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Title: Negligible Effects of TiO2 Nanoparticles at Environmentally Relevant Concentrations on the Translocation and Accumulation of Perfluorooctanoic Acid and Perfluorooctanesulfonate in Hydroponically Grown Pumpkin Seedlings (Cucurbita maxima × C. moschata)

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Abstract: Titanium dioxide nanoparticles (TiO2-NPs) are widely distributed in the environment. It has been demonstrated that TiO2-NPs could modify the environmental fate and bioavailability of organic pollutants, which affects ecological risks of TiO2-NPs and organic pollutants. In this study, the uptake, translocation and accumulation of perfluorooctanoic acid (PFOA) and perfluorooctanesulfonate (PFOS) in pumpkin plants was investigated in the presence of TiO2-NPs. We reported for the first time the negligible effects of TiO2-NPs at environmentally relevant concentrations (0.05-5 mg/L) on the uptake and accumulation of PFOA and PFOS in hydroponically grown pumpkin seedlings regardless of root, stem and leaf. This phenomenon was independent of the initial concentrations of PFOA/PFOS and TiO2-NPs in the exposure solution. Also, seedling mass and contents of chlorophyll and anthocyanin were not affected by the co-exposure. Adsorption tests demonstrated the negligible adsorption of PFOA/PFOS on TiO2-NPs in the exposure solution. Moreover, uptake of PFOA/PFOS was insensitive to aquaporin inhibitor AgNO3 but significantly inhibited by niflumic acid (anion channel blocker) and 2,4dinitrophenol (metabolic inhibitor) whereas Ti concentration in root was not affected by niflumic acid and 2,4-dinitrophenol but significantly decreased by AqNO3, indicating that transport of PFOA/PFOS and TiO2-NPs were via different routes into the pumpkin seedling. It was proposed that different pathways by which TiO2-NPs and PFOA/PFOS transported into the pumpkin seedling and negligible adsorption of PFOA/PFOS on TiO2-NPs contributed to the negligible effects of TiO2-NPs on the uptake, translocation and accumulation of PFOA/PFOS in pumpkin seedlings. In total, this work would improve our understanding of the ecological risks of TiO2-NPs in the environment.

Response to Reviewers: Dear Prof. Dr. Shuzhen Zhang,

The 'Response to Comments' has been uploaded. please see attach files. Thank you very much.

To: Prof. Dr. Shuzhen Zhang

From: Prof. Xueping Zhao

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Dear Prof. Dr. Shuzhen Zhang,

Thank you very much for handling our manuscript entitled: 'Negligible Effects of TiO_2 Nanoparticles at Environmentally Relevant Concentrations on the Translocation and Accumulation of Perfluorooctanoic Acid and Perfluorooctanesulfonate in Hydroponically Grown Pumpkin Seedlings (Cucurbita maxima × C. moschata)' (**STOTEN-D-19-03789**). We are highly grateful to you and two reviewers for taking time to review our manuscript and providing valuable comments to revise and improve the manuscript. All the suggestions have been carefully considered in the revised manuscript and the improvements have been made as shown in the 'Response to Comments' attached to this letter.

All data are original and the work has not been considered elsewhere for publication.

We sincerely hope that you will find the revised version of the manuscript satisfactory for publication. Should there be any additional improvements necessary please do not hesitate to contact us.

Once again, we appreciate your guidance and suggestions, which are valuable in improving the quality of our manuscript.

With best regards, Yours sincerely, Prof. Xueping Zhao Negligible Effects of TiO₂ Nanoparticles at Environmentally Relevant Concentrations on the Translocation and Accumulation of Perfluorooctanoic Acid and Perfluorooctanesulfonate in Hydroponically Grown Pumpkin Seedlings (*Cucurbita maxima* \times *C. moschata*)

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Graphical abstract



Highlights

1. Effect of TiO₂-NPs on accumulation of PFOA/PFOS in pumpkin seedling was explored.

- 2. Environmentally relevant concentrations of TiO_2 -NPs were applied in this work.
- 3. The negligible effects were observed.
- 4. The negligible adsorption of PFOA/PFOS on TiO_2 -NPs was proposed as one reason.
- 5. Different pathways of TiO₂-NPs and PFOA/PFOS into root were another reason.

