PROFESSOR NILS HEYNE (Orcid ID : 0000-0001-9982-5093)

Article type : Original Report

Journal: Transplant Infectious Disease

Low-dose cidofovir and conversion to mTOR-based immunosuppression in polyomavirus-associated nephropathy (PVAN)

mTOR and cidofovir in PVAN

Thomas Mühlbacher, M.D.^{1,2,3}, Robert Beck, M.D.⁴, Silvio Nadalin, Prof.⁵, Nils Heyne, Prof. ^{1,2,3}, Martina Guthoff, Assist. Prof.^{1,2,3}

¹ Dept. of Diabetology, Endocrinology, Nephrology, Section of Nephrology and Hypertension, University of Tübingen, Otfried-Müller-Str. 10, 72076 Tübingen, Germany

² Institute for Diabetes Research and Metabolic Diseases of the Helmholtz Center Munich at the University of Tübingen, Otfried-Müller-Str. 47, 72076, Tübingen, Germany.

³ German Center for Diabetes Research (DZD e.V.), Neuherberg, Germany.

⁴ Institute of Medical Virology and Epidemiology of Viral Diseases, University of Tübingen, Elfriede-Aulhorn-Str. 6, 72076 Tübingen, Germany

⁵ Dept. of General-, Visceral- and Transplant Surgery, University of Tübingen, Hoppe-Seyler-Str. 3, 72076 Tübingen, Germany

This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the <u>Version of Record</u>. Please cite this article as <u>doi:</u> 10.1111/TID.13228

Correspondence to:

Nils Heyne, M.D.

Dept. of Diabetology, Endocrinology, Nephrology, Section of Nephrology and Hypertension, University of Tübingen, Otfried-Müller-Str. 10, 72076 Tübingen, Germany

phone.: +49-7071-29-82854

fax.: +49-7071-29-3174

e-mail: nils.heyne@med.uni-tuebingen.de

Acknowledgements

None

Abstract

Background: Polyomavirus-associated nephropathy (PVAN) remains a relevant complication following kidney transplantation with allograft loss rates of up to 50%. Reduction of overall immunosuppression is a cornerstone of therapy, whereas no specific antiviral regimen has shown conclusive benefit to date. The present case series demonstrates the efficacy of a dual therapeutic approach with low-dose cidofovir and conversion to mTOR-based immunosuppression in PVAN.

Methods: Patients with biopsy-proven PVAN having received low-dose cidofovir (0.25 mg/kg) according to the Tübingen Cidofovir Protocol and been converted to mTOR-based immunosuppression were analysed retrospectively.

Results: 23 patients with a median follow-up of 2.24 [IQR 1.55-5.01] yrs were included in the analysis. Median time to PVAN diagnosis was 268 [IQR 153 – 869] days after transplantation. Polyomavirus clearance from plasma was achieved in 78% of patients after a median of 118 [IQR 76-293] days. Of the 23 patients, nine patients (39%) lost their allograft function during follow-up, but only three of these (13%) due to PVAN. 14 patients (61%) stabilized or improved allograft function. The cidofovir protocol allowed for specific antiviral therapy without adverse nephrotoxicity, even in patients with low allograft function.

Conclusions: Low-dose cidofovir and conversion to mTOR-based immunosuppression allow for effective virus clearance and preservation of allograft function in a high proportion of patients with PVAN and progressive allograft dysfunction and may prolong allograft survival in these patients.

Keywords BKV, cidofovir, mTOR, polyoma, PVAN, renal transplantation

Introduction

With the introduction of potent immunosuppressive regimens, polyomavirus-associated nephropathy (PVAN) has emerged as a relevant infectious complication, affecting 1-10% of the kidney transplant recipients ^{1,2}. Whilst reduction of the overall immunosuppression remains a mainstay of therapy, facilitating immune-mediated BK-virus clearance, previous data have shown that 30-50% of the patients will develop progressive allograft dysfunction and subsequent allograft loss when treated with reduction of immunosuppression alone ^{3,4}. Moreover, reduction of the overall immunosuppression carries the risk of acute or chronic rejection. Therefore, other treatment options including specific antiviral agents as well as modification of the immunosuppressive regimen rather than reduction of the overall immunosuppression have been investigated.

