

Evaluation of a pixelated large format CMOS sensor for x-ray microbeam radiotherapy

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Purpose: Current techniques and procedures for dosimetry in microbeams typically rely on radiochromic film or small volume ionisation chambers for validation and quality assurance in 2D and 1D, respectively. Whilst well characterised for clinical and preclinical radiotherapy, these methods are non-instantaneous and do not provide real time profile information. The objective of this work is to determine the suitability of the newly developed vM1212 detector, a pixelated CMOS (complementary metal-oxide-semiconductor) imaging sensor, for *in situ* and *in vivo* verification of x-ray microbeams.

Methods: Experiments were carried out on the vM1212 detector using a 220 kVp small animal radiation research platform (SARRP) at the Helmholtz Centre Munich. A 3 x 3 cm² square piece of EBT3 film was placed on top of a marked non-fibrous card overlaying the sensitive silicon of the sensor. 1 cm of water equivalent bolus material was placed on top of the film for build-up. The response of the detector was compared to an Epson Expression 10000XL flatbed scanner using FilmQA Pro with triple channel dosimetry. This was also compared to a separate exposure using 450 µm of silicon as a surrogate for the detector and a Zeiss Axio Imager 2 microscope using an optical microscopy method of dosimetry. Microbeam collimator slits with range of nominal widths of 25, 50, 75 and 100 µm were used to compare

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beam profiles and determine sensitivity of the detector and both film measurements to different microbeams.

Results: The detector was able to measure peak and valley profiles in real-time, a significant reduction from the 24 hour self-development required by the EBT3 film. Observed full width at half maximum (FWHM) values were larger than the nominal slit widths, ranging from 130 - 190 μm due to divergence. Agreement between the methods was found for peak-to-valley dose ratio (PVDR), peak to peak separation and FWHM, but a difference in relative intensity of the microbeams was observed between the detectors.

Conclusions: The investigation demonstrated that pixelated CMOS sensors could be applied to microbeam radiotherapy for real-time dosimetry in the future, however the relatively large pixel pitch of the vM1212 detector limit the immediate application of the results.

Key words: microbeam radiation therapy, compact microbeam sources, dosimetry, CMOS detectors

1 1. INTRODUCTION

2 1A. Microbeam Radiotherapy

3 Microbeam radiotherapy (MRT) is a novel type of spatially fractionated therapy which is defined
4 by narrow beams of radiation (typically $< 100 \mu\text{m}$) that can selectively irradiate portions of the target
5 volume [1]. To cover the entire target volume, microbeams are delivered in a grid pattern in which
6 multiple quasi-parallel rectangular beams, with typical centre-to-centre distances of 200 - 400 μm .
7 Crucially the entire target volume is not irradiated uniformly, with regions of very high dose microbeam
8 "peaks" separated by very low dose valleys.

9 Preclinical studies have indicated that this dose pattern has a greater efficacy than that of a single
10 uniform field [2]. Whilst the exact mechanism for preferential effect tumour is not fully understood and is
11 likely a combination of effects. Possible mechanisms under investigation are preferential damage to
12 vascular tissue in tumours[3–5], and radiation-induced bystander and abscopal effects [6,7].

13 1B. Current Verification Methods

14 The very small size and high dose gradients of microbeams present a significant challenge to most
15 standard detectors. That combined with high dose rates at synchrotrons adds to the complexity when
16 working towards accurate dosimetry for microbeam radiotherapy.

17 Stereotactic radiotherapy treatments (with radiation fields sizes typically between 0.4 - 30 mm[8])
18 have strict requirements on the geometrical and dosimetric accuracy from dose calculations to delivery of

19 $\pm 5\%$ ($k=2$)[9]. Microbeam irradiations are a step forward in terms of complexity and at present there is no
20 dosimetry protocol or recommendations for dosimetry of irradiations with such beam configurations[10].

21 Much of the ongoing research in the community is dedicated to optimising irradiation
22 configurations in order to obtain the best therapeutic outcomes, with peak to peak distance[11], full-
23 width at half maximum (FWHM)[12] and the peak-to-valley dose ratio (PVDR)[13,14] being of particular
24 interest.

25 Due to the very small scales involved in microbeam radiotherapy, conventional radiotherapy
26 equipment for beam profile acquisition (like small volume ionisation chambers) are unable to resolve the
27 individual microbeam peaks [9]). Scanning other types of small volume detectors through a microbeam
28 peak has been previously performed with success by using a MOSFET dosimeter [15,16] or with a
29 commercial PTW (Physikalisch-Technische Werkstätten GmbH, Freiburg, Germany) microdiamond
30 detector[17], with resolutions of $1\ \mu\text{m}$ [18]. This method has shown good agreement with radiochromic
31 film[19], however the measurements are acquired point by point and therefore the shape of the profiles
32 are not shown instantaneously which limits its use for *in vivo* dosimetry or *in situ* verification. The same
33 applies to the use of scintillating fibres, as shown by Archer *et al.*[20].

34 Various groups have developed silicon strip detectors capable of quantifying parameters of the
35 microbeam field [21–23]. Whilst hybrid strip detectors (with separate sensor and readout) can offer
36 greater resistance to radiation than monolithic pixelated detectors, strip detectors do not provide
37 detailed information about the 2D profile of the radiation field and, therefore, will be more sensitive to
38 angular misalignment.

39 A method of obtaining 2D relative dose distributions of microbeams was developed by Bartzsch *et*
40 *al.* [24] using optical microscopy and EBT3 films [25], which when using a microscope is technically
41 capable of spatial resolutions better than $1\ \mu\text{m}$. Due to film grain inhomogeneities this is reduced to $5\ \mu\text{m}$
42 in practise. This method builds on existing techniques for film dosimetry. Radiochromic films have a
43 relatively large dose range (0.5 to 20 Gy for EBT3[26]), however the analysis process is slow, requiring a
44 minimum of 24 hours for self-development post-irradiation [27]. At lower dose levels (less than 0.1
45 Gy[28,29]) noise becomes more significant. This typically necessitates two separate sets of irradiations for
46 the same set of microbeams, in order to be able to increase the accuracy of the assessment of the dose
47 distribution in the regions with lower dose range (valleys) without saturating the high dose region of the
48 microbeam peaks.

49 This investigation was carried out to evaluate the suitability of the newly developed vM1212 detector
50 for its use in the analysis of preclinical radiotherapy microbeams, using the custom built multi-slit
51 collimator at the Helmholtz Zentrum München, Germany. The objective was to quantify microbeam

52 parameters and to compare to results of the analysis of the same deliveries to EBT3 films, using the
53 optical microscopy method[24].

54 2. MATERIALS AND METHODS

55 2A. vM1212 Pixelated Detector

56 The vM1212 pixelated detector is a large format CMOS (complementary metal-oxide-
57 semiconductor) imaging sensor with 50 μm pixel pitch originally designed for medical and scientific x-ray
58 imaging by the CMOS Sensor Design Group at the Rutherford Appleton Laboratory [30] and is now
59 licenced and manufactured into a full detector assembly by vivaMOS Ltd. The active area of the vM1212
60 detector is approximately 6 x 6 cm^2 (1204 x 1248 pixels) and is sufficiently large to capture the entire
61 radiation field of the microbeam multislit collimator in a single instance.

62 The small pixel pitch and predicted resistance to damage caused by high levels of ionising
63 radiation justified a proof of principle investigation to determine the response of the detector to
64 microbeam radiation.

65 2B. Methodology

66 A SARRP (Small Animal Radiation Research Platform) x-ray irradiator at the Helmholtz Zentrum
67 München was used for this investigation. The irradiation parameters were set to 220 kVp (0.67 mm Cu
68 HVL); 2.8 mA; and fine focus (effective beam source size of 0.4 mm[31]).

69 The tungsten microbeam multislit collimator consisted of three levels of fifty one 100- μm slits (7 mm
70 total thickness), with a slit-to-slit separation of 400 μm . The first and third level are in a fixed alignment,
71 whilst the second central level is controlled by two motorised translation stages. When fully open, the
72 transmission gap is 100 μm , but the finest step resolution of the piezoelectric pistons enables variable slit
73 widths between 0 - 100 μm to be investigated to an accuracy of 0.5 μm . The collimator was mounted at a
74 distance of 21.2 cm from the source, with additional lead shielding to prevent radiation damage to the
75 electronics as shown in Figure 1a.

76 In order to obtain robust and safe positioning, the vM1212 detector had to be mounted at a
77 source to surface distance (SSD) of 29 cm, 6.8 cm from the surface of the microbeam collimator. To
78 achieve conditions similar to the ones used for small animal irradiations a 1 cm slab of tissue-equivalent
79 flexible bolus material with density of 1.03 g/cm^3 [32] (trimmed to 7 x 7 cm^2) was placed on top of the
80 EBT3 film. The vM1212 detector was used without scintillating material to maximise the potential spatial
81 resolution. To enable a direct comparison between the EBT3 film and the vM1212 detector, EBT3 film

82 pieces were placed on top of the active area of the sensor, separated by a thin layer of a non-fibrous card
83 which had been marked for repeatable alignment (Figure 1b).

84 The EBT3 films irradiated simultaneously to the vM1212 detector were scanned using an Epson
85 Expression 10000XL flatbed scanner (1400 dpi) and calibrated using FilmQA Pro with triple channel
86 dosimetry[33,34]. Due to time constraints during the experiment, it was not possible to irradiate a second
87 set of films for their analysis with optical microscopy. Those irradiations were performed in an
88 independent experiment following the same irradiation conditions: source-surface distance, same bolus
89 material and non-fibrous card, but using 450 μm of silicon simulating the thickness of the detector. This
90 second set of films was scanned using a ZEISS Axio Imager 2 optical microscope[35] on 5X magnification
91 for a pixel resolution of 1.29 μm .

