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Supporting Information

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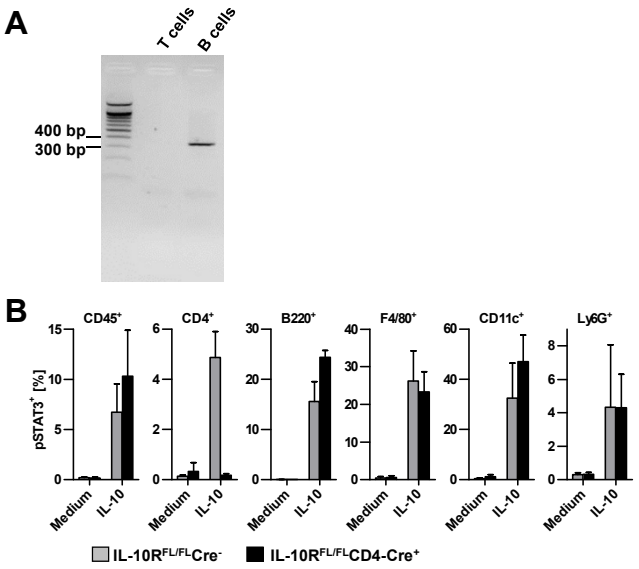
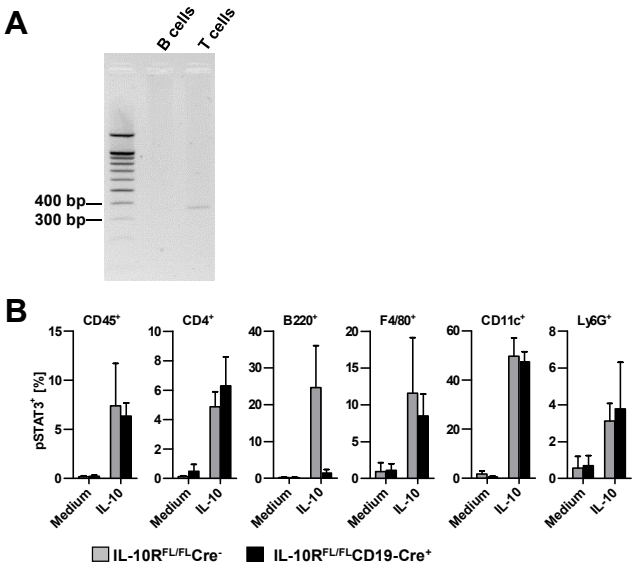
IL-10 signaling in dendritic cells is required for tolerance induction in a murine model of allergic airway inflammation



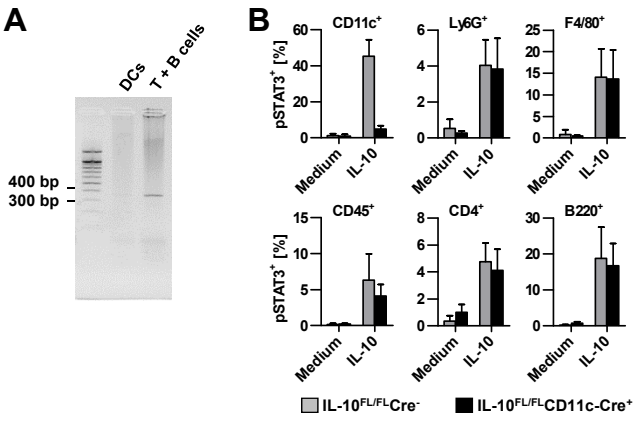
Suppl. Figure 1:
T cell specific deletion of IL-10Rα and IL-10 signaling in IL-10R$^{FL/FL}$CD4-Cre^{+} mice
(**A**) PCR analysis for IL-10Rα from DNA of sorted T cells (CD3^{+}) and B cells (CD19^{+}) of IL-10R$^{FL/FL}$CD4-Cre^{+} mice. Intact IL-10Rα results in a PCR product of 373 bp length. One exemplary experiment of four with 1-2 mice is shown. (**B**) Flow cytometric analysis of STAT3 phosphorylation upon *in vitro* IL-10 stimulation in CD45^{+} cells, T cells (CD4^{+}), B cells (B220^{+}), macrophages (F4/80^{+}), DCs (CD11c^{+}) and neutrophils (Ly6G^{+}) from spleens of IL-10R$^{FL/FL}$ CD4-Cre^{+} mice and Cre^{-} littermates. Data of 6 mice per group are shown as mean+SD.

