

Accepted Manuscript

Extensive processing of sediment pore water dissolved organic matter during anoxic incubation as observed by high-field mass spectrometry (FTICR-MS)

Juliana Valle, Michael Gonsior, Mourad Harir, Alex Enrich-Prast, Philippe Schmitt-Kopplin, David Bastviken, Ralf Conrad, Norbert Hertkorn



PII: S0043-1354(17)30931-4

DOI: [10.1016/j.watres.2017.11.015](https://doi.org/10.1016/j.watres.2017.11.015)

Reference: WR 13345

To appear in: *Water Research*

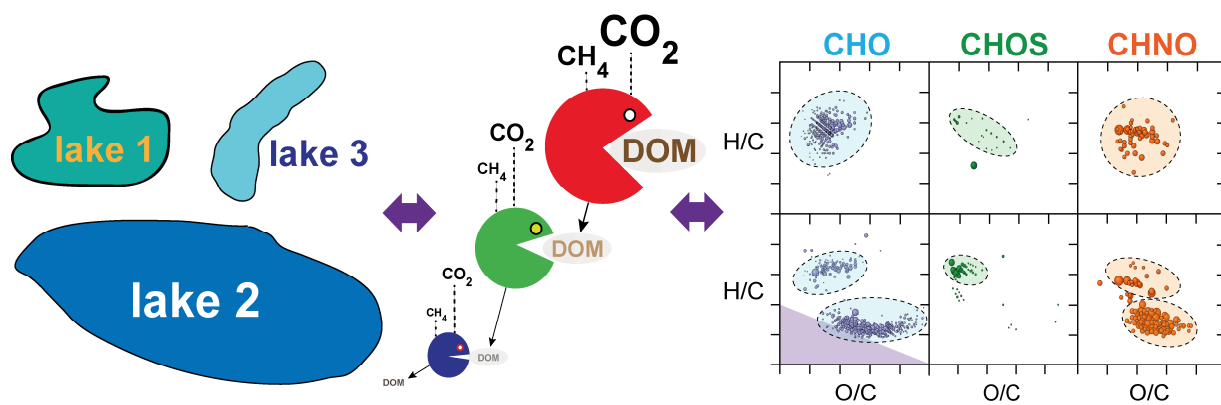
Received Date: 19 May 2017

Revised Date: 14 September 2017

Accepted Date: 5 November 2017

Please cite this article as: Valle, J., Gonsior, M., Harir, M., Enrich-Prast, A., Schmitt-Kopplin, P., Bastviken, D., Conrad, R., Hertkorn, N., Extensive processing of sediment pore water dissolved organic matter during anoxic incubation as observed by high-field mass spectrometry (FTICR-MS), *Water Research* (2017), doi: 10.1016/j.watres.2017.11.015.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



1 **Extensive processing of sediment pore water dissolved organic matter during anoxic**
2 **incubation as observed by high-field mass spectrometry (FTICR-MS)**

3

4 *Juliana Valle^a, Michael Gonsior^b, Mourad Harir^{a,c}, Alex Enrich-Prast^{d,e*}, Philippe Schmitt-Kopplin*
5 *^{a,c}, David Bastviken^d, Ralf Conrad^f and Norbert Hertkorn^{a*}*

6

7 ^a Helmholtz Zentrum Munich, German Research Center for Environmental Health, Neuherberg,
8 Germany

9 ^b University of Maryland Center for Environmental Science, Chesapeake Biological Laboratory,
10 Solomons, USA

11 ^c Department for Chemical-Technical Analysis, Research Center Weihenstephan for Brewing and
12 Food Quality, Technische Universität München, Freising-Weihenstephan, Germany

13 ^d Linköping University, Department of Thematic Studies – Environmental Change, Linköping,
14 Sweden

15 ^e Federal University of Rio de Janeiro, Department of Botany, Rio de Janeiro, Brazil

16 ^f Max Planck Institute for Terrestrial Microbiology, Marburg, Germany

17

18 *corresponding authors: e-mail: hertkorn@helmholtz-muenchen.de, alex.enrich.prast@liu.se

19

20 **Abstract**

21 Dissolved organic matter (DOM) contained in lake sediments is a carbon source for many microbial
22 degradation processes, including aerobic and anaerobic mineralization. During anaerobic degradation,
23 DOM is partially consumed and transformed into new molecules while the greenhouse gases methane
24 (CH₄) and carbon dioxide (CO₂) are produced. In this study, we used ultrahigh resolution mass
25 spectrometry to trace differences in the composition of solid-phase extractable (PPL resin) pore water
26 DOM (SPE-DOM) isolated from surface sediments of three boreal lakes before and after 40 days of
27 anoxic incubation, with concomitant determination of CH₄ and CO₂ evolution. CH₄ and CO₂
28 production detected by gas chromatography varied considerably among replicates and accounted for

29 fractions of $\sim 2\text{-}4 \times 10^{-4}$ of sedimentary organic carbon for CO_2 and $\sim 0.8\text{-}2.4 \times 10^{-5}$ for CH_4 . In contrast,
30 the relative changes of key bulk parameters during incubation, such as relative proportions of
31 molecular series, elemental ratios, average mass and unsaturation, were regularly in the percent range
32 (1-3% for compounds decreasing and 4-10% for compounds increasing), i.e. several orders of
33 magnitudes higher than mineralization alone. Computation of the average carbon oxidation state in
34 CHO molecules of lake pore water DOM revealed rather non-selective large scale transformations of
35 organic matter during incubation, with depletion of highly oxidized and of highly reduced CHO
36 molecules, and formation of rather non-labile fulvic acid type molecules. In general, proportions of
37 CHO compounds slightly decreased. Nearly saturated CHO and CHOS lipid-like substances declined
38 during incubation: these rather commonplace molecules were less specific indicators of lake sediment
39 alteration than the particular compounds, such as certain oxygenated aromatics and carboxyl-rich
40 alicyclic acids (CRAM) found more abundant after incubation. There was a remarkable general
41 increase in many CHNO compounds during incubation across all lakes. Differences in DOM
42 transformation between lakes corresponded with lake size and water residence time. While in the
43 small lake Svarttjärn, carboxyl-rich alicyclic molecules (CRAM) increased during incubation, lignin-
44 and tannin-like compounds were enriched in the large lake Bisen, suggesting selective preservation of
45 these rather non-labile aromatic compounds rather than recent synthesis. SPE-DOM after incubation
46 may represent freshly synthesized compounds, leftover bulk DOM which is primarily composed of
47 intrinsically refractory molecules and/or microbial metabolites which were not consumed in our
48 experiments. In spite of a low fraction of the total DOM being mineralized to CO_2 and CH_4 , the more
49 pronounced change in molecular DOM composition during the incubation indicates that diagenetic
50 modification of organic matter can be substantial compared to complete mineralization.

51

52 **Keywords:** solid phase extraction, DOM, FTICR-MS, sediment, methane, anaerobic respiration

53

54 1. Introduction

55 Dissolved organic matter (DOM) represents one of the largest active carbon pools and also one of the
56 most complex mixtures of organic molecules on Earth (Hertkorn et al., 2007). Organic matter cycling

57 in lakes is now considered relevant for global carbon budgets because of substantial processing of
58 incoming DOM from the catchment, leading to burial of refractory organic matter in lake sediments
59 simultaneously accompanied by large-scale formation and emissions of carbon dioxide (CO₂) and
60 methane (CH₄) from lakes (Tranvik et al., 2009). It has been proposed that allochthonous
61 (terrigenous) dissolved organic carbon (DOC) input is the major source of organic matter in boreal
62 lake sediments (von Wachenfeldt et al., 2008), and that up to 50% of the carbon mineralization occurs
63 in the sediments (Jonsson et al., 2001). However, many studies have only used bulk parameters such
64 as organic carbon and nitrogen contents as indicators of organic matter quality and lability (Bernardes
65 et al., 2004; Conrad et al., 2011).

66 Methanogenesis is considered the last step in microbial degradation of sediment organic matter, and
67 occurs after the depletion of other inorganic electron acceptors, such as oxygen (O₂), nitrate (NO₃⁻),
68 ferric iron (Fe³⁺), and sulfate (SO₄²⁻) (Conrad, 1996). During anaerobic organic matter degradation
69 first the major (bio)polymers are decomposed to oligomers and monomers, followed by fermentation
70 reactions which produce fatty acids, alcohols, acetate, CO₂ and hydrogen (H₂) – the last three being
71 the main substrates of methanogenesis (Conrad et al., 2011). Nevertheless, degradation of organic
72 matter under methanogenic conditions can account for a major share of total organic matter
73 metabolism (20-56 %; Bastviken, 2009). Previous studies have found a positive relationship between
74 CH₄ production rates and the content of organic carbon as well as a preferential degradation of algal
75 material (Conrad et al., 2011; Schulz and Conrad, 1995; West et al., 2012). However, the chemical
76 nature of the organic substances that are converted to CO₂ and CH₄ remain largely unknown, although
77 sequential processing of various carbon pools has been reported (Mahmoudi et al., 2017).

78 DOM plays an important role in the transfer of energy and matter in aquatic ecosystems, and the
79 anaerobic decomposition of sediment DOM can be extensive. On the example of lake Gäddtjärn,
80 Chmiel et al. (2016) found that, even when sedimentary organic carbon mineralization contributed
81 less than one fifth to the total lake CO₂ emissions, sediment mineralization still overcame carbon
82 burial about three-fold. In addition, regarding the potential for DOM under different conditions, the
83 total degradation potential of lake water DOM was found highest under oxic conditions, but about one
84 tenth of DOM was promptly and exclusively mineralized under anoxic conditions (Bastviken et al.,

85 2004). Extensive anoxic processing of DOM has also been demonstrated in a variety of soil and
86 marine conditions (Hodgkins et al., 2014, 2016; Schmidt et al., 2017; Seidel et al., 2014). A typical
87 feature of previous studies of anaerobic DOM decomposition is that they are based on formation of
88 degradation end-products, such as CO₂ or CH₄. However, transformations of DOM that does not lead
89 to formation of end-products could potentially be very important in generating significant metabolic
90 energy for food webs, as well as resulting in new DOM molecules that progressively become more
91 refractory and are later preserved in the sediments as a carbon sink. There is also a possibility that
92 microbial processing leads to a gradual shift from terrestrial plant derived DOM to more microbial
93 derived DOM (Einsiedl et al., 2007). Here we assess the relative magnitudes of the anaerobic DOM-
94 to-DOM transformations versus the complete DOM degradation to CO₂ and CH₄, hypothesizing, that
95 both are equally important. We also ask how the DOM pool is altered in terms of elemental
96 composition, oxidation state, and change in distribution of molecular formulas during decomposition,
97 under the hypothesis that the relative abundance of reprocessed or microbial refractory DOM
98 compounds should get progressively stronger over time, and hence differ between lakes having
99 different sizes and residence times.

100 Ultrahigh resolution Fourier transform ion cyclotron resonance mass spectrometry (FTICR-MS),
101 which can resolve thousands of individual molecular formulae within natural DOM samples
102 (Hertkorn et al., 2008), has been already been successfully used to characterize DOM from
103 freshwater environments (Einsiedl et al., 2007; Gonsior et al., 2016; Jaffé et al., 2012; Kellerman et al.
104 2015), to trace changes in DOM composition after exposure to photo- and biodegradation along
105 natural gradients (Gonsior et al., 2014, 2009; Medeiros et al., 2015; Ide et al., 2017) and incubation
106 experiments (Hodgkins et al., 2014; Medeiros et al., 2015; Seidel et al., 2016). FTICR mass
107 spectrometry has also been applied to investigate pore water in freshwater, estuarine and marine
108 sediments (Seidel et al., 2014; Schmidt et al., 2009, 2011, 2017; Tremblay et al., 2007) and soils
109 (D'Andrilli et al., 2010; Hodgkins et al., 2014, 2016). Mass spectrometry has also revealed changes of
110 DOM in bog and fen pore water profiles of peatlands (D'Andrilli et al., 2010; Hodgkins et al., 2016),
111 and possible relations with increased production of the greenhouse gases CH₄ and CO₂ in permafrost
112 environments (Hodgkins et al., 2014). In this study we have used negative mode electrospray

113 ionization (ESI) FTICR mass spectrometry to determine the extent and molecular selectivity of DOM
114 alterations in boreal lake sediment pore water during experimental anoxic incubation, alongside with
115 determination of CH₄ and CO₂ production.

