

1 Using gene expression to annotate cardiovascular GWAS loci

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11 Abstract

12 Genetic variants at hundreds of loci associated with cardiovascular phenotypes have been identified
13 by genome wide association studies. Most of these variants are located in intronic or intergenic
14 regions rendering the functional and mechanistic follow up difficult. These non-protein-coding
15 regions harbor regulatory sequences. Thus the study of genetic variants associated with transcription
16 – so called expression quantitative trait loci – has emerged as a promising approach to identify
17 regulatory sequence variants. The genes and pathways they control constitute candidate causal
18 drivers at cardiovascular risk loci. This review provides an overview of the expression quantitative
19 trait loci resources available for cardiovascular genetics research and the most commonly used
20 approaches for candidate gene identification.

21 Background

22 The ultimate goal of any genetic association analysis is to identify genetic variation linked to
23 variation of a phenotype and to elucidate the molecular mechanisms, which are altered by the
24 sequence variation. Genome wide association studies have been tremendously successful in
25 identifying thousands of disease-associated loci as documented by the steady growth of the
26 continuously updated GWAS catalog (MacArthur et al. 2017). This progress has also highlighted
27 hundreds of loci associated with cardiovascular phenotypes: the current GWAS catalog (Burdett et al.
28 2018) lists 249 distinct chromosomal regions associated with coronary artery disease with candidate
29 genes and pathways at many loci summarized in (Klarin et al. 2017), 138 / 115 with diastolic /
30 systolic blood pressure, 109 with QT interval, to name just the top three cardiovascular phenotypes.
31 Follow up analysis of these loci aim to establish the causal mechanisms underlying the statistical
32 associations. In classical family based linkage studies typically identifying rare variants with very
33 large effect sizes, the causal variants are typically located in the protein sequence and have a strong
34 impact on protein function (Timpson et al. 2018), for instance truncating mutations in the sarcomeric
35 protein TTN cause dilated cardiomyopathy (Siu et al. 1999; Gerull et al. 2002; Herman et al. 2012;
36 Roberts et al. 2015). In GWAS however, the identification of causal variants proved to be very
37 challenging, since the vast majority of these disease-associated variants is located either in introns of
38 genes or in intergenic regions (Burdett et al. 2018). Therefore the classical approach of identifying
39 the variant with strongest impact on protein function, such as gained stop codons is not sufficient.

40 Recent large-scale efforts have annotated a plethora of functional regulatory elements such as
 41 enhancers residing in the non-protein-coding part of the genome (ENCODE Project Consortium
 42 2012; Roadmap Epigenomics Consortium et al. 2015). Therefore an alternative mechanism might be
 43 that disease-associated regulatory variants alter the sequence and function of such regulatory
 44 elements. Indeed a systematic analysis of the location of disease-associated variants showed that they
 45 preferentially reside in regulatory elements (Maurano et al. 2012; Farh et al. 2015). Since regulatory
 46 elements are highly tissue specific, this information can even be used to identify the disease-relevant
 47 tissues (Maurano et al. 2012; Farh et al. 2015). These results from localization analysis are highly
 48 suggestive that disease-associated variants alter regulatory elements. It now remains to be shown that
 49 they indeed are altered and to identify the respective target gene whose transcription is controlled by
 50 the regulatory element.

51 Integrated analysis of the genetics of gene expression provides an elegant way of directly assessing
 52 the consequences of putative regulatory sequence variants on transcription. In this study design
 53 (Jansen and Nap 2001), a population cohort is characterized for their genome wide patterns of genetic
 54 variation and also for genome wide gene expression. Gene expression levels are treated as
 55 quantitative traits and systematically tested for associations between sequence variants and gene
 56 expression. Significant associations are called expression quantitative trait loci (eQTL). These eQTL
 57 not only identify putative regulatory variants, but also their target genes as the gene whose expression
 58 is associated with the variant (Civelek and Lusis 2014; Albert and Kruglyak 2015). Biological
 59 information processing and regulation is not limited to transcription, so this approach has also been
 60 generalized towards other intermediate molecular traits such as DNA methylation (Banovich et al.
 61 2014; Lemire et al. 2015), open chromatin (Degner et al. 2012), histone modifications (Waszak et
 62 al. 2015; Grubert et al. 2015) (Del Rosario et al. 2015), gene, exon and transcript expression levels
 63 (Montgomery et al. 2010; Pickrell et al. 2010; Lappalainen et al. 2013; GTEx Consortium et al. 2015;
 64 Battle et al. 2017) translation and protein levels (Li et al. 2016) as well as metabolites (Suhre et al.
 65 2011; Shin et al. 2014). In particular the information from the epigenome can be used to identify
 66 regulatory variants, and to characterize their role in disease (Maurano et al. 2012; Del Rosario et al.
 67 2015; Degner et al. 2012; Li et al. 2016).