92 Prior to the film irradiations, the output (Gy/min) was measured in reference conditions for
93 SARRP absolute calibration. Measurements were performed with the SARRP open field at Source Surface
94 Distance (SSD) of 33 cm and at 2 cm depth in WT1 water equivalent slab phantom, with 3 cm of
95 backscatter material. Two independent measurements of the SARRP output were performed, one with
96 the local dosimetry system (PTW 30010 ionisation chamber), traceable to the PTW-Freiburg SSDL
97 Calibration Laboratory and with a National Physical Laboratory (NPL) secondary standard system (PTW
98 30012 ionisation chamber), traceable to the NPL primary standard for medium energy x-rays. Both
99 ionisation chambers used a local PTW Unidos TW1001 electrometer for dosimetry. Following output
100 measurements and in order to obtain a calibration curve, a set of nine films were irradiated in the same
101 reference conditions, with doses ranging from 0 to 14 Gy.

102 For consistency throughout the investigation, the same integration time, 28 ms, was always used
103 on the vM1212 detector. This ensured that all the performed measurements were all in the linear
104 response region for the pixels and prevented saturation of the detector. The results obtained using the
105 vM1212 detector were corrected by averaging over a number of frames to reduce noise, subtracting a
106 dark image to account for dark current in the pixels and calibrating the pixel response values against
107 measurements with the NPL ionisation chamber under the same conditions.

108 Direct comparison between the EBT3 films and the different acquisitions with vM1212 detector
109 were carried out for 25, 50, 75 and 100 μm slit widths. All the slits were irradiated with 240 s of exposure
110 with the exception of the 25 μm slit width which was irradiated with 480 s, to increase the dose and
111 therefore to reduce the level of noise for the films measurements in such narrow beams.

112 Finally, to understand the difference in spatial response between the vM1212 detector and the
113 two methods of EBT3 film scanning, the modulation transfer function (MTF) was measured for each. The
114 modulation transfer function of the vM1212 detector was measured following BS EN 62220-1-3:2008 [36]

115 and using the COQ analysis software written by Donini *et al.* [37]. The MTF of the Epson Expression
116 10000XL scanner at 1400 dpi scanning resolution was measured using a sharp flat edge positioned over a
117 piece of unexposed EBT3 film at an angle of 4°. Again using the COQ analysis software, the edge spread
118 function was calculated allowing the modulation transfer function to be determined. The MTF of the Zeiss
119 Axio Imager 2 was measured with the Xradia resolution sample (provided by Zeiss), which contained a
120 pattern of lines with known width and separation. The largest line width on this pattern was 32 μm
121 (period = 64 μm), and as such the smallest resolution measurable with this resolution sample was 15.6
122 line pairs/mm (1/0.064 mm).

123 3. RESULTS

124 3A. Profile measurements

125 It was found that the vM1212 detector was able to capture the entire radiation field as defined by
126 the collimator, as can be seen in Figure 2b. To create the microbeam collimator slits in tungsten, 0.3 mm
127 diameter holes had to be drilled into the tungsten, allowing for wire erosion to mill out the 100 μm wide
128 slits. This detail can be recognized on both detectors (film and vM1212 detector) and was used for
129 alignment purposes. All profile comparisons presented are aligned relative to the central 26th peak. By
130 comparing vertical profiles from the EBT3 film methods with vertical profiles taken using the vM1212
131 detector we were able to observe that the alternating pattern of peaks and valleys of the microbeam
132 collimator are well correlated between the different detectors. The larger SSD required to mount the
133 vM1212 detector and the maximum scanning size of the EBT3 film possible with the microscope reduced
134 the number of peaks that could be recorded using this method to approximately 40 (reduced from 51
135 physical slits on the collimator).

136 The 100 μm slit profiles' comparison can be seen in Figure 3a, where an agreement in terms of
137 alignment of the peaks between the three detector methods can be observed. The vM1212 detector and
138 the Epson Expression 10000XL under respond in terms of peak dose by approximately 30% however there
139 is relatively good agreement of the location of the microbeam peak centre values (Figure 3b). As shown in
140 Figure 4, relative to the Zeiss Axio Imager 2, the valley doses are over reported by the Epson Expression
141 10000XL (with scanning resolution at 1400 dpi) by approximately 25% (15 mGy/min), whilst the vM1212
142 detector over reports by less than 5% (5 mGy/min). The average deviation between corresponding peak
143 centres for the vM1212 detector and the Epson Expression 10000XL measurement was 18.5 μm , whilst for
144 the Zeiss Axio Imager 2 measurement was found to be 55.3 μm . As shown in Figure 5 for the 26th central
145 peak, the profile resolved on all three detector methods appears to be Gaussian.

146 For the 25 μm slit width comparison (Figure 6) the agreement between the EBT3 films and the
147 vM1212 detector becomes worse as there is a strong disagreement for dose rate values between the scan
148 performed by the Zeiss Axio Imager 2 and the other methods. This deviation is likely due to spatial
149 averaging within the vM1212 detector and the Epson Expression 10000XL, however it is also possible that
150 this deviation was introduced by misalignment during the Zeiss Axio Imager 2 exposure as it was
151 performed at a later date. The lower measured dose rate is not consistent across the microbeam profiles
152 as shown for the central peak (Figure 8), where the dose rate measured by the vM1212 detector and
153 Epson Expression 10000XL EBT3 film is approximately 20% of the dose rate measured by the Zeiss Axio
154 Imager 2. For the Epson Expression 10000XL and the vM1212, the dose rate measured for the 27th peak
155 (Figure 9) is better but still measures only 40% relative to the Zeiss Axio Imager 2. Valley profiles for the
156 25 μm slit measured all of the detectors are again inconsistent, with approximate differences relative to
157 the Zeiss Axio Imager 2 of 40% and 20% for the vM1212 detector and Epson Expression 10000XL,
158 respectively. This peak specific under response not observed in the Zeiss Axio Imager 2 measurement is
159 suspected to be due to a combination of manufacturing tolerances on the machined microbeam slits and
160 repeatability issues of the microbeam setup.

161 Figures 8 and 9 show a profile comparisons with a Gaussian fit applied between the three
162 detectors for the 26th (central) and 27th peak, respectively. A stitching artefact between the high dose
163 valley irradiation and the low dose peak measurement can be seen in Figure 9 in the Zeiss Axio Imager
164 2 dose profile at approximately 50 μm . The centres of the 27th microbeam peak (relative to the 26th
165 central peak) can be calculated to be 550, 514 and 488 μm for the vM1212 detector, Epson Expression
166 10000XL and Zeiss Axio Imager 2 respectively.

167 The peak to peak separation could be measured across the three detection methods for all
168 measured slit widths, as shown in Table 1. It can be shown that the three methods agree within the
169 uncertainties calculated. Using the inverse square law and the differences between the measured peak to
170 peak separations, it can be estimated that the EBT3 films for the Epson Expression 10000XL and Zeiss Axio
171 Imager 2 measurements were positioned 0.5 ± 0.2 mm and 2.4 ± 0.2 mm closer respectively to the x-ray
172 source than the vM1212 detector measurement. As the measurements for the Epson Expression 10000XL
173 were taken concurrently with the vM1212 detector, this difference can be attributed to the thickness the
174 non-fibrous card which was independently measured with a digital calliper to be 0.53 ± 0.01 μm . The 2.4
175 mm deviation of the Zeiss Axio Imager 2 measurement is likely due to setup misalignment.

176 It was also found that the vM1212 detector was still able to detect and identify each of the 51
177 peaks when the microbeam collimator is fully closed (set to 0 μm slit width) (Figure 10). Profiles resulting
178 from this leakage are used in sections 3B FWHM measurements and 3C Peak and Valley Measurements.

179 Using the vM1212 detector it is possible to take real time horizontal profiles of the microbeam
180 collimator. A comparison between the methods averaged across all recorded peaks for the 100 μm slit
181 width can be seen in Figure 11, which again shows the approximately 30% under response of the vM1212
182 detector and Epson Expression 10000XL measurements relative to the Zeiss Axio Imager 2 measurement.
183 The sharp vertical peaks at 13,000 and 41,000 μm are due to the 0.3 mm diameter holes seen in Figure 2.
184 It can be seen in all three methods that the radiation intensity does not follow a smooth profile across the
185 collimator as one might expect, although it is beyond the scope of this paper to discuss any therapeutic
186 impact this may have.

187 3B. FWHM measurements

188 An averaged FWHM comparison between the Zeiss Axio Imager 2 and the vM1212 detector for
189 each of the slits can be seen in Figure 12. The error bars shown represent one standard deviation of
190 uncertainty for the microbeam peaks.

191 A linear relationship between the FWHMs is observed however there is a large deviation between
192 FWHMs within a measurement. This can be attributed to a significant trend in the FWHM as a function of
193 vertical position that was undetectable at the time of the experiment that can be seen in both the
194 vM1212 detector results (Figure 13) and the analysed EBT3 films (not shown). This is most probably due
195 to the angle of the beam after it is produced at the tungsten target, within the x-ray tube, known as heel
196 effect. This effect would have become more dominant due to the larger SSD and was not observed on
197 past measurements using the microbeam collimator.

198 Such a difference in beam FWHM across the beam profile would have had a significant impact on
199 patient outcome, as described by Serduc *et al.* [12]. For *in vivo* verification this would have been
200 impossible to diagnose in real time with EBT3 films, due to the minimum 24 hour time required for film
201 self-development. This highlights a potential application of the vM1212 detector for real time imaging of
202 microbeams.

203 A comparison of microbeam nominal slit width to the measured FWHM can be seen in Figure 14.
204 As the vM1212 detector could take multiple readings with minimal dead time between them, a repeat set
205 of measurements was performed to calculate the FWHM of the microbeams. Each time the slit width was
206 increased by 5 μm . Using this approach, it was possible to show that below 20 μm slit width, the value of
207 the measured FWHM begins to increase (in relation to the expected nominal one). This effect is well
208 documented for small fields in megavoltage x-ray beams[38] and is due to the finite size of the x-ray
209 source being partially occluded by the collimator, causing an overlapping beam penumbra. If this
210 geometrical penumbra is larger than the field size, then the FWHM of the resulting beam increases.