116

117 **2. Material and Methods**

118

119 **2.1. Site description**

120 Sediment was collected from three lakes in the Malingsbo area, Sweden (Svartjärn: 59°53'25.12"N
121 15°15'26.68"E; Grästjärn: 59°53'18.43"N 15°21'22.64"E; Bisen: 59°57'17.36"N 15°29'37.44"E), in
122 August 2012 (Fig. 1). The lakes investigated are geographically close, but vary in both size and depth
123 (Table 1). Bisen is the largest and deepest one, with 43 ha of area and 17 m of maximum depth.
124 Grästjärn and Svartjärn have an area of 9.5 ha and 0.7 ha, and a maximum depth of 8 m and 7 m,
125 respectively (see Table 1 for lake characteristics). The difference in lake area and volume causes
126 variable water residence time (Hanson et al., 2011), which in turn influences the average age of the
127 surface sediments and of the DOC found in the lakes. While lake Bisen is expected to have the longest
128 residence time, lake Svartjärn shows exceptionally high drainage ratio and short water residence time
129 (von Wachenfeldt and Tranvik, 2008).

130

131 **2.2. Sampling**

132 Five replicates of surface sediment samples (upper 10 cm) were collected from a boat approximately
133 at the deepest water depth of the lake using a small gravity corer with minimal exposure to air. The
134 samples were homogenized and transferred into 125 ml polyethylene bottles, which were completely
135 filled with sediment sample, avoiding air bubbles inside. The bottles were kept cold (4 °C), stored in
136 the dark and shipped by air to the Max Planck Institute for Terrestrial Microbiology, Marburg,
137 Germany, for incubation experiments. In the field, O₂ profiles in the water column were acquired
138 using an oximeter (Hach HQ40 optical DO sensor) with a depth resolution of 0.5 and 1.0 m.

139

140 **2.3. Incubation experiment**

141 An analogous incubation experiment has been described elsewhere (Conrad et al., 2007, 2011).
142 Approximately 9 ml of sediment were placed into 27 ml sterile glass vials and closed with rubber
143 stoppers. Vials were then flushed with nitrogen gas (N₂) and vortexed, sequentially and repeatedly,
144 ensuring that all initial gas phase was composed solely of nitrogen. Initial gas measurements proved
145 CH₄ concentration in the headspace to be zero. Then, the sediments were incubated in the dark at
146 15°C, higher than the actual sediment temperature (Table 1). The observed CH₄ and CO₂ production
147 rates represent the potential of the microbial community and allow for timescales practical in
148 laboratory experiments (Hodgkins et al., 2014). CH₄ and CO₂ concentrations were measured regularly
149 during the incubation period of 40 days (Fig. 2). Gas samples (250 µl) were taken from the headspace
150 and injected in the Gas Chromatograph (Shimadzu GC-8A, Japan). Organic carbon (OC) content in
151 the sediments was measured in one of the replicates of each sample at the end of the incubation
152 experiment. Samples were acidified (to remove possible carbonates) and dried. The carbon content
153 was analyzed at the Centre for Stable Isotope Research and Analysis (KOSI) at Göttingen University
154 (Germany) using an elemental analyzer (NA 2500, CE Instruments, Rodano, Italy).

155

156 **2.4. Chemical analyses**

157 Water samples from the lakes were analyzed for Dissolved Organic Carbon (DOC), Total Nitrogen
158 (TN) and Total Phosphorus (TP). DOC was measured with a TOC analyzer (Shimadzu 5000, Japan).
159 TN and TP were analyzed colourimetrically after persulfate oxidation using an AA3 autoanalyzer
160 (Bran Luebbe, Germany). The pore water DOM was extracted by a previously described, adapted
161 solid phase extraction (SPE) method (Dittmar et al., 2008). For each replicate, the pore water of about
162 2 ml sediment was isolated by centrifugation and the leftover sediment was then washed three times
163 with 1 ml ultra-pure water (LC-MS Chromasolv®, Sigma-Aldrich). The supernatants of the
164 centrifugation and washing steps were combined. Then, approximately 4 ml liquid sample per
165 replicate was acidified to pH 2 with formic acid and passed through a SPE cartridge (Bond Elut PPL,
166 100 mg, 1 ml, Agilent). The cartridge had been activated with 1 ml methanol (LC-MS Chromasolv®,
167 Sigma-Aldrich) and rinsed with 1 ml acidified pure water (pH 2) prior to extraction. In the end, DOM
168 was eluted with 500 µl methanol. This procedure was repeated for all five replicates, before and after

169 the incubation experiment (total of ten samples per lake). The eluates were stored in the freezer (-20
170 °C) until mass spectrometry analysis.

171

172 **2.5. FTICR Mass Spectrometry**

173 Negative electrospray ionization Fourier transform ion cyclotron resonance [ESI(-)] FTICR mass
174 spectra were acquired using a 12T Bruker Solarix mass spectrometer (Bruker Daltonics, Bremen,
175 Germany) and an Apollo II electrospray ionization (ESI) source in negative mode (Hertkorn et al.,
176 2016). Nebulizer gas pressure, drying gas pressure and the source heater temperature were 138 kPa,
177 103 kPa and 200°C, respectively. The spectra were acquired with a time domain of 4 MW and five
178 hundred scans were accumulated for each mass spectrum. Further information can be found in
179 Support Online Information.

180

181 **2.6. Statistical analysis**

182 Hierarchical Cluster Analysis (HCA) was performed using the Hierarchical Clustering Explorer 3.0
183 (HCE; <http://www.cs.umd.edu/hcil/multi-cluster/>), with clustering of the dataset using the Average
184 Linkage (UPGMA) method and the Pearson correlation coefficient (Pearson's r) as the
185 similarity/distance measure. Based on the HCA, we used the "profile search" tool from HCE 3.0,
186 choosing a search method (model-based), a distance measure (Pearson's r) and a threshold (0.9). With
187 these settings, we selected mass peaks that were increased in relative abundance in a chosen subset,
188 but decreased in another one. All molecular compositions which were attributed to have increased and
189 decreased in relative abundance had significantly changed between the different incubation states ($p <$
190 0.05 , Mann Whitney test, GraphPad Prism 5.0). Principal Component Analysis (PCA) was done with
191 the software SIMCA-P 9.0 (Umetrics AB, Sweden).

192

193 **3. Results and Discussion**

194

195 **3.1. CH_4 and CO_2 production rates**

196 Anoxic incubation of lake sediment samples resulted in continually increasing concentrations of CH₄
197 and CO₂ in the gas phase (Fig. 2). Release of CH₄ started without lag phase, indicating that reduction
198 of nitrate, ferric iron, sulfate, etc., was minimal, and that methanogenesis was the dominant
199 degradation process of organic matter (Conrad, 2007). Considerable variability in gas production rates
200 was observed among replicate sediment cores. The production rates of CH₄ varied also among the
201 different lakes. The highest CH₄ production rate ($11.8 \pm 5.8 \text{ nmol d}^{-1} \text{ g}_{\text{dw}}^{-1}$) was found in sediments of
202 Svarttjärn, followed by Grästjärn ($8.9 \pm 3.5 \text{ nmol d}^{-1} \text{ g}_{\text{dw}}^{-1}$) and Bisen ($5.9 \pm 3.1 \text{ nmol d}^{-1} \text{ g}_{\text{dw}}^{-1}$). Rates
203 of CO₂ production, however, were more similar in Svarttjärn, Grästjärn and Bisen (77.6 ± 5.9 , $113.5 \pm$
204 11.6 and $107.2 \pm 19.5 \text{ nmol d}^{-1} \text{ g}_{\text{dw}}^{-1}$, respectively). The CH₄ production rates were low when
205 compared with tropical and temperate lake sediments (Conrad et al., 2009, 2010, 2011). Duc et al.
206 (2010) have measured CH₄ production in lake Svarttjärn and found similar values ($9 \text{ nmol d}^{-1} \text{ g}_{\text{dw}}^{-1}$) at
207 10°C. Lake sediments in Alaska (Lofton et al., 2015) also presented variable CH₄ production, ranging
208 from 1.7 up to 118 $\text{nmol d}^{-1} \text{ g}_{\text{dw}}^{-1}$.

209 Cumulative release of gaseous CO₂ during incubation accounted for small fractions of $\sim 2\text{-}4 \times 10^{-4}$ of
210 total sedimentary carbon. Evolution of CH₄ accounted for even lower fractions of only $\sim 0.8\text{-}2 \times 10^{-5}$
211 (Fig. 2). While complete degradation of organic matter under exclusive methanogenic conditions
212 should result in about equal amounts of CO₂ and CH₄, organic matter degradation here occurred to a
213 larger extent by oxidation to CO₂. Such excess oxidation to CO₂ is only possible if an oxidant is
214 available. If O₂, nitrate, sulfate, ferric iron and other inorganic electron acceptors were not available
215 (as indicated by the instantaneous production of CH₄), organic compounds (e.g. humic acids) must
216 have served as oxidants (Corbett et al., 2015; Klüpfel et al., 2014; Yavitt and Seidmann-Zager, 2006).

217

218 **3.2. Compositional characteristics of SPE-DOM**

219 Molecular compositions of lake sediment porewater SPE-DOM before and after incubation were
220 identified by virtue of high mass accuracy and mass resolution typical of FTICR mass spectrometry
221 (see Supporting Online Information for comments on the depiction of molecular diversity of SPE-
222 DOM by mass spectrometry). Common mass peaks that were present in all five replicate samples
223 before and after incubation exceeded 3000 counts in sediment from Grästjärn and 4000 counts for

224 Svarttjärn and Bisen (Table 2). With the exception of CHNOS compounds, which were comparatively
225 less abundant, the counts of CHO, CHNO and CHOS molecular series between individual replicates
226 showed larger relative variance of incubated compared with original SPE-DOM samples (Table S1,
227 S2). The relative variance commonly changed in the order CHNOS > CHNO \approx CHOS > CHO
228 molecular series (Table S2).

229 On average, ~60-70% of all assigned molecular compositions were common to all five replicate
230 sediment cores in each lake (Table 2). The presence of unique compounds in each replicate
231 corroborates an appreciable chemical diversity and heterogeneity of carbon cycling in these sediments
232 (Krause et al., 2009; Siljanen et al., 2011), which possibly contributes to the high observed variability
233 of CH₄ and CO₂ production rates.

234 Hierarchical Cluster Analysis (HCA) of FTICR mass spectra showed a clear separation according to
235 the different lakes (minimum similarity: 0.55), consistent with studies on the molecular diversity of
236 DOM in aquatic systems (Ide et al., 2017) (Fig. S1). DOM compositions before and after incubation
237 differed considerably for all three lakes (minimum similarity: 0.83), suggesting turnover of DOM by
238 combined effects of microbial production and degradation or chemical transformations during the
239 incubation experiment. However, lake-specific characteristics (Table 1) were more influential for
240 DOM composition than changes during the incubation, and differences among the replicates were
241 always larger for incubated versus original samples (Fig. S2).

242 Analogous results were obtained from Principal Component Analysis (PCA). Again, clustering of
243 SPE-DOM compositions according to lakes was more pronounced than clustering according to
244 incubation or replication (Fig. S2). The relative variance of incubated samples always exceeded that
245 of original samples, while the fraction of molecular compositions common to all five replicates was
246 always smaller for original than for incubated samples (Table 2). Both trends were related and
247 suggested increasing chemical diversity caused by incubation (Fig. S1, S2).

