

Persistent Organic Pollutants in Human Breast Milk and Associations with Maternal Thyroid Hormone Homeostasis

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1 Persistent Organic Pollutants in Human Breast Milk
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3 Homeostasis

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19 ABSTRACT

20 Epidemiological studies have indicated the thyroid-disrupting effects of persistent organic
21 pollutants (POPs). However, the associations of low-exposure POPs with thyroid hormones (THs)
22 remain unclear. Here we aim to assess the associations of low exposure of POPs, including
23 polybrominated diphenyl ethers (PBDEs), polychlorinated biphenyls (PCBs), polychlorinated
24 dibenzo-*p*-dioxins and furans (PCDD/Fs), and polybrominated dibenzo-*p*-dioxins and furans
25 (PBDD/Fs), with THs (total _L-thyroxine (TT₄), total 3,3',5-triiodo-_L-thyronine (TT₃), and total
26 3,3',5'-triiodo-_L-thyronine (TrT₃)) measured in human breast milk. Ninety-nine breast milk
27 samples were collected from the LUPE cohort (2015–2016, Bavaria, Germany). Fourteen PBDEs,
28 17 PCBs, and 5 PCDD/Fs had quantification rates of > 80%. Nonmonotonic associations were
29 observed. In adjusted single-pollutant models: (1) TT₄ was inversely associated with BDE-99, -
30 154, and -196; (2) TT₃ was inversely associated with BDE-47, -99, -100, -197, -203, -207, and
31 OCDD; (3) TrT₃ was inversely associated with BDE-47, -99, -183, and -203. Multipollutant
32 analysis using principal component analysis and hierarchical clustering revealed inverse
33 associations of PBDEs (BDE-28, -47, -99, -100, -154, -183, and -197) with TT₄ and TrT₃. These
34 results indicate that POPs at low levels might be related to reduced THs. This study shows that
35 human breast milk might be an appropriate specimen to evaluate the thyroid-disruption of POPs.

36

37 1. Introduction

38 Persistent organic pollutants (POPs) are a group of chemicals with environmental persistence,
39 bioaccumulation, and toxicity. They occur as a result of industrial and commercial applications,
40 incomplete incineration, traffic, and industrial processes ¹. Common POPs include polybrominated
41 diphenyl ethers (PBDEs), polychlorinated biphenyls (PCBs), polychlorinated dibenzo-*p*-dioxins
42 and furans (PCDD/Fs), and polybrominated dibenzo-*p*-dioxins and furans (PBDD/Fs). Humans
43 are exposed to POPs through diet, air, house dust, and drinking water. Although many POP
44 congeners have been strictly limited or banned, exposure to these compounds continues because
45 of their long half-lives ². POPs have been detected in the environment and humans all over the
46 world ^{3,4}.

47 Certain POPs have chemical structures similar to thyroid hormones (THs) leading to concerns
48 about their potential of thyroid disruption. TH homeostasis is crucial for down-stream
49 physiological processes such as metabolism, growth, bone remodeling, cardiac function and
50 mental status ⁵. POPs can interact with any aspect of the hypothalamus-pituitary-thyroid (HPT)
51 axis, TH biosynthesis, metabolism, and release, feedback regulation, transport, agonist or
52 antagonist thyroid hormone receptor (TR), and regulation of uridine diphosphate
53 glucuronosyltransferases (UDPGTs) and sulfotransferases (SULTs) ⁵⁻⁷. POP exposures may
54 partially contribute to the rapid increasing incidence of thyroid diseases such as hypothyroidism,
55 hyperthyroidism, and thyroid cancer ⁸⁻¹⁰.

56 *In vitro* and animal studies have proved TH disruption following POP exposures ^{11, 12}. Human
57 studies also observed associations between THs and POPs using peripheral/cord blood ¹³⁻¹⁶ and
58 placenta ^{17, 18}. Conflicting results regarding the direction of associations have been reported ^{15, 19-}
59 ²¹. Possible reasons include the low-dose effects and non-monotonic effects of endocrine-

60 disrupting chemicals (EDCs) ²². For example, positive association between PBDEs and THs was
61 found in a high-exposure population (median Σ PBDEs: 38.4 ng/g lipid weight (lw)) ²¹, whereas
62 negative association was reported in a low-exposure population (median Σ PBDEs: 3.49 ng/g lw)
63 ²³. The thyroid-disrupting effects of POPs at low levels are of concern since most of the current
64 studies were conducted in high-exposure populations. However, the detection of POPs in blood of
65 low-exposure population requires high sensitivity or large sample volume to obtain sufficient
66 detection frequencies (DFs) ²⁴, which can be limiting for certain age groups.

67 Human breast milk is a complex and constantly changing mixture of endogenous and exogenous
68 substances including THs and POPs ^{3, 25}. Due to its high lipid content, breast milk has been
69 considered as an appropriate specimen to provide improved sensitivity for POP monitoring ^{3, 26}.
70 Besides, the serum TH homeostasis may be evaluated by examining THs in breast milk because
71 of the significant positive correlations between milk THs and serum THs ²⁷. Several studies have
72 assessed the associations between POPs in milk and serum TH parameters ^{28, 29}. Darnerud *et al.*
73 found that low chlorinated PCBs in breast milk were inversely associated with total 3,3',5-triiodo-
74 _L-thyronine (TT₃) in serum of 3-week old children, while PCDD/Fs in breast milk showed negative
75 associations with maternal serum TT₃ ²⁹. However, no study has been conducted to evaluate the
76 associations of THs with POPs both measured in human breast milk.

