Elevated levels of interleukin-6 and CRP predict the need for mechanical ventilation in COVID-19

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1	Elevated levels of interleukin-6 and CRP predict the need for mechanical ventilation in
2	COVID-19
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#### 31 Author contributions

T.H. and T.W. conceived and designed the study. T.H., C.A., M. K. and T.W. were responsible for clinical care and collected patient data. J.C.H. was responsible for the ethical approval of the study. Statistical analysis was conducted by V.J.. M.v.B.-B. supervised all aspects of the study. B.L. corrected and helped write the manuscript and added important aspects to the analysis. T.H. and T.W. wrote the first draft. All authors contributed to data interpretation, critical revision of the manuscript and approved the final version of the manuscript.

#### 39 Competing interests

B. L. reports grants and personal fees from Sanofi, AstraZeneca, and Teva; reports personal
fees from Cipla, Glenmark, and Lupin; reports grants, personal fees, and other from Chiesi,
outside the submitted work; and reports that his son is an employee of AstraZeneca.

M.v.B-B. is the local principal investigator of the currently conducted COVACTA-Trial (A
Study to Evaluate the Safety and Efficacy of Tocilizumab in Patients With Severe COVID-19
Pneumonia; NCT04320615, Sponsor: Hoffmann-La Roche). He has previously received
honoraria and research funding from Hoffman-La Roche unrelated to this project.

- 47 M. K received speakers' fees from BioMerieux and served on the advisory board of
- 48 BioMerieux.
- 49 The other authors declare no conflict of interest.
- 50

Journal Prevention

#### 51 Abstract

Background: COVID-19 can manifest as a viral induced hyperinflammation with multi-organ involvement. Such patients often experience rapid deterioration and need for mechanical ventilation. Currently, no prospectively validated biomarker of impending respiratory failure is available.

56 **Objective**: We aimed to identify and prospectively validate biomarkers that allow the 57 identification of patients in need of impending mechanical ventilation.

58 **Methods**: Patients with COVID-19 hospitalized from February 29<sup>th</sup> to April 09<sup>th</sup>, 2020 were 59 analyzed for baseline clinical and laboratory findings at admission and during the disease. 60 Data from 89 evaluable patients were available for the purpose of analysis comprising an 61 initial evaluation cohort (n=40) followed by a temporally separated validation cohort (n=49).

Results: We identified markers of inflammation, LDH and creatinine as most predictive 62 variables of respiratory failure in the evaluation cohort. Maximal interleukin-6 (IL-6) levels 63 64 before intubation showed the strongest association with the need of mechanical ventilation followed by maximal CRP. Respective AUC values for IL-6 and CRP in the evaluation cohort 65 66 were 0.97 and 0.86 and similar in the validation cohort 0.90 and 0.83. The calculated optimal cutoff values in the course of disease from the evaluation cohort (IL-6> 80 pg/ml and CRP> 67 97 mg/l) both correctly classified 80% of patients in the validation cohort regarding their risk 68 69 of respiratory failure.

Conclusion: Maximal levels of IL-6 followed by CRP were highly predictive of the need for
 mechanical ventilation. This suggests the possibility of using IL-6 or CRP levels to guide
 escalation of treatment in patients with COVID-19 related hyperinflammatory syndrome.

Clinical Implications: IL-6 followed by CRP strongly predicted patients at risk of respiratory
 deterioration and might be pivotal for risk-adapted escalation of treatment.

Capsule summary: We studied laboratory parameters as predictors of impending
respiratory failure in COVID-19. Maximum levels of interleukin-6 over the course of disease,
followed by CRP, were the best predictors of respiratory failure in two separate cohorts.

Key words: Interleukin-6, IL-6, CRP, COVID-19, respiratory failure, mechanical ventilation,
prediction, hyperinflammation

Abbreviations: COVID-19: Coronavirus Disease 2019; SARS-CoV-2: Severe Acute 81 Respiratory Syndrome coronavirus 2; SARS: severe acute respiratory syndrome; H7N9: 82 avian-origin influenza; H1N1: influenza A; BAL: Bronchoalveolar lavage; ROC: Receiver 83 operating characteristic; AUC: Area under the curve; CI: Confidence interval; BMI: Body 84 mass index; CT: Computed Tomography; CRP: C-Reactive Protein; WBC: White blood cell 85 count; LDH: Lactate Dehydrogenase; PCT: Procalcitonin; IL6: Interleukin-6; gSOFA score: 86 quick sequential organ failure assessment score - predicts mortality in sepsis; CURB-65 87 score: predicts mortality in community-acquired pneumonia; MuLBSTA score: predicts 88 mortality in patients with viral pneumonia; q-values represent the Benjamini-Hochberg 89 adjusted p-values 90

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#### 93 Introduction

The pandemic Coronavirus-disease 19 (COVID-19) is characterized by a highly variable course. While most patients experience only mild symptoms, a relevant proportion develops severe disease progression up to respiratory failure. Interestingly, many patients do not show signs of respiratory distress, despite severe hypoxemia in blood gas analysis <sup>1</sup>. About 5% of patients require intensive care including mechanical ventilation <sup>2, 3</sup>.

99 Recently published large retrospective analyses provide a detailed characterization of 100 COVID-19 and identify variables associated with disease severity and high mortality <sup>4, 5</sup>. One 101 of the largest studies so far shows that age, quick sequential organ failure assessment score 102 (qSOFA score) and D-Dimer correlate with in-hospital death in a multivariate analysis <sup>2</sup>. 103 Another group showed a correlation of obesity and increased inflammatory markers in the 104 blood with respiratory failure <sup>6</sup>.

In many aspects, severe COVID-19 may be regarded as a viral induced hyperinflammatory condition with multi-organ involvement due to a cytokine cascade <sup>7</sup>. Of these various cytokines, the presence of raised circulating levels of interleukin-6 (IL-6) appears to be key and is closely connected to disease severity not only in COVID-19 <sup>8</sup> but also in avian-origin H7N9 influenza infections <sup>9</sup> and the common seasonal H1N1 influenza A <sup>10</sup>.

110 While these studies identify the correlation of parameters with disease severity, prospective 111 factors predicting impending deterioration of patients are not yet established. The broad 112 spectrum of the disease courses and silent hypoxia make identification of patients at risk 113 difficult. We aimed to identify variables that allow the prediction of COVID-19 patients with a 114 high risk of respiratory failure.

115

## 117 Methods

#### 118 Patients and study design

All patients with PCR proven COVID-19 hospitalized at our institution from February 29<sup>th</sup> to April 09<sup>th</sup>, 2020 (n=115) were screened and analyzed for baseline clinical and laboratory findings. In total, 26 patients were excluded from the study and the depicted cohort consisted of 89 patients (Table 1). Patients with palliative treatment (n=3) or hospitalization due to other medical reasons and nosocomial Sars-CoV2-infection on the ward (n=13) were excluded from this study. Additionally, patients already mechanically ventilated at admission (n=8) and those receiving anti-IL-6 antibody treatment (n=2) were excluded (Figure 1).

Of the 89 evaluable patients, 40 were part of an initial evaluation cohort hospitalized from February 29<sup>th</sup> to March 27<sup>th</sup>, 2020 (Supplementary Table E1). This cohort was used to identify predictive markers of respiratory failure.