248 When comparing the FTICR mass spectra of sediment pore water SPE-DOM from the three boreal
249 lakes before and after incubation, the compounds declining in relative abundance during incubation
250 were less abundant (~1-3%) than those increasing in relative abundance (~4-10%)(Fig. S3). Here,
251 Svarttjärn and in particular Bisen showed substantial mass range (Fig. S3) and bandwidth of

252 molecular compositions (Fig. 3) increasing in relative abundance after incubation. CO₂ and CH₄ are
253 unambiguously defined molecules and easily quantified; these are likely to result from mineralization
254 of carbon present in original organic matter, mediated by activity of microorganisms (Bastviken,
255 2009; Tranvik et al., 2009). In our experiments, the relative turnover of carbon ranged from $2-4 \times 10^{-4}$
256 (CO₂), which was likely an underestimate of total produced CO₂ because bicarbonate was not
257 considered, and $<1-2 \times 10^{-5}$ (CH₄) of total sedimentary organic carbon (Fig. 2). In contrast, the relative
258 change during incubation of key bulk parameters, such as relative proportions of molecular series,
259 elemental ratios, average mass and unsaturation, was regularly in the percent range (Table 2; Fig. S3);
260 i.e. several orders of magnitudes higher than mineralization alone. This implies that during incubation,
261 the SPE-DOM transformation was far more extensive than the readily observed mineralization, which
262 is commonly regarded as a measure of microbial activity.

263 Although the consistent increase of apparent chemical diversity during incubation in all three lakes
264 contrasts with a simple notion of degradation of organic matter from larger to smaller molecules as
265 expressed in the familiar size-reactivity continuum model (Amon and Benner, 1996; Benner and
266 Amon, 2014), analogous trends were reported for anoxic marine sediments (Seidel et al., 2014). While
267 the original SPE-DOM samples represent highly processed biogeochemical materials with smooth
268 mass peak distributions resulting from the massive projection of many isomers on one given
269 molecular composition (Hertkorn et al., 2007, 2008), the incubated samples probably contained higher
270 proportions of remnant biomolecules originating from microbial transformations. Although common
271 polydisperse biogeochemical materials such as organic matter are arguably much more complex
272 mixtures than typical biological samples (Hertkorn et al., 2007), the latter may include large
273 proportions of microbial metabolites that might produce recognizable patterns in the mass spectra. In
274 our study, patterning might apply to (the rather few) compounds decreasing during incubation (Fig.
275 S3), suggesting preferential removal of specific CHO- and CHOS-based lipid-like compounds (Fig. 4;
276 Table S3).

277 In general, more than 60% of the detected mass peaks were assigned to CHO compounds, followed by
278 CHNO molecules (17-28%), CHOS molecules (8-15%) and a few CHNOS compounds (0.4-1.3%),
279 respectively. Despite the substantial molecular diversity of DOM, the chemical diversity of lipid-like

280 (H/C ratio >1.7; O/C ratio <0.3) and carbohydrate-like DOM (H/C ratio ~1.5; O/C ratio >0.7) was
281 limited. This likely resulted from activity of hydrolytic microorganisms (Shakeri Yekta et al., 2012;
282 Walker and Wilson, 1991), which is expected for lake sediments anaerobic degradation, since
283 hydrolysis is the first step in methanogenesis and other organic matter degradation processes
284 (Bastviken, 2009; Conrad, 1999).

285 While the computed average O/C ratios of SPE-DOM uniformly declined during incubation, the
286 respective H/C values stayed near constant in Svarttjärn, whereas those in Grästjärn and Bisen
287 increased considerably (Table 2). Microbial activity has been suggested as responsible for analogous
288 trends in the sediments of the Black Sea (Schmidt et al., 2017), in peat soils in Sweden (Hodgkins et
289 al., 2016), and in biodegradation experiments in the Amazon River (Medeiros et al., 2015; Seidel et
290 al., 2016). Reactive functional groups of organic matter, which often contain oxygen, are expected to
291 react more frequently than unfunctionalized aliphatics, and the oxygen-rich molecules H₂O and CO₂
292 are commonly released during organic matter processing (Einsiedl et al., 2007), likely contributing to
293 the observed relative depletion of oxygen in organic matter during incubation (Evans and Fuchs,
294 1988; Ghattas et al., 2017).

295 Van Krevelen and KMD/z* diagrams for original and incubated sediment pore water SPE-DOM from
296 the three boreal lakes were determined for initial and final samples and for respective molecular
297 compositions growing or declining in abundance during incubation (Fig. 3, 4, S4). These diagrams
298 demonstrated coverage of a rather large area of the CHO, CHNO and CHOS compositional space for
299 initial and final SPE-DOM, indicative of a remarkable molecular diversity of these organic molecules
300 which exceeds that of common DOM present in the water column of boreal lakes (Gonsior et al.,
301 2013). Furthermore, production of CHNO compounds with average H/C and O/C ratios of ~0.5-2.0
302 and ~0.2-0.8, respectively, was found in all three boreal lake sediments (Fig. 4). Production of N-
303 containing compounds might be related to protein degradation and formation during anaerobic
304 degradation of biomass-derived substrates (Schmidt et al., 2011; Shakeri Yekta et al., 2012), or from
305 microbial biomass (Seidel et al., 2016).

306 While CHNO and CHNOS compounds covered a smaller area in the van Krevelen diagrams than
307 CHO and CHOS compounds, CHNO compounds, specifically, were mostly limited to average H/C

308 and O/C ratios of ~ 0.5-2 and ~0.2-0.8, respectively (Fig. 4). In addition, CHO compounds covered a
309 relatively large mass range (~200-850 Da). In comparison, CHNO and CHNOS compounds were
310 smaller than ~700 Da, and CHOS compounds were almost exclusively restricted to <600 Da. The
311 relative gain of heteroatom-containing molecules during incubation seemed to be a common feature
312 among the three lakes. Increased proportions of CHNO and CHOS compounds were also observed in
313 Swedish peat soils at an advanced stage of decomposition (Hodgkins et al., 2014, 2016). In our study,
314 lower mass compounds ($m/z < 500$) showed higher H/C ratios (i.e. were more aliphatic) than higher
315 mass compounds (Fig. 3) among consumed and produced compounds. The average mass of DOM
316 molecules in general declined during incubation (Table 2). Nevertheless, a number of DOM
317 molecules formed during incubation were of high mass (Fig. 3, S3, S4, S5).

318 Summary van Krevelen, mass-edited H/C ratio and KMD/z* diagrams of original and incubated SPE-
319 DOM showed extensive coverage and overlap of the CHO, CHNO and CHOS compositional space,
320 precluding clear assessment of molecular changes caused by incubation (Fig. 3, S4). KMD/z*
321 diagrams revealed extended ($n \sim 12-20$) CH₂-based homologous series for original and incubated
322 samples for CHO compounds and shorter ones ($n \sim 6-10$) for CHNO and CHOS compounds (Fig. S4).
323 Anoxic incubation of sediment caused some uniform progression of key bulk parameters (Table 2):
324 the average mass of SPE-DOM decreased, in line with greenhouse gas release from organic matter.
325 The proportions of CHO compounds consistently decreased after incubation whereas the proportions
326 of CHNO compounds uniformly increased. Higher abundances of CHNO compounds might be an
327 indication of intensive microbial activity (Hodgkins et al., 2016; Roth et al., 2014; Seidel et al., 2014).
328 Rather non labile CHNO compounds were present after incubation; hence, incubation affected the
329 coupled carbon and nitrogen cycles in lake sediments (Hedges and Oades, 1997). Oxygen content as
330 well as O/C ratios of the residual SPE-DOM decreased following incubation, partially because
331 oxidized substrates H₂O and CO₂ had been liberated. In contrast, average carbon, average double
332 bond equivalent (DBE) and DBE/C values were less affected, indicating that C-based molecular
333 skeletons were rather modified by corresponding functional groups than fundamentally rearranged
334 during incubation.

335

336 3.3. Changes according to CHO-, CHNO-, CHOS- and CHNOS molecular series in SPE-DOM

337 Two stages of sorting were required to enable a meaningful assessment of lake sediment processing
338 by anoxic incubation. First, the exclusive recognition of molecular compositions common to all five
339 respective replicates alleviated the considerable intrinsic variance of the individual incubation
340 experiments (which likely reflected the variance of organic matter molecules and that of the temporal
341 evolution of microorganisms; Fig. S5, S6; Table S1, S2) and enabled the detection of specific organic
342 matter transformation within lake sediments. Second, the separate display of CHO, CHNO, CHOS
343 and CHNOS molecular series common to all five replicates removed projection and clearly revealed
344 distinct trends for each of the three lake sediments investigated in this study. Summary van Krevelen,
345 mass-edited H/C ratio and KMD/z* diagrams for presumably consumed and produced compounds
346 during the anoxic incubation experiments were computed for three selected individual samples each
347 (Fig. S5, S6), and for common molecular compositions present in all five replicates of the six
348 incubation experiments (Fig. 3, 4, S4, Table 2, S1, S2).

349 In addition, a display for individual CHO, CHNO, CHOS and CHNOS molecular series in van
350 Krevelen and KMD/z* diagrams was provided for common molecular compositions found in all five
351 replicates (Fig. 4, S4). While the figures displaying common molecular compositions appeared more
352 distinct and “less noisy” than those resulting from individual SPE-DOM samples, a visual comparison
353 of SPE-DOM samples before and after incubation still could not retrieve relevant information in any
354 of these diagrams. The high abundance of CHO molecules in particular, and the broad coverage of the
355 CHO, CHNO and CHOS chemical space at average H/C and O/C ratios (Hertkorn et al., 2007) by
356 SPE-DOM both prior to and after incubation precluded easy determination of changes in composition
357 by visual inspection. Finally, relative changes of normalized mass peak amplitudes were computed by
358 means of “profile search” within HCA, providing depiction of compounds decreasing and increasing
359 in relative abundance during incubation. When CHO, CHNO and CHOS molecular series were shown
360 separately, distinct DOM processing during incubation according to lake was clearly revealed in van
361 Krevelen and KMD/z* diagrams (Fig. 4, S6). In general, the following changes were observed (all
362 Tables):

- 363 • Loss of certain compounds during incubation was mainly restricted to near saturated CHO and
364 CHOS lipid-like compounds (H/C ratio > 1.8) and seemed less significant than gain of specific
365 compounds (Fig. 3, 4, S3).
- 366 • Svarttjärn showed formation of CHO and CHNO-based CRAM molecules (carboxylic-rich
367 alicyclic molecules) (Fig. 4) (Hertkorn et al., 2006) which are considered exemplary microbial
368 products (Bianchi, 2011). A minor suite of common CHOS organic matter molecules of near
369 average H/C and O/C ratio was also produced.
- 370 • Grästjärn showed related but less specific trends: two groups of oxygen-deficient lipid-like CHOS
371 compounds with variable extent of unsaturation (H/C ratio ~1.5; O/C ratio < 1.1) and an extended
372 group of unsaturated CHNO compounds had appeared following incubation. A small proportion of
373 supposedly aromatic CHOS compounds (H/C ratio < 1.1; O/C ratio < 0.2) was observed in
374 Grästjärn after incubation (Fig. 4). Analogous CHOS compounds have also been found in SPE-
375 DOM samples from Amazonian rivers and German mine pit lake pore water (Gonsior et al., 2016;
376 Herzprung et al., 2010). These compounds could result from sulfurization of aromatic CHO
377 compounds by biological sulfide production under anoxic conditions (Shakeri Yekta et al., 2012).
- 378 • Bisen exhibited the most unequivocal chemical evolution of sediment organic matter. Here, an
379 extended set of CHO- and CHNO-based CRAM had been generated (although less distinct than in
380 Svarttjärn), and a particular set of unsaturated sulfolipid-like compounds. Most important,
381 however, was a very specific array of rather unsaturated (H/C ratio < 1) lignin- and tannin-like
382 CHO compounds which was found more abundant in Bisen after incubation. Another, peculiar set
383 of unsaturated CHNO compounds (H/C ratio < 1.1; mass range $m/z \sim 300-500$; Fig. 4) with
384 unclear attribution at present deserves further observance.
- 385 • It is noteworthy that both Grästjärn and Bisen showed substantial depletion of oxygen after
386 incubation, which was almost exclusively restricted to their CHOS compounds. This most likely
387 indicates reduction of sulfur in CHOS compounds, probably originating from sulfate reduction
388 (Table S4).
- 389 Compounds found more abundant after incubation may represent molecules synthesized during
390 incubation or, alternatively, compounds being less efficiently degraded during incubation. In case of

391 Bisen, general considerations about relative persistence of lake organic matter compound classes
392 (Kellerman et al., 2015; Mahmoudi et al., 2017) suggest that the lignin- and tannin-like CHO
393 compounds found at elevated relative abundance after incubation represent less efficiently degraded
394 compounds under anoxic conditions (Seidel et al., 2014) which were already present in original lake
395 Bisen sediments (leftovers of incubation) rather than newly synthesized ones. Tannin-like compounds
396 have been found characteristic of forest soil leachates (Roth et al., 2014), and the evolution of SPE-
397 DOM conforms to the land coverage present in Bisen catchment, which shows ~84% of coniferous
398 and mixed forest.