77 The primary goal of the current study was to evaluate the associations of POPs (PBDEs, PCBs,
78 PCDD/Fs, and PBDD/Fs) with THs (total _L-thyroxine (TT₄), TT₃, and total 3,3',5'-triiodo-_L-
79 thyronine (TrT₃)) measured in human breast milk. Samples were collected from the LUPE cohort
80 (2015–2016, Bavaria, Germany), which is exposed to low levels of POPs from a global
81 perspective. Single-pollutant and multipollutant models were applied to evaluate the relationship
82 between THs and POPs.

83 2. Materials and methods

84 2.1 Sample collection

85 We included 99 human breast milk samples in this study. Approximately 150 mL of sample was
86 collected from each participating woman within 10 months after delivery. Samples were collected
87 into sample cups (AVENT VIA) using a manual breast pump (AVENT ISIS) after breastfeeding.
88 Afterwards, samples were transported to the Bavarian Health and Food Safety Authority (Munich,
89 Germany) for POP determination. An aliquot of 2 mL was delivered to the Helmholtz Center
90 Munich (Munich, Germany) for TH analysis. Samples were stored at -80 °C until processing.

91 The ethics committee of the Bavarian Chamber of Physician approved this study. Informed
92 written consent was obtained from each participant.

93 2.2 POP analysis

94 Detailed analytical methods regarding POP quantification are available elsewhere^{30, 31}. The
95 materials are shown in the Supporting Method. Briefly, milk lipid was extracted with *n*-
96 hexane/propane-2-ol and applied on a column composed of Isolute HM-N/sodium chloride. The
97 concentrated lipid extract was dried on an anhydrous sodium sulphate column and extracted with
98 *n*-pentane. After further automated clean-up and fractionation with DEXTech (3 columns setup),
99 the final extracts were analyzed by two gas chromatographs/high resolution mass spectrometer
100 (2GC/HRMS) on a Thermo DFS system with three different columns. The World Health
101 Organization Toxicant Equivalent Quotient (WHO₂₀₀₅-TEQ) of dioxins and dioxin-like PCBs (dl-
102 PCBs) was calculated³². The average method quantification limits (MQLs) were 0.125 pg/g lw
103 for PCDD/Fs, 2.63 pg/g lw for dl-PCBs, 4.71 pg/g lw for non-dl-PCBs, 4.16 pg/g lw for PBDD/Fs,
104 and 3.99 pg/g lw for PBDEs. The recoveries of these POPs ranged overall from 50% to 140% and
105 comply with the requirements of Regulation (EU) No. 589/2014.

106 2.3 TH measurement

107 Total levels of T₄, T₃, rT₃, 3,3'-diiodo-L-thyronine (3,3'-T₂), 3,5-diiodo-L-thyronine (3,5-T₂), 3-
108 iodo-L-thyronine (T₁) and 3-iodothyronamine (3-T₁AM) were targeted for analysis in breast milk
109 using isotope-dilution liquid chromatography tandem mass spectrometry (LC-MS/MS). The
110 method was based on our previous technology with some modifications³³. Complete details can
111 be found in the Supporting Method, Table S1, and Fig. S1-2. The method detection limits (MDLs)
112 and MQLs were 0.01–0.13 ng/mL and 0.10–0.42 ng/mL, respectively. The matrix effects were
113 between -9.67% and 14.7%. The overall recoveries ranged from 102% to 125%. The spike-
114 recoveries were in the range of 98.4%–122%. The intra-day and inter-day variations were 0.47%–
115 6.91% and 1.37%–7.71%, respectively (Table S2).

116 2.4 Statistics

117 The statistical analyses were conducted on POP congeners with DF of > 80%, measurements
118 below the LOQ were replaced by LOQ × DF³⁴. Normality was tested using Shapiro-Wilk test.
119 The distributions of biomarkers were log-normal and therefore transformed by the natural
120 logarithm. We examined the bivariate associations between biomarkers and a set of demographic
121 variables using t-test or analysis of variance (ANOVA). Afterwards, Spearman's rank correlation
122 was applied to evaluate the correlation of biomarkers. Statistical analyses were conducted using R
123 (version 3.4.2; R Foundation for Statistical Computing, Vienna, Austria) and DAGitty v2.3³⁵ for
124 constructing directed acyclic graph (DAG). Statistical significance was defined as *p*-value < 0.05.

125 Potential confounders considered for inclusion in models were maternal age, educational level,
126 parity, smoking, diet, infant gender, infant age at sampling. Data on most covariates were
127 complete. Confounders were identified based on previous reports and a DAG framework (Fig. S3).

128 Body mass index (BMI) was not controlled because BMI might be a consequence of thyroid
129 dysfunction^{36,37}.

130 Single-pollutant models were conducted to investigate the associations between THs and each
131 POP congener. Generalized additive models (GAM) were used to examine the linearity of the
132 relationship between POPs and THs. Some of the POP congeners showed significant non-linear
133 associations with THs (data not shown), thus we modeled all exposure biomarkers in categories
134 defined by tertiles.

135 ***Principal component analysis***

136 Due to the structural and biological similarity within and across the classes, interpretation of the
137 effect of individual POP congeners can be misleading. We assessed the multiple collinearity by
138 the eigen values of the correlation and the variable inflation factor (VIF). Principal component
139 analysis (PCA) was then conducted to convert the correlated variables into a small number of
140 principal components (PCs). Afterwards, varimax rotation was applied to calculate factor scores
141 for each participant. The number of factors was decided based on the scree plot³⁸. The factor
142 scores were categorized into tertiles and included in the regression models. Regressions were
143 performed including factors simultaneously and separately.

