

Increased mitochondrial respiration of adipocytes from metabolically unhealthy obese compared to healthy obese individuals

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Supplemental legends

Figure S1: Potential underlying pathomechanisms, n=4/group; mean \pm SEM; mRNA expression normalized to RPS13 of genes pivotal for: lipogenesis (panel A), lipolysis (B), β -oxidation (C), respiratory chain enzymes of complexes I-IV (D), and mitochondrial biogenesis (E), analyzed in adipocytes. Panel F – citrate synthase activity in preadipocytes and differentiated adipocytes.

Figure S2: Western Blots for protein levels of selected genes, n=4/group, if not stated otherwise; relative band intensity values (mean \pm SD) are normalized to tubulin; proteins include important proteins for A) Complex II, B) Complex III (n=3 vs. 4), and C+D) mitochondrial biogenesis. On the right side, representative original western blots are shown.

Figure S3: mitochondrial DNA encoded genes; mean \pm SD; mRNA expression normalized to RPS13; n=2 vs.3; ND1 representing complex I, COI = complex IV, ATP6 = complex V.

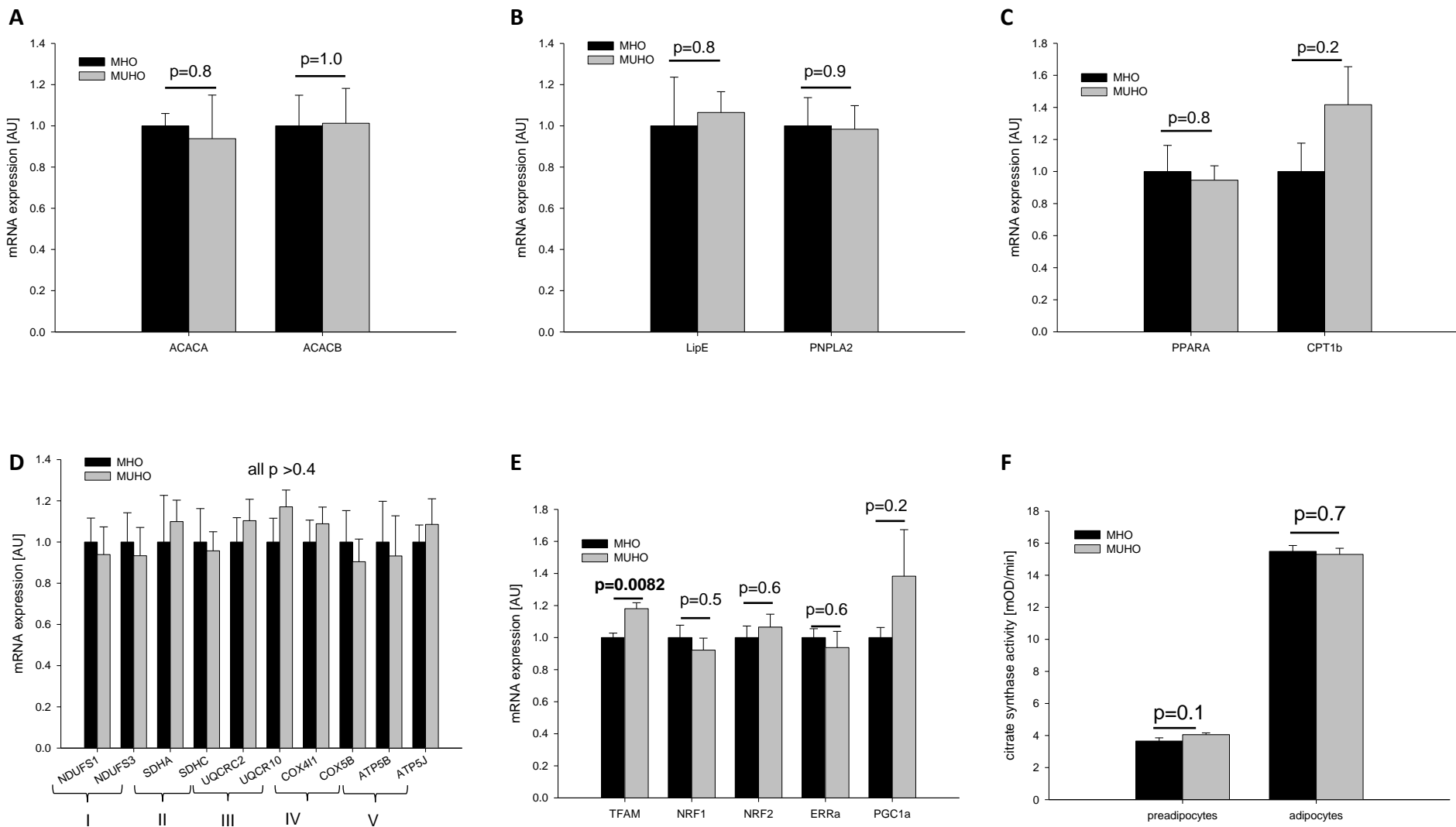
Figure S4: additional pathomechanistic hypotheses; panel A) lipolysis, B) cAMP, and C) ROS; mean \pm SD; each n=2 vs. 3; A) and c) normalized to protein.