211 Differences between the two vM1212 detector measurements are attributed to subtle differences when
212 repositioning the detector and uncertainties in the reproducibility of the collimator movements, however
213 this effect appears to be minimal.

214 The larger FWHM for all measurements can be attributed to the finite size of the x-ray source. As
215 shown in Figure 15, for a finite source size (S), collimator slit width (w), source-collimator distance (A), and
216 collimator-projection distance (B); the projected beam width can be approximated using equation (1).

217
$$L = \left[\frac{B}{A} + 1 \right] w + \frac{BS}{A} \quad (1)$$

218 For this approximation and to simplify the scatter effects, we assumed that the collimator is infinitely thin
219 and consists of only one layer instead of the three that comprise the actual and previously described
220 design of the collimator. With the previous assumptions we are considering the calculated projected
221 beam size as an approximation of the FWHM of the microbeam peak. Using the values described
222 previously for A , B and S , the values for the theoretical resolvable slit size were plotted on Figure 14 for
223 comparison with measured results. With equation 1, the smallest microbeam peak FWHM created by the
224 collimator that could be possible to resolve would be equal to $128.3 \pm 13.0 \mu\text{m}$ (assuming 10% uncertainty
225 of x-ray source size), whilst using the extrapolated results from the vM1212 detector the minimum is
226 calculated to be $126.0 \pm 0.7 \mu\text{m}$. The differences in the slope between the derived (geometric
227 approximation) and measured (vM1212 detector repeat linear fit) FWHMs are likely to be due to the
228 numerous approximations and would need full Monte Carlo simulation with an accurate model of the
229 geometry and scatter conditions.

230 **3C. Peak and Valley Measurements**

231 By fitting Gaussians to each of the peaks in both the vM1212 detector and EBT3 film profiles, the
232 Peak-to-Valley Dose Ratio (PVDR) can be estimated and compared to results reported in the literature
233 (Figure 16). The values calculated for the PVDR were comparable to what one might expect for this
234 microbeam collimator when comparing to previous measurements in a similar collimator by Bartzsch *et*
235 *al.* (where 15.5 ± 1.5 was measured at 10 mm depth) [39], especially when considering the significantly
236 larger SSD of this investigation.

237 The PVDRs obtained using the Epson Expression 10000XL for the 25 and 50 μm slit widths were
238 found to be significantly larger than both predicted by literature and as reported by the vM1212 detector
239 and the Zeiss Axio Imager 2 measurements. This can be attributed to a significant under response of the
240 Epson Expression 10000XL to the microbeam valleys, as shown in Figure 7. It is possible that the two film
241 method used for optical microscopy could be applied to compensate for this and record a more accurate
242 dose profile, however this was not within the scope of the investigation.

243 Using the vM1212 detector it was possible to rapidly calculate the PVDR for a large number of slit
244 widths. As shown in Figure 10, radiation leak is present through the collimator at slit width 0 μm from
245 which a PVDR could be calculated. The decrease in PVDR below 20 μm is consistent with the increase in
246 FWHM as observed in Figure 14 which was attributed to an increased proportion of the radiation resulting
247 from scatter with decreasing slit width.

248 3D. Modulation Transfer Measurements

249 The results of modulation transfer measurements are shown in Figure 17. It can be shown that while
250 the spatial resolution of the vM1212 detector is better than the Epson Expression 10000XL scanner, the
251 Zeiss Axio Imager 2 microscope is superior to both.

252

253 4. DISCUSSION

254 In comparison to dedicated facilities such as the European Synchrotron Radiation Facility (ESRF), the
255 x-ray source used for this investigation was not optimised for microbeam radiotherapy with the dose rate
256 measured after the collimator to be less than 0.05 Gy/s. This is substantially less than the dose rate used
257 at synchrotrons for microbeam radiotherapy (often exceeding 100 Gy/s)[40]. The microbeam FWHMs
258 delivered in this investigation are significantly larger than the 25 μm wide beams capable at the ESRF and
259 as such, further research of the vM1212 detector under such beam conditions is required. The mean
260 energy of this investigation (approximately 95 keV as calculated by the x-ray emission spectra calculation
261 software SpekCalc[41–43]) was comparable to that of dedicated synchrotrons[44,45], however
262 undoubtedly the effect of the different spectra must be considered.

263 A comparison of the three microbeam detection methods evaluated in this work can be seen in Table
264 2. Whilst the vM1212 detector has demonstrated the feasibility of a CMOS sensor for microbeams
265 measurement in this investigation, significant deviations to established dosimetry methods were
266 observed and further studies comparing to Monte Carlo simulations for relative dosimetry are still
267 necessary. The Zeiss Axio Imager 2 remains a suitable readout method for commissioning and situations
268 where maximum precision is required however, this method is relatively young and validated protocols
269 and workflows need to be established to allow wider uptake for this method among microbeam
270 community. The use of the Epson Expression 10000XL for microbeam measurements is not recommended
271 due to the (relatively) poor spatial resolution.

272 The vM1212 detector does not possess the spatial resolution necessary for accurate microbeam
273 dosimetry with its relatively large 50 μm pixels, compared to other quality assurance mechanisms
274 discussed previously (such as the PTW microdiamond) with ~ 1 μm resolution. In addition, well

275 established characteristics of other detection methods necessary for routine quality assurance such as
276 dose rate and beam quality dependence have not been taken into account. The vM1212 detector
277 operates using a "rolling shutter" frame acquisition method which does not present an issue for static or
278 slow moving microbeam sources such as the type used in this investigation but may not be ideal for fast
279 scanned microbeam spots. Additionally the maximum full field refresh rate of 34 fps may cause temporal
280 blurring, however this effect could be minimised by binning pixels together or recording only a smaller
281 region of interest. This refresh rate is still considerably lower than that of commercial radiotherapy
282 electrometers (such as the Unidos webline with 1 kHz sampling rate[46]). Whilst the dose delivered to the
283 films scanned by the Epson Expression 10000XL is relatively low (average peak dose of > 1 Gy) for EBT3
284 film standards, it must be noted that the vM1212 detector is capable of obtaining similar or better quality
285 images in less than 2 mGy per frame, highlighting its potential for real-time microbeam verification.

286 Looking forward, CMOS sensors resistant to ionising radiation have been developed for other harsh
287 radiation environments (such as space), achieving pixel pitches of less than 10 μm [47,48] in size. The use
288 of such sensors in the future could obtain real-time microbeam profile information surpassing even that
289 of the Zeiss Axio Imager 2, however making these sensors large enough to cover the same field of view as
290 the vM1212 detector could become prohibitively expensive due the number of pixels required and sensor
291 yield losses.

292 5. CONCLUSION

293 Microbeam radiotherapy is a rapidly developing method of cancer treatment with significant
294 therapeutic improvements over conventional radiotherapy [50,51]. The dosimetric challenges associated
295 with the high dose gradients in microbeam radiotherapy prevent the use of well-established dosimetry
296 equipment used in radiotherapy and (to date), all novel techniques for monitoring microbeams have only
297 obtained one dimensional profile information; limiting their clinical viability.

298 In this study we have demonstrated the capacity of the two dimensional vM1212 pixelated detector
299 to discriminate individual microbeams peaks with FWHM between 130 and 190 μm . The high dynamic
300 range of the vM1212 detector allows the signal detection of both the high dose peaks and the low dose
301 valleys (of microbeams with less than 20 PVDR) to be measured in real-time, which provides a significant
302 advantage over EBT3 films requiring at least 24 hours post-irradiation processing. Observed peak-to-valley
303 dose ratios and peak to peak separations measured by the vM1212 detector were comparable those
304 obtained using the optical microscopy method employing Zeiss Axio Imager 2 microscope. The use of
305 pixelated sensors for in-vivo beam monitoring in conventional radiotherapy beams is already being

306 researched by multiple groups[52,53] and as the technology behind the sensors matures, it is anticipated
307 that future CMOS detectors will have all of the required characteristics for microbeam dosimetry.
308

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314

315 CONFLICT OF INTEREST DISCLOSURE

316 Due to the prototype nature of the device, the manufacturer of the vM1212 detector, vivaMOS
317 Ltd, has been involved in data collection providing advice and technical support throughout the
318 investigation.