1 Abstract

2	Titanium dioxide nanoparticles (TiO ₂ -NPs) are widely distributed in the environment. It has been
3	demonstrated that TiO ₂ -NPs could modify the environmental fate and bioavailability of organic pollutants, which
4	affects ecological risks of TiO ₂ -NPs and organic pollutants. In this study, the uptake, translocation and
5	accumulation of perfluorooctanoic acid (PFOA) and perfluorooctanesulfonate (PFOS) in pumpkin plants was
6	investigated in the presence of TiO ₂ -NPs. We reported for the first time the negligible effects of TiO ₂ -NPs at
7	environmentally relevant concentrations (0.05-5 mg/L) on the uptake and accumulation of PFOA and PFOS in
8	hydroponically grown pumpkin seedlings regardless of root, stem and leaf. This phenomenon was independent of
9	the initial concentrations of PFOA/PFOS and TiO ₂ -NPs in the exposure solution. Also, seedling mass and contents
10	of chlorophyll and anthocyanin were not affected by the co-exposure. Adsorption tests demonstrated the negligible
11	adsorption of PFOA/PFOS on TiO2-NPs in the exposure solution. Moreover, uptake of PFOA/PFOS was
12	insensitive to aquaporin inhibitor AgNO ₃ but significantly inhibited by niflumic acid (anion channel blocker) and
13	2,4-dinitrophenol (metabolic inhibitor) whereas Ti concentration in root was not affected by niflumic acid and
14	2,4-dinitrophenol but significantly decreased by AgNO ₃ , indicating that transport of PFOA/PFOS and TiO ₂ -NPs
15	were via different routes into the pumpkin seedling. It was proposed that different pathways by which TiO_2 -NPs
16	and PFOA/PFOS transported into the pumpkin seedling and negligible adsorption of PFOA/PFOS on TiO2-NPs
17	contributed to the negligible effects of TiO ₂ -NPs on the uptake, translocation and accumulation of PFOA/PFOS in
18	pumpkin seedlings. In total, this work would improve our understanding of the ecological risks of TiO ₂ -NPs in the
19	environment.

20

21 Keywords

22 TiO₂ Nanoparticles; Pumpkin seedling; Perfluoroalkyl substances; Uptake; Accumulation

1. Introduction

25	In recent decades, titanium dioxide nanoparticles (TiO ₂ -NPs) have been extensively used in commercial
26	products such as functional textiles, personal care products, food storage materials, house appliances and
27	semiconductors due to the special physiochemical properties (Ai-Kattan et al., 2014; Shandilya et al., 2015;
28	Morsella et al., 2016; Kaegi et al., 2017), which results in an accidental or intentional discharge of TiO ₂ -NPs into
29	the natural environment during the production, usage and disposal of these products (Gondikas et al., 2014;
30	Shandilya et al., 2015; Kaegi et al., 2017). On the basis of material flow models, predicted concentrations of
31	TiO_2 -NPs in the environment ranged from tens $\mu g/L$ (e.g., water) to tens mg/kg (e.g., sediment and soil) (Sun et al.,
32	2016; 2017). TiO ₂ -NPs have physicochemical properties including high insolubility and large surface area, which
33	implies that they are prone to interact with other substances such as organic chemicals. With the co-existence of
34	organic pollutants in the natural environment, TiO2-NPs would modify the environmental fate and bioavailability
35	of organic pollutants. The enhanced bioaccumulation of bisphenol A, perfluoroctanesulfonate (PFOS) and
36	decabromodiphenyl ether in zebrafish was observed in the presence of TiO ₂ -NPs at environmentally relevant
37	concentrations, and TiO ₂ -NPs acted as a carrier of chemicals due to the adsorption of chemicals on TiO ₂ -NPs
38	(Wang et al., 2014; Qiang et al., 2015; Fang et al., 2016). To date, nevertheless few studies investigated the effect
39	of TiO ₂ -NPs on the uptake, translocation and bioaccumulation of pollutants in plants. Given precautionary
40	principles, some questions arise inevitably: What is the influences of TiO ₂ -NPs at environmentally relevant
41	concentrations on the translocation and bioaccumulation of organic chemicals in plants? Can TiO ₂ -NPs at
42	environmentally relevant concentrations enhance or moderate the uptake of organic chemicals in plant? And thus
43	what is the potential reason for the phenomenon?
44	Perfluoroalkyl substances are a large group of anthropogenic persistent organic pollutants that have been
45	attracted wide attention as global contaminants. Among them, perfluorooctanoic acid (PFOA) and PFOS are two
46	of the most widely and extensively detected homologous in the natural environment (Butt et al., 2010;