Following an interim analysis of the initial evaluation cohort<sup>11</sup>, we performed a power analysis 129 130 to estimate the number of patients needed to validate our findings. Assuming the need of mechanical ventilation to be 20% in the validation cohort and the risks for mechanical 131 132 ventilation to be 70% and 20% in the high-risk and the low-risk group, respectively, the total sample size for a two-sided test was determined to be 40. We defined an additional safety 133 134 margin of 10%. This subsequent validation cohort consisted of patients hospitalized from March 27<sup>th</sup> to April 09<sup>th</sup>, 2020 (n=49) (Supplementary Table E2). Follow up for all patients 135 was complete through April 12<sup>th</sup>, 2020. A comparison of both cohorts is shown in 136 Supplementary Table E3. 137

Use of compassionate medication was low in the study cohort before mechanical ventilation
(5 patients received lopinavir/ritonavir, 8 patients received hydroxychloroquine).

140 Decision on endotracheal intubation was made following internationally accepted 141 recommendations (PaO2/FiO2 <150mmHg or <200mmHg in case of anticipated difficult 142 airway)<sup>12</sup>.

Patients are part of the COVID-19 Registry of the Ludwig-Maximilian-University Hospital Munich (CORKUM). Patient data were anonymized for analysis and the study was approved by the local ethics committee (Ethics committee of the LMU Munich, No: 20-245).

#### 146 **IL-6 and CRP measures**

The fully automated Elecsys® system on a cobas e801 platform (Roche Diagnostics, Switzerland) was used to measure single levels of IL-6, as described previously <sup>13, 14</sup>. The Elecsys® IL-6 immunoassay has been standardized against the NIBSC 1st IS 89/548 Standard. CRP values were measured on a cobas c702 platform using the Tina-quant® C-Reactive Protein assay (Roche Diagnostics, Switzerland).

## 152 Statistical analysis

All variables with less than 50% of missing data in the initial cohort were tested for the 153 association with respiratory failure. Categorical variables were tested with the  $\chi^2$  test, and 154 numerical variables with the Mann-Whitney U test. When appropriate, a paired test was 155 156 performed. All tests were two-sided. The p-values were adjusted for multiple testing with the Benjamini-Hochberg-method to avoid inflating the alpha error. An adjusted p-value (q-value) 157 158 of  $\leq 0.05$  was considered significant. We constructed receiver operating characteristic (ROC) curves and calculated the area under the curve (AUC) to compare the predictive ability of 159 160 continuous variables. The AUC can be interpreted as the probability that the predictor's value for a randomly chosen patient requiring intubation will be higher than its value for a randomly 161 chosen patient not requiring intubation. The optimal cut off was defined as the one 162 maximizing the Youden's Index <sup>15</sup>. Statistical analyses were performed using the R software 163 package (version 3.6.2). Figures were drawn using Graphpad Prism® (Version 6.0). 164

165

### 167 Results

#### 168 Initial identification of IL-6 and CRP as strongest predictors of respiratory failure

To initially evaluate predictors of respiratory failure, 40 patients with confirmed COVID-19 were recruited from February 29<sup>th</sup> to March 27<sup>th</sup>, 2020 and served as an evaluation cohort (Figure 1). Thirteen (32.5%) patients deteriorated during hospitalization and required mechanical ventilation. The time from hospital admission to intubation varied from less than two hours to 9 days (median 2 days). Patients requiring mechanical ventilation did not differ in age, comorbidities, radiological findings, respiratory rate or qSofa score (Supplementary Table E1).

Heart rate, markers of inflammation, LDH and creatinine at admission were significantly associated with respiratory failure (Supplementary Table E1). Elevated IL-6 showed the strongest association with the need for mechanical ventilation (Figure 2A,  $p=1.2x10^{-5}$ ).

In addition to values at first assessment, follow-up data were available for laboratory 179 180 variables. These follow-up data were used to test if there are critical laboratory values that are associated with respiratory failure once they have been reached during disease course. 181 182 For each patient, we assessed the maximum level of each parameter during disease (for patients requiring ventilation, only values before intubation were used). The maximal values 183 184 were correlated with respiratory failure (Table 2). Maximal IL-6 levels predicted respiratory failure with highest accuracy (Figure 2, AUC=0.97, CI [0.93, 1.0]), followed by CRP (Figure 3 185 AUC=0.86, CI [0.74, 0.98]) and creatinine (AUC=0.85, CI [0.74, 0.97]). The optimal cutoff for 186 maximal IL-6 was 80 pg/ml. After reaching an IL-6 value of 80 pg/ml, the median time to 187 mechanical ventilation was 1.5 days (range 0-4 days). The optimal cutoff for maximal CRP 188 was 97 mg/l, with the median time to mechanical ventilation of 0 days after reaching the 189 cutoff (range 0-4 days). 190

## 191 Prospective validation of calculated cutoffs for IL-6 and CRP

A cohort of 40 patients was estimated to have an adequate power to validate our findings 192 (see Methods). The validation cohort prospectively recruited 49 patients from March 27<sup>th</sup> to 193 194 April 09<sup>th</sup>, 2020, of which 19 (39%) required mechanical ventilation. As in the initial cohort, creatinine, LDH, and several markers of inflammation were significantly elevated in patients 195 requiring intubation (Table 2 and Supplementary Table E2). Again, IL-6 at assessment was 196 strongly associated with respiratory failure (Figure 2B), and maximal IL-6 was the best 197 predictor of future respiratory failure among all parameters (Figure 2D, AUC 0.90, CI [0.81, 198 0.98], Table 2). CRP values at initial assessment were significantly associated with 199 respiratory failure (Figure 2F and Figure 3 AUC=0.86, CI [0.75, 0.96]). Follow-up values of 200 CRP during the disease course did not improve the prediction of respiratory failure in the 201 validation cohort (Table 2, AUC=0.83, CI [0.72, 0.95]). 202

203 To validate our findings from the initial cohort, we analyzed the number of patients correctly classified regarding their need of mechanical respiratory support by the determined cutoffs of 204 205 IL-6 and CRP at presentation and in the course of disease (Table 3). At presentation, IL-6 >35 pg/ml as well as CRP >32.5 mg/l showed high sensitivity to detect patients at risk for 206 respiratory failure (84% and 95%) with moderate specificity (63% for both parameters). 207 Measuring IL-6 and CRP values in the course of disease (cutoffs 80 pg/ml and 97 mg/l) 208 209 increased the specificity for both parameters (83% and 77%) accompanied with a decrease in sensitivity (74% vs. 84%). In detail, nineteen (39%) patients exceeded the calculated 210 maximal IL-6 cutoff (>80 pg/ml) in the validation cohort, compared to 23 (47%) patients 211 exceeding the CRP cutoff (>97mg/l). Of these patients, 74% and 70% were correctly 212 classified by IL-6 and CRP, respectively, as being at risk for respiratory failure (positive 213 214 predictive value). Of the 30 patients with values below the IL-6 cutoff, 83% did not require mechanical ventilation, while this was the case for 88% of the 26 patients remaining below 215 216 the CRP cutoff of 97 mg/l (negative predictive value). In total, the calculated cutoffs for 217 maximal IL-6 and CRP both correctly classified 80% of patients regarding their risk of respiratory failure (Table 3), while values at assessment show poorer predictor properties 218 owing to the moderate specificity (correct classification of 71% for IL-6 and 76% for CRP) 219

Taken together, while both values have a strong sensitivity at assessment, specificity is gained when examining values in the course of disease. The risk ratios for the cutoffs of IL-6 and CRP were 4.4 and 6.0 in the validation cohort, with corresponding p-values of 0.00022 and 0.00011. The optimal cut point in the validation cohort was slightly lower for IL-6 (60 pg/ml) and identical for CRP (97 mg/l).