399

400

401 **3.4. Average oxidation state of carbon in CHO compounds during incubation**

402 An alternative projection of the CHO-compositional space is the depiction of the average oxidation
403 state (OS_{av}) according to $OS_{av} = 2 \times O/C - H/C$ (Kroll et al., 2011) which has been computed for
404 individual CHO compounds as well as for the consolidated MS-derived intensity weighted average of
405 all detected CHO compounds, before and after incubation (Fig. 5). Alternative indices like NOSC
406 (nominal oxidation state of carbon; LaRowe and Van Cappellen, 2011) were not considered because
407 these would have required severe estimates about (average) oxidation states of the heteroatoms sulfur
408 and nitrogen, respectively, which are not warranted at present. Here, these OS_{av} values have been
409 sorted in ranks of 0.1 width. All organic matter molecules displayed average oxidation states between
410 -1.8 and +1.3, i. e. a range smaller than that between the endmembers CH₄ (OS_{av} = -4) and CO₂
411 (OS_{av} = +4). In addition, the difference “original minus incubated” was shown to demonstrate
412 relative losses and gains of compounds according to redox status. Remarkably, OS_{av} of the
413 consolidated complement of all CHO compounds became more reduced after incubation (change
414 towards more negative values in the order Svartjärn > Grästjärn > Bisen; Fig. 5). The apparent
415 reduction of the residual organic matter was most probably an indirect consequence of the loss of (the
416 highly oxidized molecules) H₂O (in case of exclusive methanogenesis, H₂O would be taken up instead
417 of being released) and CO₂ during incubation. Interestingly, oxidized CHO molecules, which
418 contained several oxygen-containing functional groups, became preferentially depleted after anoxic
419 incubation. This behavior resembles that of aerobic processing of organic matter in a karst
420 environment in which some reduced organic carbon compounds with relatively small O/C and high
421 H/C ratio were formed, although the bulk of the organic matter became oxidized (Einsiedl et al.,
422 2007). Remarkably, in addition to high-energy substrates of considerable saturation (OS_{av} ~ -1.7 -
423 1.1; possibly bioavailable lipids), molecules with elevated OS_{av} (OS_{av} ~ -0 - 1.2; organic acids)
424 were also preferentially depleted. Such molecules will necessarily contain several oxygen-containing
425 functional groups, making those more polar and reactive than common aliphatic compounds (Burdige,
426 2007).

427 Microbial redox reactions rely on transfer of electrons from electron donor substrates to terminal
428 electron acceptors and must be thermodynamically favorable (LaRowe and Van Cappellen, 2011;

429 Thullner et al., 2007). Reduced carbon in the form of organic matter is often the ultimate source of
430 electrons in sediments, and environmental redox conditions progressively change during degradation
431 or incubation. The bioenergetic potential of organic matter and of its individual molecular constituents
432 is related to its oxidation state as expressed by OS_{av}. The Gibbs ΔG_{ox} energy for oxidation half
433 reactions of organic molecules depends on their molecular composition and structures, and is
434 commonly inversely related with the oxidation state, even when ΔG_{ox} is normalized to the number of
435 electrons released (LaRowe and Van Cappellen, 2011). Hence, the removal of electrons from an
436 organic compound becomes thermodynamically more favorable with increasing OS_{av} (Kroll et al.,
437 2011; LaRowe and Van Cappellen, 2011). In Svarttjärn and Grästjärn, potentially easily bioavailable
438 lipid-like compounds as well as oxygenated compounds were preferentially depleted during
439 incubation (cf. above; Fig. 5) whereas Bisen showed similar trends with however lesser distinction.
440 These underlying fundamental trends were not disclosed by the differential mass analysis shown
441 above (sections 3.2 and 3.3), which had revealed lake-specific organic matter transformations based
442 on relative mass peak amplitude differences exceeding a certain threshold (e.g. for CHO compounds;
443 Fig. 4). It is therefore highly probable that a directional redox evolution of organic matter during
444 incubation as expressed by OS_{av} will apply to fundamentally different chemical structures which
445 then may occupy diverging sections within van Krevelen and mass-edited H/C ratio diagrams.
446 Notably, compounds with similar OS_{av} as found in fulvic acids (OS_{av} ~ -0.52; Kroll et al., 2011;
447 LaRowe and Van Cappellen, 2011) had grown in abundance after incubation (Fig. 5). Fulvic acids
448 represent the highly processed, water soluble complement of humic substances (Hertkorn et al., 2002),
449 It appears that the extent of relative changes progressively decreased with ever increasing lake size
450 and water residence time. Here, the increase of molecular diversity in pore water DOM (Seidel et al.,
451 2014) could be explained due to the accumulation of non-labile DOM in the sediments, when
452 compared to the water column (Kellerman et al., 2014).

453

454 **4. Conclusion**

455 This study revealed rather non-selective large scale transformations of organic matter during
456 incubation and lake-specific, extensive molecular level alterations during anaerobic degradation of
457 sediment pore water DOM obtained from boreal lakes. A dependable assessment of the general and
458 the lake-specific, complex chemical transformations happening during anaerobic DOM processing
459 required mass spectrometry-based assignment of molecular compositions, computation of the average
460 carbon oxidation state in CHO molecules, averaging across replicates, consideration of gain and loss
461 of relative mass peak amplitude before and after incubation, and depiction of individual CHO, CHOS
462 and CHNO molecular series. Here, such assessments revealed that anaerobic processing and
463 transformation of pore water DOM in sediments was much more extensive than revealed just from
464 formation of the degradation end-products CO₂ and CH₄. Molecular evidence for the gradual
465 transformation from clearly reduced and oxidized DOM (fatty acid like and carboxylic rich DOM,
466 respectively) to fulvic acid and tannin like DOM with intermediate oxidation state was obtained.
467 There were also clear signs of increasing the relative abundance of nitrogen-containing DOM of likely
468 microbial origin. This study highlights the importance to link biogeochemical process studies with
469 studies of molecular composition using high resolution non-target approaches. Further studies could
470 clarify effects of study design on DOM processing in natural and artificial systems.

471

472

473 **Author contributions**

474 J.V. collected samples from Swedish lakes in August 2012; D.B. generated funding in support of this
475 research. J.V. performed gas evolution measurements at the facilities of R.C. A.P., D.B. and R.C.
476 contributed to the study design. J.V., M.G. and M.H. performed the FTICR/MS acquisition, data
477 analyses and participated in data interpretation. P.S.-K. provided support for the FTICR/MS analyses
478 and general data interpretations. Data interpretation was performed by all authors. J.V., M.H., M.G.,
479 D.B., and N.H. actively participated in the writing of the manuscript; all authors provided significant
480 input on the final manuscript.

481

482

483

484 **Acknowledgments**

485 The authors acknowledge the financial support from the Swedish Research Councils VR (2012-
486 00048) and STINT (2012-2085), and ERC (grant no. 725546). We are grateful to Peter Claus (Max
487 Planck Institute, Marburg), who assisted in the experiment logistics. J.V. thanks the Brazilian
488 National Counsel of Technological and Scientific Development (CNPq, 290004/2014-4) and the
489 Alexander von Humboldt Foundation for fellowship and financial support (Research Group Linkage
490 Brazil - Germany: Connecting the diversity of dissolved organic matter and CO₂ and CH₄ production
491 in tropical lakes). A.P. is a research fellow from CNPq and Cientista do Estado from FAPERJ
492 (Fundação de Amparo à Pesquisa do Estado do Rio de Janeiro) and uses financial resources from
493 these two funding agencies and also from CAPES (Coordenação de Aperfeiçoamento de Pessoal de
494 Nível Superior) and STINT (The Swedish Foundation for International Cooperation in Research and
495 Higher Education) for this research. M.H. thanks DAAD for the PPP exchange program with the
496 Federal University of Rio de Janeiro.

497

498

499 **References**

- 500 Amon, R.M.W., Benner, R., 1996. Bacterial utilization of different size classes of dissolved organic
501 matter. *Limnol. Oceanogr.* 41, 41–51. doi:10.4319/lo.1996.41.1.0041.
- 502 Bastviken, D., Persson, L., Odham, G., Tranvik, L., 2004. Degradation of dissolved organic matter in
503 oxic and anoxic lake water. *Limnol. Oceanogr.* 49, 109–116. doi:10.4319/lo.2004.49.1.0109.
- 504 Bastviken, D., 2009. Methane, in: Likens, G.E. (Ed.), *Encyclopedia of Inland Waters*. Elsevier,
505 Oxford, pp. 783–805.
- 506 Benner, R., Amon, R.M.W., 2014. The Size-Reactivity Continuum of Major Bioelements in the
507 Ocean. *Ann. Rev. Mar. Sci.* 1–21. doi:10.1146/annurev-marine-010213-135126.
- 508 Bernardes, M.C., Martinelli, L.A., Krusche, A.V., Gudeman, J., Moreira, M., Victoria, R.L., Ometto,
509 J.P.H.B., Ballester, M.V.R., Aufdenkampe, A.K., Richey, J.E., Hedges, J.I., 2004. Riverine Organic
510 Matter Composition as a Function of Land Use Changes, Southwest Amazon. *Ecol. Appl.* 14, 263–
511 279. doi:10.1890/01-6028.
- 512 Bianchi, T.S., 2011. The role of terrestrially derived organic carbon in the coastal ocean: a changing
513 paradigm and the priming effect. *Proc. Natl. Acad. Sci. U. S. A.* 108, 19473–81.
514 doi:10.1073/pnas.1017982108.
- 515 Burdige, D.J., 2007. Preservation of Organic Matter in Marine Sediments: Controls, Mechanisms, and
516 an Imbalance in Sediment Organic Carbon Budgets? *Chem. Rev.* 107, 467–485.
517 doi:10.1021/cr050347q.
- 518 Chmiel, H.E., Kokic, J., Denfeld, B.A., Einarsdóttir, K., Wallin, M.B., Koehler, B., Isidorova, A.,
519 Bastviken, D., Ferland, M.-È., Sobek, S., 2016. The role of sediments in the carbon budget of a small
520 boreal lake. *Limnol. Oceanogr.* 61, 1814–1825. doi:10.1002/lno.10336.
- 521 Conrad, R., 1996. Soil Microorganisms as Controllers of Atmospheric Trace Gases (H₂, CO, CH₄,
522 OCS, N₂O, and NO). *Microbiol. Rev.* 60, 609–640. doi:0146-0749/96/\$04.00?0.