Suppl. Figure 1



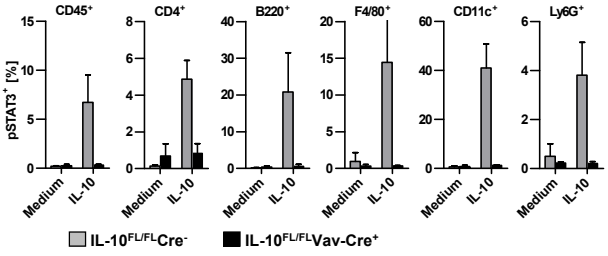
Suppl. Figure 2:
B cell specific deletion of IL-10Rα and IL-10 signaling in IL-10R$^{FL/FL}$CD19-Cre^{+} mice
(**A**) PCR analysis for IL-10Rα from DNA of sorted T cells (CD3^{+}) and B cells (CD19^{+}) of IL-10R$^{FL/FL}$CD19-Cre^{+} mice. Intact IL-10Rα results in a PCR product of 373 bp length. One exemplary experiment of four with 1-2 mice is shown. (**B**) Flow cytometric analysis of STAT3 phosphorylation upon *in vitro* IL-10 stimulation in CD45^{+} cells, T cells (CD4^{+}), B cells (B220^{+}), macrophages (F4/80^{+}), DCs (CD11c^{+}) and neutrophils (Ly6G^{+}) from spleens of IL-10R$^{FL/FL}$ CD19-Cre^{+} mice and Cre^{-} littermates. Data of 4-6 mice per group are shown as mean+SD.

Suppl. Figure 2



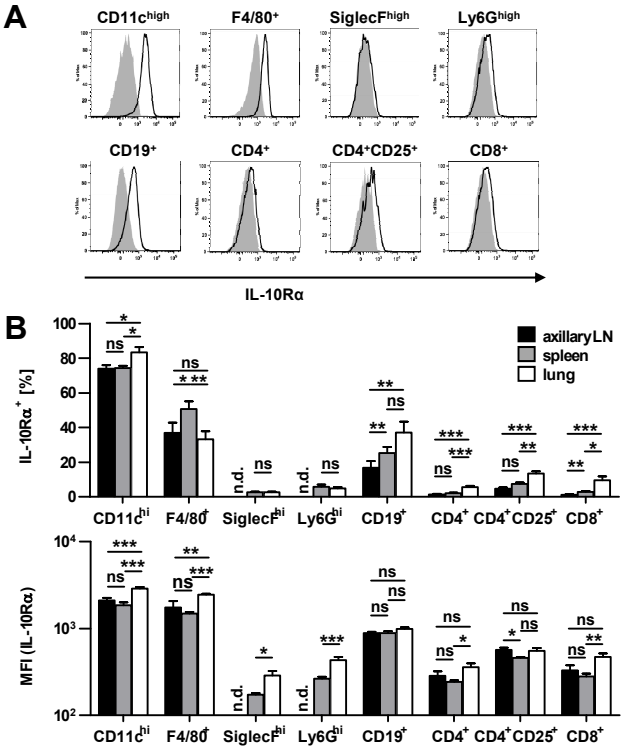
Suppl. Figure 3:
IL10Rα-deficiency and IL-10 signaling in IL10R$^{FL/FL}$CD11c-Cre^{+} mice
(A) PCR analysis for IL-10Rα from DNA of sorted DCs (CD11c^{+}MHCII^{+}) and T and B cells (CD3^{+} and CD19^{+} cells) of IL-10R$^{FL/FL}$CD11c-Cre^{+} mice. Intact IL-10Rα results in a PCR product of 373 bp length. One exemplary experiment of two with 3 mice is shown (B) Flow cytometric analysis of STAT3 phosphorylation upon *in vitro* IL-10 stimulation in DCs (CD11c^{+}), neutrophils (Ly6G^{+}), macrophages (F4/80^{+}), CD45^{+} cells, T cells (CD4^{+}) and B cells (B220^{+}) from spleens of IL-10R$^{FL/FL}$ CD11c-Cre^{+} mice and Cre^{-} littermates. Data of n=6-13 mice/group are shown as mean+SD.