144 ***Hierarchical clustering***

145 We used the partial least squares (PLS) regression to evaluate the impact of all POPs and
146 covariates on THs simultaneously. Only variables with variable importance to projection (VIP)
147 values > 0.4 were included in the final model to reduce data and increase the model predictive
148 ability¹⁴. The score of each participant on PC1 was included in multiple linear regression models
149 as a common vector to avoid collinearity while adjusting for these factors. In order to minimize
150 the number of POPs to be included in linear regression models, we conducted hierarchical

151 clustering analysis of POPs based on correlations (method: complete linkage). Groupings
152 according to clusters were subsequently performed by simple addition of POP concentrations.

153 ***Sensitivity analyses***

154 Previous studies measuring serum POPs typically adjusted for lipid content. However, there is
155 controversy regarding the best approach³⁹. In this study we performed the analyses including POPs
156 in units of ng/g lipid. In sensitivity analysis we repeated the analyses with POPs in units of ng/L
157 milk while controlling for lipid content as a covariate. Additional sensitivity analysis included the
158 adjustment of BMI.

159 **3. Results**

160 **3.1 Biomarker concentrations and their correlations**

161 An LC-MS/MS method was optimized and validated for TH quantification in human breast milk.
162 The mean \pm SD concentrations of TT₄, TT₃, and TrT₃ were 0.57 ± 0.20 , 0.13 ± 0.03 , and $0.02 \pm$
163 0.01 ng/mL, respectively (Table S3).

164 As shown in Tables 1 & S4, 14 PBDEs had DFs of $> 80\%$. The median concentrations of these
165 PBDEs were 8.25–440 pg/g lw, in which BDE-209 was the dominating congener, followed by
166 BDE-153, -47, -197, -99, -207, -100, -28, -206, -183, -208, -196, -203, and -154. Seventeen PCBs
167 were detected in $> 80\%$ of the samples with median concentrations in the range of 0.14–3619 pg/g
168 lw. The dominating congener was PCB-118 followed by PCB-156, -167, -105, -157, -114, -189, -
169 123, -153, -126, -138, -169, -180, -77, -28, -101, and -52. Five PCDD/Fs were quantified in $> 80\%$
170 of the samples with median concentrations between 1.03 and 17.0 pg/g lw. The dominating
171 congener was OCDD followed by 2,3,4,7,8-PeCDF, 1,2,3,6,7,8-HxCDD, 1,2,3,4,6,7,8-HpCDD,
172 and 1,2,3,6,7,8-HxCDF. The DFs of all the PBDD/Fs were $\leq 38\%$ and therefore not included in
173 the statistical analyses. As shown in Table S5, the highest WHO₂₀₀₅-TEQ levels in dl-PCBs and

174 PCDD/Fs were found for PCB-126 (1.81 pg/g lw) and 2,3,4,7,8-PeCDF (1.12 pg/g lw),
 175 respectively. The median values of Σ mono-ortho PCBs, Σ non-ortho PCBs, Σ PCBs, Σ PCDD/Fs,
 176 Σ PBDD/Fs, and Σ POPs were 2.78, 0.33, 3.11, 4.37, 0.93, and 8.22 pg/g lw, respectively. Table
 177 S6-S8 show the comparison of POP levels in human breast milk reported here with recent studies
 178 from different regions. POP levels in this study were generally lower, especially compared with
 179 those measured in North America.

180 **Table 1** Descriptive statistics of PBDEs, PCBs, PCDD/Fs, and PBDD/Fs with DFs > 80% in
 181 human breast milk from LUPE study (2015–2016, Bavaria, Germany).

POPs	N (%)	Mean (pg/g lw)	Range (pg/g lw)	Q1 (pg/g lw)	Median (pg/g lw)	Q3 (pg/g lw)
BDE-28	95 (96)	31.7	<LOQ–122	20.2	29.3	37.7
BDE-47	99 (100)	307	61.6–2419	136	204	299
BDE-99	98 (99)	85.8	<LOQ–419	46.1	62.5	93.9
BDE-100	97 (98)	73.7	<LOQ–364	31.3	54.3	92.5
BDE-153	99 (100)	460	112–1979	304	377	545
BDE-154	82 (83)	9.42	<LOQ–28.5	5.58	8.25	11.1
BDE-183	97 (98)	33.7	<LOQ–182	19.4	28.4	42.2
BDE-196	89 (90)	22.4	<LOQ–146	12.4	16.8	24.1
BDE-197	99 (100)	83.2	19.3–224	53.6	73.1	103
BDE-203	91 (92)	22.8	<LOQ–265	12.8	16.7	24.4
BDE-206	86 (87)	176	<LOQ–3545	19.1	28.9	57.3
BDE-207	98 (99)	147	<LOQ–2842	39.7	56.3	82.0
BDE-208	98 (99)	67.5	<LOQ–1540	13.3	19.3	34.0
BDE-209	95 (96)	4444	<LOQ–104000	287	440	1074
PCB-28	99 (100)	0.99	0.24–4.36	0.61	0.81	1.16