47	Zareitalabad et al., 2013; Wang et al., 2015). Both PFOA and PFOS have shown high toxicity to aquatic
48	organisms and plants (Du et al., 2010; Hagenaars et al., 2013; Nordén et al., 2016), and even neural toxicity was
49	observed in recent (Yin et al., 2018). Plants such as pumpkin, soybean and wheat could accumulate PFOA and
50	PFOS (Zhao et al., 2013; Bizkarguenaga et al., 2016; Xiang et al., 2018), being a potential risk for humans
51	through food chains (de Vos et al., 2008; van Asselt et al., 2011). Foods of plant origin (e.g. fruit and vegetables)
52	have been observed to be the most important for the dietary exposure to PFOA in some European countries
53	despite relative low concentrations measured in these foods (Klenow et al., 2013). Obviously, it is of importance
54	to understand whether uptake, translocation and accumulation of PFOA and PFOS in plants would be affected by
55	TiO ₂ -NPs at environmentally relevant concentrations. Yet, no data on this issue has been available to date.
56	In the present study, herein, a hydroponically grown pumpkin seedling (<i>Cucurbita maxima</i> \times <i>C. moschata</i>) was
57	used as a model plant to investigate the effect of TiO ₂ -NPs at environmentally relevant concentrations on the
58	uptake, translocation and accumulation of PFOA and PFOS in plants. Three concentrations of TiO ₂ -NPs and two
59	different dose of PFOA/PFOS were examined. Besides quantification the amount of titanium, PFOA and PFOS in
60	different organs including root, stem and leaf, biomass and contents of chlorophyll as well as anthocyanin were
61	determined. According to the results of adsorption experiments and effects of channel blockers and metabolic
62	inhibitors on the uptake of PFOA and PFOS, we further proposed potential reasons for the observed phenomenon.
63	2. Materials and methods
64	2.1 Chemicals
65	Sodium hypochlorite, silver nitrate, niflumic acid, hydrochloric acid, 2,4-dinitrophenol, ethanol and chloroform
66	were purchased from Sinopharm Chemical Reagent Co. Ltd (Shanghai, China). Perfluorooctanesulfonic acid
67	potassium salt (KPFOS >98%) and perfluorooctanoic acid (PFOA >96%) obtained from CNW Technologies
68	GmbH (Shanghai, China) were used for hydroponic exposure experiments. Analytical reference standards of

69 PFOA, PFOS, ${}^{13}C_8$ -PFOS, ${}^{13}C_4$ -PFOA and ${}^{13}C_8$ -PFOA were purchased from Wellington Laboratories (Guelph,