#### 225 Predictive values of the combined cohort

To further evaluate positive and negative predictive values (PPV/NPV) of IL-6 and CRP we 226 combined the two cohorts (Table 1). We calculated predictive values across the range of all 227 228 possible cutoffs. The PPV of CRP was consistently lower compared to IL-6 in the overall study cohort (Figure 4). In other words, increased CRP misclassified more patients as being 229 at risk for respiratory failure than IL-6. However, the predictive values strongly depend on the 230 231 selected cutoff (Figure 4). For cutoffs <50 pg/ml for IL-6 and <40 mg/l for CRP (dotted line), 232 the risk of intubation for patients with sub-threshold levels is roughly zero, while patients with levels above these values show a dramatic increase in the risk of respiratory failure. The risk 233 for respiratory failure in patients with IL-6 levels exceeding 210 pg/ml was 100% (dashed 234 line). The NPV of IL-6 and CRP parameters was comparable. In the combined cohort, the 235 optimal threshold value (maximal Youden index<sup>15</sup>) is highest at 65 pg/ml for IL-6 and for CRP 236 at 97 mg/l (corresponding risk ratio of 18.1 and 6.9). 237

Furthermore, we analyzed the time lag from reaching the cutoff values to intubation in the combined cohort. Patients reached the cutoff of IL-6 (>65 ng/ml) and CRP (>97 mg/l) at a median of 23.2 and 15.7 hours before intubation, resulting in a significant time difference between the two values of 7.5 hours in favor for IL-6 (Figure 5; p=0.014).

242

#### 244 Discussion

245 Our study in hospitalized patients with COVID-19 has provided three key findings: First, circulating levels of IL-6 as well as CRP were highly predictive of the need for invasive 246 ventilation, with corresponding AUC values of 0.97 and 0.90 for IL-6 and 0.86 and 0.83 for 247 CRP in the first and the second cohorts, respectively. Secondly, we defined cutoffs for IL-6 248 249 (at presentation >35 pg/ml; maximal value >80 pg/ml) and CRP (at presentation >32.5 mg/l; maximal value >97 mg/l) in the evaluation cohort. Cutoff values at assessment correctly 250 classified 71% (for IL-6) and 76% (for CRP) of patients in the validation cohort with a further 251 increase when measuring maximal values in the course of disease (80% for both 252 253 parameters). Thirdly, elevated IL-6 levels in the course of disease predicted respiratory failure significantly earlier than CRP (23.2 vs. 15.7 hours). Therefore, IL-6 and CRP are 254 255 useful markers that predict impending respiratory failure with high accuracy and can help physicians correctly allocate patients who might benefit from early treatment escalation, for 256 257 example using anti-cytokine strategies. We believe that having these data reproduced across the two separate cohorts enhances the strength of our conclusions. It is important to note 258 that the commercial diagnostic IL-6 assay used in our study allows the measurement of II-6 259 in a comparable time scale as CRP. Since it uses the broadly available Cobas platform it can 260 261 be implemented in most laboratories.

262 Our study also has several limitations. It is still unclear whether elevated inflammatory markers merely represent an epiphenomenon or a causal pathogenic element of severe 263 COVID-19<sup>16</sup>. It is likely that elevated IL-6 reflects the cytokine mediated hyperinflammatory 264 state as evidenced by the similarly predictive values for CRP. Further, even though IL-6 and 265 CRP levels are significantly elevated in patients requiring ventilation, they are relatively low 266 compared to levels observed in patients with septic shock <sup>17</sup>. However, earlier studies in 267 268 severe acute respiratory syndrome (SARS) or H7N9 influenza patients show that inflammatory cytokines are highly expressed in lung tissues. Autopsy reports from SARS 269 patients showed a high amount of inflammatory cytokines in cells expressing angiotensin-270

converting enzyme 2<sup>18</sup>, the functional receptor for SARS-CoV and in even higher affinity for 271 SARS-CoV2<sup>19</sup>. Bronchoalveolar lavage (BAL) in H7N9 influenza patients showed 10<sup>3</sup> times 272 273 higher concentrations of different cytokines including IL-6 compared to plasma levels, hinting towards a massively increased local concentration of inflammatory cytokines in the diseased 274 lung<sup>9</sup>. Recent preprints provide detailed single cell RNA-sequencing data from immune cells 275 in peripheral blood as well as BAL from COVID-19 patients. The authors report that 276 peripheral monocytes did not substantially express proinflammatory cytokines <sup>20</sup>, while there 277 was high expression in monocyte derived macrophages in BAL<sup>21</sup>. Taken together, these 278 data possibly suggest that circulating levels of IL-6 might be a putative surrogate for the 279 burden of lung tissue damage and provide a "window" into the lung <sup>9</sup>. 280

IL-6 and CRP have been associated with severity of COVID-19 (in most cases defined by the 281 Chinese National Health Commission) and mortality before <sup>22-24</sup>. However, to our knowledge 282 our study is the first to demonstrate a prospective prediction of the end point "mechanical 283 284 ventilation", which is of high clinical relevance not only for patient treatment but also for resource planning. Very recent publications provide additional data that strengthen the role of 285 IL-6 and CRP in COVID-19 as predictive markers <sup>22, 23</sup>. Unfortunately, these studies did not 286 include a prospective validation cohort and sometimes did not mention analysis platforms<sup>22</sup>. 287 A further difference between our and other studies is the dramatic discrepancy in mortality of 288 severely diseased patients. We are not able to analyze mortality as an end point because 289 only two patients had died until April 12<sup>th</sup>. This number has only increased by one until May 290 6<sup>th</sup> (overall mortality 3.4%). While still some patients are in critical condition and the mortality 291 rate in our cohort is likely to increase in the next weeks it will be significantly below those 292 reported. We can only speculate about the reasons for this huge difference but argue that 293 overwhelmed hospitals and patient selection might have contributed to the increased 294 mortality observed in other studies. As we did not perform sequential CT-scans after 24-48 295 296 hours in our patients due to radiation hygiene, we are not able to precisely calculate severity of COVID-19 according the Chinese National Health Commission classification to compare 297 our patient cohort to the cohorts of the mentioned studies. However, our validation cohort at 298

least exists of 63% of severe patients due to the available parameters (2% with mild and
35% with moderate symptoms), which exceeds the recently published cohorts <sup>22, 23</sup>.