Although in-vitro drug effects on the course of BKV-replication have been demonstrated for leflunomide, fluoroquinolones and intravenous immunoglobulins ⁵⁻⁷, clinical data have not shown convincing results for a broad implementation of these strategies. The nucleotide analogue cidofovir has also demonstrated in-vitro activity against polyomavirus, whereas various clinical case series and retrospective studies have shown inconsistent results following administration of cidofovir at variable doses (0.25 mg/kg – 1 mg/kg), duration and treatment cycles ⁸⁻¹¹. The rationale for the use of cidofovir is based on the intratubular uptake and activity at the site of viral replication, however, this fact being also the cause of the most common side effect, nephrotoxicity.

A number of studies have shown benefits of an mTOR-inhibitor based immunosuppression regarding the risk of BK-viremia compared to the standard of care tacrolimus-based regimen ^{12,13}, however limited data exist on effectiveness of this strategy in case of a biopsy proven polyomavirus-nephropathy. The rationale for this treatment approach relies on the interaction of BK-virus replication with components of the mTOR pathway as well as data showing an increased differentiation of virus-specific CD8⁺-memory T-cells ¹⁴⁻¹⁶ following mTOR-inhibition.

We now present data from our kidney transplant center focusing on probability and time course of BK-virus clearance as well as allograft function in kidney transplant recipients with biopsy proven PVAN, treated with a novel dual therapeutic approach consisting of low-dose cidofovir together with conversion to mTOR-based immunosuppression.

Materials and Methods

We performed a retrospective analysis of the center protocol in kidney transplant recipients from the Tübingen Collaborative Transplant Center between 04/2006 and 03/2018 with biopsy-proven PVAN. The analysis was conducted in accordance with the Declaration of Helsinki and approved by the institutional review board (651/2016BO2). As a retrospective analysis, no written informed consent was required. All patients had shown BK-viremia measured by quantitative PCR (qPCR) as well as deterioration of renal function prior to confirmation of the diagnosis PVAN in kidney biopsy. Screening for BKV-viremia was historically a clinical decision, mostly incidence-based upon worsening of allograft function. With the advent of routine monitoring, a surveillance protocol was implemented for a standardized approach starting at the first visit after transplantation, 3 and 9 months post-transplantation, as well as incidence-based. PCR analysis was performed using the LightMix Kit Polyomaviruses JC and BK and was replaced by the Realstar BKV PCR Kit 1.0 in December 2014. Patients with BK-viremia without histological evidence of PVAN were not included in the analysis.

The application of low-dose cidofovir was performed in an inpatient clinic according to the Tübingen Cidofovir Protocol, developed to effectively deliver therapeutic drug concentrations at limited nephrotoxicity (*Table 1*). Adequate intravenous hydration aiming at a urine output > 100 ml/h is used in order to achieve a calculated tubular passage time for reduction of toxicity. No probenecid is administered to allow for tubular uptake of cidofovir via the basolateral membrane human renal organic anion transporter 1 (hOAT1) and obtain therapeutic drug concentrations at the site of viral replication. A single cidofovir application was performed. The necessity of subsequent applications was evaluated clinically and based on the course of BKV replication rate and allograft function.

In most cases, conversion to mTOR-based immunosuppression was performed via reduction of calcineurin inhibitor (CNI) trough levels and stopping as soon as targeted mTOR inhibitor trough levels were achieved. In few patients with high immunological risk, the CNI was continued with cessation of mycophenolate mofetil (MMF).

Data are given as median [interquartile range]. Comparison between groups was tested using Wilcoxon-Test. Results with two-sided $p \le 0.05$ were considered statistically

significant. The JMP (Version 14.0, SAS Institute, Cary, NC) statistical software package was used.