319

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- 322 [1] Slatkin D N, Spanne P, Dilmanian F A and Sandborg M 1992 Microbeam radiation therapy *Med. Phys.* **19** 1395–400
- 323 [2] Bouchet A, Serduc R, Laissue J A and Djonov V 2015 Effects of microbeam radiation therapy on normal and tumoral
324 blood vessels *Phys. Medica* **31** 634–41
- 325 [3] Serduc R, Christen T, Laissue J, Farion R, Bouchet A, Sanden B van der, Segebarth C, Bräuer-Krisch E, Le Duc G, Bravin A,
326 Rémy C and Barbier E L 2008 Brain tumor vessel response to synchrotron microbeam radiation therapy: a short-term *in*
327 *vivo* study *Phys. Med. Biol.* **53** 3609–22
- 328 [4] Crosbie J C, Anderson R L, Rothkamm K, Restall C M, Cann L, Ruwanpura S, Meachem S, Yagi N, Svalbe I, Lewis R A,
329 Williams B R G and Rogers P A W 2010 Tumor Cell Response to Synchrotron Microbeam Radiation Therapy Differs
330 Markedly From Cells in Normal Tissues *Int. J. Radiat. Oncol.* **77** 886–94
- 331 [5] Bouchet A, Serduc R, Laissue J A and Djonov V 2015 Effects of microbeam radiation therapy on normal and tumoral
332 blood vessels *Phys. Medica* **31** 634–41
- 333 [6] Prise K M, Schettino G, Folkard M and Held K D 2005 New insights on cell death from radiation exposure *Lancet Oncol.* **6**
334 520–8
- 335 [7] Fernandez-Palomo C, Bräuer-Krisch E, Laissue J, Vukmirovic D, Blattmann H, Seymour C and Mothersill C 2015 Use of
336 synchrotron medical microbeam irradiation to investigate radiation-induced bystander and abscopal effects *in vivo* *Phys.*
337 *Medica* **31** 584–95
- 338 [8] Bagheri H, Soleimani A, Gharehaghaji N, Mesbahi A, Manouchehri F, Shekarchi B, Dormanesh B and Dadgar H 2017 An
339 overview on small-field dosimetry in photon beam radiotherapy: Developments and challenges *J. Cancer Res. Ther.* **13**

- 340 175
- 341 [9] Das I J, Downes M B, Kassaei A and Tochner Z 2000 Choice of Radiation Detector in Dosimetry of Stereotactic
342 Radiosurgery-Radiotherapy *J. Radiosurgery* **3** 177–86
- 343 [10] Bräuer-Krisch E, Serduc R, Siegbahn E A, Le Duc G, Prezado Y, Bravin A, Blattmann H and Laissue J A 2010 Effects of
344 pulsed, spatially fractionated, microscopic synchrotron X-ray beams on normal and tumoral brain tissue *Mutat. Res. -
345 Rev. Mutat. Res.* **704** 160–6
- 346 [11] Regnard P, Le Duc G, Bräuer-Krisch E, Troprès I, Siegbahn E A, Kusak A, Clair C, Bernard H, Dallery D, Laissue J A and
347 Bravin A 2008 Irradiation of intracerebral 9L gliosarcoma by a single array of microplanar x-ray beams from a
348 synchrotron: Balance between curing and sparing *Phys. Med. Biol.* **53** 861–78
- 349 [12] Serduc R, Bouchet A, Bräuer-Krisch E, Laissue J A, Spiga J, Sarun S, Bravin A, Fonta C, Renaud L, Boutonnat J, Siegbahn E
350 A, Estève F and Le Duc G 2009 Synchrotron microbeam radiation therapy for rat brain tumor palliation—influence of the
351 microbeam width at constant valley dose *Phys. Med. Biol.* **54** 6711–24
- 352 [13] Bräuer-Krisch E, Adam J-F, Alagoz E, Bartzsch S, Crosbie J, DeWagter C, Dipuglia A, Donzelli M, Doran S, Fournier P, Kalef-
353 Ezra J, Kock A, Lerch M, McErlean C, Oelfke U, Petasecca M, Povoli M, Rosenfeld A, Siegbahn E A, Sporea D and Stugu B
354 2015 Medical physics aspects of the synchrotron radiation therapies: Microbeam radiation therapy (MRT) and
355 synchrotron stereotactic radiotherapy (SSRT) *Phys. Medica* **31** 568–83
- 356 [14] Annabell N, Yagi N, Umetani K, Wong C and Geso M 2012 Evaluating the peak-to-valley dose ratio of synchrotron
357 microbeams using PRESAGE fluorescence *J. Synchrotron Radiat.*
- 358 [15] Rosenfeld A B, Kaplan G I, Kron T, Allen B J, Dilmanian A, Orion I, Ren B, Lerch M L F and Holmes-Siedle A 1999 MOSFET
359 dosimetry of an X-ray microbeam *IEEE Trans. Nucl. Sci.* **46** 1774–80
- 360 [16] Kaplan G I, Rosenfeld A B, Allen B J, Booth J T, Carolan M G and Holmes-Siedle A 2000 Improved spatial resolution by
361 MOSFET dosimetry of an x-ray microbeam *Med. Phys.* **27** 239–44
- 362 [17] Livingstone J, Stevenson A W, Butler D J, Häusermann D and Adam J F 2016 Characterization of a synthetic single crystal
363 diamond detector for dosimetry in spatially fractionated synchrotron x-ray fields *Med. Phys.* **43** 4283–93
- 364 [18] PTW Freiburg 2013 IONIZING RADIATION DETECTORS: Including Codes of Practice 100
- 365 [19] Rosenfeld A B 2006 Electronic dosimetry in radiation therapy *Radiat. Meas.* **41** S134–53
- 366 [20] Archer J, Li E, Davis J, Cameron M, Rosenfeld A and Lerch M 2019 High spatial resolution scintillator dosimetry of
367 synchrotron microbeams *Sci. Rep.* **9** 6873
- 368 [21] Lerch M L F, Dipuglia A, Cameron M, Fournier P, Davis J, Petasecca M, Cornelius I, Perevertaylo V and Rosenfeld A B 2017
369 New 3D Silicon detectors for dosimetry in Microbeam Radiation Therapy *J. Phys. Conf. Ser.* **777** 012009
- 370 [22] Davis J A, Paino J R, Dipuglia A, Cameron M, Siegele R, Pastuovic Z, Petasecca M, Perevertaylo V L, Rosenfeld A B and
371 Lerch M L F 2018 Characterisation and evaluation of a PNP strip detector for synchrotron microbeam radiation therapy
372 *Biomed. Phys. Eng. Express* **4** 044002
- 373 [23] Povoli M, Alagoz E, Bravin A, Cornelius I, Bräuer-Krisch E, Fournier P, Hansen T-E E, Kok A, Lerch M, Monakhov E, Morse
374 J, Petasecca M, Requardt H, Rosenfeld A B, Röhrich D, Sandaker H, Salomé M and Stugu B 2015 Thin silicon strip
375 detectors for beam monitoring in Micro-beam Radiation Therapy *J. Instrum.* **10** P11007–P11007
- 376 [24] Bartzsch S, Lott J, Welsch K, Bräuer-Krisch E and Oelfke U 2015 Micrometer-resolved film dosimetry using a microscope
377 in microbeam radiation therapy *Med. Phys.* **42** 4069–79
- 378 [25] Anon GAFCHROMIC™ DOSIMETRY MEDIA, TYPE EBT-3
- 379 [26] Anon Garchromic EBT Films - GAFchromic™

- 380 [27] McLaughlin W L, Yun-Dong C, Soares C G, Miller A, Van Dyk G and Lewis D F 1991 Sensitometry of the response of a new
381 radiochromic film dosimeter to gamma radiation and electron beams *Nucl. Instruments Methods Phys. Res. Sect. A Accel.*
382 *Spectrometers, Detect. Assoc. Equip.* **302** 165–76
- 383 [28] Li Y, Chen L, Zhu J and Liu X 2017 The combination of the error correction methods of GAFCHROMIC EBT3 film *PLoS One*
384 **12** 1–17
- 385 [29] Tagiling N, Ab Rashid R, Azhan S N A, Dollah N, Geso M and Rahman W N 2018 Effect of scanning parameters on dose-
386 response of radiochromic films irradiated with photon and electron beams *Heliyon* **4** e00864
- 387 [30] Sedgwick I, Das D, Guerrini N, Marsh B and Turchetta R 2013 LASSENA: A 6.7 Megapixel, 3-sides Buttable Wafer-Scale
388 CMOS Sensor using a novel grid- addressing architecture *Proc. Int. Image Sens. Work.* 3–6
- 389 [31] Tryggestad E, Armour M, Iordachita I, Verhaegen F and Wong J W 2009 A comprehensive system for dosimetric
390 commissioning and Monte Carlo validation for the small animal radiation research platform. *Phys. Med. Biol.* **54** 5341–57
- 391 [32] Anon ExaFlex™ - MacroMedics
- 392 [33] Micke A, Lewis D F and Yu X 2011 Multichannel film dosimetry with nonuniformity correction *Med. Phys.* **38** 2523–34
- 393 [34] Mayer R R, Ma F, Chen Y, Miller R I, Belard A, McDonough J and O'Connell J J 2012 Enhanced dosimetry procedures and
394 assessment for EBT2 radiochromic film *Med. Phys.* **39** 2147–55
- 395 [35] Anon ZEISS Axio Imager 2 for Life Science Research
- 396 [36] Anon 2015 *BSI Standards Publication Medical electrical equipment — Characteristics of digital x-ray imaging devices Part*
397 *1-1 : Determination of the detective quantum efficiency — Detectors used in*
- 398 [37] Donini B, Rivetti S, Lanconelli N and Bertolini M 2014 Free software for performing physical analysis of systems for
399 digital radiography and mammography
- 400 [38] Das I J, Ding G X and Ahnesjö A 2007 Small fields: Nonequilibrium radiation dosimetry *Med. Phys.* **35** 206–15
- 401 [39] Bartzsch S, Cummings C, Eismann S and Oelfke U 2016 A preclinical microbeam facility with a conventional x-ray tube
402 *Med. Phys.* **43** 6301–8
- 403 [40] Eling L, Bouchet A, Nemoz C, Djonov V, Balosso J, Laissue J, Bräuer-Krisch E, Adam J F and Serduc R 2019 Ultra high dose
404 rate Synchrotron Microbeam Radiation Therapy. Preclinical evidence in view of a clinical transfer *Radiother. Oncol.* **139**
405 56–61
- 406 [41] Poludniowski G G and Evans P M 2007 Calculation of x-ray spectra emerging from an x-ray tube. Part I. Electron
407 penetration characteristics in x-ray targets *Med. Phys.* **34** 2164–74
- 408 [42] Poludniowski G G 2007 Calculation of x-ray spectra emerging from an x-ray tube. Part II. X-ray production and filtration
409 in x-ray targets *Med. Phys.* **34** 2175–86
- 410 [43] Poludniowski G, Landry G, DeBlois F, Evans P M and Verhaegen F 2009 *SpekCalc* : a program to calculate photon spectra
411 from tungsten anode x-ray tubes *Phys. Med. Biol.* **54** N433–8
- 412 [44] Martínez-Rovira I, Sempau J and Prezado Y 2012 Development and commissioning of a Monte Carlo photon beam model
413 for the forthcoming clinical trials in microbeam radiation therapy *Med. Phys.* **39** 119–31
- 414 [45] Livingstone J, Stevenson A W, Häusermann D and Adam J-F 2018 Experimental optimisation of the X-ray energy in
415 microbeam radiation therapy *Phys. Medica* **45** 156–61
- 416 [46] PTW 2010 *User Manual: UNIDOS webline Type 10021, Type 10022 and Type 10023*
- 417 [47] Sellier C, Gambart D, Perrot N, Garcia-Sanchez E, Virmondois C, Moullem W and Bardoux A 2019 Development and
418 qualification of a miniaturised CMOS camera for space applications (3DCM734/3DCM739) *International Conference on*
419 *Space Optics — ICSO 2018* vol 11180, ed N Karafolas, Z Sodnik and B Cugny (SPIE) p 106

