

An *in vivo* high-throughput screening for riboswitch ligands using a reverse reporter gene system

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**Supplementary table 1.** Criteria: (1) mean RLU/OD > 4 x  $\sigma_{\min}$  control (t=3 h); (2) raw RLU > mean RLU + 3 x  $\sigma_{\min}$  control ; (3) mean RLU/OD > 4 x  $\sigma_{\min}$  control (t=3.5 h); (4) low auto luminescence at t<sub>0</sub>; (5) no edge effect. "+" means the measurements meet the criterion. "-" means that the compound was excluded due to this criterion.

Compound	Criteria 1 (>407 RLU/OD)	Criteria 2	Criteria 3	Criteria 4	Criteria 5
1	429.38	-			
2	783.51	-			
3	417.18	-			
4	828.40	-			
5	449.70	-			
6	790.96	-			
7	965.52	-			
8	418.54	-			
9	473.87	-			
10	456.46	-			
11	504.98	-			
12	434.24	-			
13	503.14	-			
14	410.26	-			
15	1133.86	-			
16	771.80	-			
17	<b>845.28</b>	+	+	+	+
18	<b>2223.17</b>	+	+	+	+
19	<b>1107.94</b>	+	+	+	+
20	<b>1546.12</b>	+	+	+	+
21	483.38	-			
22	1495.33	-			
23	413.03	+	-		
24	517.60	+	-		
25	650.11	-			
26	416.22	+	-		
27	602.69	+	+	+	-
28	1058.27	+	+	-	

**Supplementary table 2.** *E. coli* and *B. subtilis* strains used in this study.

XL1 blue	<i>endA1 gyrA96(nalR) thi-1 recA1 relA1 lac glnV44 F[Tn10 proAB+ lacIq</i>	Stratagene
	<i>Δ(lacZ)M15] hsdR17(rK- mK+) tetR</i>	
DH5α	F- <i>Φ80lacZΔM15 Δ(lacZYA-argF)</i> U169 <i>recA1 endA1 hsdR17 (rK-, mK+) phoA supE44 λ-thi-1 gyrA96 relA1</i>	laboratory stock
W168	wild type, <i>trpC2</i>	laboratory stock
BS2	W168 <i>amyE::pSB<sub>BS</sub>1C-P<sub>blaP</sub>-lacZ</i>	this paper
BS41	W168 <i>amyE::pSB<sub>BS</sub>1C-P<sub>blaP</sub>-lux</i>	17
BS44 ( <i>xpt RS lacZ</i> )	W168 <i>amyE::pSB<sub>BS</sub>1C-P<sub>blaP</sub>-lacZ thrC::pXT-B.ant_xptRS-bla/</i>	this paper
BS47 ( <i>xpt RS lux</i> )	W168 <i>thrC::pXT-B.ant_xptRS-bla/ amyE::pSB<sub>BS</sub>1C-P<sub>blaP</sub>-lux</i>	this paper
BS118 ( <i>Δ RS lux</i> )	W168 <i>amyE::pSB<sub>BS</sub>1C-P<sub>blaP</sub>-lux thrC::pXT-P<sub>xyr</sub>SD<sub>B.ant</sub>xptRS-bla/</i>	this paper
BS140 ( <i>Δ RS lacZ</i> )	W168 <i>amyE::pSB<sub>BS</sub>1C-P<sub>blaP</sub>-lacZ thrC::pXT-P<sub>xyr</sub>SD<sub>B.ant</sub>xptRS-bla/</i>	this paper

**Supplementary Table 3:** Plasmids used in this study.

Name	Description	Construction / Reference
#157	pSB <sub>BS</sub> 1C-P <sub>blaP</sub> - <i>lacZ</i>	17
#197	pSB <sub>BS</sub> 1C-P <sub>blaP</sub> - <i>gfp</i>	17
#209	pSB <sub>BS</sub> 1C-P <sub>blaP-</sub> <i>luxABCDE</i>	17
#171	pXT-BS-RS- <i>bla/</i>	17
#207	W168 <i>thrC::pXT-B.ant_xptRS-bla/</i>	plasmid #171 amplified with primer pair o301/o302 and o259 cloned with golden gate cloning ( <i>Bsal</i> )
#253	<i>thrC::pXT-P<sub>xyr</sub>SD<sub>B.ant</sub>xptRS-bla/</i>	pXT backbone amplified with o301/o363, cloned with golden gate cloning ( <i>Bsal</i> )

**Supplementary Table 4.** Primers and oligos used in this study. Restriction enzyme recognition sites are in italics, restriction sites are underlined.

o259	BaXptRS	TATGGTCTCA <u>ATCCAATAAATAGTTAGCTACACTCATATAATCGCGGGGATATGGC</u> CTGCAAGTTCTACCGAAGTACCGTAA <u>ACTTTGACTATGAGTGAGGACGAATAT</u> ATTGCTTGTTAGCATT <u>CTTTTGCGAAACTCCAAAAGCGCGTCTCTCACTTGTA</u> ACGAGTGGTGGCGG <u>CTTTGGAGTTTTTATTGCATAAGAGGGGGAACAAACAT</u> <u>GAAGAGACCATT</u>
o301	pXT- <i>bla</i> GGCFwd	TATGGTCT <u>CAATGAAAAAAATACCTCAAATCTCTGATGC</u>
o302	pXT- <i>P<sub>xyr</sub></i> GGCRev	TATGGTCT <u>CAGGATCCTCTAGAGTCGACCTGC</u>
o363	dRSSD <sub>xptB.ant</sub> _ctr_rev	TATGGTCT <u>CATCATGTTGTC</u> CCCCTCTGGATCCTCTAGAGTCGAC