Results

Patient characteristics:

A total of 23 kidney transplant recipients met the criteria of biopsy proven PVAN and treatment according to the protocol with low-dose cidofovir and conversion to an mTOR-based immunosuppression. The cohort consisted of 19 males and 4 females with a median age of 54 [40-59] yrs at diagnosis of PVAN. 10 patients had received a living donor kidney transplantation, whereas 13 patients had a kidney from a deceased donor. Median eGFR MDRD at PVAN diagnosis was 30.1 [24.7-38.2] ml/min/1.73m² with a median time to diagnosis of 268 [153-869] days after kidney transplantation. At diagnosis of PVAN, all patients were on a maintenance immunosuppression comprising tacrolimus and mycophenolate mofetil (MMF). Corticosteroids were used in 16 patients, whereas 7 patients were on corticosteroid-free maintenance immunosuppression. Median BK-virus replication rate in plasma at the time point of diagnosis was 65500 [25500 – 200250] copies/ml plasma. Patient characteristics are shown in *Table 2*.

Therapeutic interventions:

All patients received low-dose cidofovir according to the Tübingen Cidofovir Protocol (*Table 1*). Median cumulative dose was 25.0 [18.6-30.6] mg of cidofovir. Median eGFR at the time-point of first cidofovir administration was 28.8 [24.3-36.0] ml/min/1.73m². All patients were converted to mTOR-based immunosuppression. In 18 patients, the mTOR-inhibitor sirolimus was used and replaced tacrolimus, whereas four patients were converted to everolimus replacing either tacrolimus, azathioprine or MMF. In one patient, MMF was replaced with leflunomide and afterwards changed to an immunosuppressive regimen containing sirolimus, leflunomide and tacrolimus. The mTOR inhibitor had to be discontinued in five patients (22%) after a median of 112 [60-211] days due to aphthous lesions, cellular rejection or development of donor-specific antibodies.

The application of low-dose cidofovir was not associated with any reported side effects. While 13 (57%) patients received only a single dose seven (30%) patients required a second dose and three (13%) patients received three doses of cidofovir, respectively. Further information regarding cidofovir applications and mTOR treatment are summarized in *Table 3*.

BKV clearance and allograft function:

Clearance of BK-viremia, defined as a reduction of BKV-DNA levels below the threshold of 1000 copies/ml plasma in qPCR, was achieved in 18 (78%) patients. The median duration until BKV-clearance was 118 [76-293] days (*Figure 1*). No association between time since transplantation, cidofovir dose, tacrolimus-free immunosuppression or highest viral replication rate with time to clearance of BK-viremia could be identified (p > 0.05). However, those allograft recipients achieving clearance of BK-viremia had significantly lower viral replication rates (38 000 [16 500-87 000] copies/ml plasma) at diagnosis compared to recipients not achieving clearance (300 000 [150 000-870 000] copies/ml plasma) (p = 0.03). The course of BK-viremia after diagnosis of PVAN is displayed in *Figure 2* (patients with quantitative PCR (n=20) only).

Interestingly, when comparing the eGFR at initiation of therapy, the patients achieving viral clearance had worse allograft function than those not achieving viral clearance (28.2 [23.8-35.7] ml/min/1.73m² vs. 41.8 [32.5-45.4], p = 0.087).

Within the median follow-up period of 2.24 [1.55-5.01] yrs, 14 (61%) kidney transplant recipients stabilized allograft function, with a median eGFR at clearance of BKV-replication of 25.1 [19.0 - 26.3] ml/min/1.73m². The remaining nine (39%) kidney transplant recipients developed allograft failure, but only three (13%) patients lost their allograft function due to the PVAN, as shown in *Figure 3*. From diagnosis of PVAN, time to allograft loss due to PVAN was 3.6 [3.5-13.6] months, time to allograft loss due to other causes 50.0 [27.0-67.4] months. Overall allograft survival in the group with loss due to PVAN was 12.0 [IQR 11.7-18.3] months, whereas overall allograft survival with loss due to other causes was 74.2 [IQR 34.9-109.7] months. Other causes of allograft loss included cardiovascular complications with subsequent cardiorenal syndrome (n = 1), recurrence of underlying glomerulonephritis (n = 1), allograft rejection (n = 1) and death with functioning graft (n = 1), whereas in 2 patients the reason for allograft loss was unknown.

Discussion

When discussing therapeutic options in polyoma virus-associated nephropathy, three treatment goals have to be taken into consideration: 1) clearance of viremia, 2) preservation of allograft function and 3) prevention of rejection episodes. Our data show that the dual therapeutic approach to PVAN with low-dose cidofovir and conversion to an mTOR based immunosuppression is feasible, even in patients with low allograft function. Furthermore, our approach allows for clearance of viremia and preservation of allograft function of patients and carries a low risk of rejection compared to the standard of care reduction of the overall immunosuppression.