- 523 Conrad, R., 1999. Contribution of hydrogen to methane production and control of hydrogen
524 concentrations in methanogenic soils and sediments. *FEMS Microbiol. Ecol.* 28, 193–202.
525 doi:10.1111/j.1574-6941.1999.tb00575.x.
- 526 Conrad, R., 2007. Microbial ecology of methanogens and methanotrophs, in: *Advances in Agronomy*.
527 Elsevier Inc., pp. 1–63.
- 528 Conrad, R., Chan, O.-C., Claus, P., Casper, P., 2007. Characterization of methanogenic Archaea and
529 stable isotope fractionation during methane production in the profundal sediment of an oligotrophic
530 lake (Lake Stechlin, Germany). *Limnol. Oceanogr.* 52, 1393–1406. doi:10.4319/lo.2007.52.4.1393.
- 531 Conrad, R., Claus, P., Casper, P., 2009. Characterization of stable isotope fractionation during
532 methane production in the sediment of a eutrophic lake, Lake Dagow, Germany. *Limnol. Oceanogr.*
533 54, 457–471. doi:10.4319/lo.2009.54.2.0457.
- 534 Conrad, R., Klose, M., Claus, P., Enrich-Prast, A., 2010. Methanogenic pathway, ^{13}C isotope
535 fractionation, and archaeal community composition in the sediment of two clear-water lakes of
536 Amazonia. *Limnol. Oceanogr.* 55, 689–702. doi:10.4319/lo.2010.55.2.0689.
- 537 Conrad, R., Noll, M., Claus, P., Klose, M., Bastos, W.R., Enrich-Prast, A., 2011. Stable carbon
538 isotope discrimination and microbiology of methane formation in tropical anoxic lake sediments.
539 *Biogeosciences* 8, 795–814. doi:10.5194/bg-8-795-2011.
- 540 Corbett, J.E., Tfaily, M.M., Burdige, D.J., Glaser, P.H., Chanton, J.P., 2015. The relative importance
541 of methanogenesis in the decomposition of organic matter in northern peatlands. *J. Geophys. Res.*
542 *Biogeosciences* 120, 280–293. doi:10.1002/2016JC011790.
- 543 D'Andrilli, J., Chanton, J.P., Glaser, P.H., Cooper, W.T., 2010. Characterization of dissolved organic
544 matter in northern peatland soil porewaters by ultra high resolution mass spectrometry. *Org.*
545 *Geochem.* 41, 791–799. doi:10.1016/j.orggeochem.2010.05.009.

- 546 Dittmar, T., Koch, B., Hertkorn, N., Kattner, G., 2008. A simple and efficient method for the solid-
547 phase extraction of dissolved organic matter (SPE-DOM) from seawater. *Limnol. Oceanogr. Methods*
548 6, 230–235. doi:10.4319/lom.2008.6.230.
- 549 Duc, N.T., Crill, P., Bastviken, D., 2010. Implications of temperature and sediment characteristics on
550 methane formation and oxidation in lake sediments. *Biogeochemistry* 100, 185–196.
551 doi:10.1007/s10533-010-9415-8.
- 552 Einsiedl, F., Hertkorn, N., Wolf, M., Frommberger, M., Schmitt-Kopplin, P., Koch, B.P., 2007. Rapid
553 biotic molecular transformation of fulvic acids in a karst aquifer. *Geochim. Cosmochim. Acta* 71,
554 5474–5482. doi:10.1016/j.gca.2007.09.024.
- 555 Evans, W.C., Fuchs, G., 1988. Anaerobic Degradation of Aromatic Compounds. *Annu. Rev.*
556 *Microbiol.* 42, 289–317. doi:10.1146/annurev.mi.42.100188.001445.
- 557 Ghattas, A.-K., Fischer, F., Wick, A., Ternes, T., 2017. Anaerobic biodegradation of (emerging)
558 organic contaminants in the aquatic environment. *Water Res.* 116, 268–295.
559 doi:10.1016/j.watres.2017.02.001.
- 560 Gonsior, M., Peake, B.M., Cooper, W.T., Podgorski, D., D’Andrilli, J., Cooper, W.J., 2009.
561 Photochemically Induced Changes in Dissolved Organic Matter Identified by Ultrahigh Resolution
562 Fourier Transform Ion Cyclotron Resonance Mass Spectrometry. *Environ. Sci. Technol.* 43, 698–703.
563 doi:10.1021/es8022804.
- 564 Gonsior, M., Schmitt-Kopplin, P., Bastviken, D., 2013. Depth-dependent molecular composition and
565 photo-reactivity of dissolved organic matter in a boreal lake under winter and summer conditions.
566 *Biogeosciences* 10, 6945–6956. doi:10.5194/bg-10-6945-2013.
- 567 Gonsior, M., Hertkorn, N., Conte, M.H., Cooper, W.J., Bastviken, D., Druffel, E., Schmitt-Kopplin,
568 P., 2014. Photochemical production of polyols arising from significant photo-transformation of
569 dissolved organic matter in the oligotrophic surface ocean. *Mar. Chem.* 163, 10–18.
570 doi:10.1016/j.marchem.2014.04.002.

- 571 Gonsior, M., Valle, J., Schmitt-Kopplin, P., Hertkorn, N., Bastviken, D., Luek, J., Harir, M., Bastos,
572 W., Enrich-Prast, A., 2016. Chemodiversity of Dissolved Organic Matter in the Amazon Basin.
573 *Biogeosciences Discuss.* 1–21. doi:10.5194/bg-2016-72.
- 574 Hanson, P.C., Hamilton, D.P., Stanley, E.H., Preston, N., Langman, O.C., Kara, E.L., 2011. Fate of
575 allochthonous dissolved organic carbon in lakes: A quantitative approach. *PLoS One* 6.
576 doi:10.1371/journal.pone.0021884.
- 577 Hedges, J., Oades, J., 1997. Comparative organic geochemistries of soils and marine sediments. *Org.*
578 *Geochem.* 27, 319–361. doi:10.1016/S0146-6380(97)00056-9.
- 579 Hertkorn, N., Claus, H., Schmitt-Kopplin, P., Perdue, E.M., Filip, Z., 2002. Utilization and
580 Transformation of Aquatic Humic Substances by Autochthonous Microorganisms. *Environ. Sci.*
581 *Technol.* 36, 4334–4345. doi:10.1021/es010336o.
- 582 Hertkorn, N., Claus, H., Schmitt-Kopplin, P., Perdue, E.M., Filip, Z., 2002. Utilization and
583 Transformation of Aquatic Humic Substances by Autochthonous Microorganisms. *Environ. Sci.*
584 *Technol.* 36, 4334–4345. doi:10.1021/es010336o.
- 585 Hertkorn, N., Benner, R., Frommberger, M., Schmitt-Kopplin, P., Witt, M., Kaiser, K., Kettrup, A.,
586 Hedges, J., 2006. Characterization of a major refractory component of marine dissolved organic
587 matter. *Geochim. Cosmochim. Acta* 70, 2990–3010. doi:10.1016/j.gca.2006.03.021.
- 588 Hertkorn, N., Ruecker, C., Meringer, M., Gugisch, R., Frommberger, M., Perdue, E.M., Witt, M.,
589 Schmitt-Kopplin, P., 2007. High-precision frequency measurements: indispensable tools at the core of
590 the molecular-level analysis of complex systems. *Anal. Bioanal. Chem.* 389, 1311–1327.
591 doi:10.1007/s00216-007-1577-4.
- 592 Hertkorn, N., Frommberger, M., Witt, M., Koch, B.P., Schmitt-Kopplin, P., Perdue, E.M., 2008.
593 Natural Organic Matter and the Event Horizon of Mass Spectrometry. *Anal. Chem.* 80, 8908–8919.
594 doi:10.1021/ac800464g.

- 595 Hertkorn, N., Harir, M., Koch, B.P., Michalke, B., Schmitt-Kopplin, P., 2013. High-field NMR
596 spectroscopy and FTICR mass spectrometry: powerful discovery tools for the molecular level
597 characterization of marine dissolved organic matter., *Biogeosciences*, 10, 1583-1624. doi:10.5194/bg-
598 10-1583-2013.
- 599 Hertkorn, N., Harir, M., Cawley, K.M., Schmitt-Kopplin, P., Jaffé, R., 2016. Molecular
600 characterization of dissolved organic matter from subtropical wetlands: A comparative study through
601 the analysis of optical properties, NMR and FTICR/MS. *Biogeosciences* 13, 2257–2277.
602 doi:10.5194/bg-13-2257-2016.
- 603 Herzsprung, P., Hertkorn, N., Friese, K., Schmitt-Kopplin, P., 2010. Photochemical degradation of
604 natural organic sulfur compounds (CHOS) from iron-rich mine pit lake pore waters--an initial
605 understanding from evaluation of single-elemental formulae using ultra-high-resolution mass
606 spectrometry. *Rapid Commun. mass Spectrom.* 24, 2909–2924. doi:10.1002/rcm.
- 607 Hodgkins, S.B., Tfaily, M.M., McCalley, C.K., Logan, T. a, Crill, P.M., Saleska, S.R., Rich, V.I.,
608 Chanton, J.P., 2014. Changes in peat chemistry associated with permafrost thaw increase greenhouse
609 gas production. *Proc. Natl. Acad. Sci. U. S. A.* 111, 5819–24. doi:10.1073/pnas.1314641111.
- 610 Hodgkins, S.B., Tfaily, M.M., Podgorski, D.C., McCalley, C.K., Saleska, S.R., Crill, P.M., Rich, V.I.,
611 Chanton, J.P., Cooper, W.T., 2016. Elemental composition and optical properties reveal changes in
612 dissolved organic matter along a permafrost thaw chronosequence in a subarctic peatland. *Geochim.*
613 *Cosmochim. Acta* 187, 123–140. doi:10.1016/j.gca.2016.05.015.
- 614 Ide, J., Ohashi, M., Takahashi, K., Sugiyama, Y., Piirainen, S., Kortelainen, P., Fujitake, N., Yamase,
615 K., Ohte, N., Moritani, M., Hara, M., Finér, L., 2017. Spatial variations in the molecular diversity of
616 dissolved organic matter in water moving through a boreal forest in eastern Finland. *Sci. Rep.* 7,
617 42102. doi:10.1038/srep42102.
- 618 Jaffé, R., Yamashita, Y., Maie, N., Cooper, W.T., Dittmar, T., Dodds, W.K., Jones, J.B., Myoshi, T.,
619 Ortiz-Zayas, J.R., Podgorski, D.C., Watanabe, A., 2012. Dissolved Organic Matter in Headwater

- 620 Streams: Compositional Variability across Climatic Regions of North America. *Geochim.*
621 *Cosmochim. Acta* 94, 95–108. doi:10.1016/j.gca.2012.06.031.
- 622 Jonsson, A., Meili, M., Bergström, A.-K., Jansson, M., 2001. Whole-lake mineralization of
623 allochthonous and autochthonous organic carbon in a large humic lake (Örträsket, N. Sweden).
624 *Limnol. Oceanogr.* 46, 1691–1700. doi:10.4319/lo.2001.46.7.1691.
- 625 Kellerman, A.M., Dittmar, T., Kothawala, D.N., Tranvik, L.J., 2014. Chemodiversity of dissolved
626 organic matter in lakes driven by climate and hydrology. *Nat. Commun.* 5, 1–8.
627 doi:10.1038/ncomms4804.
- 628 Kellerman, A.M., Kothawala, D.N., Dittmar, T., Tranvik, L.J., 2015. Persistence of dissolved organic
629 matter in lakes related to its molecular characteristics. *Nat. Geosci.* 8, 454–457.
630 doi:10.1038/ngeo2440.
- 631 Klüpfel, L., Piepenbrock, A., Kappler, A., Sander, M., 2014. Humic substances as fully regenerable
632 electron acceptors in recurrently anoxic environments. *Nat. Geosci.* 7, 195–200.
633 doi:10.1038/NGEO2084.
- 634 Krause, S., Lüke, C., Frenzel, P., 2009. Spatial heterogeneity of methanotrophs: A geostatistical
635 analysis of pmoA-based T-RFLP patterns in a paddy soil. *Environ. Microbiol. Rep.* 1, 393–397.
636 doi:10.1111/j.1758-2229.2009.00044.x.
- 637 Kroll, J.H., Donahue, N.M., Jimenez, J.L., Kessler, S.H., Canagaratna, M.R., Wilson, K.R., Altieri,
638 K.E., Mazzoleni, L.R., Wozniak, A.S., Bluhm, H., Mysak, E.R., Smith, J.D., Kolb, C.E., Worsnop,
639 D.R., 2011. Carbon oxidation state as a metric for describing the chemistry of atmospheric organic
640 aerosol. *Nat. Chem.* 3, 133–139. doi:10.1038/nchem.948.
- 641 LaRowe, D.E., Van Cappellen, P., 2011. Degradation of natural organic matter: A thermodynamic
642 analysis. *Geochim. Cosmochim. Acta* 75, 2030–2042. doi:10.1016/j.gca.2011.01.020.