PCB-52	99 (100)	0.18	0.06–1.62	0.11	0.14	0.19
PCB-77	82 (83)	3.55	<LOQ–17.2	2.37	2.97	3.83
PCB-101	99 (100)	0.37	0.10–6.20	0.18	0.25	0.34
PCB-105	99 (100)	666	177–2067	459	591	785
PCB-114	99 (100)	251	54.8–810	159	224	330
PCB-118	99 (100)	3955	1044–9635	2821	3619	4881
PCB-123	97 (98)	42.5	<LOQ–110	28.7	37.6	55.1
PCB-126	99 (100)	20.1	4.40–58.5	14.1	18.1	23.5
PCB-138	99 (100)	15.2	3.93–35.6	10.1	14.0	18.2
PCB-153	99 (100)	26.3	5.76–70.0	17.6	23.3	34.7
PCB-156	99 (100)	2668	526–8664	1633	2128	3573
PCB-157	99 (100)	393	78.3–1143	241	344	528
PCB-167	99 (100)	688	157–1481	447	662	877
PCB-169	98 (99)	12.6	<LOQ–34.1	7.88	10.9	15.2
PCB-180	99 (100)	15.7	2.69–91.8	8.76	13.2	21.3
PCB-189	99 (100)	251	36.1–1339	133	219	351
1,2,3,6,7,8- HxCDD	87 (88)	3.26	<LOQ–14.2	2.22	2.94	3.99
1,2,3,4,6,7,8- HpCDD	91 (92)	3.26	<LOQ–14.7	1.96	2.62	3.89
OCDD	98 (99)	21.2	<LOQ–75.0	12.9	17.0	24.4
2,3,4,7,8- PeCDF	94 (95)	4.21	<LOQ–10.8	3.01	3.80	5.15
1,2,3,6,7,8- HxCDF	80 (81)	1.18	<LOQ–7.86	0.72	1.03	1.41
ΣPBDEs	99 (100)	5753	511–112998	1123	1731	2727
ΣPCBs	99 (100)	9090	2222–20225	6238	8322	11211
ΣPCDD/Fs	99 (100)	35.7	0.00–115	24.3	30.2	41.6

ΣPBDD/Fs	99 (100)	43.2	0.00–1352	0.00	4.53	11.5
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182 Abbreviations: LOQ, limit of quantification. lw, lipid weight. Q1, Q3: first and third quantile.

183 As shown in Fig. S4, THs showed weak negative to weak positive correlations with most of the
 184 POPs (T_4 : -0.25–0.17, T_3 : -0.34–0.01, rT_3 : -0.28–0.11). The intragroup correlations of PBDEs,
 185 PCBs, and PCDD/Fs were -0.05–0.90, 0.0001–0.98, and 0.18–0.68, respectively.

186 3.2 Population characteristics

187 Table S9 summarizes the sociodemographic characteristics of all the participants. The mean \pm
 188 SD age was 33.9 ± 4.4 years. Among them, 84 (84.8%) of them were > 30 years old; 66 (66.7%)
 189 had a BMI value of < 25 kg/m²; majority (95.0%) did not smoke; 45 (45.5%) were nullipara. The
 190 mean \pm SD infant age at sampling was 114 ± 57 days. As shown in Tables S9-S12, we observed
 191 significant correlations between demographic variables and biomarkers.