Although in-vitro data and small clinical trials have shown effectiveness of low-dose cidofovir against BKV-replication, inconsistent doses as well as frequencies of application have been used in these trials ^{8,9,11}. Following our protocol, application of a single dose of 0.25 mg/kg bodyweight cidofovir was safe without any side effects or signs of nephrotoxicity, even in allograft recipients with a higher degree of renal functional impairment. The higher single and cumulative cidofovir dose used in the above mentioned studies in Belgium and Taiwan, did not result in any nephrotoxicity, but had led to skin rashes and three patients developed severe anterior uveitis after 6-8 applications of cidofovir and permanent visual impairment in two patients ¹¹. One might argue that our dose was too low to achieve therapeutic plasma concentrations. Measurements of peak concentrations after administration of 0.5-1.0 mg/kg bodyweight cidofovir in another study have shown that the peak concentration only reaches one-tenth of the in vitro 50% effective concentration (EC 50) against BK-virus and even only onetwentieth of the corresponding 50% inhibitory concentration (IC50) 9,17. However, the plasma concentration does not reflect to the intratubular concentration of cidofovir. A study on new Zealand white rabbits could show that intrarenal concentrations of radioactive-labeled cidofovir were about 10 times higher than in plasma ¹⁸. Therefore, we do not think measurement of plasma levels is helpful, but - in our opinion - achieving sufficient intratubular cidofovir concentrations accomplished by omission of probenecid is essential for the antiviral efficacy of cidofovir. In fact, 57% of our patients achieved clearance of BK-viremia after a single cidofovir application. Our finding, that patients with higher eGFR at diagnosis had a lower probability of clearance of BK-viremia has led to the notion, that in patients with higher eGFR, repetitive cidofovir applications may be

beneficial. We can only speculate that, having a higher eGFR, more cidofovir is lost in urine due to better glomerular filtration and saturated tubular uptake.

Even though studies have consistently shown lower rates of BKV-infection following an mTOR-based immunosuppression ^{2,12,13,19}, results on the course of PVAN following conversion to an mTOR-based immunosuppression in manifest disease are lacking. Invitro data have shown a reduction in BKV-replication with mTOR inhibition, which is explained by dependency of early BKV-replication on the mTOR pathway ¹⁴, as well as by an improved immune response through regulation of the differentiation of memory CD8⁺ T-cells ¹⁶. Interestingly, the inhibitory effect of mTOR inhibition on BKV-replication seems to be abolished after adding tacrolimus. The mechanistic background has been explained by Hirsch et al., with FK binding protein 12kda (FKBP-12) being the pivotal protein. Tacrolimus and sirolimus exert opposite effects on FKBP-12 and thereby on BKV-replication in renal tubular epithelial cells with tacrolimus even promoting BKVreplication ¹⁴. Therefore, if possible, a tacrolimus-free immunosuppressive regimen should be preferred. Interestingly, this effect is not present with cyclosporine ¹⁴. In our study, in 18 out of 23 patients, tacrolimus was replaced with sirolimus and in one patient with everolimus which facilitated clearance of viremia. No significant correlation between tacrolimus-free immunosuppression and probability BK-virus clearance could be identified, but two of the three patients with allograft loss due to PVAN were on ongoing concomitant tacrolimus, pointing towards an advantage of a tacrolimus-free, mTORbased immunosuppression in PVAN.