- 643 Lofton, D.D., Whalen, S.C., Hershey, A.E., 2015. Vertical sediment distribution of methanogenic
644 pathways in two shallow Arctic Alaskan lakes. *Polar Biol.* 38, 815–827. doi:10.1007/s00300-014-
645 1641-4.
- 646 Mahmoudi, N., Beaupré, S.R., Steen, A.D., Pearson, A., 2017. Sequential bioavailability of
647 sedimentary organic matter to heterotrophic bacteria. *Environmental Microbiology* 19, 2629-2644.
648 doi: 10.1111/1462-2920.13745.
- 649 Medeiros, P.M., Seidel, M., Ward, N.D., Carpenter, E.J., Gomes, H.R., Niggemann, J., Krusche, A.
650 V., Richey, J.E., Yager, P.L., Dittmar, T., 2015. Fate of the Amazon River dissolved organic matter in
651 the tropical Atlantic Ocean. *Global Biogeochem. Cycles* 667–690. doi:10.1002/2015GB005115.
- 652 Roth, V.-N., Dittmar, T., Gaupp, R., Gleixner, G., 2014. Ecosystem-Specific Composition of
653 Dissolved Organic Matter. *Vadose Zone J.* 13, 0. doi:10.2136/vzj2013.09.0162.
- 654 Schmidt, F., Elvert, M., Koch, B.P., Witt, M., Hinrichs, K.-U., 2009. Molecular characterization of
655 dissolved organic matter in pore water of continental shelf sediments. *Geochim. Cosmochim. Acta* 73,
656 3337–3358. doi:10.1016/j.gca.2009.03.008.
- 657 Schmidt, F., Koch, B.P., Elvert, M., Schmidt, G., Witt, M., Hinrichs, K.-U., 2011. Diagenetic
658 transformation of dissolved organic nitrogen compounds under contrasting sedimentary redox
659 conditions in the Black Sea. *Environ. Sci. Technol.* 45, 5223–9. doi:10.1021/es2003414.
- 660 Schmidt, F., Koch, B.P., Goldhammer, T., Elvert, M., Witt, M., Lin, Y.S., Wendt, J., Zabel, M.,
661 Heuer, V.B., Hinrichs, K.-U., 2017. Unraveling signatures of biogeochemical processes and the
662 depositional setting in the molecular composition of pore water DOM across different marine
663 environments. *Geochim. Cosmochim. Acta* 207, 57–80. doi:10.1016/j.gca.2017.03.005.
- 664 Schulz, S., Conrad, R., 1995. Effect of algal deposition on acetate and methane concentrations in the
665 profundal sediment of a deep lake (Lake Constance). *FEMS Microbiol. Ecol.* 16, 251–260.
666 doi:10.1111/j.1574-6941.1995.tb00289.x.

- 667 Seidel, M., Beck, M., Riedel, T., Waska, H., Suryaputra, I.G.N.A., Schnetger, B., Niggemann, J.,
668 Simon, M., Dittmar, T., 2014. Biogeochemistry of dissolved organic matter in an anoxic intertidal
669 creek bank. *Geochim. Cosmochim. Acta* 140, 418–434. doi:10.1016/j.gca.2014.05.038.
- 670 Seidel, M., Dittmar, T., Ward, N.D., Krusche, A.V., Richey, J.E., Yager, P.L., Medeiros, P.M., 2016.
671 Seasonal and spatial variability of dissolved organic matter composition in the lower Amazon River.
672 *Biogeochemistry* 131, 281–302. doi:10.1007/s10533-016-0279-4.
- 673 Shakeri Yekta, S., Gonsior, M., Schmitt-Kopplin, P., Svensson, B.H., 2012. Characterization of
674 Dissolved Organic Matter in Full Scale Continuous Stirred Tank Biogas Reactors Using Ultrahigh
675 Resolution Mass Spectrometry: A Qualitative Overview. *Environ. Sci. Technol.* 46, 12711–12719.
676 doi:10.1021/es3024447.
- 677 Siljanen, H.M.P., Saari, A., Krause, S., Lensu, A., Abell, G.C.J., Bodrossy, L., Bodelier, P.L.E.,
678 Martikainen, P.J., 2011. Hydrology is reflected in the functioning and community composition of
679 methanotrophs in the littoral wetland of a boreal lake. *FEMS Microbiol. Ecol.* 75, 430–445.
680 doi:10.1111/j.1574-6941.2010.01015.x.
- 681 Thullner, M., Regnier, P., van Cappellen, P., 2007. Modeling Microbially Induced Carbon
682 Degradation in Redox-Stratified Subsurface Environments: Concepts and Open Questions.
683 *Geomicrobiol. J.* 24, 139–155. doi:10.1080/01490450701459275.
- 684 Tranvik, L.J., Downing, J.A., Cotner, J.B., Loiselle, S.A., Striegl, R.G., Ballatore, T.J., Dillon, P.,
685 Finlay, K., Fortino, K., Knoll, L.B., Kortelainen, P.L., Kutser, T., Larsen, S., Laurion, I., Leech, D.M.,
686 McCallister, S.L., McKnight, D.M., Melack, J.M., Overholt, E., Porter, J.A., Prairie, Y., Renwick,
687 W.H., Roland, F., Sherman, B.S., Schindler, D.W., Sobek, S., Tremblay, A., Vanni, M.J., Verschoor,
688 A.M., von Wachenfeldt, E., Weyhenmeyer, G.A., 2009. Lakes and reservoirs as regulators of carbon
689 cycling and climate. *Limnol. Oceanogr.* 54, 2298–2314. doi:10.4319/lo.2009.54.6_part_2.2298.
- 690 Tremblay, L.B., Dittmar, T., Marshall, A.G., Cooper, W.J., Cooper, W.T., 2007. Molecular
691 characterization of dissolved organic matter in a North Brazilian mangrove porewater and mangrove-
692 fringed estuaries by ultrahigh resolution Fourier Transform-Ion Cyclotron Resonance mass

- 693 spectrometry and excitation/emission spectroscopy. *Mar. Chem.* 105, 15–29.
694 doi:10.1016/j.marchem.2006.12.015.
- 695 von Wachenfeldt, E., Sobek, S., Bastviken, D., Tranvik, L.J., 2008. Linking allochthonous dissolved
696 organic matter and boreal lake sediment carbon sequestration: The role of light-mediated flocculation.
697 *Limnol. Oceanogr.* 53, 2416–2426. doi:10.4319/lo.2008.53.6.2416.
- 698 von Wachenfeldt, E., Tranvik, L.J., 2008. Sedimentation in Boreal Lakes—The Role of Flocculation
699 of Allochthonous Dissolved Organic Matter in the Water Column. *Ecosystems* 11, 803–814.
700 doi:10.1007/s10021-008-9162-z.
- 701 Walker, L.P., Wilson, D.B., 1991. Enzymatic hydrolysis of cellulose: An overview. *Bioresour.*
702 *Technol.* 36, 3–14. doi:10.1016/0960-8524(91)90095-2.
- 703 West, W.E., Coloso, J.J., Jones, S.E., 2012. Effects of algal and terrestrial carbon on methane
704 production rates and methanogen community structure in a temperate lake sediment. *Freshw. Biol.* 57,
705 949–955. doi:10.1111/j.1365-2427.2012.02755.x.
- 706 Yavitt, J.B., Seidmann-Zager, M., 2006. Methanogenic conditions in northern peat soils.
707 *Geomicrobiol. J.* 23, 119–127. doi:10.1080/01490450500533957.

**Extensive processing of sediment pore water
dissolved organic matter during anaerobic incubation as observed
by high-field mass spectrometry (FT-ICR-MS)**

Juliana Valle^a, Michael Gonsior^b, Mourad Harir^{a,c}, Alex Enrich-Prast^{d,e}, Philippe Schmitt-Kopplin^{a,c}, David Bastviken^d, Ralf Conrad^f and Norbert Hertkorn^{a*}*

^a Helmholtz Zentrum Munich, German Research Center for Environmental Health, Neuherberg, Germany

^b University of Maryland Center for Environmental Science, Chesapeake Biological Laboratory, Solomons, USA

^c Department for Chemical-Technical Analysis, Research Center Weihenstephan for Brewing and Food Quality, Technische Universität München, Freising-Weihenstephan, Germany

^d Linköping University, Department of Thematic Studies – Environmental Change, Linköping, Sweden

^e Federal University of Rio de Janeiro, Department of Botany, Rio de Janeiro, Brazil

^f Max Planck Institute for Terrestrial Microbiology, Marburg, Germany

*corresponding authors: e-mail: hertkorn@helmholtz-muenchen.de, alex.enrich.prast@liu.se

Tables and Captions

Table 1. GPS coordinates, physico-chemical parameters from water column and sediments, and greenhouse gases production rates (\pm SE) from three Swedish lakes studied.

item	Svartjärn	Grästjärn	Bisen
position	59°53'26.50" N 15°15'27.00" E	59°53'23.30" N 15°21'17.50" E	59°57'20.60" N 15°21'38.00" E
area [km ²]	0.007	0.09	0.43
maximum depth [m]	7	8	17
pH	4.8	4.9	6.1
DOC lake water [mg l ⁻¹]	25.7	17.4	7.3
lake water total phosphorus TP [μ g l ⁻¹]	15.1	8.7	6.5
lake water total nitrogen TN [μ g l ⁻¹]	502	335	281
sedimentary organic carbon [%]	8.5	17.4	13.0
sedimentary organic nitrogen [%]	0.4	1.1	0.9
CH ₄ evolution [nmol g dw ⁻¹ d ⁻¹]	11.8 \pm 2.6	8.9 \pm 1.5	5.9 \pm 1.4
CO ₂ evolution [nmol g dw ⁻¹ d ⁻¹]	77.6 \pm 75.9	113.5 \pm 11.6	107.2 \pm 19.5
depth / temperature	6.5 m / 5.3°C	4.5 m / 10.9°C	13 m / 6.8°C
O ₂ saturation %	1.2	7.6	3.6
mg/L oxygen	0.15	0.82	0.42

Table 2. Counts of common mass peaks between five replicates of sediment pore water SPE-DOM in the three studied lakes (Svartjärn, Grästjärn, and Bisen) before and after incubation as computed from negative electrospray (ESI) 12 T FTICR mass spectra for singly charged ions.