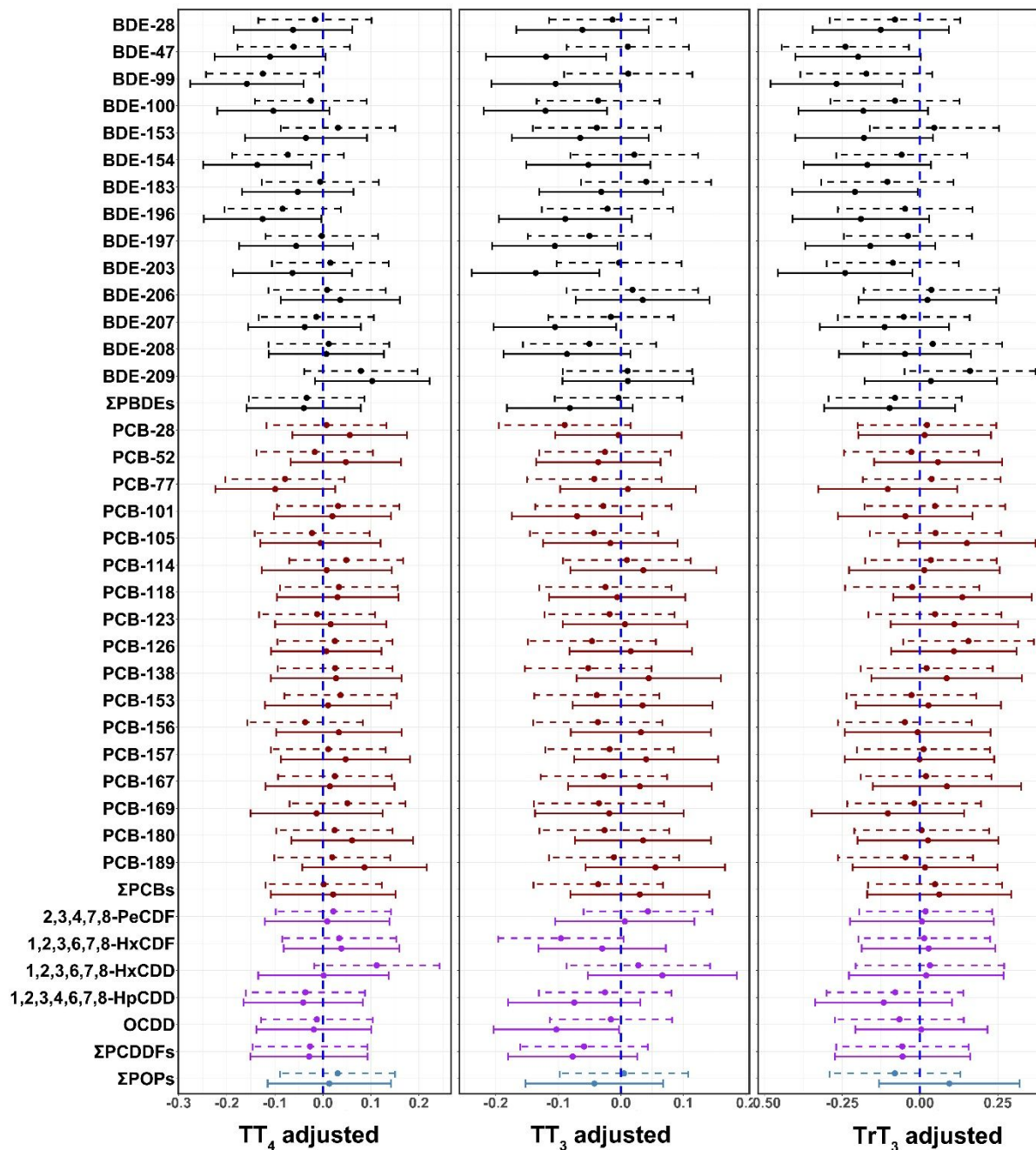
192 3.3 Single-pollutant model

193 As shown in Fig. 1 and Table S13, single-pollutant, crude models for the 36 POPs showed a
 194 significant decrease in TT_4 with increasing exposure to BDE-99, -154, -169, -196, -203, PCB-169,
 195 and 1,2,3,6,7,8-HxCDD. After adjustment, TT_4 showed significant inverse associations with BDE-
 196 99 [adjusted (adj) β tertile 2 vs. 1: -0.12; 95% CI: -0.24, -0.01. adj β tertile 3 vs. 1: -0.16; 95% CI:
 197 -0.28, -0.04], BDE-154 (adj β tertile 3 vs. 1: -0.14; 95% CI: -0.25, -0.02), and BDE-196 (adj β
 198 tertile 3 vs. 1: -0.13; 95% CI: -0.25, -0.003).

199 Single pollutant, crude models revealed a significant decrease in TT_3 with increasing BDE-47, -
 200 100, -197, -203, -207, -208, ΣPBDEs, PCB-101, -156, -169, OCDD, ΣPCDD/Fs, and ΣPOPs
 201 (Table S13). After adjustment, TT_3 showed significant negative associations with BDE-47 (adj β
 202 tertile 3 vs. 1: -0.12; 95% CI: -0.22, -0.02), BDE-99 (adj β tertile 3 vs. 1: -0.10; 95% CI: -0.21, -
 203 0.002), BDE-100 (adj β tertile 3 vs. 1: -0.12; 95% CI: -0.22, -0.02), BDE-197 (adj β tertile 3 vs. 1:
 204 -0.11; 95% CI: -0.21, -0.01), BDE-203 (adj β tertile 3 vs. 1: -0.14; 95% CI: -0.24, -0.03), BDE-

205 207 (adj β tertile 3 vs. 1: -0.11; 95% CI: -0.20, -0.01), and OCDD (adj β tertile 3 vs. 1: -0.10; 95%
206 CI: -0.20, -0.003) (Fig. 1).

207 Single-pollutant, crude models showed a significant decrease in TrT₃ with increasing BDE-47,
208 -99, -100, -154, -183, -203, and 1,2,3,4,6,7,8-HpCDD (Table S13). In adjusted models, TrT₃ was
209 significantly inversely associated with BDE-47 (adj β tertile 2 vs. 1: -0.24; 95% CI: -0.44, -0.04),
210 BDE-99 (adj β tertile 3 vs. 1: -0.27; 95% CI: -0.48, -0.06), BDE-183 (adj β tertile 3 vs. 1: -0.21;
211 95% CI: -0.41, -0.01), and BDE-203 (adj β tertile 3 vs. 1: -0.24; 95% CI: -0.46, -0.02) (Fig. 1).



212

Beta (95% CI)213 **Fig. 1** Adjusted single pollutant models show the associations between exposure to tertiles of 36

214 POPs and THs in human breast milk. Dashed lines represent the associations of tertile 2 vs. 1 while

215 the straight lines represent the associations of tertile 3 vs. 1. The estimated effects and

216 corresponding confidence intervals (95% CI) are shown by dots and error bars, respectively.

217 3.4 Multi-pollutant model

218 *Factor analysis*

219 Using PCA on the 36 POPs, we generated five factors that sufficiently accounted for the total
 220 variance inherent in the data. Table S14 presents the factor loadings. As shown in Table 2, in the
 221 model that simultaneously included all five factors, exposure to tertile 3 of factor 3 (highly loaded
 222 with BDE-28, -47, -99, -100, -154, -183, and -197) was associated with significant decreases in
 223 TT₄ (adj β : -0.16; 95% CI: -0.29, -0.04) and TrT₃ (adj β : -0.29; 95% CI: -0.52, -0.06). However,
 224 TT₃ demonstrated a nonsignificant decrease (adj β : -0.10; 95% CI: -0.22, 0.01) in tertile 3 of factor
 225 3. Similar results were observed in single-factor models, in which TT₄ (adj β : -0.12; 95% CI: -
 226 0.23, 0.00) and TrT₃ (adj β : -0.21; 95% CI: -0.41, 0.00) were significantly negatively associated
 227 with factor 3 in tertile 3, whereas nonsignificant association was found for TT₃ (adj β : -0.10; 95%
 228 CI: -0.19, 0.00). Besides, exposure to factor 4 (highly loaded with PCB-28, -105, -118, -123, -126,
 229 1,2,3,4,6,7,8-HpCDD, OCDD, 2,3,4,7,8-PeCDF, and 1,2,3,6,7,8-HxCDF) was significantly
 230 positively associated with TT₄ (adj β : 0.13; 95% CI: 0.00, 0.26) and TrT₃ (adj β : 0.26; 95% CI:
 231 0.03, 0.49) in tertile 2.

232 **Table 2** Associations between exposure to tertiles of five factors from principal component
 233 analysis and TH levels based on single- and multiple-factor models.