70	Ontario, Canada). Acetonitrile and methanol of HPLC-MS grade were purchased from Fisher (Waltham, MA,
71	USA). All reagents used in this study were of analytical or HPLC grade and used as received without any further
72	treatment. Aqueous solutions were freshly prepared with ultrapure water which was from a Direct-Q-system
73	(Millipore, Billerica, USA) with a resistivity of 18.2 M Ω /cm. Florisil SPE cartridges (1000 mg, 6 mL) were
74	purchased from Applied Separations (Allentown, PA, USA). Oasis HLB SPE cartridges (60 mg, 3cc) were from
75	Waters (Wexford, Ireland). Supelclean ENVI-Carb 120/140 was from Supelco (Bellefonte, PA, USA).
76	TiO ₂ -NPs (Aeroxide P25, 21 nm) were from Sigma-Aldrich (St. Luis, USA). The ζ potential and
77	hydrodynamic size of TiO ₂ -NPs were determined using a Malvern Zeta sizer Nano Series dynamic light scattering
78	(DLS) instrument. The morphology of TiO ₂ -NPs was observed through a JEOL JEM-2100F high-resolution
79	transmission electron microscope (HRTEM) at 200 kV.
80	2.2 Materials
81	Pumpkin seeds (Cucurbita maxima × C. moschata, Taigu Yinong Seed Co., Ltd., Shanxi Province, China) were
82	surface-sterilized by 70% (v/v) of ethanol for 5 min and then were soaked in 5% (v/v) sodium hypochlorite
83	solution for 15 min. Afterward, seeds were washed thoroughly five times with ultrapure water and then soaked in
84	ultrapure water for 8 h at 25°C. Then, seeds were placed in Petri dishes in the dark at 25°C to germinate for 3 d.
85	The 3-day old pumpkin seedlings were then transferred to a vessel with wet sterile quartz sand and cultivated in
86	an illumination growth chamber at 25°C for a 16 h light period and at 22°C for an 8 h dark period. Light was
87	supplied by sets of fluorescent lighting with a light intensity of 150 - 200 μ mol m ⁻² s ⁻¹ . After incubation of 7 d,
88	seedling stems grew to 5~6 cm height, and healthy seedlings with similar strength were collected. The roots of
89	seedlings were rinsed with ultrapure water before hydroponic exposures.
90	2.3 Hydroponic exposures

The reactors were 50 mL polypropylene Corning tubes. Two pumpkin seedlings were cultivated in each reactor,
and each reactor was filled with 40 mL of ultrapure water with desired concentrations of TiO₂-NPs, PFOA or

93	PFOS. Three concentrations (0.05, 0.5 and 5 mg/L) of TiO ₂ -NPs and two dose (0.05 and 0.5 mg/L) of PFOA or
94	PFOS were examined. After cultivation, two pumpkin plants in each reactor were combined into one sample for
95	analysis. There were three parallel samples in each exposure group. Besides chemical exposures, plant blanks
96	(with seedlings but without chemicals, three reactors), unplanted controls (without seedlings but with chemicals,
97	three reactors) and TiO ₂ -NPs controls (seedlings single exposure to TiO ₂ -NPs, three reactors) were set-up
98	simultaneously. The pumpkin seedlings were cultivated in an illumination growth chamber at 25°C for a 16 h light
99	period and at 22°C for an 8 h dark period. Light was supplied by sets of fluorescent lighting with a light intensity
100	of 150 - 200 μ mol m ⁻² s ⁻¹ . The exposure was lasted for 8 d. All tubes were wrapped with aluminum foil to support
101	root growth in the dark and to avoid possible photolysis of PFOA or PFOS. Here it should be noted that the plant
102	was fixed by predrilled polypropylene foam snugly to make only roots emerge into the hydroponic solutions.
103	Water loss via evaporation of water itself was determined by weighing the unplanted controls every day,
104	indicating the negligible daily loss of water via volatilization. Also, water loss via transpiration from pumpkin
105	seedlings was determined gravimetrically and replenished every day. After cultivation of 8 d, all pumpkin
106	seedlings were manually collected and each seedling was sectioned into root, stem and leaf. Roots were
107	ultrasonically rinsed with ultrapure water to remove possible chemicals adsorbed on the surface. The rinse water
108	was combined with the exposure solution for the determination of chemicals in view of mass balance. All samples
109	except those for chlorophyll and anthocyanin measurements were freeze-dried and stored at -20°C before further
110	treatment. The measurements of chlorophyll, anthocyanin, titanium, PFOA and PFOS were described in the
111	Supplementary Material (Text S1, S2, S3 and S4 and Table S1).
112	In addition, to understand the kinetics of chemicals uptake by pumpkin seedlings, samples were also collected
113	at certain intervals (e.g., 0 h, 4 h, 14 h, 1 d, 2 d, 3 d, 5 d and 8 d). Also, to examine potential uptake pathways of
114	chemicals, different channel blockers such as AgNO ₃ (0.5 mmol/L, water-channel blocker), niflumic acid (0.5

115 mmol/L, anion-channel blocker) and 2,4-dinitrophenol (0.5 mmol/L, metabolic inhibitor) were added into

exposure solutions separately before cultivation (Wagatsuma et al., 1983; Wu et al., 2011; Devi et al., 2012). After
4 h exposure, samples were manually collected and treated according to the above-mentioned method. All the

treatments were performed in three replicates.