members of molecular series	Svartjärn original	Svartjärn incubated	Grästjärn original	Grästjärn incubated	Bisen original	Bisen incubated
consolidated count of assigned mass peaks detected in any of the five replicates	6555	6695	4821	4898	6401	7090
total count of assigned mass peaks common to all five replicates	4571	4203	3316	3018	4473	4592
CHO compounds	3100 (68%)	2693 (64%)	2299 (69%)	2083 (69%)	2919 (65%)	2900 (63%)
CHOS compounds	686 (15%)	614 (15%)	352 (11%)	240 (8%)	420 (9%)	418 (9%)
CHNO compounds	784 (17%)	895 (21%)	664 (20%)	695 (23%)	1132 (25%)	1263 (28%)
CHNOS compounds	1 (0.02%)	1 (0.02%)	1 (0.03%)	0 (0%)	2 (0.04%)	11 (0.24%)
proportion of formulas shared in all five replicates	70%	63%	69%	62%	70%	65%
average H [%]	43.7	43.9	45.1	47.6	40.2	42.2
average C [%]	37.7	37.8	37.3	37.1	37.9	37.7
average O [%]	18.3	17.9	17.2	14.6	21.5	19.5
average N [%]	0.1	0.2	0.2	0.2	0.2	0.2
average S [%]	0.2	0.2	0.2	0.5	0.1	0.3
computed average H/C ratio	1.16	1.16	1.21	1.28	1.06	1.12
computed average O/C ratio	0.49	0.47	0.46	0.39	0.57	0.52
computed average C/N ratio	292	188	187	182	174	169
computed average C/S ratio	165	166	169	68	448	113
average carbon oxidation state of CHO compounds	-0.183	-0.214	-0.215	-0.219	0.059	-0.056
average DBE	9.0	8.8	8.1	7.4	9.8	9.2
average DBE/C	0.5	0.5	0.5	0.4	0.5	0.5
mass weighted average [Da]	400.3	388.6	369.7	358.3	412.9	399.7

**Extensive processing of sediment pore water
dissolved organic matter during anaerobic incubation as observed
by high-field mass spectrometry (FT-ICR-MS)**

Juliana Valle^a, Michael Gonsior^b, Mourad Harir^{a,c}, Alex Enrich-Prast^{d,e},*

Philippe Schmitt-Kopplin^{a,c}, David Bastviken^d, Ralf Conrad^f and Norbert Hertkorn^{a}*

^a Helmholtz Zentrum Munich, German Research Center for Environmental Health, Neuherberg, Germany

^b University of Maryland Center for Environmental Science, Chesapeake Biological Laboratory, Solomons, USA

^c Department for Chemical-Technical Analysis, Research Center Weihenstephan for Brewing and Food Quality, Technische Universität München, Freising-Weihenstephan, Germany

^d Linköping University, Department of Thematic Studies – Environmental Change, Linköping, Sweden

^e Federal University of Rio de Janeiro, Department of Botany, Rio de Janeiro, Brazil

^f Max Planck Institute for Terrestrial Microbiology, Marburg, Germany

*corresponding authors: e-mail: hertkorn@helmholtz-muenchen.de, alex.enrich.prast@liu.se

Figures and Captions

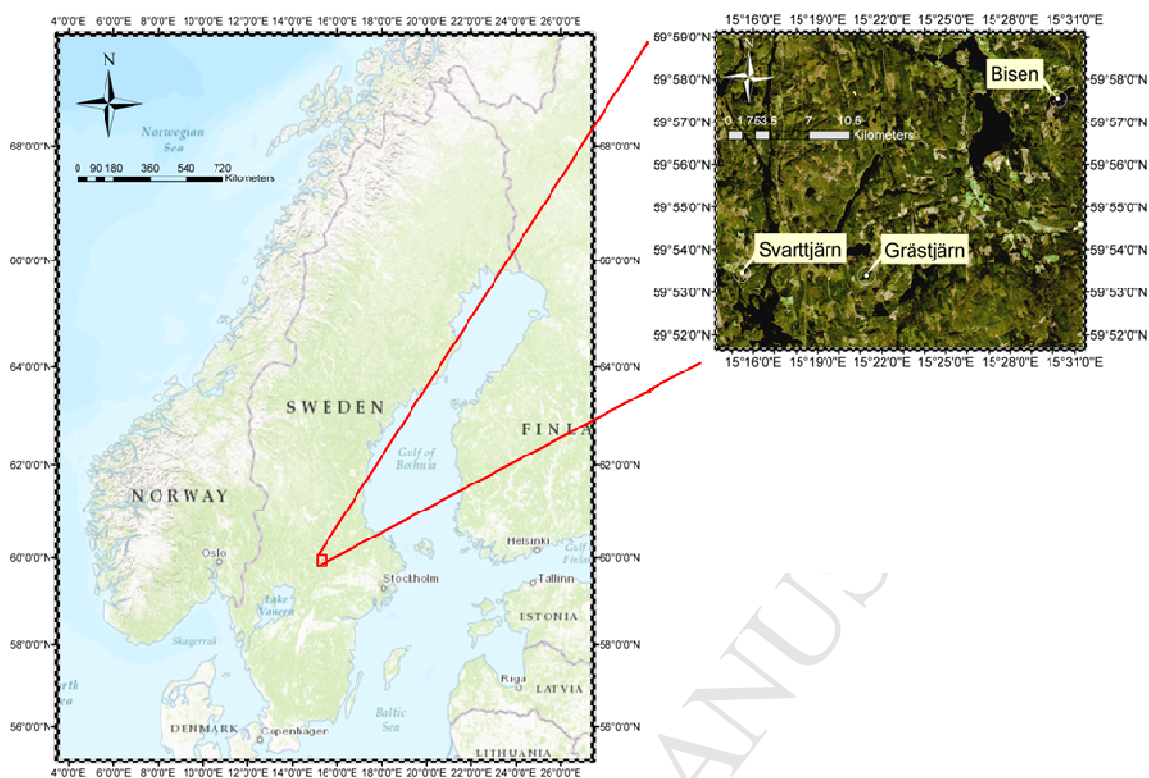


Figure 1. Maps of sampling areas.

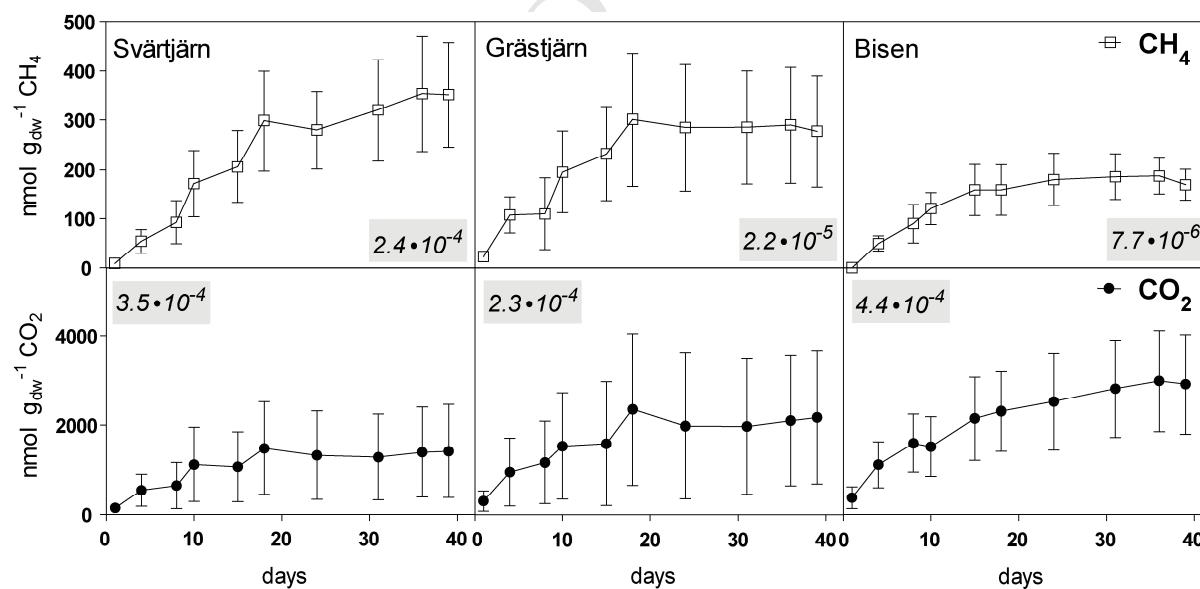


Figure 2. CH₄ and CO₂ concentrations during the anaerobic incubation experiment. Values shown are average from five replicates and error bars represent standard error. Gray shaded values denote fraction of carbon evolved after 40 days of incubation in the form of CO₂ and CH₄ with respect to organic carbon present in dry sediment after incubation.

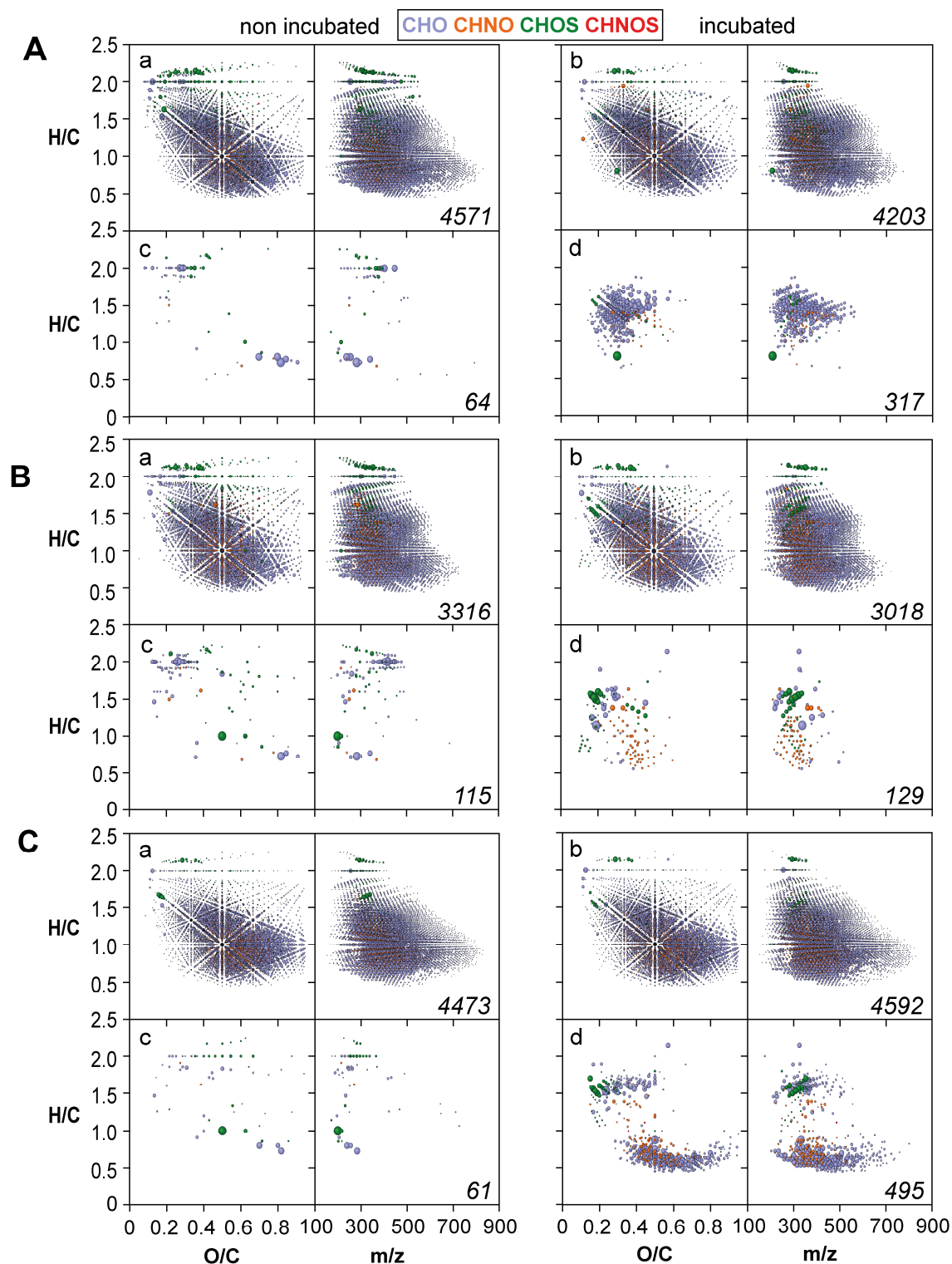


Figure 3. FTMS-derived van Krevelen diagrams of common molecular compositions observed in all five replicates of original and incubated sediment pore water SPE-DOM from boreal lakes (A) Svattjärn, (B) Grästjärn and (C) Bisen, provided with total counts of molecular compositions (overlay) and color-coded according to CHO (blue), CHOS (green), CHNO (orange), and CHNOS (red) molecular series. Panel a: all compounds observed in the original sample; panel b: all compounds observed in the incubated sample; panel c: compounds declining in abundance during the incubation (presumably consumed compounds); panel d: compounds increasing in abundance during the incubation (presumably produced compounds). Van Krevelen diagrams derived from individual SPE-DOM samples (Figures S5 and S6) show individual characteristics and variance, resulting in larger counts and nominal chemical diversity of observed molecular compositions (Table 2), thereby obscuring common trends of SPE-DOM evolution associated with incubation.