In our cohort, clearance of BK-viremia was achieved in 18 (78%) patients after a median time until clearance of BK-viremia of 118 [76-293] days. In a Canadian study cohort, the time until clearance with reduction of immunosuppression alone and in part addition of leflunomide was markedly longer with 266 [116-398] days ²⁰. A prospective, randomized controlled trial on 40 patients with BK-viremia or -viruria but without PVAN compared MMF dose reduction to conversion to everolimus and showed that after 3 months, only 50% of the patients treated with everolimus and 33% of the patients with a reduction in MMF achieved clearance of viremia ²¹. Taken together, our approach with the combination of low-dose cidofovir and mTOR-based immunosuppression results in better viral clearance than conventional approaches. Regarding allograft survival, only three

(13%) patients lost their allograft function due to PVAN, whereas 14 (61%) patients stabilized allograft function within the median follow-up of 2.24 years. Other studies on PVAN have presented huge differences in allograft loss rates but altogether higher percentages ranging from 11-71% ²²⁻²⁶. The reason for the large differences in allograft survival rates is most likely the differences in allograft function at time of diagnosis and in severity of PVAN. A recent study in 105 patients with BK-viremia has shown a higher allograft survival rate of 84% with reduction of immunosuppression alone ²⁷, yet large differences of the study population have to be taken in consideration: First, only 5% of these patients had biopsy proven PVAN, showing that this study population might have a lower degree of injury caused by polyomavirus. Second, the allograft function of these patients at baseline was markedly better with a median eGFR MDRD of 46-51 ml/min compared to our baseline eGFR MDRD of 30.1 ml/min.

Our data do have limitations. We present retrospective data, albeit from an ongoing protocol, strictly and consistently followed in all patients. Patients underwent two simultaneous interventions, precluding a distinction whether antiviral therapy of conversion to mTOR-based immunosuppression plays the leading role. Nonetheless, we do believe that the combination of these two strategies with the intratubular, antiviral effects of cidofovir together with the beneficial effects on the antiviral immune response allowed by mTOR-inhibition is key. The strength of our study is the standardized approach to patients with biopsy proven PVAN as well as the long follow-up time.

In conclusion, we demonstrate that our novel dual therapeutic approach to PVAN with application of low-dose cidofovir and conversion to an mTOR based immunosuppression allows for clearance of BK-viremia and preservation of allograft function in a high proportion of patients. Randomized, prospective studies on this issue are highly warranted.

Conflict of Interest Statement

The authors declare that there is no conflict of interest associated with this manuscript.

Authors' Contributions

T.M.: designed the work, acquired data, interpreted results, drafted manuscript, approved the final version

R.B.: acquired data, revised manuscript, approved the final version

S.N.: designed the work, revised manuscript, approved the final version

N.H.: designed the work, revised manuscript, approved the final version

M.G.: designed the work, acquired data, interpreted results, provided intellectual content of critical importance, revised manuscript, approved the final version

References

4.

5.

6.

8.

- Hirsch HH, Knowles W, Dickenmann M, et al. Prospective study of polyomavirus type BK replication and nephropathy in renal-transplant recipients. *N Engl J Med.* 2002;347(7):488-496.
- Dharnidharka VR, Cherikh WS, Abbott KC. An OPTN analysis of national registry data on treatment of BK virus allograft nephropathy in the United States. *Transplantation.* 2009;87(7):1019-1026.
- 3. Randhawa PS, Finkelstein S, Scantlebury V, et al. Human polyoma virusassociated interstitial nephritis in the allograft kidney. *Transplantation*. 1999;67(1):103-109.
 - Vasudev B, Hariharan S, Hussain SA, Zhu YR, Bresnahan BA, Cohen EP. BK virus nephritis: risk factors, timing, and outcome in renal transplant recipients. *Kidney Int.* 2005;68(4):1834-1839.
 - Bernhoff E, Tylden GD, Kjerpeseth LJ, Gutteberg TJ, Hirsch HH, Rinaldo CH. Leflunomide inhibition of BK virus replication in renal tubular epithelial cells. *J Virol.* 2010;84(4):2150-2156.
 - Sharma BN, Li R, Bernhoff E, Gutteberg TJ, Rinaldo CH. Fluoroquinolones inhibit human polyomavirus BK (BKV) replication in primary human kidney cells. *Antiviral Res.* 2011;92(1):115-123.
 - Randhawa PS, Schonder K, Shapiro R, Farasati N, Huang Y. Polyomavirus BK neutralizing activity in human immunoglobulin preparations. *Transplantation*. 2010;89(12):1462-1465.
 - Andrei G, Snoeck R, Vandeputte M, De Clercq E. Activities of various compounds against murine and primate polyomaviruses. *Antimicrob Agents Chemother.* 1997;41(3):587-593.
 - Kuypers DR, Vandooren AK, Lerut E, et al. Adjuvant low-dose cidofovir therapy for BK polyomavirus interstitial nephritis in renal transplant recipients. *Am J Transplant.* 2005;5(8):1997-2004.
- Wu SW, Chang HR, Lian JD. The effect of low-dose cidofovir on the long-term outcome of polyomavirus-associated nephropathy in renal transplant recipients. *Nephrol Dial Transplant.* 2009;24(3):1034-1038.

