Supplementary Figures:



50 mM But

Supplementary Figure 1. WT mice orally treated with indicated butyrate concentrations for 21 days. The frequency of Foxp3⁺CD4⁺ cells was determined by flow cytometry analysis. For the statistical analysis, the one-way analysis of variance (ANOVA) was used. Bars represent the mean \pm SEM; n.s. = not significant. **, P = 0.001-0.01.

100 mM But.



Supplementary Figure 2. Naïve CD4⁺CD62L⁺GFP⁻ T cells were sorted from LN and spleen of DEREG mice and differentiated into Tregs in the presence of indicated concentrations of sodium butyrate. The frequencies of GFP⁺ (Foxp3⁺) cells was determined by FACS analysis on day 5 of cell culture. Two similar experiment were performed.



Supplementary Figure 3. (A and B) $CD4^+$ T cells were differentiated into Tregs in the presence of increasing concentrations of sodium butyrate (0, 0.25 mM, 0.5 mM and 1 mM). At day 6 of the cell culture, the intracellular staining for IL-4 and IL-17A was performed. Bars represent the mean \pm SEM of IL-17A⁺ (A) and IL-4⁺ (B) T cells, respectively. Three independent experiments were performed.



Supplementary Figure 4. (**A** and **B**) The percentage of Foxp3⁺ Tregs within splenic CD4⁺ T cell population in WT, *Ffar2^{-/-}Ffar3^{-/-}* and *Slc5a8^{-/-}* mice was analysed by FACS analysis. Two similar experiments were performed. (**C**) CD4⁺ T cells isolated from LNs and spleens of WT, *Ffar2^{-/-}Ffar3^{-/-}* and *Slc5a8^{-/-}* mice were cultured under optimal Treg conditions (2 ng/ml TGF- β 1 and 100 U/ml rhIL-2) for three days. The frequency of Foxp3⁺ cells was analysed by flow cytometry. Bars represent the mean ± SEM. (**D**) WT, *Ffar2^{-/-}Ffar3^{-/-}* and *Slc5a8^{-/-}* CD4⁺ T cells were cultured under Treg-inducing conditions (1 ng/ml TGF- β 1 and 100 U/ml rhIL-2) and the percentage of Foxp3⁺ cells was determined by FACS analysis. Bars represent the mean ± SEM. Two experiments were performed.



Supplementary Figure 5. (A) CD4⁺ T cells were isolated from WT mice and cultured under Treg-inducing conditions in the presence or absence of 0.25 mM butyrate for 3 days. Western blot analysis shows the pan-acetylation of histones H3. Two experiments were performed. (B and C) ChIP analysis of acetylated state of H3 at the promotor region of *Ifn* γ , *Tbx21* and *Foxp3* in CD4⁺ T cells cultured under Treg-inducing conditions in the presence of 0.25 mM butyrate (*Ifn* γ and *Tbx21*) or 0.25 and 1 mM butyrate (*Foxp3*) was performed using an anti-acetyl-H3 antibody. For the ChIP analysis at the *Foxp3* promoter, naïve CD4⁺ T cells were used as the control. *, P = 0.01-0.05, **, P = 0.001-0.01, n.s. = not significant. Two experiments were performed.



Supplementary Figure 6. Representative dot blots showing IFN- γ and IL-17A expression in CD4⁺ T cells cultured under Th1 conditions for 6 days in the presence of indicated butyrate concentrations. Three independent experiments were performed.



Supplementary Figure 7. (A) Impact of butyrate on the expression of *Ifn* γ and *Tbx21* in nonpolarized and polarized WT CD4⁺ T cells was anylased by RT-PCR. (B) WT and *Tbx21^{-/-}* CD4⁺ T cells were cultured under Th17- or Th2-polarizing conditions for 6 days in the absence or presence of butyrate. After 6 days of the cell culture, the qRT-PCR analysis of *Roryt* (Th17 cells) and *Gata3* (Th2 cells) was performed. Data are displayed as the mean ± SEM from two experiments; **, P = 0.001-0.01, ***, P <0.001., n.s. = not significant.



Supplementary Figure 8. (A) WT mice were orally given 2% DSS into the drinking water for 5 days. Weight loss was monitored each day throughout the course of the experiment; *, P = 0.01-0.05. Two experiments were performed. (B) The percentage of T-bet⁺ (left) and IFN- γ^+ (right) cells within the colonic CD4⁺ T cell population was analysed by flow cytometry. Two independent experiments were performed; *, P = 0.01-0.05

Supplemental Table 1

Colonic content	GF mice	GF mice + DSS	<i>GF</i> + <i>DSS</i> + 100
			mM Butyrate
Acetate	-	-	-
Propionate	-	-	-
Butyrate	-	-	$3,8 \pm 0.48$

Supplemental Table 1. Colonic contents (µmol/g luminal content) measured in the indicated experimental groups.