- 420 [48] Kim W T, Park C, Lee H, Lee I and Lee B G 2019 A high fullwell capacity CMOS image sensor for space applications
421 *Sensors (Switzerland)* **19**
- 422 [49] Borca V C, Pasquino M, Russo G, Grosso P, Cante D, Sciacero P, Girelli G, Porta M R La and Tofani S 2013 Dosimetric
423 characterization and use of GAFCHROMIC EBT3 film for IMRT dose verification *J. Appl. Clin. Med. Phys.* **14** 158
- 424 [50] Schültke E, Balosso J, Breslin T, Cavaletti G, Djonov V, Esteve F, Grotzer M, Hildebrandt G, Valdman A and Laissue J 2017
425 *Microbeam radiation therapy-grid therapy and beyond: a clinical perspective*
- 426 [51] Prezado Y, Jouvion G, Patriarca A, Nauraye C, Guardiola C, Juchaux M, Lamirault C, Labiod D, Jourdain L, Sebrie C,
427 Dendale R, Gonzalez W and Pouzoulet F 2018 Proton minibeam radiation therapy widens the therapeutic index for high-
428 grade gliomas *Sci. Rep.* **8** 16479
- 429 [52] Page R F, Abbott N L, Davies J, Dyke E L, Randles H J, Velthuis J J, Fletcher S, Gregory S D, Hall C, John A M, Lawrence H,
430 Stevens P H, Hugtenburg R P and Tunbridge V 2014 Using a monolithic active pixel sensor for monitoring multileaf
431 collimator positions in intensity modulated radiotherapy *IEEE Trans. Nucl. Sci.* **61** 74–8
- 432 [53] Bartoli A, Scaringella M, Baldi A, Greto D, Scoccianti S, Masi L, Pallotta S, Bruzzi M and Talamonti C 2018 PO-0873: 2D
433 pixelated diamond detector for patient QA in advanced radiotherapy treatments *Radiother. Oncol.* **127** S459–60
- 434

Figure 1: Experimental set up: (a) vM1212 detector with 1 cm of water equivalent build-up, (b) vM1212 detector with aligned EBT3 Film.

- i. Lead shield to protect collimator electronics
- ii. Microbeam collimator
- iii. 1 cm of water equivalent bolus
- iv. Cable for microbeam collimator
- v. vM1212 detector
- vi. Ribbon cables for vM1212 detector
- vii. Non-fibrous card with alignment points
- viii. 3 x 3 cm² square of EBT3 film

Figure 2: (a) Photograph of microbeam collimator slits. (b) vM1212 detector image (cropped). (c) Scan of exposed EBT3 film using the Epson Expression 10000XL scanner (100 μm slit width). (d) Scan of exposed EBT3 film using Zeiss Axio Imager 2.

Figure 3: (a) 100 μm slit width profile comparison, (b) Microbeam peak deviation between the vM1212 detector and the two EBT3 film methods.

Figure 4: 100 μm slit width valley profile comparison.

Figure 5: 100 μm slit width profile comparison of the 26th central peak.

Figure 6: 25 μm slit width profile comparison.

Figure 7: 25 μm slit width valley profile comparison.

Figure 8: 25 μm slit width peak profile comparison of the 26th central peak.

Figure 9: 25 μm slit width peak profile comparison of the 27th peak.

Figure 10: Radiation leakage through the collimator at 0 μm slit width as measured by the vM1212 detector.

Figure 11: Horizontal profile of the 100 μm slit width.

Figure 12: FWHM comparison between Zeiss Axio Imager 2 and the vM1212 detector. A 1:1 ratio has been added to guide the eye.

Figure 13: FWHM trend as measured by the vM1212 detector.

Figure 14: Comparing the microbeam slit width to observed FWHM.

Figure 15: Geometric setup of the microbeam collimator, resulting in the larger full width at half maximum (FWHM).

Figure 16: Comparison of PVDR for different slit widths. The PVDR measurements for the 25 and 50 μm slit width Epson Expression 10000XL are omitted.

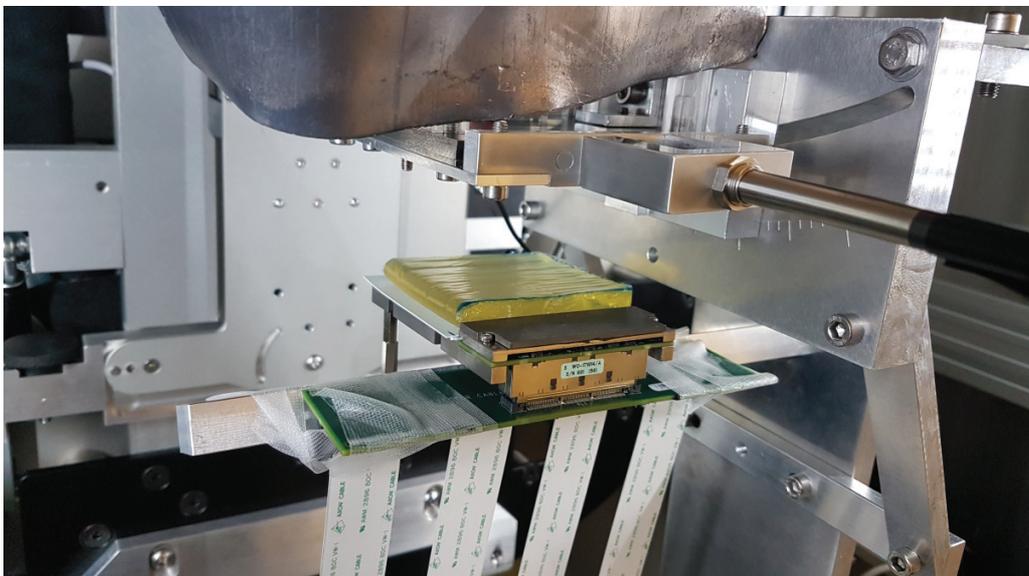
Figure 17: Comparison of MTF for different measurement techniques.

Table 1: Measured peak to peak separation as measured on the three detectors. Statistical uncertainty corresponds to one standard deviation.

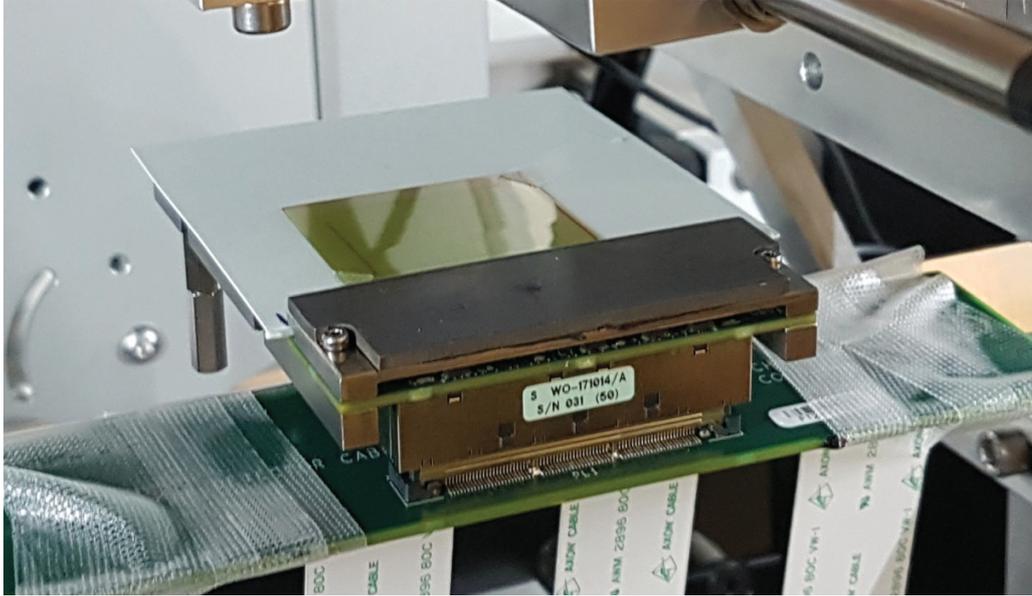
Nominal slit width (μm)	Measured peak to peak separation (μm)		
	vM1212 detector	Epson Expression 10000XL	Zeiss Axio Imager 2
25	513.4 ± 13.9	512.0 ± 11.3	508.3 ± 9.9
50	512.9 ± 10.1	511.7 ± 9.7	508.9 ± 9.1
75	512.6 ± 9.2	511.9 ± 10.1	508.3 ± 8.6
100	512.4 ± 9.5	511.8 ± 9.6	508.5 ± 9.8

Table 2: Comparison of the different microbeam detection methods evaluated in this work.