- Kuypers DR, Bammens B, Claes K, Evenepoel P, Lerut E, Vanrenterghem Y. A single-centre study of adjuvant cidofovir therapy for BK virus interstitial nephritis (BKVIN) in renal allograft recipients. *J Antimicrob Chemother.* 2009;63(2):417-419.
- Tedesco Silva H, Jr., Cibrik D, Johnston T, et al. Everolimus plus reducedexposure CsA versus mycophenolic acid plus standard-exposure CsA in renaltransplant recipients. *Am J Transplant.* 2010;10(6):1401-1413.
- Moscarelli L, Caroti L, Antognoli G, et al. Everolimus leads to a lower risk of BKV viremia than mycophenolic acid in de novo renal transplantation patients: a single-center experience. *Clin Transplant.* 2013;27(4):546-554.
- Hirsch HH, Yakhontova K, Lu M, Manzetti J. BK Polyomavirus Replication in Renal Tubular Epithelial Cells Is Inhibited by Sirolimus, but Activated by Tacrolimus Through a Pathway Involving FKBP-12. *Am J Transplant.* 2016;16(3):821-832.
- Jouve T, Rostaing L, Malvezzi P. Place of mTOR inhibitors in management of BKV
 infection after kidney transplantation. *J Nephropathol.* 2016;5(1):1-7.
- Araki K, Turner AP, Shaffer VO, et al. mTOR regulates memory CD8 T-cell differentiation. *Nature*. 2009;460(7251):108-112.
- Farasati NA, Shapiro R, Vats A, Randhawa P. Effect of leflunomide and cidofovir on replication of BK virus in an in vitro culture system. *Transplantation*. 2005;79(1):116-118.
- 18. Cundy KC, Li ZH, Lee WA. Effect of probenecid on the distribution, metabolism, and excretion of cidofovir in rabbits. *Drug Metab Dispos.* 1996;24(3):315-321.
- 19. Tedesco-Silva H, Pascual J, Viklicky O, et al. Safety of Everolimus With Reduced Calcineurin Inhibitor Exposure in De Novo Kidney Transplants: An Analysis From the Randomized TRANSFORM Study. *Transplantation*. 2019.
- Simard-Meilleur MC, Bodson-Clermont P, St-Louis G, et al. Stabilization of renal function after the first year of follow-up in kidney transplant recipients treated for significant BK polyomavirus infection or BK polyomavirus-associated nephropathy. *Transpl Infect Dis.* 2017;19(3).
- Wojciechowski D, Chandran S, Webber A, Hirose R, Vincenti F. Mycophenolate Mofetil Withdrawal With Conversion to Everolimus to Treat BK Virus Infection in Kidney Transplant Recipients. *Transplant Proc.* 2017;49(8):1773-1778.

- Trofe J, Gaber LW, Stratta RJ, et al. Polyomavirus in kidney and kidney-pancreas
 transplant recipients. *Transpl Infect Dis.* 2003;5(1):21-28.
- Mengel M, Marwedel M, Radermacher J, et al. Incidence of polyomavirusnephropathy in renal allografts: influence of modern immunosuppressive drugs. *Nephrol Dial Transplant.* 2003;18(6):1190-1196.
- 24. Ramos E, Drachenberg CB, Papadimitriou JC, et al. Clinical course of polyoma virus nephropathy in 67 renal transplant patients. *J Am Soc Nephrol.* 2002;13(8):2145-2151.
- 25. Dall A, Hariharan S. BK virus nephritis after renal transplantation. *Clin J Am Soc Nephrol.* 2008;3 Suppl 2:S68-75.
- 26. Li RM, Mannon RB, Kleiner D, et al. BK virus and SV40 co-infection in polyomavirus nephropathy. *Transplantation.* 2002;74(11):1497-1504.
- 27. Bischof N, Hirsch HH, Wehmeier C, et al. Reducing calcineurin inhibitor first for treating BK polyomavirus replication after kidney transplantation: long-term outcomes. *Nephrol Dial Transplant.* 2018.