68 **eQTL resources for cardiovascular genetics**

69 Regulatory elements and also the effects of variants on those elements can be highly tissue specific,
 70 therefore it is key to investigate the tissue relevant for the disease (Maurano et al. 2012; Grundberg et
 71 al. 2012; Farh et al. 2015; GTEx Consortium et al. 2015). Because biopsies of tissues relevant for
 72 cardiovascular diseases, in particular of the heart are very difficult to obtain from humans, it is not
 73 surprising, that early applications of eQTL analysis to identify candidate genes for cardiovascular
 74 phenotypes were reported in animal models (Monti et al. 2008). To understand the regulatory impact
 75 of sequence variants in humans, samples of disease relevant tissues are often obtained during surgery,
 76 from organ donors or from post-mortem sections. As a consequence of these practical considerations,
 77 the transcriptome data might be confounded by differences in tissue composition (Heinig et al. 2017)
 78 or ischemic time of post-mortem samples (GTEx Consortium et al. 2015). Therefore additional care
 79 has to be taken in data analysis accounting for observed and hidden confounders (Stegle et al. 2010).
 80 Current reviews provide an overview of recent human eQTL studies (Albert and Kruglyak 2015;
 81 Vandiedonck 2018). The most comprehensive study to date is the Genotype tissue expression (GTEx)
 82 project, which aims to characterize regulatory sequence variants across 44 distinct tissues from post-
 83 mortem sections (Battle et al. 2017). This includes cardiac tissues: left ventricle, atrial appendage;
 84 vascular tissues: aorta, tibial artery, coronary artery; as well as metabolic tissues: liver, subcutaneous
 85 and viscelar adipose tissue (Table 1). In terms of sample size and coverage of tissues of interest, the

86 eQTL data generated in the STARNET consortium is currently the most comprehensive resource
 87 (Franzén et al. 2016). It focuses on vascular and metabolic tissues in patients with coronary artery
 88 disease. It has been shown that eQTL are sometimes dependent on the disease context (Heinig et al.
 89 2017). This observation is also supported by the finding that more eQTLs associated with disease
 90 SNP can be found in diseased populations (Franzén et al. 2016). Formation of atherosclerotic plaques
 91 is an inflammatory process, therefore also immune cells such as monocytes or macrophages are
 92 considered disease relevant tissues and have been extensively profiled (Zeller et al. 2010). Since the
 93 disease relevant tissues are not always known a priori efforts are currently underway to establish
 94 cohorts of induced pluripotent stem cell that can potentially be differentiated into any cell type for
 95 genetic mapping (Kilpinen et al. 2017). These eQTL projects are complemented by large scale
 96 projects aimed at creating a reference map of regulatory elements across an exhaustive set of 111
 97 human cell types and tissues (Roadmap Epigenomics Consortium et al. 2015) by annotation with
 98 epigenetic markers of regulatory elements and recent developments of sequencing based methods
 99 (e.g. Hi-C) to study chromosomal architecture (Davies et al. 2017) in a wide variety of human tissues
 100 (Schmitt et al. 2016) including heart, liver and aorta. These techniques can identify promoter –
 101 enhancer interactions and have already been used successfully to identify IRX3 as the causal gene
 102 underlying an obesity GWAS hit located in the intron of the FTO gene (Smemo et al. 2016).