	Single-factor model β (95% CI)			Multi-factor model β (95% CI)		
	TT ₄	TT ₃	TrT ₃	TT ₄	TT ₃	TrT ₃
Factor 1						
1	Reference	Reference	Reference	Reference	Reference	Reference
2	0.02 (-0.10– 0.14)	0.04 (-0.07– 0.14)	0.00 (-0.21– 0.21)	0.02 (-0.10– 0.14)	0.05 (-0.06– 0.15)	-0.02 (-0.24– 0.20)

3	0.05 (-0.08– 0.19)	0.07 (-0.04– 0.19)	0.02 (-0.22– 0.26)	0.05 (-0.09– 0.19)	0.08 (-0.04– 0.20)	-0.02 (-0.28– 0.23)
Factor 2						
1	Reference	Reference	Reference	Reference	Reference	Reference
2	0.00 (-0.12– 0.12)	-0.01 (-0.12– 0.09)	0.02 (-0.20– 0.24)	0.03 (-0.10– 0.17)	0.00 (-0.12– 0.12)	0.12 (-0.12– 0.37)
3	0.01 (-0.11– 0.14)	-0.04 (-0.15– 0.06)	-0.05 (-0.27– 0.17)	0.09 (-0.04– 0.22)	0.00 (-0.12– 0.11)	0.08 (-0.16– 0.32)
Factor 3						
1	Reference	Reference	Reference	Reference	Reference	Reference
2	-0.03 (-0.14– 0.09)	-0.02 (-0.12– 0.08)	-0.08 (-0.29– 0.12)	-0.07 (-0.19– 0.06)	-0.04 (-0.14– 0.07)	-0.16 (-0.38– 0.07)
3	-0.12 (-0.23– 0.00)*	-0.10 (-0.19– 0.00)#	-0.21 (-0.41– 0.00)*	-0.16 (-0.29– -0.04)*	-0.10 (-0.22– 0.01)#	-0.29 (-0.52– -0.06)*
Factor 4						
1	Reference	Reference	Reference	Reference	Reference	Reference
2	0.09 (-0.03– 0.21)	0.04 (-0.06– 0.15)	0.20 (-0.01– 0.42)#	0.13 (0.00– 0.26)*	0.06 (-0.06– 0.17)	0.26 (0.03– 0.49)*
3	-0.02 (-0.14– 0.10)	-0.04 (-0.14– 0.06)	0.10 (-0.11– 0.30)	-0.01 (-0.13– 0.11)	-0.03 (-0.14– 0.07)	0.12 (-0.10– 0.33)
Factor 5						
1	Reference	Reference	Reference	Reference	Reference	Reference
2	-0.05 (-0.17– 0.07)	-0.07 (-0.18– 0.03)	-0.04 (-0.26– 0.17)	-0.03 (-0.15– 0.09)	-0.05 (-0.16– 0.06)	-0.02 (-0.24– 0.21)
3	0.03 (-0.09– 0.15)	0.01 (-0.10– 0.11)	0.08 (-0.13– 0.29)	0.01 (-0.11– 0.13)	-0.01 (-0.11– 0.10)	0.05 (-0.17– 0.27)

234 All models were adjusted for maternal age, education level, parity, ethnicity, smoking, diet, and
 235 breastfeeding duration. Factor 1 loaded with PCB-114, -138, -153, -156, -157, -167, -169, -180, -
 236 189, BDE-153, and 1,2,3,6,7,8-HxCDD. Factor 2 loaded with BDE-196, -203, -206, -207, -208,
 237 and -209. Factor 3 loaded with BDE-28, -47, -99, -100, -154, -183, and -197. Factor 4 loaded with
 238 PCB-28, -105, -118, -123, -126, 1,2,3,4,6,7,8-HpCDD, OCDD, 2,3,4,7,8-PeCDF, and 1,2,3,6,7,8-
 239 HxCDF. Factor 5 loaded with PCB-52, -77, and -101.

240 ***Hierarchical clustering***

241 POPs were categorized into four groups using hierarchical clustering (Table 3 & Fig. S5). Group
 242 1 included PCB-114, -138, -153, -156, -157, -167, -169, -180, and -189; Group 2 included PCB-
 243 28, -118, -126, 1,2,3,6,7,8-HxCDD, 1,2,3,4,6,7,8-HpCDD, OCDD, 2,3,4,7,8-PeCDD, and
 244 1,2,3,6,7,8-HxCDF; Group 3 included BDE-28, -47, -99, -100, -153, -154, -183, and -197; Group
 245 4 included BDE-196, -203, -207, and -208. Multiple linear regression models demonstrated that
 246 TT₄ (adj β: -0.13; 95% CI: -0.23, -0.02) and TrT₃ (adj β: -0.21; 95% CI: -0.42, 0.00) were
 247 significantly negatively associated with Group 3 in tertile 3. Besides, TT₃ was significantly
 248 inversely associated with Group 4 in tertile 3 (adj β: -0.12; 95% CI: -0.23, -0.01).

249 **Table 3** Associations between POPs and THs. POPs were categorized based on hierarchical
 250 clustering.

	TT ₄ β (95% CI)	TT ₃ β (95% CI)	TrT ₃ β (95% CI)
Group 1			
1	Reference	Reference	Reference
2	0.01 (-0.12–0.13)	-0.02 (-0.13–0.08)	-0.04 (-0.25–0.18)
3	0.07 (-0.08–0.22)	0.08 (-0.04–0.21)	-0.02 (-0.28–0.24)
Group 2			
1	Reference	Reference	Reference
2	0.04 (-0.09–0.17)	-0.02 (-0.13–0.09)	-0.04 (-0.26–0.18)
3	0.05 (-0.10–0.19)	0.02 (-0.10–0.15)	0.17 (-0.08–0.42)
Group 3			
1	Reference	Reference	Reference
2	-0.04 (-0.16–0.08)	-0.02 (-0.12–0.09)	-0.06 (-0.27–0.15)
3	-0.13 (-0.23–0.02)*	-0.06 (-0.17–0.04)	-0.21 (-0.42–0.00)*
Group 4			
1	Reference	Reference	Reference