Since the start of the pandemic, hundreds of research articles on COVID-19 have been 301 published <sup>25</sup>. To our knowledge, we report the first predictive marker for respiratory failure 302 303 that was prospectively validated in an independent cohort. Although our sample sizes were 304 small, the large difference in risk for respiratory failure between the high-risk and the low-risk group made it possible to successfully validate our findings. Interestingly, a study of 134 305 patients with avian-origin H7N9 influenza in 2013 also showed a strong correlation of IL-6 306 and disease severity. In analogy to our findings, this study reports that IL-6 plasma levels 307 308 >80 pg/ml were found in all patients with lethal outcome compared to only 8.3% in surviving patients<sup>9</sup>. The combined cohort (n=89) produced an only slightly lower cutoff for IL-6 (65 309 310 pg/ml) while the cutoff for CRP levels remained the same at 97 mg/l when calculated from the combined cohort. However, even the combined sample size is probably too small to 311 312 determine an optimal cutoff value. Furthermore, the acceptable proportion of falsely identified low-risk patients, and therefore the set threshold, is largely dictated by the availability of 313 health care resources. Future prospective studies with larger sample sizes are needed to 314 formally address this issue. We want to stress that IL-6 and CRP should be used as a 315 316 predictor not an indication for invasive respiratory support, as mechanical ventilation per se has several unintended adverse consequences and may support inflammation of distal 317 airways in COVID-19 patients. 318

Immunologically, CRP and IL-6 are closely intertwined. IL-6 is known to induce gene expression and release of CRP from the liver <sup>26, 27</sup> and also from immune cells <sup>28</sup>. A functional connection has been shown in different trials using IL-6 inhibition, in which CRP-levels rapidly normalized after blocking IL-6 <sup>29</sup>. In analogy, we found that IL-6 levels predicted respiratory failure significantly earlier than CRP-levels, which is essential for a predictive marker. While inhibition of inflammatory pathways represents a promising approach to treat hyperinflammatory COVID-19 patients, inhibition of IL-6 could be detrimental in the immune

response to virus-induced pneumonias <sup>30, 31</sup>. Thus, our study does not facilitate any recommendations for or against IL-6 inhibition. Ongoing randomized controlled clinical trials of IL-6-antibodies in the treatment of COVID-19 will shed light on this question (e.g. NCT04320615 and NCT04331795). More importantly, in times of missing established therapeutic options, best supportive care is essential <sup>32</sup>.

In summary, we were able to validate our finding that IL-6 and CRP levels serve as strong predictors of patients in need of ventilator support. In the current situation with overwhelmed intensive care units and overcrowded emergency rooms, correct identification of patients in need of intensive care is crucial. Assessing these parameters to identify patients at risk of respiratory failure at an early stage might be helpful for triage planning and timely allocation of critically ill patients as well as a guide to escalation of treatment strategies in COVID-19 patients.

338

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		Journal Pre-proof								
347	7 References									
348	1.	Xie J, Tong Z, Guan X, Du B, Qiu H, Slutsky AS. Critical care crisis and some								
349		recommendations during the COVID-19 epidemic in China. Intensive Care Med 2020.								
350	2.	Zhou F, Yu T, Du R, Fan G, Liu Y, Liu Z, et al. Clinical course and risk factors for								
351		mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort								
352		study. Lancet 2020; 395:1054-62.								
353	3.	Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, et al. Clinical Characteristics of 138								
354		Hospitalized Patients With 2019 Novel Coronavirus-Infected Pneumonia in Wuhan,								
355		China. JAMA 2020.								
356	4.	Grasselli G, Zangrillo A, Zanella A, Antonelli M, Cabrini L, Castelli A, et al. Baseline								
357		Characteristics and Outcomes of 1591 Patients Infected With SARS-CoV-2 Admitted								
358		to ICUs of the Lombardy Region, Italy. JAMA 2020.								
359	5.	Petrilli CM, Jones SA, Yang J, Rajagopalan H, O'Donnell LF, Chernyak Y, et al.								
360		Factors associated with hospitalization and critical illness among 4,103 patients with								
361		COVID-19 disease in New York City. medRxiv 2020.								
362	6.	Dreher M, Kersten A, Bickenbach J, Balfanz P, Hartmann B, Cornelissen C, et al.								
363		Charakteristik von 50 hospitalisierten COVID-19-Patienten mit und ohne ARDS.								
364		Dtsch Arztebl International 2020; 117:271-8.								
365	7.	Lipworth B, Chan R, Lipworth S, RuiWen Kuo C. Weathering the Cytokine Storm in								
366		Susceptible Patients with Severe SARS-CoV-2 Infection. J Allergy Clin Immunol Pract								
367		2020.								
368	8.	Chen X, Zhao B, Qu Y, Chen Y, Xiong J, Feng Y, et al. Detectable serum SARS-CoV-								
369		2 viral load (RNAaemia) is closely correlated with drastically elevated interleukin 6 (IL-								
370		6) level in critically ill COVID-19 patients. Clin Infect Dis 2020.								
371	9.	Wang Z, Zhang A, Wan Y, Liu X, Qiu C, Xi X, et al. Early hypercytokinemia is								
372		associated with interferon-induced transmembrane protein-3 dysfunction and								
373		predictive of fatal H7N9 infection. Proc Natl Acad Sci U S A 2014; 111:769-74.								

		Journal Pre-proof
374	10.	Hagau N, Slavcovici A, Gonganau DN, Oltean S, Dirzu DS, Brezoszki ES, et al.
375		Clinical aspects and cytokine response in severe H1N1 influenza A virus infection.
376		Crit Care 2010; 14:R203.
377	11.	Herold T, Jurinovic V, Arnreich C, Hellmuth JC, von Bergwelt-Baildon M, Klein M, et
378		al. Level of IL-6 predicts respiratory failure in hospitalized symptomatic COVID-19
379		patients. medRxiv 2020.
380	12.	Bellani G, Laffey JG, Pham T, Fan E, Brochard L, Esteban A, et al. Epidemiology,
381		Patterns of Care, and Mortality for Patients With Acute Respiratory Distress
382		Syndrome in Intensive Care Units in 50 Countries. JAMA 2016; 315:788-800.
383	13.	Fischer SK, Williams K, Wang L, Capio E, Briman M. Development of an IL-6 point-of-
384		care assay: utility for real-time monitoring and management of cytokine release
385		syndrome and sepsis. Bioanalysis 2019; 11:1777-85.
386	14.	Jekarl DW, Lee SY, Lee J, Park YJ, Kim Y, Park JH, et al. Procalcitonin as a
387		diagnostic marker and IL-6 as a prognostic marker for sepsis. Diagn Microbiol Infect
388		Dis 2013; 75:342-7.
389	15.	Youden WJ. Index for rating diagnostic tests. Cancer 1950; 3:32-5.
390	16.	Hirano T, Murakami M. COVID-19: a new virus, but an old cytokine release
391		syndrome. Immunity 2020; pre-proof print.
392	17.	Surbatovic M, Popovic N, Vojvodic D, Milosevic I, Acimovic G, Stojicic M, et al.
393		Cytokine profile in severe Gram-positive and Gram-negative abdominal sepsis. Sci
394		Rep 2015; 5:11355.
395	18.	He L, Ding Y, Zhang Q, Che X, He Y, Shen H, et al. Expression of elevated levels of
396		pro-inflammatory cytokines in SARS-CoV-infected ACE2+ cells in SARS patients:
397		relation to the acute lung injury and pathogenesis of SARS. J Pathol 2006; 210:288-
398		97.
399	19.	Shang J, Ye G, Shi K, Wan Y, Luo C, Aihara H, et al. Structural basis of receptor
400		recognition by SARS-CoV-2. Nature 2020.