103 **Candidate identification strategies**

104 *cis* eQTL candidate genes

105 Overlapping eQTL and GWAS SNPs is the most straightforward approach to identify candidate
 106 genes for GWAS hits. If a GWAS SNP is also an eQTL for a close by gene or in tight LD with an
 107 eQTL, it is conceivable that the SNP indeed affects a regulatory element controlling the expression
 108 the respective gene. These genes are typically called *cis*-eQTL when the distance between gene and
 109 variant is not further than 500kb – 1Mb, as opposed to *trans*-eQTL, where the distances are greater or
 110 the variant and gene are located on different chromosomes. Cardiovascular candidate genes such as
 111 SORT1 (Musunuru et al. 2010) and LIPA (Wild et al. 2011) have been identified as *cis*-eQTL. It has
 112 been demonstrated that these candidate genes frequently are not the genes located closest to the
 113 GWAS SNP for heart related traits (Heinig et al. 2017) and also more generally for any GWAS trait
 114 (GTEx Consortium et al. 2015; Battle et al. 2017). Nowadays, this candidate annotation approach is
 115 becoming a standard analysis included in many GWAS papers and can be performed conveniently
 116 using the online software FUMA (Watanabe et al. 2017). For instance a recent GWAS on CAD (van
 117 der Harst and Verweij 2018) identified eQTL for 196 genes at 97 of the 161 CAD loci found in the
 118 analysis from GTEx and other eQTL data bases. This result already demonstrates one caveat of the
 119 approach: several candidate genes might emerge for a locus and might be inconsistent between
 120 tissues or GWAS variants might also associate with eQTL by chance (Battle et al. 2017). In this
 121 particular example 36 loci have unique candidate genes and additional 24 loci have candidate genes
 122 detected consistently across tissues, so 60 loci can be annotated confidently. Overall a highly
 123 significant enrichment of trait associated SNPs can be observed among eQTLs as demonstrated for
 124 heart related traits (Heinig et al. 2017). Less frequently also *trans*-eQTL are considered for the
 125 annotation of GWAS SNPs, as they do not readily provide a clear mechanistic explanation.
 126 Nevertheless, it has been shown in a systematic analysis of GWAS variants, that they frequently also
 127 associate with expression levels of genes distant to the GWAS locus (Westra et al. 2013).

128 An important limitation of the overlap-based strategy is that it cannot be used to establish causality.
 129 Strictly speaking the experimental design does only allow inferring causality in a statistical sense. In
 130 genetic associations the direction of causality is always fixed (Figure 1 (A)). To establish a causal

131 chain between genetic variation, gene expression and the disease phenotype in the strict sense, an
 132 interventional experiment would be required, where all other confounding factors that could
 133 determine the phenotype are fixed and only the gene expression level would be manipulated to test
 134 an effect on the phenotype. If gene expression is indeed causal for the phenotype, any change of the
 135 gene expression necessarily would cause a change in the phenotype. In the concept of Mendelian
 136 randomization (MR) one is considering a genetic variant as instrumental variable controlling the
 137 levels of gene expression and observes its effect on the phenotypic outcome (Davey Smith and
 138 Hemani 2014). In analogy to randomized control trials, individuals get assigned to a group based on
 139 their genotype. Because the direction of causality between genetic variant and gene expression is
 140 fixed and the genetic variant is robustly associated with expression levels, one group will receive a
 141 higher dose of gene expression. Assuming that the genotype is independent of confounding factors
 142 (Figure 1 (A)) changes in phenotypic outcome can be attributed to the changes in gene expression.

