## **Supplemental information**



A GlastCreER<sup>T2</sup>; GFP; Tfam<sup>wt/wt</sup> and GlastCreER<sup>T2</sup>; GFP; Tfam<sup>fl/fl</sup>

## Supplemental Figure 1: Confirmation of *Tfam* deletion in astrocytes

(A) Experimental paradigm used for (B). (B) Genotyping PCR of GFP-expressing astrocytes isolated by FACS from distinct brain regions of 10 months old  $Tfam^{ctrl}$  and  $Tfam^{cko}$  mice (3 animals per genotype).  $Tfam^{ctrl}$  animals exclusively carried the  $Tfam^{wt}$  band (404 bp); positive control (Tfam<sup>wt</sup> lane) was DNA isolated from tail clips of  $Tfam^{ctrl}$  mice. Recombination of the Tfam locus in astrocytes isolated from diverse brain regions of  $Tfam^{cko}$  mice was indicated by  $Tfam^{cko}$  band (330bp);  $Tfam^{floxed}$  band (437bp); DNA of  $Tfam^{cko}$  lane derived from tail clips of  $Tfam^{cko}$  mice. OB = olfactory bulb, SVZ = subventricular zone, CB = cerebellum, HC = hippocampus, CTX = cortex. (C) Experimental scheme used in (D-E). (D) Confocal image of HTNCre transduced astrocytes derived from GFP;  $Tfam^{fl/fl}$  animals; Immunohistochemistry against the GFP reporter (green) indicates recombined cells; Cytochrome C (CytC, white) labels mitochondria; recombined astrocytes (GFP<sup>+</sup>) lost expression of Tfam (red); non-recombined astrocytes (GFP<sup>-</sup>) expressed Tfam. (E) Genotyping PCR of astrocytes transduced

with different amounts of HTNCre or control solution. Increasing recombination efficiency of the *Tfam* locus occurred with increasing concentration of Cre protein. Band size: *Tfam*<sup>floxed</sup> band = 437bp, *Tfam*<sup>cko</sup> band = 330bp. Scale bar =  $10\mu m$ .



Supplemental Figure 2: *Tfam*-deficient astrocytes survive under physiological and injury conditions

(A-C) Survival of astrocytes in  $Tfam^{ctrl}$  and  $Tfam^{cko}$  mice was measured by counting recombined astrocytes (A) under physiological conditions 4 months post-recombination, (B) one year post-recombination; (C) upon PIT-induced injury 4 months post-recombination. (A)  $n_{ctrl} = 3$  animals,  $n_{cko} = 4$  animals; (B)  $n_{ctrl} = 4$  animals,  $n_{cko} = 4$  animals; (C)  $n_{ctrl} = 5$  animals,  $n_{cko} = 5$  animals. Data represented as mean  $\pm$  SEM; t-test (B, C) and Mann-Whitney test (A) were performed to determine significance.



## Supplemental Figure 3: Cell death in the contralateral and PIT-lesioned cortex

(A-B) Confocal images of a coronal cortical section from  $Tfam^{ctrl}$  and  $Tfam^{cko}$  mice upon PIT; immunohistochemistry against Casp3 (red) to identify dying cells; nuclei stained with DAPI (blue). Contralateral hemisphere shown on the right; PIT lesioned hemisphere (left) with lesion core containing Casp3<sup>+</sup> cells and damaged tissue. Very few cell undergo cell death in the contralateral hemisphere. (C) High magnification confocal image of perilesional area; immunostaining against Casp3<sup>+</sup> in red (cell death marker; arrows); GFP (labeling recombined cells, green); NeuN<sup>+</sup> (neuronal marker; white), and DAPI (nulcei; blue) showing a dying neuron (Casp3<sup>+</sup>/GFP<sup>-</sup>/NeuN<sup>+</sup>). (D-F) Confocal images and quantification of Casp3 immunostaining (red) of the contralateral hemispheres; GFP<sup>+</sup> shows recombined cells (green); NeuN labels neurons (white); DAPI indicates cell nuclei. No difference in neuronal cell death was detected contralaterally between  $Tfam^{ctrl}$  and  $Tfam^{cko}$  mice (F). (D-F) n<sub>ctrl</sub> = 5 animals, n<sub>cko</sub> = 5 animals. Scale bars = 100µm (A-B), and = 20µm (C-E). Data represented as mean ± SEM; t-test was performed to determine significance.