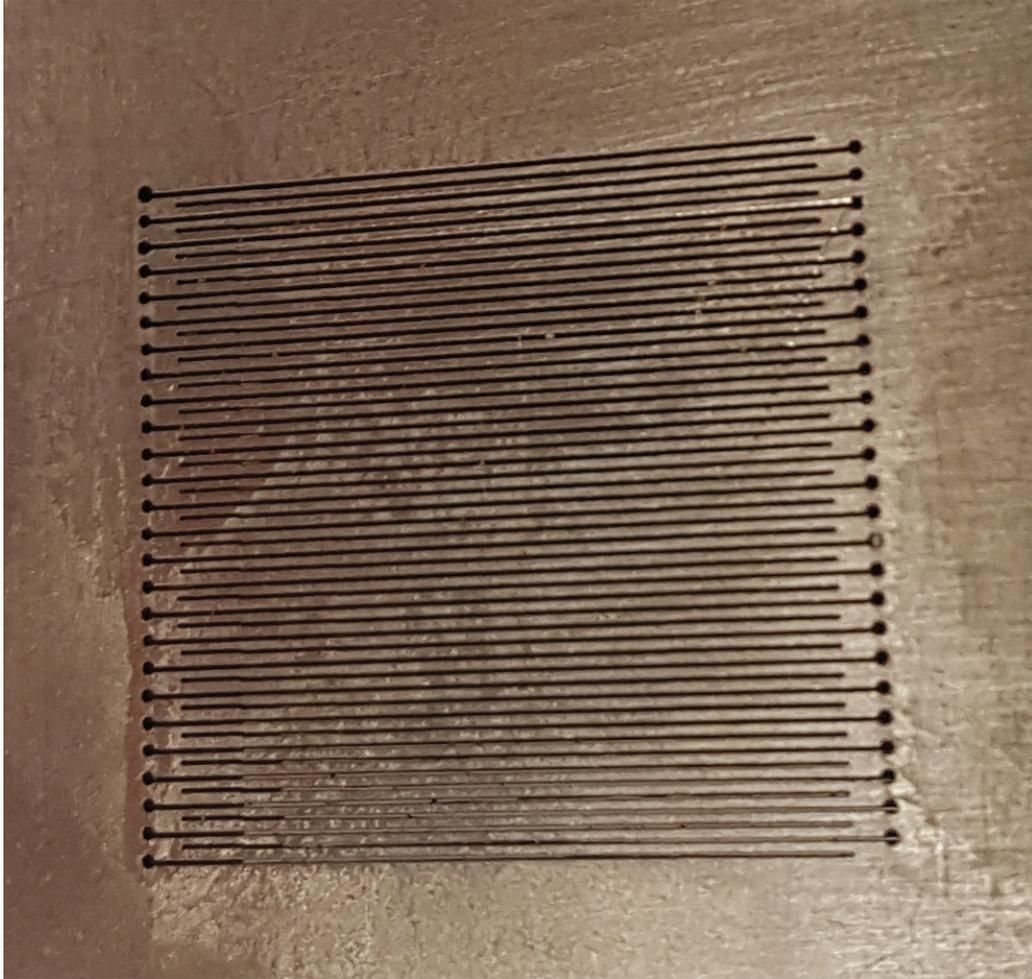
		Microbeam detection method			
		vM1212 detector	Zeiss Imager (+ EBT3 film)	Axio 2	Epson Expression 10000XL (+ EBT3 film)
Advantages	Real time measurement and analysis		Highest resolution	spatial	Lower cost
	Short exposure is sufficient to obtain accurate profile information		No dose rate dependence[49]		Established clinical workflow No dose rate dependence[49]
Disadvantages	Limited life expectancy due to cumulative radiation damage		24 hours self-development	self-	24 hours self-development
	Spatial resolution limited by 50 μm pixel pitch		Complex and time consuming analysis process	and	Poorest spatial resolution ad hence limited suitability for microbeam applications