_		Journal Pre-proof
401	20.	Wilk AJ, Rustagi A, Zhao NQ, Roque J, Martinez-Colon GJ, McKechnie JL, et al. A
402		single-cell atlas of the peripheral immune response to severe COVID-19. medRxiv
403		2020.
404	21.	Liao M, Liu Y, Yuan J, Wen Y, Xu G, Zhao J, et al. The landscape of lung
405		bronchoalveolar immune cells in COVID-19 revealed by single-cell RNA sequencing.
406		medRxiv 2020.
407	22.	Liu F, Li L, Xu M, Wu J, Luo D, Zhu Y, et al. Prognostic value of interleukin-6, C-
408		reactive protein, and procalcitonin in patients with COVID-19. J Clin Virol 2020;
409		127:104370.
410	23.	Zhu Z, Cai T, Fan L, Lou K, Hua X, Huang Z, et al. Clinical value of immune-
411		inflammatory parameters to assess the severity of coronavirus disease 2019. Int J
412		Infect Dis 2020.
413	24.	Velavan TP, Meyer CG. Mild versus severe COVID-19: laboratory markers. Int J
414		Infect Dis 2020.
415	25.	London AJ, Kimmelman J. Against pandemic research exceptionalism. Science 2020.
416	26.	Castell JV, Gomez-Lechon MJ, David M, Andus T, Geiger T, Trullenque R, et al.
417		Interleukin-6 is the major regulator of acute phase protein synthesis in adult human
418		hepatocytes. FEBS Lett 1989; 242:237-9.
419	27.	Castell JV, Andus T, Kunz D, Heinrich PC. Interleukin-6. The major regulator of
420		acute-phase protein synthesis in man and rat. Ann N Y Acad Sci 1989; 557:87-99;
421		discussion 100-1.
422	28.	Sehgal PB. Interleukin-6: a regulator of plasma protein gene expression in hepatic
423		and non-hepatic tissues. Mol Biol Med 1990; 7:117-30.
424	29.	Nishimoto N, Terao K, Mima T, Nakahara H, Takagi N, Kakehi T. Mechanisms and
425		pathologic significances in increase in serum interleukin-6 (IL-6) and soluble IL-6
426		receptor after administration of an anti-IL-6 receptor antibody, tocilizumab, in patients
427		with rheumatoid arthritis and Castleman disease. Blood 2008; 112:3959-64.

		Journal Pre-proof
428	30.	Yang ML, Wang CT, Yang SJ, Leu CH, Chen SH, Wu CL, et al. IL-6 ameliorates
429		acute lung injury in influenza virus infection. Sci Rep 2017; 7:43829.
430	31.	Lauder SN, Jones E, Smart K, Bloom A, Williams AS, Hindley JP, et al. Interleukin-6
431		limits influenza-induced inflammation and protects against fatal lung pathology. Eur J
432		Immunol 2013; 43:2613-25.
433	32.	Clinical management of severe acute respiratory infection when Covid-19 is
434		suspected.: World Health Organization; Updated March 13, 2020. Accessed April 14,
435		2020.] Available from https://www.who.int/publications-detail/clinical-management-of-
436		severe-acute-respiratory-infection-when-novel-coronavirus-(ncov)-infection-is-
437		suspected.
438	33.	Seymour CW, Liu VX, Iwashyna TJ, Brunkhorst FM, Rea TD, Scherag A, et al.
439		Assessment of Clinical Criteria for Sepsis: For the Third International Consensus
440		Definitions for Sepsis and Septic Shock (Sepsis-3). JAMA 2016; 315:762-74.
441	34.	Lim WS, van der Eerden MM, Laing R, Boersma WG, Karalus N, Town GI, et al.
442		Defining community acquired pneumonia severity on presentation to hospital: an
443		international derivation and validation study. Thorax 2003; 58:377-82.
444	35.	Guo L, Wei D, Zhang X, Wu Y, Li Q, Zhou M, et al. Clinical Features Predicting
445		Mortality Risk in Patients With Viral Pneumonia: The MuLBSTA Score. Front
446		Microbiol 2019; 10:2752.

## 448 Tables

## 449 Table 1: Combined Cohort

Variable	Evaluable	Median	Mechanic	al ventilation	p-value	q-value	
		(range) / n (%)	No (n = 57)	Yes (n = 32)	-		
Baseline Characteristics	S *	1			1		
Age (years)	89	61 (18 - 84)	58 (18 - 84)	65 (45 - 81)	0.031	0.067	
Respiratory rate (/min)	74	18 (11 - 40)	17 (13 - 39)	25 (11 – 40)	0.0024	0.0073	
Heart rate (/min)	66	86 (54 - 130)	85 (54 - 130)	89 (64 - 112)	0.32	0.47	
BMI	71	26.9 (18.1 – 45.7)	26.0 (18.1 - 36.2)	27.6 (18.3 – 45.7)	0.074	0.15	
Male gender	89	62 (70)	33 (58)	29 (91)	0.0029	0.0073	
Any comorbidities	87	70 (80)	43 (77)	27 (87)	0.38	0.53	
Hypertension	86	45 (52)	25 (45)	20 (65)	0.14	0.25	
Diabetes mellitus	86	13 (15)	7 (13)	6 (19)	0.61	0.68	
Coronary heart disease	85	7 (8)	4 (7)	3 (10)	>0.99	>0.99	
Chronic obstructive lung disease	86	9 (10)	7 (13)	2 (6)	0.54	0.67	
Computed Tomography	#			·	·		
Consolidation	78	46 (59)	30 (59)	16 (59)	>0.99	>0.99	
Ground glass opacity	78	72 (92)	47 (92)	25 (93)	>0.99	>0.99	
Bilateral infiltration	78	70 (90)	44 (86)	26 (96)	0.32	0.47	
Scores <sup>§</sup>							
qSOFA score 33	71	30 (42)	13 (28)	17 (68)	0.0028	0.0073	
CURB-65 score <sup>34</sup> ≥ 1	47	22 (47)	11 (41)	11 (55)	0.50	0.67	
MuLBSTA score <sup>35</sup>	68	11 (0 - 15)	9 (0 - 15)	11 (5 - 15)	0.090	0.17	
Laboratory	Evaluable	Median	Mechanic	al ventilation	p-value	q-value	
parameters		(range)	No (n = 57)	Yes (n = 32)	-		
Lymphocyte count	67	0.92 (0.20 – 2.84)	0.85 (0.31	0.94 (0.20 – 2.84)	0.60	0.68	
CRP (mg/l)	89	36 (0 - 369)	20 (0 - 315)	93 (16 - 369)	1.9-10 <sup>-7</sup>	2.6-10 <sup>-6</sup>	
Bilirubin (mg/dl)	84	0.5 (0.2 – 1.9)	0.5 (0.2 – 1.2)	0.6 (0.2 – 1.9)	0.19	0.32	
WBC (G/I)	89	5.86 (0.15 – 308)	5 (1.92 – 12.4)	7.26 (0.15 - 308)	0.0024	0.0073	
LDH (U/I)	88	311 (153 - 1121)	278 (153 - 619)	462 (240 - 1121)	1.5·10 <sup>-6</sup>	0.000010	
PCT (ng/ml)	87	0 (0 - 5)	0 (0 – 0.6)	0.2 (0 - 5)	8.7·10 <sup>-7</sup>	8.1·10 <sup>-6</sup>	
IL-6 (pg/ml)	86	34 (0 - 430)	23.2 (0 - 209)	95.4 (14.2 - 430)	2.3-10 <sup>-9</sup>	6.5-10 <sup>-*</sup>	
Thrombocyte count (G/I)	89	194 (0.12 - 450)	194 (0.27 - 383)	202 (0.12 - 450)	0.55	0.67	
Troponin T (ng/ml)	78	0 (0 – 0.178)	0 (0 – 0.143)	0 (0 – 0.178)	0.0001 0	0.00047	
Creatinine (mg/dl)	89	0.9 (0.4 – 7)	0.9 (0.4 – 5.6)	1.1 (0.8 – 7)	5.2 <b>-10</b> <sup>-</sup>	0.000029	
D-Dimer	76	0.7 (0 – 35.2)	0.6 (0 – 35)	0.9 (0 – 35.2)	0.0079	0.018	
Ferritin (ng/ml)	79	703 (30 - 3577)	545 (30 - 2578)	1392 (237 - 3577)	0.0002 3	0.00092	