143 Classically, MR and similar approaches to statistically establish causality (Schadt et al. 2005)
 144 (Millstein et al. 2009) require to measure all variables in the same population Figure 1 (B). This is
 145 often not feasible, as gene expression profiling in each and every disease cohort is prohibitively
 146 expensive. In practice GWAS SNPs and eQTLs are identified in separate populations. Because of
 147 data privacy regulations, often a researcher only has access to the full individual level data of one
 148 population and the summary statistics of the other population. Depending on which full data set is
 149 available there exist several methods allowing to directly integrate the measured data with summary
 150 statistics (Pickrell et al. 2016; Hormozdiari et al. 2016; Gusev et al. 2016; Zhu et al. 2016). A
 151 Bayesian co-localization approach based on summary statistics (Giambartolomei et al. 2014) is
 152 testing whether the co-localization of two association signals is compatible with a common
 153 underlying causal variant and has been successfully applied to blood lipid traits and liver eQTL. An
 154 alternative approach is to impute gene expression levels (Manor and Segal 2013) into a GWAS
 155 population (Gamazon et al. 2015; Gusev et al. 2016) using eQTL summary statistics from an eQTL
 156 reference population. Subsequently the imputed gene expression can be correlated to the disease
 157 phenotype to identify candidate genes (Gamazon et al. 2015; Gusev et al. 2016). Alternatively the
 158 transcriptome wide association study (TWAS) method (Gusev et al. 2016) and other methods
 159 (Barbeira et al. 2017) can also work completely without individual level data by indirectly
 160 associating expression and phenotype using eQTL and GWAS summary statistics and the LD
 161 structure between SNPs. The TWAS approach showed superior power compared to colocalization
 162 analysis and simple overlap based analysis in cases where the causal variants are not directly
 163 observed, or when multiple causal variants affecting expression and phenotype exist. Consistent with
 164 other candidate identification strategies, analysis of obesity related traits with TWAS showed that
 165 66% of identified trait associated genes were not the closest gene (Gusev et al. 2016). Summary data-
 166 based Mendelian Randomization (SMR) is a method that can be used if only summary statistics are
 167 available from both eQTL and GWAS results. The method makes use of standard two-sample MR
 168 (Pierce and Burgess 2013) to identify causal or pleiotropic effects of sequence variants on gene
 169 expression and phenotypes and distinguishes this situation from overlapping independent causal
 170 variants in LD using a test on multiple SNPs (Zhu et al. 2016). Similar to results from TWAS
 171 analyses, the application of this method to five common diseases showed that only 60% of the
 172 identified candidate genes are the closest gene to the GWAS SNP.

173 *Network based analysis*

174 Genes are not acting in isolation, but rather form functionally related pathways and networks.
 175 Pathways are usually defined based on curated prior knowledge about well-studied processes such as
 176 biochemical reactions and signaling pathways (KEGG, Reactome, GO). Pathways can be represented

177 as sets of genes of the same process or as networks preserving the topological information which
178 genes are connected to one another, for instance by catalyzing adjacent steps in a metabolic pathway.
179 Alternatively, networks can be derived from high-throughput experiments such as transcriptome
180 profiling (co-expression network) or protein-protein interaction (PPI) screening (PPI network).
181 Pathways and networks defined either from prior knowledge or from data can subsequently be used
182 for the interpretation of disease associations derived from GWAS. Representing pathways as sets of
183 genes, one can ask, whether a set of genes shows higher evidence of association to disease than
184 random gene sets of the same size. Because GWAS test individual SNPs and not genes, a mapping
185 between SNPs and genes is required, for instance based on genomic positions. Methods such as SNP
186 set enrichment analysis (Zhong, Beaulaurier, et al. 2010; Zhong, Yang, et al. 2010) can then be used
187 to test the statistical significance of the association between gene sets and the GWAS results by
188 comparing the distribution of GWAS P-values of SNPs within the pathway to a background
189 distribution. These methods have been applied to show the association between CAD and pathways
190 for lipid metabolism, coagulation, immunity (Mäkinen et al. 2014).

191 Since eQTL experiments require transcriptome profiling in large cohorts, it is natural to use this data
192 to define data driven gene co-expression networks and gene sets, so called co-expression modules.
193 These gene sets are then annotated according to their gene function or cell type specificity and then
194 related to disease via GWAS results using SNP set enrichment analysis. The link between genes and
195 SNPs can naturally be established via cis-eQTLs of the genes of a co-expression module. This
196 approach was also used in the CAD study mentioned above (Mäkinen et al. 2014). It is important to
197 note that co-expression modules are not necessarily fully overlapping with biochemical pathways
198 although they might represent the same disease process. For instance the modules might contain
199 transcriptional regulators and parts of a biochemical process that they control.