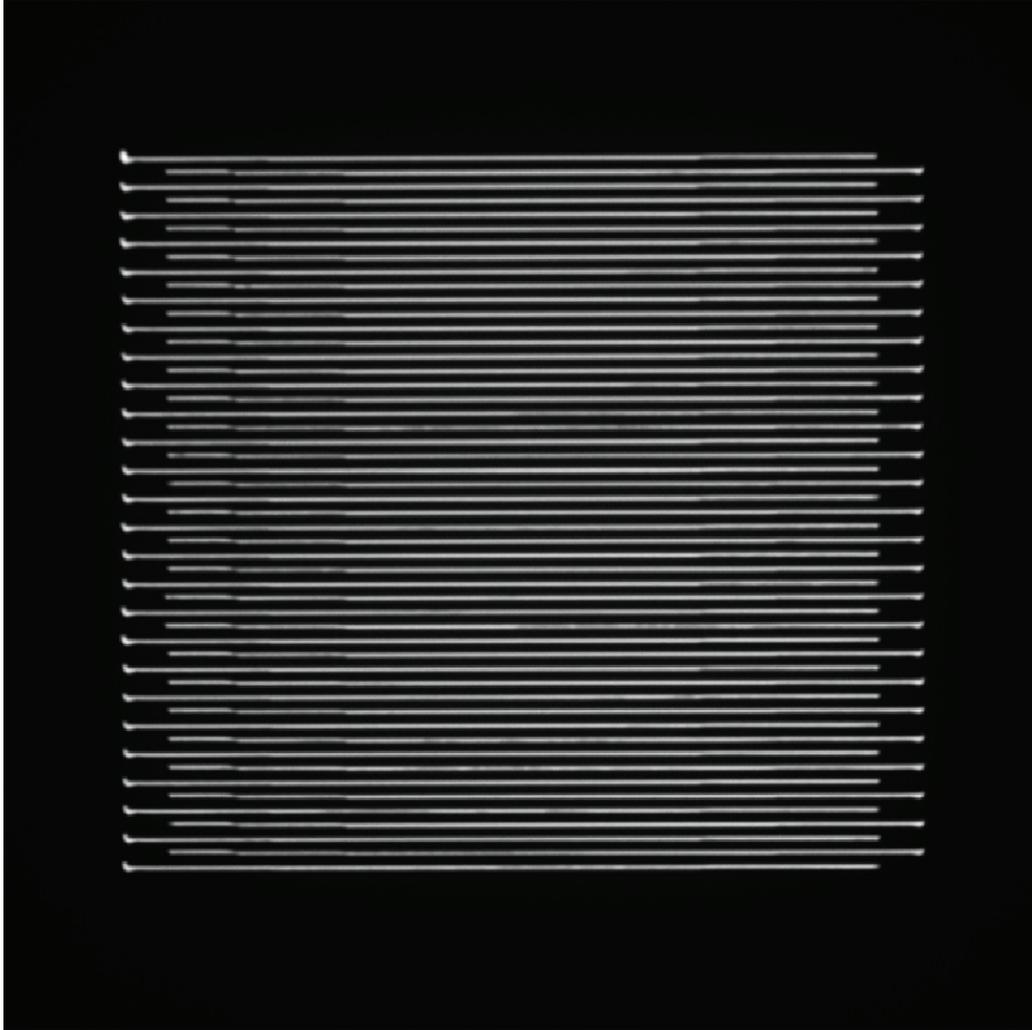
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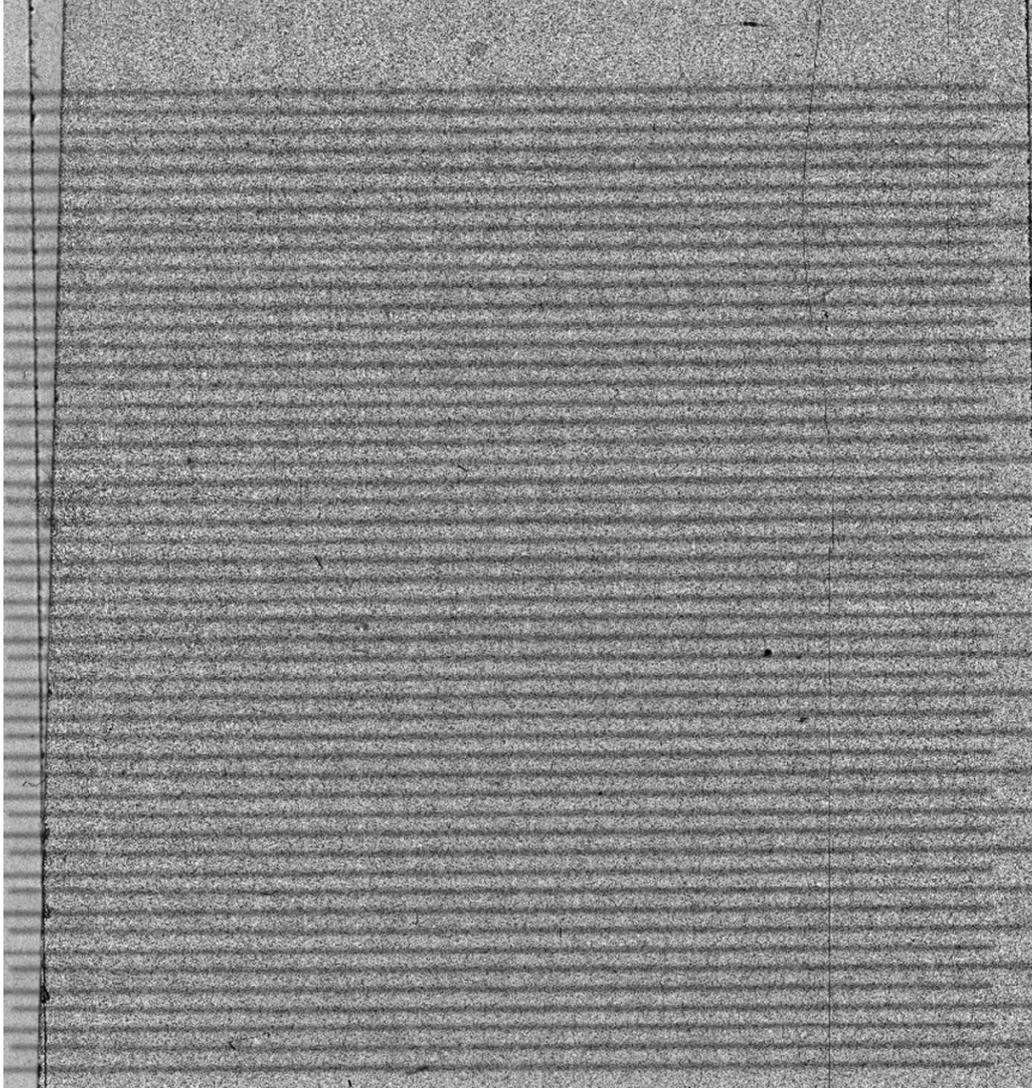
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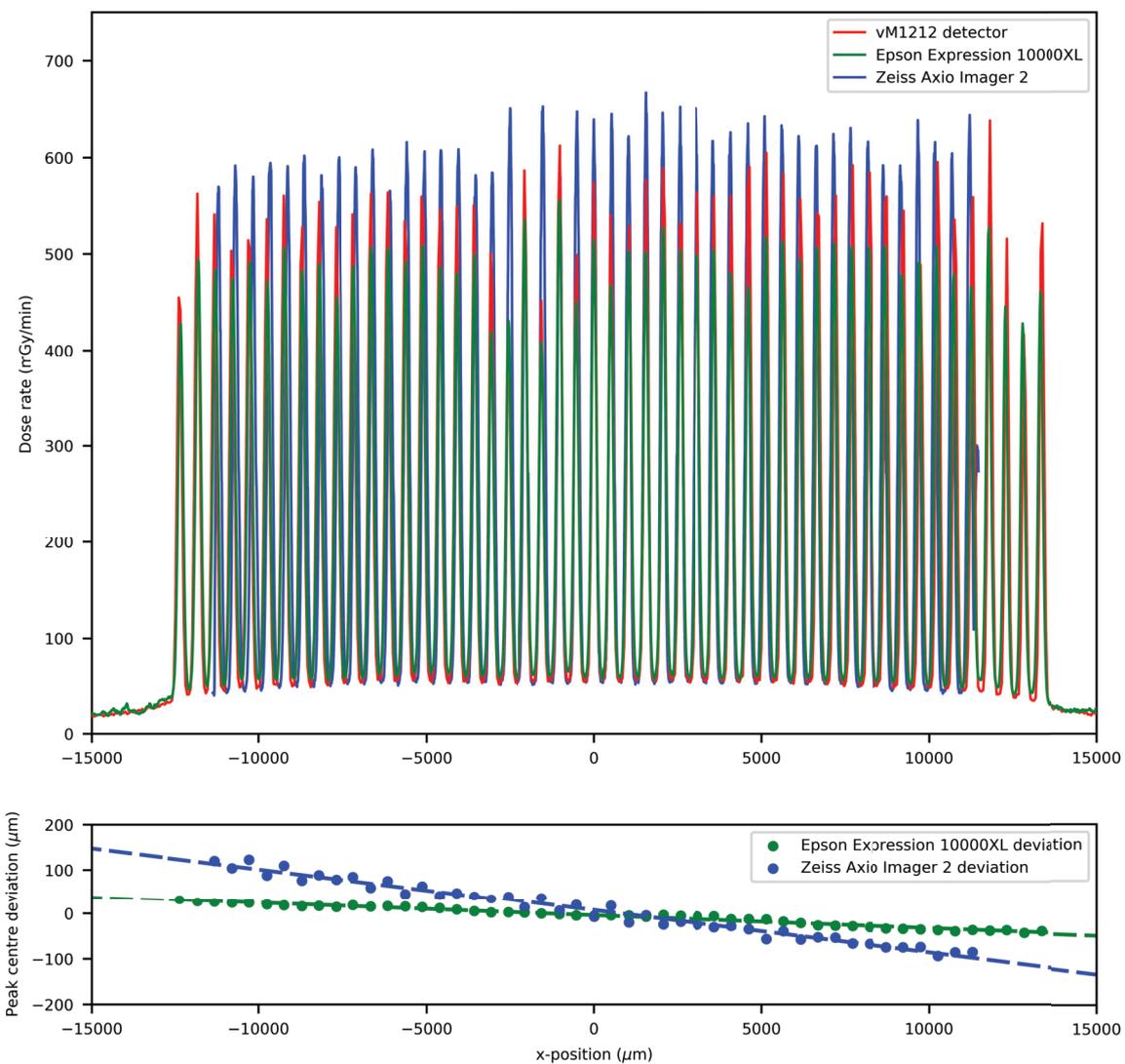
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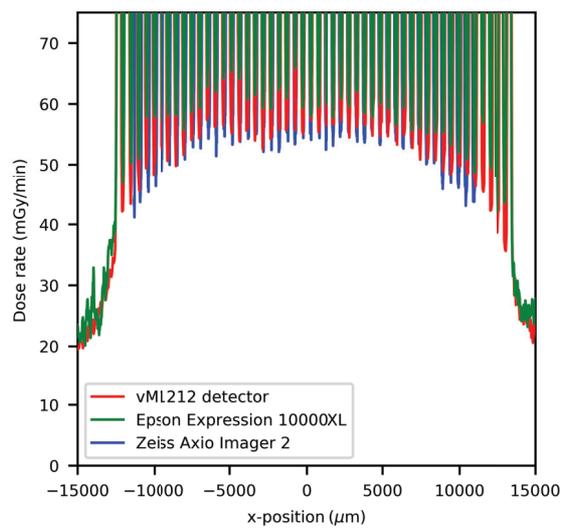
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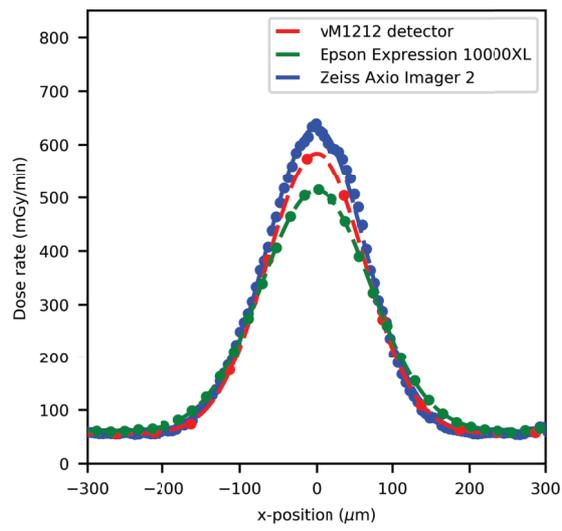
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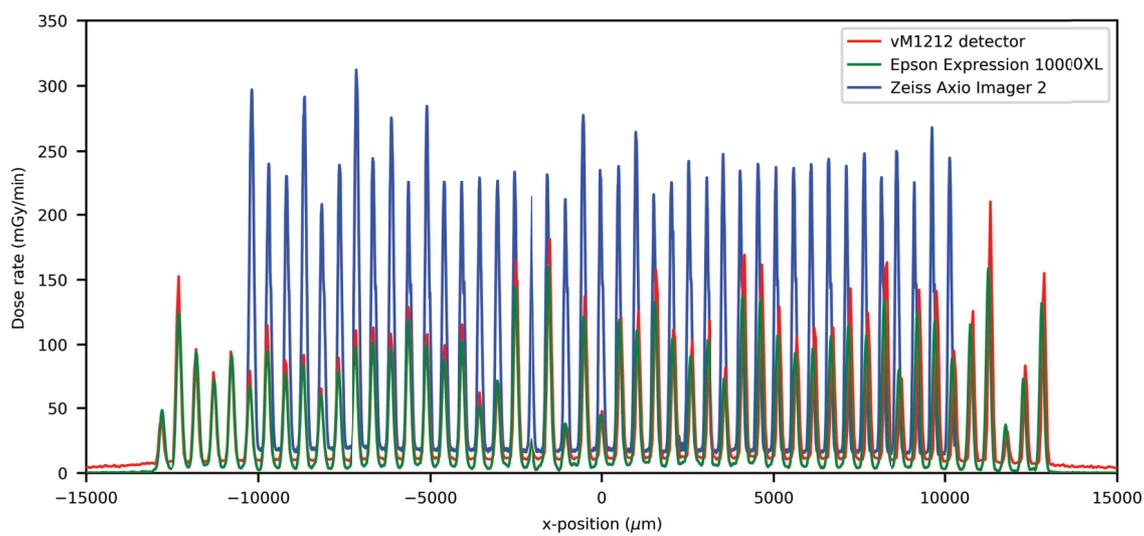
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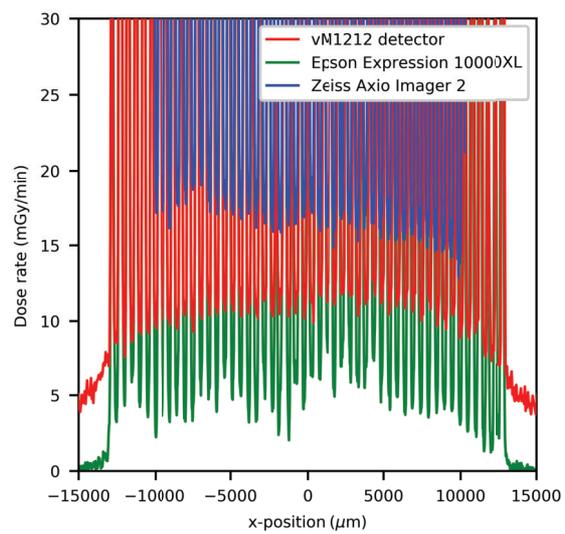
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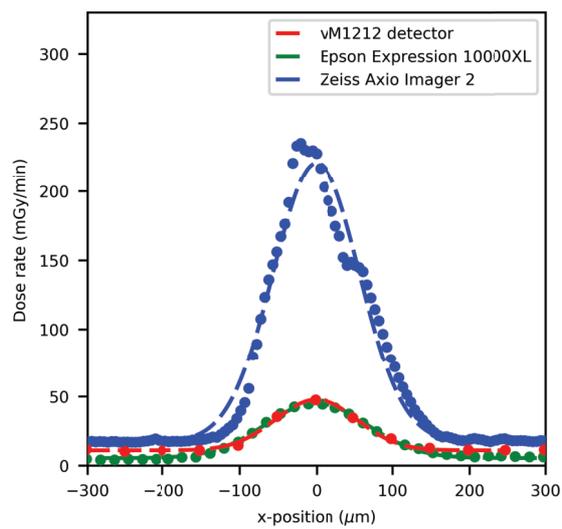
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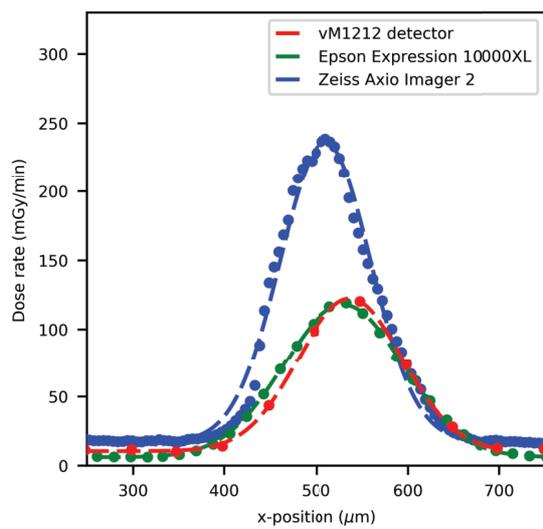