450	* respiratory rate and heart rate and BMI (Body mass index) were measured at admission;
451	existing comorbidities were evaluated by patient history at admission; $^{*}$ CT-scans and
452	laboratory parameters at admission; $\$$ scores were calculated at admission. CRP = C-
453	Reactive Protein; WBC= White blood cell count; LDH = Lactate Dehydrogenase; PCT =
454	Procalcitonin; IL6 = Interleukin-6; qSOFA score = predicts mortality in sepsis, CURB-65
455	score = predicts mortality in community-acquired pneumonia, MuLBSTA score = predicts
456	mortality in patients with viral pneumonia; q-values represent the Benjamini-Hochberg
457	adjusted p-values
458	
459	

	Evaluation set					Validation set					Combined cohort							
Variable	At presentation			Maximal			At presentation		Maximal		At presentation			Maximal				
	p-value	AUC [CI]	Cutoff	p-value	AUC [CI]	Cutoff	p-value	AUC [CI]	Cutoff	p-value		Cutoff	p-value	AUC [CI]	Cutoff	p-value	AUC [CI]	Cutoff
IL-6 pg/ml	0.000012	0.94 [0.86, 1.00]	35	5.4·10 <sup>-8</sup>	0.97 [0.93, 1.00]	80	0.000076	0.84 [0.73, 0.95]	48.9	4.9·10 <sup>-7</sup>	0.90 [0.81, 0.98]	60	2.3·10 <sup>-9</sup>	0.89 [0.81, 0.96]	48.9	2.6·10 <sup>-11</sup>	0.93 [0.88, 0.98]	65
CRP mg/l	0.0031	0.79 [0.65, 0.93]	32.5	0.00027	0.86 [0.74, 0.98]	97	0.000032	0.86 [0.75, 0.96]	32.5	0.000097	0.83 [0.72, 0.95]	97	1.9·10 <sup>-7</sup>	0.83 [0.75, 0.92]	32.5	7.0·10 <sup>-8</sup>	0.85 [0.76, 0.93]	97
PCT ng/ml	0.0043	0.74 [0.58, 0.90]	0.05	0.0084	0.74 [0.57, 0.91]	0.25	0.000073	0.81 [0.69, 0.93]	0.05	0.00015	0.80 [0.67, 0.93]	0.25	8.7·10 <sup>-7</sup>	0.78 [0.68, 0.88]	0.05	4.2·10 <sup>-6</sup>	0.78 [0.67, 0.88]	0.25
LDH U/I	0.00062	0.83 [0.70, 0.97]	320	0.071	0.68 [0.50, 0.86]	590	0.00032	0.81 [0.67, 0.95]	410	0.0076	0.73 [0.60, 0.89]	440	1.4·10 <sup>-6</sup>	0.81 [0.72, 0.91]	410	0.0015	0.70 [0.59, 0.82]	380.5
WBC G/I	0.0028	0.80 [0.66, 0.93]	4920	0.010	0.75 [0.58, 0.93]	9860	0.13	0.63 [0.45, 0.81]	6190	0.30	0.59 [0.41, 0.77]	10510	0.0024	0.69 [0.57, 0.81]	6190	0.015	0.66 [0.53, 0.78]	9860
Creatinine mg/dl	0.00051	0.84 [0.72, 0.96]	0.95	0.00028	0.85 [0.74, 0.97]	1.05	0.0023	0.76 [0.63, 0.89]	0.95	0.026	0.69 [0.54, 0.84]	1.05	5.2·10 <sup>-6</sup>	0.79 [0.70, 0.88]	0.95	0.000070	0.75 [0.65, 0.86]	1.05
Troponin ng/ml	0.0053	0.72	0.005	0.0079	0.72 [0.55, 0.90]	0.005	0.0078	0.72 [0.57, 0.87]	0.005	0.020	0.69 [0.54, 0.85]	0.005	0.00010	0.73	0.005	0.00027	0.72	0.005
Ferritin ng/ml	0.064	0.72 [0.52, 0.91]	766	0.12	0.68 [0.47, 0.89]	530	0.0026	0.76 [0.62, 0.90]	1285	0.010	0.72 [0.58, 0.87]	1510	0.00023	0.75 [0.64, 0.86]	1285	0.0024	0.71 [0.59, 0.83]	1610

CRP = C-Reactive Protein; WBC= White blood cell count; LDH = Lactate Dehydrogenase; PCT = Procalcitonin; IL6 = Interleukin-6; AUC = area under the curve; CI = confidence interval

- 460 Table 3: Contingency table for high-risk and low-risk groups as defined by IL-6 and
- 461 **CRP in the validation cohort**

Variable	Value	Mechanic	462 p-value	
		No	Yes	463
IL-6 at presentation	≤35	19	3	0.0030
	>35	11	16	-0-
	•			465
Maximal IL-6	≤80	25	5	0.00022
	>80	5	14	.0
	•	L		467
CRP at presentation	≤32.5	19	1	468 0.00019
·	>32.5	11	18	469
Maximal CRP	≤97	23	3	470 0.00011
	>97	7	16	
	0)			

#### 471 Figure legends

#### 472 **Figure 1: Consort Diagram:**

473 Consort Diagram. DNR/DNI: do-not-resuscitate and do-not-intubate order.

474

# Figure 2: IL-6 at presentation, maximal IL-6 levels before mechanical ventilation and ROC-analysis of different parameters in the evaluation and validation cohort

Box plots showing IL-6 levels at first assessment (A, B) and maximal IL-6 levels before mechanical ventilation (C, D) in the evaluation cohort and in the validation cohort; dashed lines represents the cutoff calculated from the evaluation cohort (IL-6 at initial assessment >35 pg/ml, maximal IL-6 >80 pg/ml). Mean  $\pm$  SD is shown. Receiver operating characteristic (ROC) curve of maximal follow-up levels before mechanical ventilation in the evaluation (E) and validation cohorts (F).

483

## Figure 3: CRP levels at presentation and maximal CRP levels before mechanical ventilation

Box plot showing CRP levels at first assessment (A, B) and maximal IL-6 levels before
mechanical ventilation (C, D) in the evaluation cohort and in the validation cohort; dashed
lines represents the cutoff calculated from the training cohort (CRP at assessment >32.5
mg/l, maximal CRP>97 mg/l). Mean ± SD is shown.