119 **2.4 Quality assurance and quality control (QA/QC)**

To avoid contamination, polytetrafluoroethylene (PTFE) materials were not used. Fresh polypropylene tubes were used during the experiment. The isotope dilution method was applied to quantify PFOA and PFOS. Before sample extraction, internal standards of ${}^{13}C_8$ -PFOS and ${}^{13}C_8$ -PFOA were spiked. Before sample injection,

123 ${}^{13}C_4$ -PFOA was added. The ion ratios of qualifier to quantifier were checked for native compounds in each sample

124 during the peak integration. The ratio was set as 26.5% for PFOA and 51.9% for PFOS with a maximum tolerance

125 of $\pm 30\%$ (SANCO/12571/2013). The recoveries of ${}^{13}C_8$ -PFOS, ${}^{13}C_8$ -PFOA, PFOS and PFOA were in the ranges

126 of 72-83%, 76-82%, 81-97% and 85-102%, respectively. The limit of detection (LOD) was defined as a value

127 corresponding to a signal-to-noise ratio of 3 (S/N=3). The LODs of PFOA and PFOS were both 2 µg/kg. At least

128 one procedure blank sample was prepared and analyzed with each batch of samples, indicating there was no

129 contamination during the sample preparation. The levels of PFOA or PFOS found in the plant blanks and

130 TiO₂-NPs controls were shown in Table S2. Since PFOA and PFOS concentrations in plant blank samples and

131 TiO₂-NPs control samples were comparable, the reported results in this work were subtracted the plant blanks.

132 One blank solvent was injected every 10 samples and no memory effect was observed. The amount of PFOA or

133 PFOS recovered from exposure solutions and seedlings were 86-110% of their initial amount in the hydroponic

134 solution. Titanium was not detected in plant blanks.

135 **2.5 Statistical analysis**

In this study, concentrations of PFOA, PFOS and Ti were shown on a dry weight basis. The normality of data was tested by using the Shapiro-Wilk Test (Origin 8.5), and the results showed that the data set for each treatment was normally distributed (p < 0.05). Then, student's t test (Origin 8.5) was used to evaluate possible differences of 139 statistical significance between treatments. Statements of significant differences were based on p < 0.05.

3. Results and discussion

3.1 Characterization of TiO₂-NPs

142	According to HRTEM images, the average size of TiO ₂ -NPs used in this study was 24.3 ± 1.7 nm (Figure S1 in
143	Supplementary Material), which is comparable to the size (21 nm) declared by the manufacturer (Sigma-Aldrich).
144	DLS measurement showed the TiO ₂ -NPs (e.g., 5 mg/L) hydrodynamic size of 130 ± 3.2 nm, with a polydispersity
145	index of 0.44 \pm 0.21. Also, TiO ₂ -NPs showed a strongly negative charge, with a ζ potential of -33.3 \pm 14 mV.
146	Although TiO_2 is well-known to be relatively insoluble, dissolution kinetics as a function of exposure time (e.g., 0
147	h, 4 h, 14 h, 1 d, 2 d, 3 d, 5 d and 8 d) were examined in the TiO ₂ -NPs alone or mixture of TiO ₂ -NPs and
148	PFOA/PFOS. No dissolved Ti was determined through the exposure period (data not shown), documenting the
149	high stability of TiO ₂ -NPs in the aqueous solution, which is well consistent with the finding in previous studies
150	(Bourgeault et al., 2015; Brzicova et al., 2019).
151	3.2 Effect of TiO ₂ -NPs on the uptake, translocation and accumulation of PFOA in pumpkin seedlings
152	The concentration of PFOA in pumpkin seedlings increased with the increasing concentration of PFOA in the
153	exposure solution regardless of seedlings exposed to PFOA alone or mixture of PFOA and TiO ₂ -NPs (Figure 1a).
154	However, no significant differences in concentrations of PFOA in seedlings were observed in the absence and
155	presence of TiO ₂ -NPs (Figure 1a), suggesting negligible effects of TiO ₂ -NPs on the uptake and accumulation of
156	PFOA in pumpkin seedlings. Moreover, this phenomenon was not affected by the initial concentrations of PFOA
157	and TiO ₂ -NPs (Figure 1a, b). For example, the co-exposed seedlings showed comparable concentrations of PFOA
158	regardless of the co-existed TiO ₂ -NPs at a concentration of 0.05, 0.5 or 5 mg/L (Figure 1b). This is different from
159	the finding in a recent study about the effect of TiO ₂ -NPs on the accumulation of Pb in hydroponically grown rice
160	seedlings. Cai et al. (2017) found that TiO2-NPs at a concentration of 1000 mg/L reduced the uptake and
161	bioaccumulation of Pb probably due to the obstruction effects of TiO ₂ -NP aggregates in the root tissues. Clearly,