Suppl. Figure 3



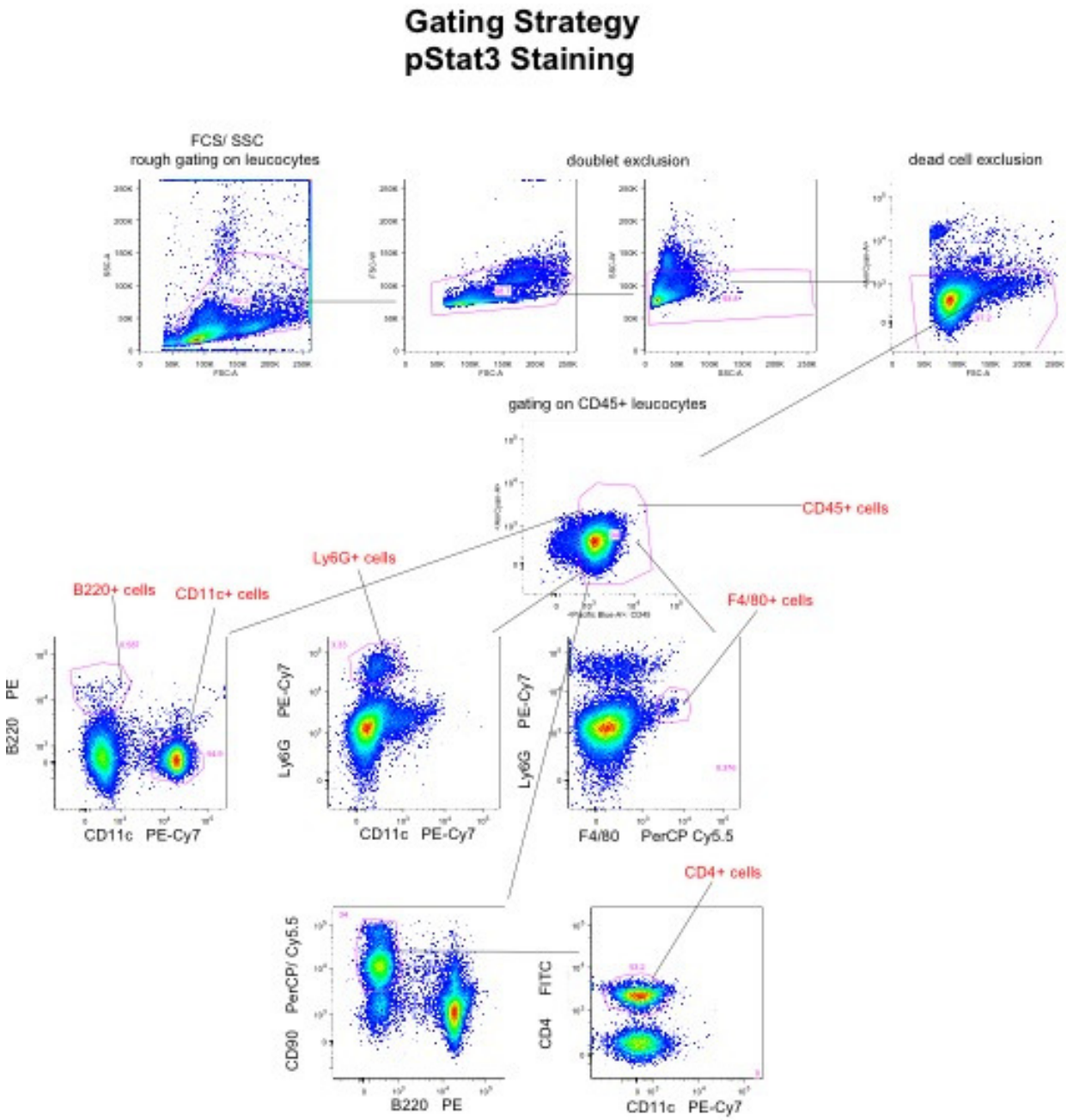
Suppl. Figure 4:
Lack of IL-10 signaling in IL-10R$^{FL/FL}$Vav-Cre^{+} mice
Flow cytometric analysis of STAT3 phosphorylation upon *in vitro* IL-10 stimulation in CD45^{+} cells, T cells (CD4^{+}), B cells (B220^{+}), macrophages (F4/80^{+}), DCs (CD11c^{+}) and neutrophils (Ly6G^{+}) from spleens of IL-10R$^{FL/FL}$ Vav-Cre^{+} mice and Cre^{-} littermates. Data of 4-7 mice per group are shown as mean+SD.