490

491 Figure 4: Cutoffs and predictive values of maximal IL-6 and CRP values in the 492 combined cohort

Box plots depicting the maximal values of IL-6 and CRP in the overall cohort (A, B); dashed line represents the validated cutoff; dotted line represents the calculated improved cutoff from all patients (applicable only for IL-6). Positive predictive value (PPV) and negative 496 predictive value (NPV) as a function of different cutoffs is shown for IL-6 (C) and CRP (D)
497 values (dotted line represents cutoff for perfect NPV; dashed line represents cutoff for perfect
498 PPV)

499

Figure 5: Time from exceeding the maximal cutoff value of IL-6 or CRP to intubation in
 the combined cohort

- 502 Box plot depicting the time from exceeding the IL-6 (>65 ng/ml) and CRP (>97 mg/l) cutoff to
- 503 intubation in hours in the combined cohort. Median  $\pm$  min/max is shown.

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## Supplementary

Elevated levels of interleukin-6 and CRP predicts the need for mechanical ventilation in COVID-19

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## Table E1: Evaluation Cohort

Variable	Evaluable	Median (range) /	Mechanica	I ventilation	p-value	q-value	
		n (%)	No (n = 27)	Yes (n = 13)	-		
Baseline Characteristics *		1	1	1			
Age (years)	40	57 (19 - 81)	54 (19 - 80)	64 (45 - 81)	0.15	0.29	
Respiratory rate (/min)	34	18 (14 - 40)	18 (14 - 32)	23 (15 - 40)	0.066	0.14	
Heart rate (/min)	32	81 (54 - 112)	77 (54 - 111)	94 (80 - 112)	0.0069	0.022	
BMI	30	25.9 (19.0 – 45.7)	23.7 (19.0 – 34.7)	30.5 (24.8 – 45.7)	0.0030	0.014	
Male gender	40	29 (72)	16 (59)	13 (100)	0.020	0.051	
Any comorbidities	39	32 (82)	20 (77)	12 (92)	0.46	0.81	
Hypertension	38	19 (50)	10 (40)	9 (69)	0.17	0.32	
Diabetes mellitus	38	3 (8)	1 (4)	2 (15)	0.55	0.82	
Coronary heart disease	36	3 (8)	3 (12)	0 (0)	0.52	0.82	
Chronic obstructive lung disease	37	3 (8)	2 (8)	1 (8)	>0.99	>0.99	
Computed Tomography <sup>#</sup>							
Consolidation	36	21 (58)	14 (61)	7 (54)	0.95	>0.99	
Ground glass opacity	36	31 (86)	20 (87)	11 (85)	>0.99	>0.99	
Bilateral infiltration	36	33 (92)	21 (91)	12 (92)	>0.99	>0.99	
Scores <sup>§</sup>				1	1		
qSOFA score <sup>1</sup>	32	12 (37)	7 (32)	5 (50)	0.55	0.82	
CURB-65 score <sup>2</sup> ≥ 1	24	7 (29)	5 (31)	2 (25)	>0.99	>0.99	
MuLBSTA score <sup>3</sup>	29	9 (4 - 15)	9 (4 - 13)	7 (5 - 15)	0.89	>0.99	
Laboratory <sub>#</sub>	Evaluable	Median (range)	Mechanica	I ventilation	p-value	q-value	
parameters"			No (n = 27)	Yes (n = 13)	-		
Lymphocyte count G/I	31	0.99 (0.45 –	0.99 (0.45 –	0.95 (0.57 –	0.92	>0.99	
CRP (mg/l)	40	28 (0 – 315)	17 (0 – 315)	77 (16 – 171)	0.0031	0.014	
Bilirubin (mg/dl)	37	0.5 (0.2 – 1.9)	0.5 (0.2 – 1.2)	0.5 (0.4 – 1.9)	0.78	>0.99	
WBC (G/I)	40	5.04 (2.12 - 308)	4.67 (2.12 – 10.8)	7.38 (4.67 - 308)	0.0028	0.014	
LDH (U/I)	39	285 (153 - 1078)	258 (153 - 619)	381 (252 - 1078)	0.00062	0.0058	
PCT (ng/ml)	38	0 (0 - 5)	0 (0 – 0.6)	0.1 (0 - 5)	0.0043	0.017	
IL-6 (pg/ml)	37	27.1 (0 - 430)	19.6 (0 – 76.5)	121 (19.2 - 430)	0.000012	0.00034	
Thrombocyte count (G/I)	40	161 (0.12 - 440)	162 (0.27 - 334)	160 (0.12 - 440)	0.74	>0.99	
Troponin T (ng/ml)	34	0 (0 – 0.032)	0 (0 – 0.022)	0 (0 – 0.032)	0.0053	0.019	
Creatinine (mg/dl)	40	0.9 (0.4 – 2.1)	0.9 (0.4 – 1.3)	1.0 (0.9 – 2.1)	0.00051	0.0058	
D-Dimer	31	0.7 (0 – 2.9)	0.6 (0 – 2.2)	1.1 (0.6 – 2.9)	0.019	0.051	
Ferritin (ng/ml)	31	626 (46 - 2153)	553 (46 - 1748)	, 810 (431 - 2153)	0.064	0.14	

\* respiratory rate and heart rate and BMI (Body mass index) were measured at admission; existing comorbidities were evaluated by patient history at admission; <sup>#</sup> CT-scans and laboratory parameters at admission; <sup>§</sup> scores were calculated at admission. CRP = C-Reactive Protein; WBC= White blood cell count; LDH = Lactate Dehydrogenase; PCT = Procalcitonin; IL6 = Interleukin-6; qSOFA score = predicts mortality in sepsis, CURB-65 score = predicts mortality in community-acquired pneumonia, MuLBSTA score = predicts mortality in patients with viral pneumonia; q-values represent the Benjamini-Hochberg adjusted p-values

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## Table E2: Validation cohort