2	0.01 (-0.12–0.14)	-0.02 (-0.12–0.09)	-0.01 (-0.23–0.22)
3	-0.05 (-0.18–0.09)	-0.12 (-0.23–0.01)*	-0.16 (-0.39–0.07)

251 All models were adjusted for maternal age, education level, parity, ethnicity, smoking, diet, and
 252 breastfeeding duration. Group 1 included PCB-114, -138, -153, -156, -157, -167, -169, -180, and
 253 -189. Group 2 included PCB-28, -118, -126, 1,2,3,6,7,8-HxCDD, 1,2,3,4,6,7,8-HpCDD, OCDD,
 254 2,3,4,7,8-PeCDD, and 1,2,3,6,7,8-HxCDF. Group 3 included BDE-28, 47, -99, -100, -153, -154, -
 255 183, and -197. Group 4 included BDE-196, -203, -207, and -208.

256 3.5 Sensitivity analysis

257 Similar results were obtained when POPs were modeled in ng/g lw or in ng/L milk. The models
 258 kept robust when BMI was further controlled. As shown in Table S15, in multifactor model,
 259 exposure to tertile 3 of factor 3 was significantly inversely associated with TT₄ (adj β: -0.16; 95%
 260 CI: -0.29, -0.03) and TrT₃ (adj β: -0.30; 95% CI: -0.53, -0.07). Similar results were observed in
 261 single-factor models, in which TT₄ (adj β: -0.12; 95% CI: -0.24, 0.00) and TrT₃ (adj β: -0.21; 95%
 262 CI: -0.41, 0.00) showed significant negative associations with factor 3 in tertile 3. Exposure to
 263 factor 4 was significantly positively associated with TT₄ (adj β: 0.15; 95% CI: 0.01, 0.29) and TrT₃
 264 (adj β: 0.26; 95% CI: 0.02, 0.51) in tertile 2.

265 4. Discussion

266 THs were quantified in human breast milk using LC-MS/MS for the first time. TT₄ and TT₃
 267 levels measured here were similar to a previous report measured in preterm breast milk using
 268 radioimmunoassay (RIA), while higher TT₄ was found in term breast milk (see Table S3)²⁵. This
 269 is probably due to the differences in the time of sampling. However, our results might be more
 270 reliable and accurate since IA technology is prone to nonspecific interferences. Additionally, due
 271 to their low concentrations, THs in human breast milk is not an adequate source for infants with
 272 congenital hypothyroidism²⁵.

273 Compared with BAMBI data from 2007–2008, this cohort had substantially lower exposure to
 274 POPs, with PCDD/Fs and dl-PCBs decreased for 52% and 44%, respectively³⁰. Besides, the POP

275 levels were generally lower compared with values reported in other regions. For example, PBDE
276 and PCB levels measured here were lower than those reported in the North America ³, while
277 PCDD/Fs were lower than those measured in India ⁴⁰. Therefore, our study represents a low-
278 exposure population.

279 **4.1 Associations of PBDEs with THs**

280 We observed the highest thyroid-disrupting potencies for PBDEs among the POPs examined.
281 Single pollutant models revealed significant inverse associations of THs with BDE-47, -99, -100,
282 -154, -183, -196, -197, -203, and -207. Similar and robust results were obtained in multipollutant
283 models using PCA and hierarchical clustering, which proved a significant association of increasing
284 PBDEs (including BDE-28, -47, -99, -100, -154, -183, and -197) with depressed TT₄ and TrT₃.
285 These findings are consistent with previous epidemiologic studies ^{13, 20, 41}. Animals studies using
286 rats ⁴², fish ⁴³, kestrels ⁴⁴, and minks ⁴⁵ also proved decreased THs following PBDE exposure.
287 Putative mechanisms include the interference of PBDEs with TH transport and metabolism. For
288 example, *in vitro* studies demonstrated that lower-brominated OH-PBDEs are structurally similar
289 to THs and can competitively bind with TR ^{46, 47}. Enhanced TH metabolism, in combination with
290 elevated cytochrome P450 enzymes 2B (CYP2B) expression, deiodinase I (DIO1) enzyme
291 activity, as well as the gene expression of Cyp2b1/2, dio1, and hepatic efflux transporters were
292 observed in rats following DE-71 (predominately composed of BDE-47, 99, -100, -153, and -154)
293 exposure ⁴⁸. In addition, PBDEs may disrupt the HPT axis by interfering with the TSH β expression
294 ⁴³.

295 In contrast, others observed positive or nonsignificant associations, and the associations might
296 be differed by sex ^{15, 17, 49}. The inconsistency is probably the results of random error given the
297 intraindividual and inter-individual variability in TH set-points. For example, Stapleton *et al.* ¹⁵

298 and Zota *et al.* ⁴⁹ employed serum samples from women during pregnancy when marked
299 fluctuations in HPT axis homeostasis occur ⁵⁰. Our samples were collected post-partum when HPT
300 axis tends to be more stable ⁵⁰.

301 Another possibility is that the relationship between PBDEs and THs may vary by exposure level.
302 THs act at quite low concentrations (free serum T₄ (FT₄) level: 8–20 ng/L ⁵¹) while low-dose
303 effects and non-monotonic responses are remarkably common in studies of EDC (low-dose cutoff
304 of BDE-99: 0.3 mg/kg d) ²². Abdelouahab *et al.* observed significant decreases in TT₄ and TT₃ in
305 lambs following low-dose exposure of BDE-47 ⁵². A meta-analysis suggested that the relationship
306 between THs and PBDEs might be an inverted U-shape curve ¹⁹. Inverse association was found in
307 populations with low- ^{23, 41} and high-dose PBDE exposures ²⁰, while positive association was
308 reported in a population exposed with middle-level PBDEs ⁵³. This is generally in agreement with
309 our findings because of the low exposure levels of POPs in this study. Our results proved negative
310 associations of low level PBDE exposure with THs. However, caution should be taken when
311 comparing our results to previous findings, given variation in study design, population
312 susceptibility, exposure level, biomarker measurement, as well as sample type which contain
313 different contents of lipid, TH-binding proteins, and enzymes. Further studies are warranted to
314 clarify the underlying mechanisms.