Suppl. Figure 4



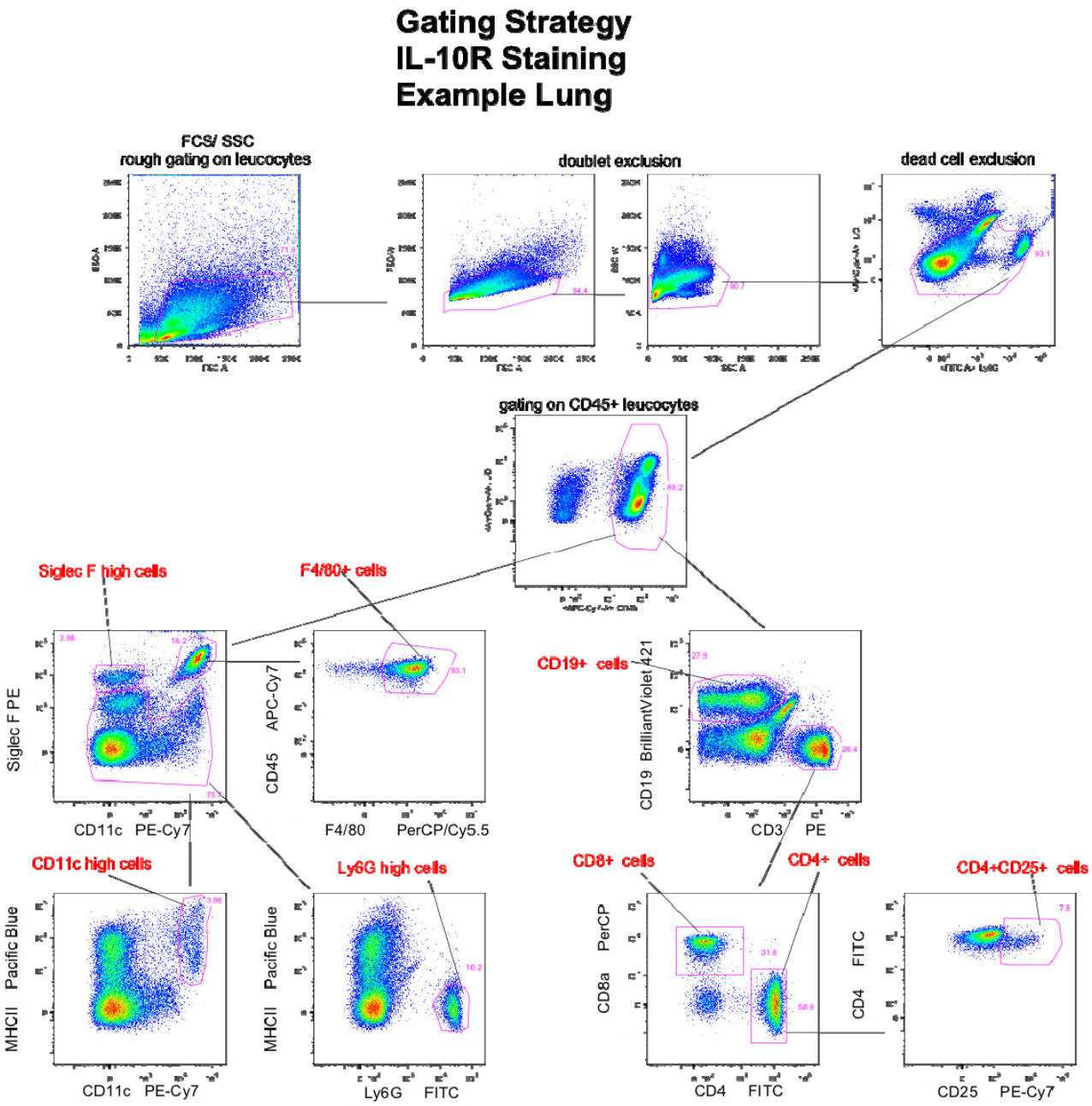
Suppl. Figure 5:
IL-10Rα expression of immune cells
IL-10Rα expression on different immune cells from lung, lymph node and spleen of naïve mice was analyzed by flow cytometry. (A) Representative histograms of IL-10Rα staining (black line) compared to isotype control (grey) in DCs ($CD11c^{high}$), macrophages ($F4/80^+$), eosinophils ($SiglecF^{high}$), neutrophils ($Ly6G^{high}$), B cells ($CD19^+$), $CD4^+$ T cells, $CD4^+CD25^+$ T cells and $CD8^+$ T cells from lung tissue. (B) Percentage of IL-10Rα$^+$ cells of the indicated populations from axillary lymph nodes (LN), spleen and lung (upper panel) and corresponding mean fluorescence intensity (MFI) of IL-10Rα staining (lower panel). Pooled data (n=6 mice/group and experiment; mean+SEM, Mann-Whitney U test) of 3 independent experiments are shown. * $p<0.05$, ** $p<0.01$, *** $p<0.001$

Suppl. Figure 5



Suppl. Figure 6:
Gating strategy for pStat3 staining
Gating strategy used for flow cytometric analysis of STAT3 phosphorylation upon in vitro IL-10 stimulation of spleen cells

Suppl. Figure 6



Suppl. Figure 7:
Gating strategy for IL-10R staining
Gating strategy used for flow cytometric analysis of IL-10Rα expression on different immune cells. One representative lung cell sample is shown.

Suppl. Figure 7