Table 1: Tübingen Cidofovir Protocol

| i.v. hydration until urine output > 100 ml/h | calculated tubular passage time for reduced toxicity |
|--|--|
| cidofovir, 0.25 mg/kg i.v. over 6 h | low-dose concept for limited side effects |
| no application of probenecid | allows for tubular uptake (site of viral replication) of cidofovir via hOAT1 |
| single dose | virus clearance takes several weeks |

Table 2: Patient characteristics

| | | n = 23 |
|--------------------|---|----------------------|
| gende | er, f/m (n) | 4 / 19 |
| decea | sed/living kidney donation (n) | 13/10 |
| age at | t diagnosis (yrs) | 54 [40-59] |
| time to | o diagnosis (days) | 268 [153-869] |
| eGFR | ^a MDRD at diagnosis | 30.1 [24.7-38.2] |
| BKV ^b a | at diagnosis (copies/ml plasma) | 65500 [25500-200250] |
| | | |
| mainte | enance immunosuppression at diagnosis, (n | |
| - | Tac ^b /MMF ^d /Pred ^e | 16 |
| | Tac ^b /MMF ^d | 7 |

Data are given as median [interquartile range]

^aeGFR: estimated glomerular filtration rate, ^bBKV: BK-viremia, ^cTac: tacrolimus, ^dMMF: mycophenolate mofetil, ^ePred: prednisone

| | n = 23 |
|--|------------------|
| cidofovir application (n) | |
| - once | 13 |
| - two times | 7 |
| - three times | 3 |
| cidofovir cumulative dose (mg) | |
| - single application | 18.8 [15.5-20] |
| - double application | 30.0 [29.5-33.6] |
| - triple application | 56.9 [54.5-61] |
| - cumulative dose all patients | 25.0 [18.6-30.6] |
| interval between cidofovir applications (days) | 58 [44-109] |
| change in immunosuppression (n) | |
| - tacrolimus to sirolimus | 18 |
| - MMF ^d to leflunomide and sirolimus | 1 |
| - MMF ^d to everolimus | 1 |
| - MMF ^d to sirolimus | 1 |
| - tacrolimus to everolimus | 1 |
| - azathioprine to everolimus | 1 |
| discontinuation of mTOR inhibitor (n) | 5 |
| - due to aphtous lesions | 2 |
| - due to cellular rejection | 1 |
| - due to DSA ^a and refractory PVAN ^e | 1 |
| - gastrointestinal side-effects | 1 |
| | _ |

Table 3: Therapeutic interventions

| time until discontinuation of mTOR-inhibitor (days) | 112 [60-211] |
|--|------------------|
| eGFR ^b MDRD ^c at cidofovir application | 28.8 [24.3-36.0] |
| eGFR ^b MDRD ^c 30 days after cidofovir | 24.4 [18.8-30.0] |
| eGFR ^b MDRD ^c 90 days after cidofovir | 25.8 [19.4-36.5] |
| eGFR ^b MDRD ^c at BKV-clearance | 25.1 [19.0-26.3] |

Data are given as median [interquartile range]

^aDSA: donor specific antibodies, ^beGFR: estimated glomerular filtration rate, ^cMDRD: model of end stage renal disease, ^dMMF: mycophenolate mofetil, ^ePVAN: polyomavirus-associated nephropathy

Figure legends

Figure 1: Cumulative incidence of BKV-clearance

Figure 2: Time course of BK-virema after diagnosis of PVAN. Solid line displays the group not achieving BKV-clearance, dotted line displays the group with clearance of BK-viremia (< 1000 copies/ml plasma)

Figure 3: Kaplan Meier estimate of allograft survival; solid line displays allograft loss due to all causes, dashed line displays allograft loss due to PVAN only (censored for other causes). Downward marks indicate allograft losses, upward marks indicate end of follow up for individual patients



tid_13228_f1.tif



tid_13228_f2.jpg





tid_13228_f3.jpg