315 **4.2 Associations of PCBs with THs**

316 DI-PCBs and dioxins may upregulate UDPGA by binding with aryl hydrocarbon receptor
317 (AhR), leading to enhanced excretion of T₄. Non-dl-PCBs, however, may interfere with the
318 activities of CYP enzymes (i.e., CYP 2B1 and CYP 3A1) which may also reduce circulating T₄ ⁵⁴,
319 ⁵⁵. A substantial body of animal and epidemiologic studies have reported decreased THs with
320 increasing exposure of PCBs, despite the literature is inconsistent ^{18, 56-58}. Langer *et al.* reported

321 negative associations of FT₄ and TT₃ with PCBs at low exposure levels (serum level: < 530 ng/g
322 lw), but positive relations at higher levels ⁵⁹. A recent study also observed inverse associations
323 between low-dose PCBs and TT₃, Free T₃, and FT₄ in a Chinese population ⁸. These findings are
324 in line with our results of single-pollutant crude models, which revealed significant inverse
325 associations of PCB-101, -156, and -169 with THs, although they were insignificant after
326 adjustment. In contrast, multipollutant analysis revealed significant positive associations of TT₄
327 and TrT₃ with Factor 4, which was mainly loaded with PCBs and PCDD/Fs. This probably
328 indicates that the relationship between PCBs and THs can be influenced by PCDD/Fs. Similarly,
329 in a population highly exposed with polybrominated biphenyls (PBBs) from Michigan, serum
330 PCBs were found to be positively associated with THs ³⁶.

331 **4.3 Associations of PCDD/Fs and PBDD/Fs with THs**

332 1,2,3,6,7,8-HxCDD, OCDD, and 1,2,3,4,6,7,8-HpCDD showed significant inverse associations
333 with TT₄ and TT₃ in crude single-pollutant models. After adjustment, only the association of
334 OCDD with TT₃ remained. Our previous study observed that placental TT₄ was significantly
335 inversely associated with 2,3,7,8-TCDD, but significantly positively associated with 1,2,3,6,7,8-
336 HpCDF, TT₃ was significantly positively associated with 2,3,7,8-TCDF and 1,2,3,7,8-PeCDF,
337 while TrT₃ was significantly positively associated with 1,2,3,7,8-PeCDF and 2,3,4,6,7,8-HxCDF
338 ¹⁸. Other studies also reported inconsistent results ^{60,61}. The probable reasons include the presence
339 of different pollutant mixtures, varying timing of sampling, exposure level, and uncontrolled bias.

340 PBDD/Fs are brominated dioxins found as impurities of PBDEs and formed during the
341 incineration and degradation of brominated chemicals ⁶². Similar to previous reports ^{63, 64}, we
342 observed low DFs for PBDD/Fs. Therefore, the thyroid-disrupting properties of these chemicals
343 were not assessed here.

344 **4.4 Strengths and limitations**

345 Our study has several strengths: (1) THs in human breast milk was measured for the first time
346 to investigate the thyroid-disrupting effects of POPs. Compared with serum, we obtained much
347 higher detection frequencies for POPs. This is because it is easier to obtain large sample amount,
348 as well as the high lipid content in milk; (2) A wide variety of POPs and potential confounders
349 were measured and included in the statistical analysis, which can provide an overview of possible
350 relationships between POPs and THs. Furthermore, multipollutant approaches enabled us to
351 evaluate the integrated effects of POP mixtures. This is critical because many POP congeners show
352 similar chemical and biological properties; (3) The low exposure of POPs in this population
353 enabled us to estimate the thyroid-disrupting effects of POPs in background low-exposure
354 population. Our study also has certain limitations. For example, we did not measure serum TH
355 levels of infants that are more susceptible to thyroid disruption. Besides, with human breast milk
356 we can only assess the maternal TH homeostasis, and therefore we are not able to estimate the sex-
357 specific associations between POPs and THs. Additional limitations include the lack of thyroid-
358 binding protein levels and the OH-PBDEs and OH-PCBs, which in general show higher potencies
359 of thyroid-disruption. Lastly, this study was limited by the small sample size which may reduce
360 the statistical power.

361 **ASSOCIATED CONTENT**

362 **Supporting Information**

363 The following files are available free of charge.

364 Analytical methods of THs in human breast milk, comparison of POPs in different populations,
365 correlations between biomarkers, crude single-pollutant models, factor loadings, and directed
366 acyclic graph (PDF).

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371 **Notes**

372 The authors declare no competing financial interest.

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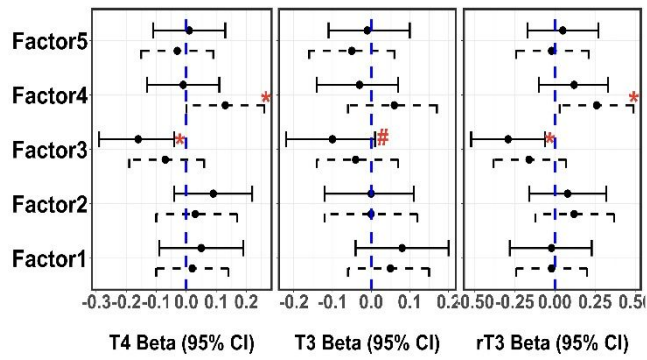
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590 Table of Contents (TOC) Graphic



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