Variable	Evaluable	Median (range) /	Mechanica	l ventilation	p-value	q-value
		n (%)	No (n = 30)	Yes (n = 19)	-	
Baseline Characteristics *		1	1	1		
Age (years)	49	64 (18 - 84)	61 (18 - 84)	65 (46 - 81)	0.18	0.31
Respiratory rate (/min)	34	18 (11 - 40)	17 (13 - 39)	26 (11 – 40)	0.027	0.083
Heart rate (/min)	34	90 (64 - 130)	94 (74 - 130)	86 (64 - 107)	0.033	0.091
BMI	41	27.5 (18.1 – 36.2)	27.6 (18.1 – 36.2)	27.0 (18.4 – 34.7)	0.58	0.71
Male gender	49	33 (67)	17 (57)	16 (84)	0.091	0.21
Any comorbidities	48	38 (79)	23 (77)	15 (83)	0.85	0.96
Hypertension	48	26 (54)	15 (50)	11 (61)	0.65	0.76
Diabetes mellitus	48	10 (21)	6 (20)	4 (22)	>0.99	>0.99
Coronary heart disease	49	4 (8)	1 (3)	3 (16)	0.31	0.46
Chronic obstructive lung disease	49	6 (12)	5 (17)	1 (5)	0.46	0.61
Computed Tomography <sup>#</sup>		·				
Consolidation	42	25 (59)	16 (57)	9 (64)	>0.99	0.98
Ground glass opacity	42	41 (98)	27 (96)	14 (100)	>0.99	>0.99
Bilateral infiltration	42	37 (88)	23 (82)	14 (100)	0.24	0.37
Scores <sup>§</sup>	1			1	1	
qSOFA score <sup>1</sup>	39	18 (46)	6 (25)	12 (80)	0.0025	0.010
CURB-65 score <sup>2</sup> ≥ 1	23	15 (65)	6 (55)	9 (75)	0.55	0.71
MuLBSTA score <sup>3</sup>	39	11 (0 - 15)	10 (0 - 15)	13 (9 - 15)	0.038	0.096
Laboratory	Evaluable	Median (range)	Mechanica	Mechanical ventilation		q-value
parameters			No (n = 30)	Yes (n = 19)	-	
Lymphocyte count G/I	36	0.80 (0.20 – 2.84)	0.73 (0.31 – 2.36)	0.94 (0.20 – 2.84)	0.43	0.60
CRP (mg/l)	49	42 (1 – 369)	22 (1 – 163)	134 (31 – 369)	0.000032	0.00068
Bilirubin (mg/dl)	47	0.5 (0.2 – 1.2)	0.4 (0.2 – 1.2)	0.6 (0.2 – 1.1)	0.16	0.30
WBC (G/I)	49	6.0 (0.15 – 25.8)	, 5.79 (1.92 – 12.4)	7.22 (0.15 – 25.8)	0.13	0.26
LDH (U/I)	49	336 (181 - 1121)	278 (181 - 502)	474 (240 - 1121)	0.00032	0.0022
PCT (ng/ml)	49	0 (0 – 2.3)	0 (0 – 0.3)	0.2 (0 – 2.3)	0.000073	0.00068
IL6 (pg/ml)	49	42.7 (0 - 272)	23.7 (0 - 209)	83.5 (14.2 - 272)	0.000072	0.00068
Thrombocyte count (G/I)	49	216 (93 - 450)	212 (112 - 383)	220 (93 - 450)	0.23	0.37
Troponin T (ng/ml)	44	0 (0 – 0.178)	0 (0 – 0.143)	0.022 (0 – 0.178)	0.0078	0.027
Creatinine (mg/dl)	49	0.9 (0.5 – 7.0)	0.9 (0.5 – 5.6)	1.1 (0.8 – 7.0)	0.0023	0.010
D-Dimer	45	0.8 (0 – 35.2)	0.6 (0 - 35)	0.9 (0 - 35.2)	0.11	0.24
Ferritin (ng/ml)	48	789 (30 - 3577)	508 (30 - 2578)	1692 (237 - 3577)	0.0026	0.010

\* respiratory rate and heart rate and BMI (Body mass index) were measured at admission; existing comorbidities were evaluated by patient history at admission; <sup>#</sup> CT-scans and laboratory parameters at admission; <sup>§</sup> scores were calculated at admission. CRP = C-Reactive Protein; WBC= White blood cell count; LDH = Lactate Dehydrogenase; PCT = Procalcitonin; IL6 = Interleukin-6; qSOFA score = predicts mortality in sepsis, CURB-65 score = predicts mortality in community-acquired pneumonia, MuLBSTA score = predicts mortality in patients with viral pneumonia; q-values represent the Benjamini-Hochberg adjusted p-values

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Supplementary Table E3:	<b>Comparison Evaluation</b>	and Validation cohort
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Variable	Cohort		p-value
	Evaluation (n = 40)	Validation (n = 49)	
Baseline Characteristics *			
Age (years)	57 (19 - 81)	64 (18 - 84)	0.15
Respiratory rate (/min)	18 (14 - 40)	18 (11 - 40)	0.76
Heart rate (/min)	81 (54 - 112)	90 (64 - 130)	0.017
BMI	25.9 (19.0 – 45.7)	27.5 (18.1 – 36.2)	0.18
Male gender	29 (72)	33 (67)	0.77
Any comorbidities	32 (82)	38 (79)	0.95
Hypertension	19 (50)	26 (54)	0.87
Diabetes mellitus	3 (8)	10 (21)	0.17
Coronary heart disease	3 (8)	4 (8)	>0.99
Chronic obstructive lung disease	3 (8)	6 (12)	0.79
Computed Tomography <sup>#</sup>			
Consolidation	21 (58)	25 (60)	>0.99
Ground glass opacity	31 (86)	41 (98)	0.14
Bilateral infiltration	33 (92)	37 (88)	0.89
Scores <sup>§</sup>			
qSOFA score <sup>1</sup>	12 (37)	18 (46)	0.62
CURB-65 score <sup>2</sup> ≥ 1	7 (29)	15 (65)	0.029
MuLBSTA score <sup>3</sup>	9 (4 - 15)	11 (0 - 15)	0.13
Laboratory parameters <sup>#</sup>	Cohort		p-value
	Evaluation (n = 40)	Validation (n = 49)	
Lymphocyte count G/I	0.99 (0.45 – 2.5)	0.8 (0.2 – 2.84)	0.27
CRP (mg/l)	28 (0 – 315)	42 (1 – 369)	0.10
Bilirubin (mg/dl)	0.5 (0.2 – 1.9)	0.5 (0.2 – 1.2)	0.71
WBC (G/I)	5.04 (2.12 - 308)	6 (0.15 – 25.8)	0.47
LDH (U/I)	285 (153 - 1078)	336 (181 - 1121)	0.18
PCT (ng/ml)	0 (0 – 5)	0 (0 – 2.3)	0.32
IL-6 (pg/ml)	27.1 (0 - 430)	42.7 (0 - 272)	0.34
Thrombocyte count (G/I)	161 (0.12 - 440)	216 (93 - 450)	0.0084
Troponin T (ng/ml)	0 (0 – 0.032)	0 (0 – 0.178)	0.016
Creatinine (mg/dl)	0.9 (0.4 – 2.1)	0.9 (0.5 – 7.0)	0.82
D-Dimer	0.7 (0 – 2.9)	0.8 (0 – 35.2)	0.57
Ferritin (ng/ml)	626 (46 - 2153)	789 (30 - 3577)	0.20

\* respiratory rate and heart rate and BMI (Body mass index) were measured at admission; existing comorbidities were evaluated by patient history at admission; <sup>#</sup> CT-scans and laboratory parameters at admission; <sup>§</sup> scores were calculated at admission. CRP = C-Reactive Protein; WBC= White blood cell count; LDH = Lactate Dehydrogenase; PCT = Procalcitonin; IL6 = Interleukin-6; qSOFA score = predicts mortality in sepsis, CURB-65 score = predicts mortality in community-acquired pneumonia, MuLBSTA score = predicts mortality in patients with viral pneumonia; q-values represent the Benjamini-Hochberg adjusted p-values

## **Supplementary References**

- Seymour CW, Liu VX, Iwashyna TJ, Brunkhorst FM, Rea TD, Scherag A, et al. Assessment of Clinical Criteria for Sepsis: For the Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3). JAMA 315, 762-774 (2016).
- Lim WS, Van der Eerden MM, Laing R, Boersma WG, Karalus N, Town GI, *et al.* Defining community acquired pneumonia severity on presentation to hospital: an international derivation and validation study. *Thorax* 58, 377-382 (2003).
- Guo L, Wei D, Wu Y, Zhou M, Zhang X, Li Q, *et al.* Clinical Features Predicting Mortality Risk in Patients With Viral Pneumonia: The MuLBSTA Score. *Front Microbiol* 10, 2752 (2019).