the concentration of TiO₂-NPs used in this study was far lower than that of the work operated by Cai et al. (2017),
which would show more credible findings in our study due to the environmentally relevant concentrations of
TiO₂-NPs.

165	Root is the compartment where accumulated the highest concentration of PFOA. This phenomenon still
166	remained in the presence of TiO ₂ -NPs (Figure 1a, b), indicating that the translocation of PFOA would not be
167	affected by the co-existed TiO ₂ -NPs. Similarly, the uptake, translocation and accumulation of Ti in pumpkin
168	seedlings were not affected by the co-existed PFOA regardless of the initial concentrations of TiO ₂ -NPs and
169	PFOA in the exposure solution (Figure 1e, f), and the highest concentrations of Ti were also observed in root.
170	The uptake and accumulation kinetics of PFOA in different parts of pumpkin seedlings were investigated,
171	showing different trends of PFOA among the root, stem and leaf (Figure 2a,b,c). The concentration of PFOA in
172	root increased within the first 48 h, reaching the highest level of 20-25 mg/kg, followed by equilibrium afterwards
173	(Figure 2a). The PFOA concentration in stem remained constant (~4 mg/kg) after a gradual initial increase within
174	the first 48 h (Figure 2b). Similarly, the concentration of PFOA in leaf gradually increased toward 72 h, with the
175	highest level of 12-14 mg/kg, and then kept constant concentration (Figure 2c). Moreover, the presence of
176	TiO ₂ -NPs appeared not to affect the uptake and accumulation kinetics of PFOA. Interestingly, chlorophyll
177	production of leaf was unaffected by the high level of PFOA accumulated (Table 1); the total chlorophyll of plant
178	blanks was 13.0 ± 1.1 mg/g, with the PFOA-exposed or TiO ₂ -NPs and PFOA co-exposed seedlings having a value
179	of 12.8 ± 1.6 or 13.3 ± 0.79 mg/g. Quantitation of anthocyanin production supported the negligible changes in
180	chlorophyll as anthocyanin was considered as defense against chlorophyll loss (Jung et al., 2004; Garriga et al.,
181	2014); exposure to PFOA alone or mixture of PFOA and TiO2-NPs showed no impact on anthocyanin content
182	(Table 1). Also, we found that seedling masses were not affected by the co-exposure (Figure S2).
183	3.3 Effect of TiO ₂ -NPs on the uptake, translocation and accumulation of PFOS in pumpkin seedlings