200 Network topology of co-expression networks is often used to prioritize candidate genes based on the
201 assumption, that genes with many network connections (so called hubs) are more important (Wang
202 et al. 2012; Shu et al. 2017; Mäkinen et al. 2014; Talukdar et al. 2016; Franzén et al. 2016). A study
203 investigating shared molecular networks and their drivers between cardiovascular diseases and type 2
204 Diabetes applied this strategy (Shu et al. 2017). Knockout mice for selected key driver genes show
205 indeed metabolic phenotypes and gene expression changes in the network neighborhood of the key
206 drivers. Similarly several studies on CAD identified key driver genes and provided evidence for their
207 functional implication in mouse (Talukdar et al. 2016) and in vitro studies (Talukdar et al. 2016;
208 Mäkinen et al. 2014).

209 **Conclusions**

210 eQTL data provides first leads towards uncovering the mechanisms underlying the statistical
211 associations observed between genetic loci and common cardiovascular diseases. Major challenges
212 for a broad applicability of this approach need to be overcome. First, regulatory elements and
213 therefore also the regulatory impact of sequence variation is highly cell type specific. The GTEx
214 project is addressing this challenge by providing a large scale cross tissue eQTL data base. However,
215 not all conceivable tissues and cell types can be systematically analyzed. In particular transient
216 developmental stages might leave a lasting phenotypic footprint. Induced pluripotent stem cells from
217 cohorts offer an elegant solution (Kilpinen et al. 2017) as they can potentially be differentiated into
218 any cell type or developmental stage (Nguyen et al. 2018) and studied for eQTLs. A second
219 challenge is posed by variability of the genetic effects on expression between different cells making
220 up a tissue and even between cells of the same cell type. eQTL mapping based on single cell
221 transcriptomic data is becoming feasible (Kang et al. 2018) and can be used to quantify and map the

222 genetic determinants of cell to cell variability of gene expression. Lastly the grand challenge is to
 223 move from correlation or co-localization towards causation. Clearly this is the most difficult task and
 224 requires on top of rigorous statistical approaches such as MR also experimental validation.

225 1 Conflict of Interest

226 The author declares that the research was conducted in the absence of any commercial or financial
 227 relationships that could be construed as a potential conflict of interest.

228 2 Author Contributions

229 MH wrote the manuscript.

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461
462 **5 Tables**

463
464 Table 1. Recent cardiovascular eQTL resources.
465

Ref	Tissue	Sample size	Population
(Sigurdsson et al. 2017)	Left Atrial wall	62	European
(Heinig et al. 2017)	Left Ventricle	205	European
(Christophersen et al. 2017)	Left Atria	329	European / African American
(Koopmann et al. 2014)	Left Ventricle	129	European
(Battle et al. 2017)	Atrial Appendage	264	European / African American
(Battle et al. 2017)	Left Ventricle	272	European / African American
(Battle et al. 2017)	Aorta	267	European / African American
(Battle et al. 2017)	Tibial artery	388	European / African American
(Battle et al. 2017)	Coronary artery	152	European / African American
(Battle et al. 2017)	Adipose - Subcutaneous	385	European / African American
(Battle et al. 2017)	Adipose - Visceral	313	European / African American
(Battle et al. 2017)	Liver	153	European / African American
(Franzen et al. 2016)	Mammary artery	600	European
(Franzen et al. 2016)	Atherosclerotic aortic root	600	European
(Franzen et al. 2016)	Visceral abdominal fat	600	European
(Franzen et al. 2016)	Skeletal muscle	600	European
(Franzen et al. 2016)	Liver	600	European

466
467 **Figure legends**

468
469 **Figure 1:** *Using eQTL data to identify causal candidate gene at GWAS loci.* Integration of eQTL and
470 GWAS data allows for the identification of candidate causal genes, where the effect of the genetic
471 variant (SNP) on the complex trait is mediated by expression levels of an RNA encoded at the locus
472 (A). Overlapping associations of gene expression and clinical trait at the same locus are however not
473 sufficient to infer causality, as they might also be explained as independent pleiotropic effects (A).
474 Depending on the availability of overlapping individual level data sets of genotypes, gene expression
475 and clinical traits there exist several statistical methods to perform causal inference from the data (B).