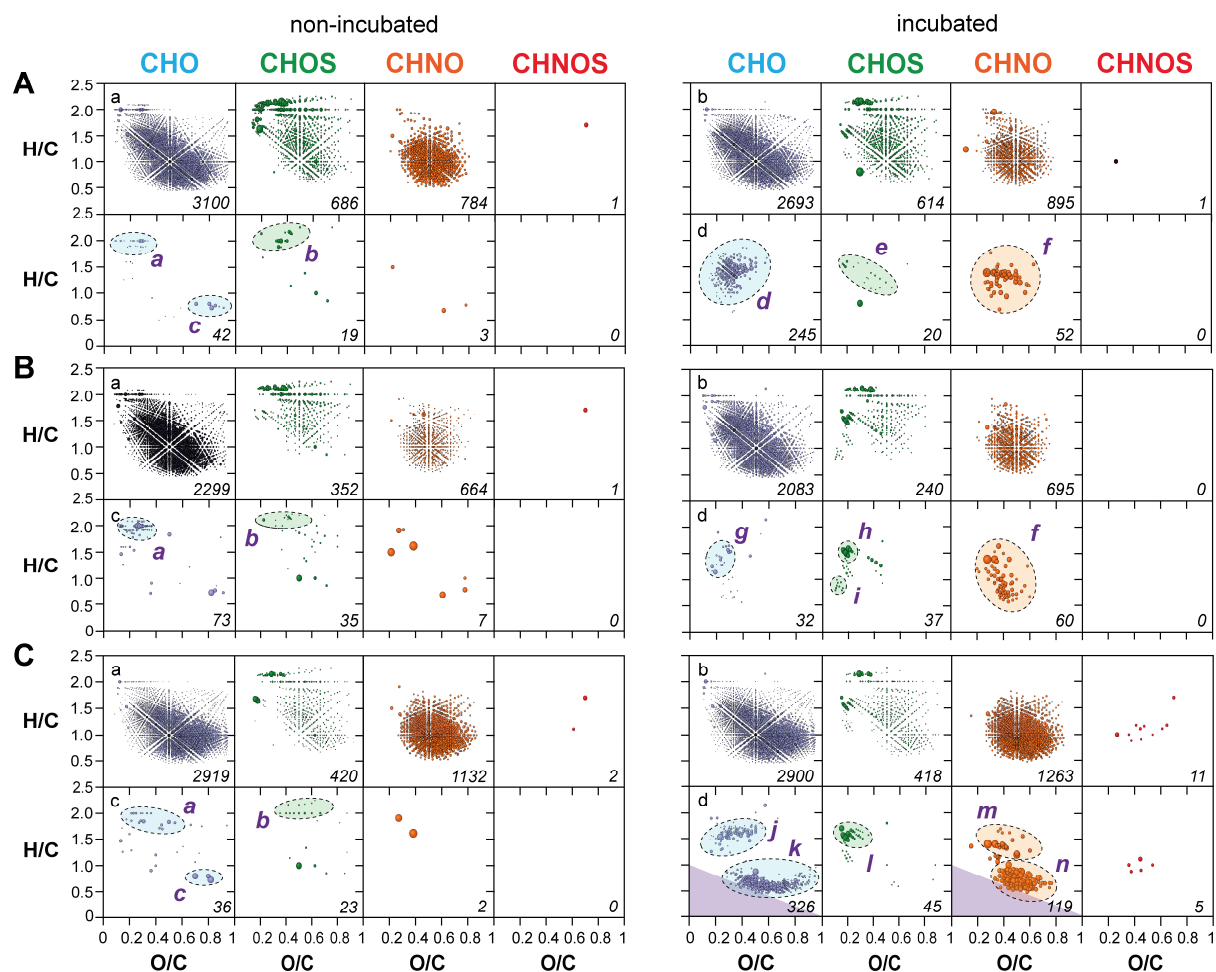


Figure 4. FTMS derived van Krevelen diagrams of common molecular compositions observed in all five replicates of original and incubated sediment pore water SPE-DOM from boreal lakes (A) Svattjärn, (B) Grästjärn and (3) Bisen, provided with counts of molecular compositions and color-coded according to CHO (blue), CHOS (green), CHNO (orange), and CHNOS (red) individual molecular series. Panel a: all compounds observed in the original sample; panel b: all compounds observed in the incubated sample; panel c: compounds declining in abundance during the incubation (presumably consumed compounds); panel d: compounds increasing in abundance during the incubation (presumably produced compounds). Van Krevelen diagrams derived from individual SPE-DOM samples (Figure S6) show individual characteristics and variance (Tables S1 and S2), resulting in larger counts and nominal chemical diversity of observed molecular compositions (Table 2), thereby obscuring common trends of SPE-DOM evolution associated with incubation. Attribution of key structures (purple, italics): *a*: aliphatic CHO-lipid like compounds; *b*: aliphatic CHOS lipid-like compounds; *c*: highly oxygenated, tannin-like compounds; *d*: carboxylic-rich alicyclic molecules (CRAM; Hertkorn et al., 2006); *e*: common CHOS organic matter; *f*: rather unsaturated and oxygenated CHNO compounds of yet unclear provenance; *g*: unsaturated CHO-lipids; *h*: unsaturated CHOS lipids; *i*: aromatic CHOS compounds; *j*: carboxylic-rich alicyclic molecules (CRAM; Hertkorn et al., 2006); *k*: lignin- and tannin-like compounds; *l*: unsaturated CHOS lipid-like compounds; *m*: common CHNO organic matter; *n*: nitrogen-containing lignin- and tannin-like compounds. The aromaticity index AI (Koch and Dittmar, 2006) provided (purple triangles) denotes presence of aromatic compounds.

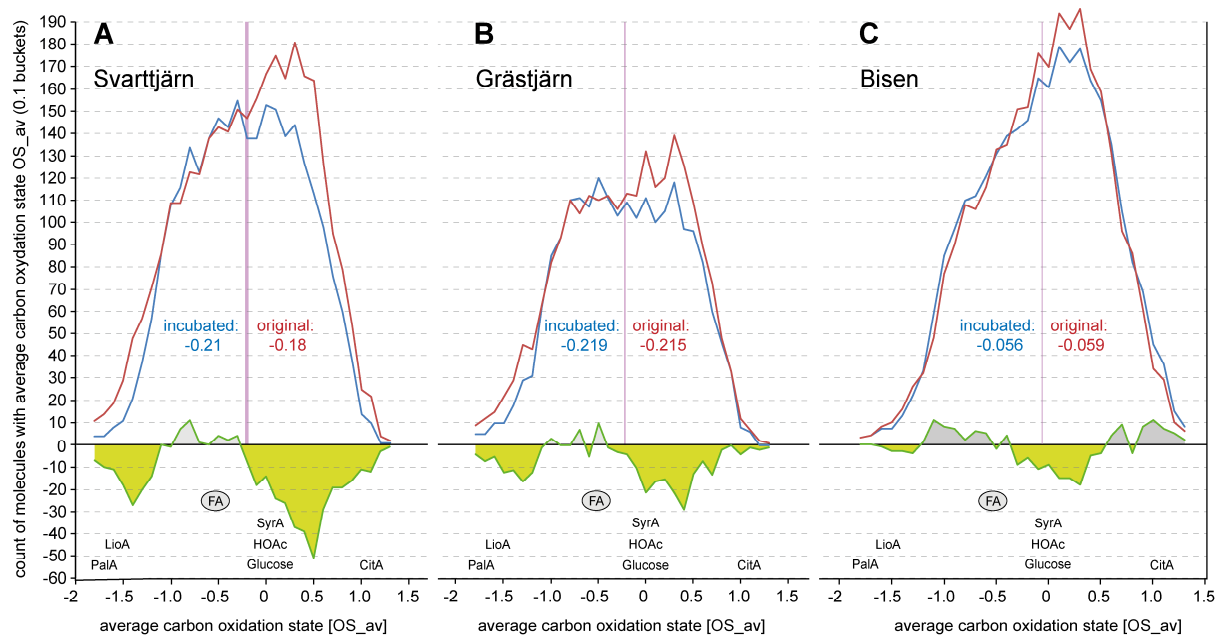


Figure 5. Distribution of average carbon oxidation state OS_{av} in CHO compounds only, sorted in increments of ± 0.1 of nominal carbon oxidation state (Kroll et al., 2011) in three lake sediments before and after incubation; color code: red, original SPE-DOM; blue: SPE-DOM after incubation; green: difference in counts: original minus incubated; yellow-green: compounds decreased during incubation; gray: compounds increased after incubation. It appears that the extent of relative changes progressively decreases with ever increasing lake size and water residence time. At the bottom, several reference compounds (common metabolites) are indicated: PalA, palmitic acid $C_{16}H_{32}O_2$, $OS_{av} = -1.75$ (lipid); linoleic acid, $C_{18}H_{32}O_2$, $OS_{av} = -1.55$ (lipid); FA, fulvic acid, $OS_{av} = -0.52$ (Le Rowe et al., 2011); SyrA, syringic acid, $C_9H_{10}O_2$, $OS_{av} = 0$ (aromatic acid); HOAc, acetic acid, $C_2H_4O_2$, $OS_{av} = 0$ (aliphatic acid); glucose, $C_6H_{12}O_6$, $OS_{av} = 0$ (carbohydrate); CitA, citric acid, $C_6H_8O_7$, $OS_{av} = 1.167$ (oxygenated aliphatic acid).

**Extensive processing of sediment pore water
dissolved organic matter during anaerobic incubation as observed
by high-field mass spectrometry (FT-ICR-MS)**

Juliana Valle^a, *Michael Gonsior*^b, *Mourad Harir*^{a,c}, *Alex Enrich-Prast*^{d,e*}, *Philippe Schmitt-Kopplin*^{a,c}, *David Bastviken*^d, *Ralf Conrad*^f and *Norbert Hertkorn*^{a*}

^a Helmholtz Zentrum Munich, German Research Center for Environmental Health, Neuherberg, Germany

^b University of Maryland Center for Environmental Science, Chesapeake Biological Laboratory, Solomons, USA

^c Department for Chemical-Technical Analysis, Research Center Weihenstephan for Brewing and Food Quality, Technische Universität München, Freising-Weihenstephan, Germany

^d Linköping University, Department of Thematic Studies – Environmental Change, Linköping, Sweden

^e Federal University of Rio de Janeiro, Department of Botany, Rio de Janeiro, Brazil

^f Max Planck Institute for Terrestrial Microbiology, Marburg, Germany

*corresponding authors: e-mail: hertkorn@helmholtz-muenchen.de, alex.enrich.prast@liu.se

Highlights

DOM processing by anaerobic incubation greatly exceeds production of greenhouse gases CO₂ and CH₄

DOM after incubation may represent microbial metabolites and recalcitrant original DOM

Lake-specific DOM processing was observed