184 Besides PFOA, PFOS exhibited similar phenomena; as shown in Figure 1c, compared to exposure to PFOS

185	alone, no significant changes in the concentration of PFOS accumulated in the seedling are observed in the
186	presence of TiO ₂ -NPs (p <0.05), suggesting that co-exposure to TiO ₂ -NPs at environmentally relevant
187	concentrations had no impacts on the uptake, translocation and accumulation of PFOS in pumpkin seedlings. This
188	phenomenon was also independent on the initial concentrations of TiO2-NPs and PFOS in the exposure solution
189	(Figure 1c,d). We found that root showed the highest level of PFOS, suggesting that root could readily accumulate
190	PFOS. Clearly, the PFOS concentrations (~200 mg/kg, Figure 1c) were an order of magnitude greater than those
191	of PFOA (~20 mg/kg, Figure 1a) accumulated in root when seedlings were exposed to the same concentrations of
192	PFOA and PFOS, indicating that PFOS would readily be accumulated by pumpkin seedlings compared to PFOA.
193	Protein and lipid have found to play an important role in the accumulation and distribution of PFOA and PFOS in
194	plants and protein would promote the root accumulation (Wen et al., 2016). The high PFOS in pumpkin root
195	compared to PFOA might be attributed to the high affinity of PFOS to plant protein (Xia et al., 2013). A previous
196	study conducted by Wen et al. (2013) also showed high levels of PFOS accumulated in the root of maize (Zea mys
197	L. cv. TY2) than those of PFOA. The highest concentration of Ti was also observed in the root for the TiO ₂ -NP
198	and PFOS co-exposure (Figure 1g, h), which was consistent with the phenomena of TiO ₂ -NP and PFOA
199	co-exposure.
200	The uptake and accumulation kinetics of PFOS in different parts of pumpkin seedlings were examined (Figure
201	2d, e, f). PFOS concentrations in root, stem and leaf followed a comparable tendency; increasing to a maximum
202	value, and then remaining constant. PFOS concentrations in root, stem and leaf were peaked at approximately 48,
203	48, and 72 h, respectively, with the highest concentrations in the ranges of 130-150, 10-15 and 50-60 mg/kg. It
204	appears that TiO ₂ -NP exposure exerted negligible influence on the kinetics of PFOS accumulation in pumpkin
205	seedlings, which is identical with observation of PFOA.
206	3.4 Potential mechanisms for negligible effects of TiO ₂ -NPs on PFOA/PFOS uptake and accumulation

207 (i) *PFOA/PFOS adsorption on TiO*₂-*NPs*. The data of this study showed that TiO₂-NPs at environmentally

208 relevant concentrations had negligible impacts on the uptake, translocation and accumulation of PFOA and PFOS

209 in the pumpkin seedlings, which is out of our expectation based on available studies. In order to understand the

210 potential mechanisms, potential roles of PFOA/PFOS adsorption on TiO₂-NPs at concentrations in the hydroponic

- 211 exposure experiments were examined, showing that negligible adsorption of PFOA/PFOS on the TiO₂-NPs in the
- tested exposure solution (Text S5 and Table S3), which might be related to the electrostatic repulsion between

213 negative PFOA/PFOS molecules and negative-charged TiO₂-NPs in the exposure solution.

218

219

(ii) Uptake channels of PFOA/PFOS and TiO₂-NPs. Effects of AgNO₃ (aquaporin inhibitor), niflumic acid

215 (anion channel blocker) and 2,4-dinitrophenol (metabolic inhibitor) on uptake and accumulation of PFOA/PFOS

216 were examined in the co-exposure solutions. Uptake of PFOA/PFOS was insensitive to aquaporin inhibitor

217 AgNO₃; compared to control, no significant differences in the concentration of PFOA/PFOS in the root were

observed (Figure 3a,c), indicating that aquaporins may not be a major channel for PFOA/PFOS transport into

pumpkin seedling root. In contrast, treatments with niflumic acid and 2,4-dinitrophenol significantly inhibited the

220 uptake of PFOA in root by 16 and 33% respectively (p < 0.05, Figure 3a). Comparably, the uptake of PFOS was

221 depressed by niflumic acid and 2,4-dinitrophenol significantly (p < 0.05, Figure 3c), and in comparison with

control, 81% and 79% of PFOS were recovered in root. Obviously, these findings suggested that transport of

223 PFOA and PFOS into pumpkin seedling was similar, and it was primarily an energy-dependent process and

224 mediated by anion channels, which is well consistent with the finding in previous study on uptake of PFOA in

maize (Wen et al., 2013). The *pKa* values of PFOA and PFOS were reported to be -0.2 and -3.27, respectively

(Brooke et al., 2004; Steinle-Darling et al., 2008), and thus PFOA and PFOS were in the anionic form in

227 hydroponic solution with pH of approximately 7.3 (data not shown). This might be the reason why the transport

228 pathway of PFOA/PFOS into pumpkin root was related with anion channels.