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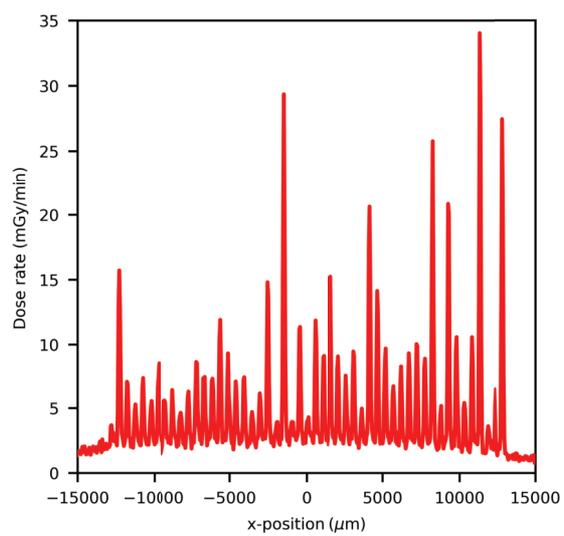
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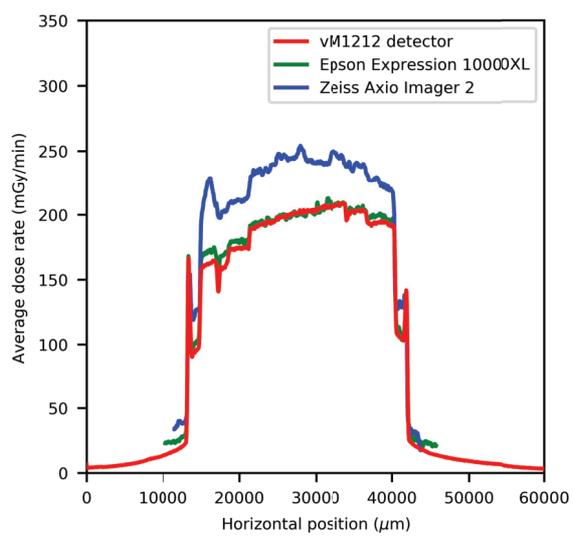
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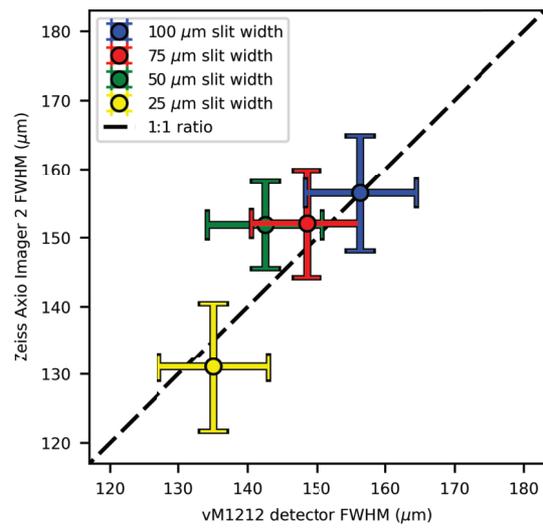
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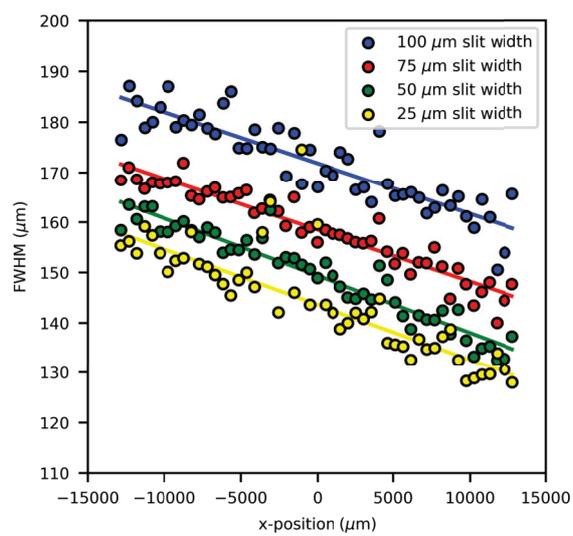
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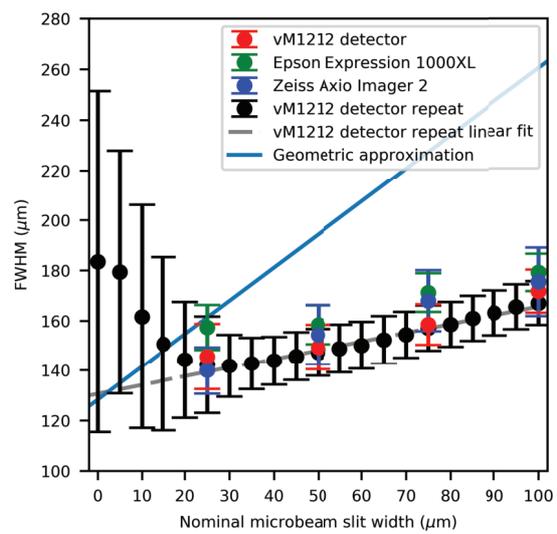
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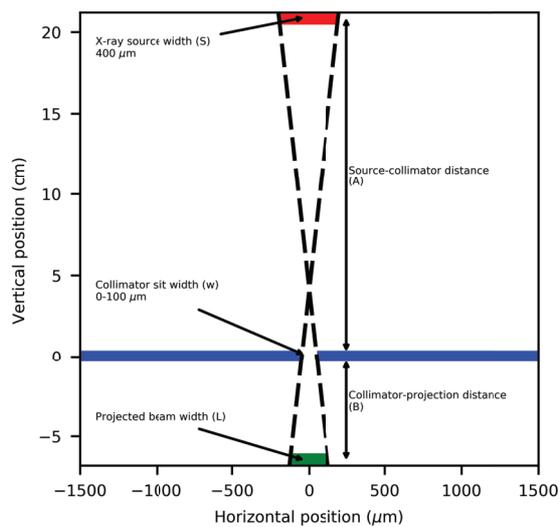
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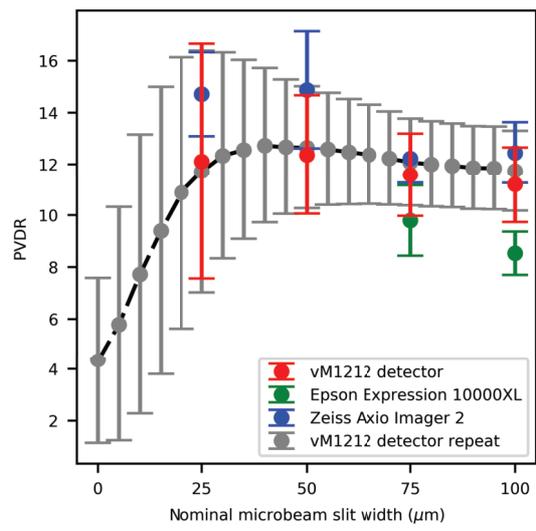
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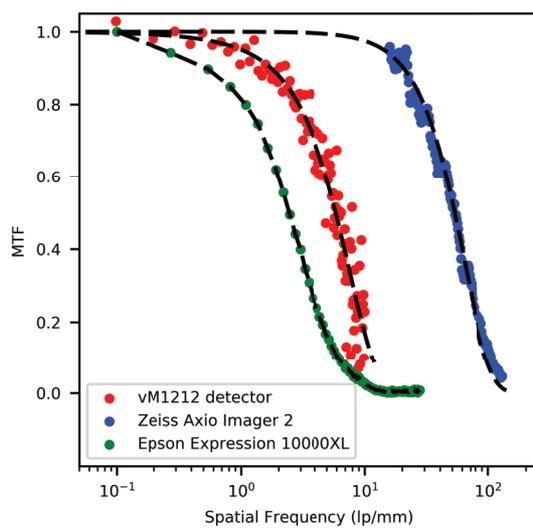
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