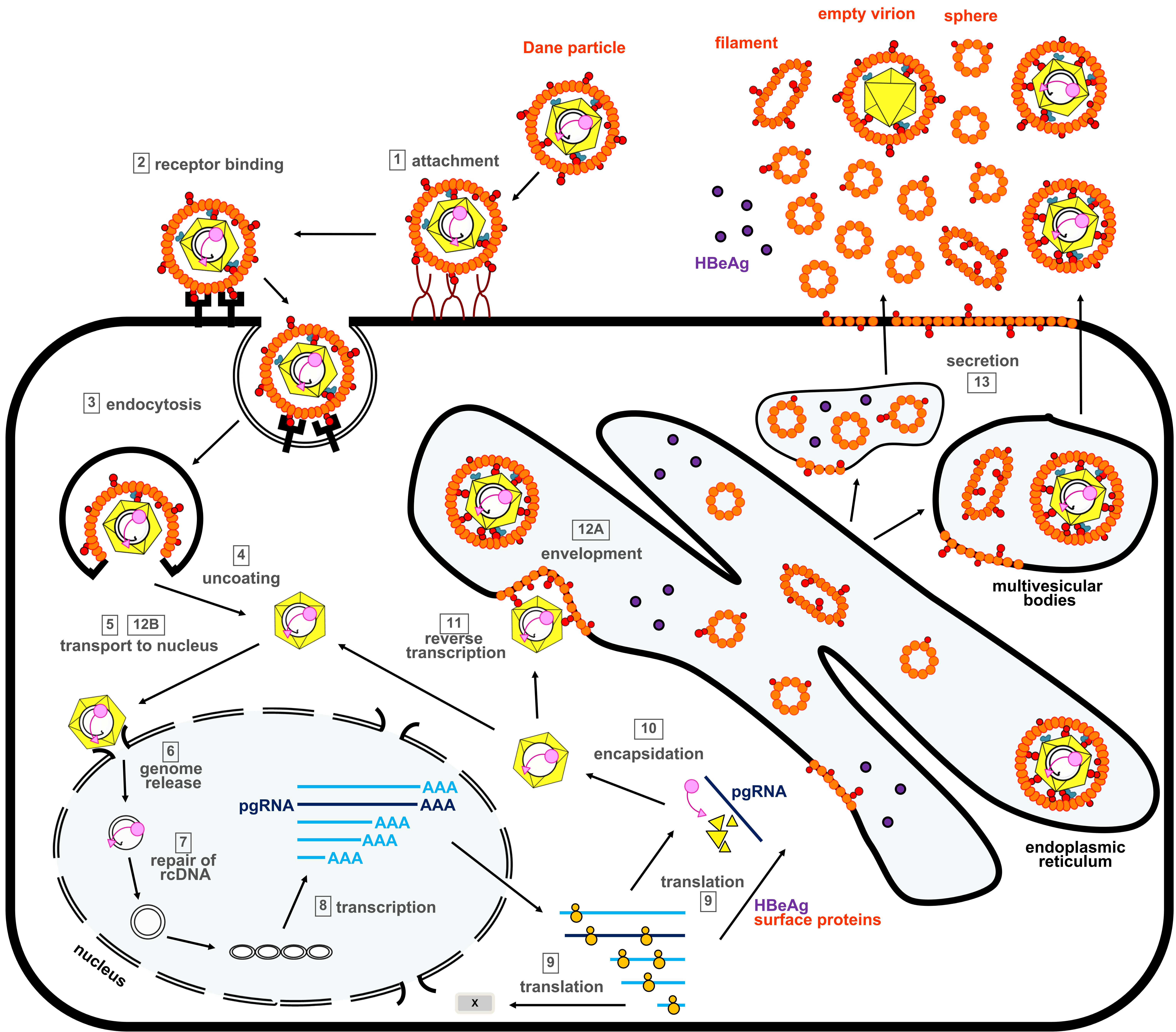
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**Figure 1. Schematic representation of the HBV life cycle:** (1) unspecific and reversible attachment of HBV to heparin sulfate proteoglycans, (2) specific binding to NTCP, (3) endocytosis, (4) uncoating and (5) intracellular trafficking of the viral capsid, (6) release of the rcDNA genome into the nucleus, (7) conversion of rcDNA to the cccDNA persistence form, (8) transcription, (9) translation, (10) encapsidation and (11) reverse transcription of a pregenomic RNA, (12A) envelopment and (12B) re-import of mature capsid into the nucleus, (13) secretion of progeny virions.



Figure 1



- |  |  |  |                  |  |                                       |  |   |
|--|--|--|------------------|--|---------------------------------------|--|---|
|  | myristoylated preS1-domain of HBV L-protein  |  | HBV core protein |  | HBx                                   |  | heparan sulfate proteoglycans                         |
|  | preS2-domain of HBV L- and M-protein         |  | HBV polymerase   |  | heat shock cognate protein 70 (Hsc70) |  | sodium taurocholate cotransporting polypeptide (NTCP) |
|  | S protein / S-domain of HBV L- and M-protein |  | HBeAg            |  | ribosome                              |  |   |



## **Highlights**

- Current antiviral treatment of HBV can control viral replication, but still is far from achieving a cure.
- A detailed understanding of the HBV life cycle and viral-host interaction might enable the development of curative therapies.
- A variety of viral and host factors are currently exploited for the development of new antivirals.
- Key viral and host targets for anti-HBV therapy and potential targeting approaches are discussed