229 The Ti concentration in root was not affected by the niflumic acid and 2,4-dinitrophenol (Figure 3b,d),

suggesting that transport of TiO₂-NPs into pumpkin seedling root might be independent of energy process and

anion channels. Furthermore, we found that aquaporin inhibitor $AgNO_3$ significantly decreased the uptake of Ti by 51-54% in root (p < 0.05, Figure 3b,d), demonstrating that the aquaporins would be involved in uptake of TiO₂-NPs into pumpkin seedling root. Clearly, transport of PFOA/PFOS and TiO₂-NPs were via different routes into the pumpkin seedling, indicating that there was no competitive uptake between PFOA/PFOS and TiO₂-NPs. Taken together, it was reasonable to understand the negligible effects of TiO₂-NPs at environmentally relevant concentrations on the uptake, translocation and accumulation of PFOA/PFOS in the pumpkin seedling.

4. Conclusions

239 The production, usage and disposal of commercial products containing TiO₂-NPs or PFOA/PFOS would result 240 in their occurrence in the aquatic environment. Although previous studies documented either the enhanced accumulation of chemicals by TiO₂-NPs in aquatic organisms like zebra fish or the moderated uptake of chemicals 241 by TiO₂-NPs in hydroponically grown plants like rice seedlings, this study reported for the first time that 242 243 TiO₂-NPs at environmentally relevant concentrations had negligible effects on the uptake, translocation and 244 accumulation of PFOA and PFOS in hydroponically grown pumpkin seedlings. The observed phenomenon was 245 probably resulted from the different pathways by which TiO₂-NPs and PFOA/PFOS transported into the pumpkin 246 seedling root and negligible adsorption of PFOA/PFOS on TiO₂-NPs. To investigate the impact of TiO₂-NPs at environmentally relevant concentrations on the chemical 247 248 accumulation in plants is of vital importance for evaluating its ecological risks. The results in the present study is 249 contradictory to previous work, which suggests that more work should be conducted on the effect of TiO₂-NPs on 250 the chemical accumulation in plants in future, including clarifying the potential mechanisms. Meanwhile, it is 251 worth investigating whether other engineered nanomaterials at environmentally relevant concentrations also have negligible impacts on the transportation and accumulation of organic pollutants in plants in future, to 252

comprehensively understand the environmental risks of engineered nanomaterials.

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374	exposure (bars: average value (n=3), lines: standard deviations). The same normal letter indicates non-significant
375	differences among treatments at 0.05 level.
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400 Figure 2. Kinetics of PFOA/PFOS accumulation in different compartments (root, stem and leaf) of pumpkin





410 the root of pumpkin seedling (p < 0.05 compared with the control). (a) and (b) results were from the co-exposure

411 of TiO_2 -NPs and PFOA. (c) and (d) results were from the co-exposure of TiO_2 -NPs and PFOS.

	Plant blanks	TiO ₂ -NPs controls	PFOA	PFOS	TiO_2 -NPs + PFOA	TiO ₂ -NPs + PFOS
Chlorophyll contents (mg/g)	13.0 ± 1.1	13.5 ± 1.4	12.8 ± 1.6	13.4 ± 1.9	13.3 ± 0.79	12.7 ±1.2
Anthocyanin contents (mg/g)	0.11 ± 0.012	0.11 ± 0.013	0.11 ± 0.012	0.11 ± 0.021	0.11 ± 0.00	0.12 ± 0.013

Table 1. Contents of chlorophyll and anthocyanin in the leaf of pumpkin seedlings collected from different treatments.^a

422 ^a Here concentrations of TiO₂-NPs, PFOA and PFOS were 5.0, 0.5 and 0.5 mg/L respectively for pumpkin seedlings exposure.

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