Figure S5: Original western blot; blots in figure S2 were taken from this blot. Black framed parts correspond to figure S2.

Figure S6: FCCP titration experiments showing the mitochondrial stress test under various FCCP concentrations (in μ M), showing that optimal conditions were used in this study; n=5.

Table S1: Primer sequences used for quantitative real-time PCR.

fig. S1



Western Blots

fig. S2

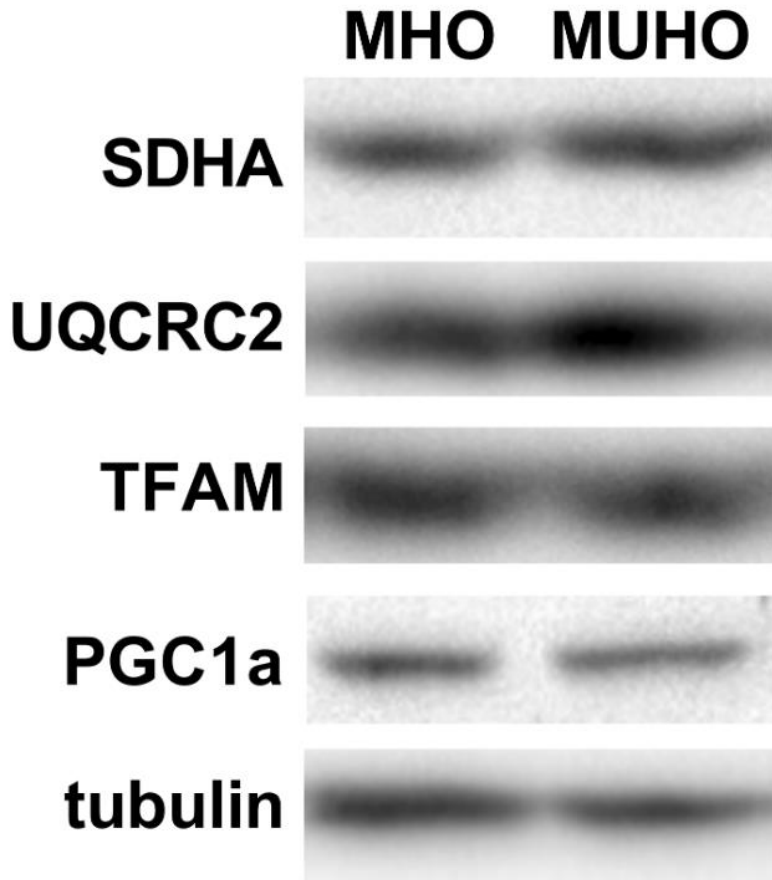
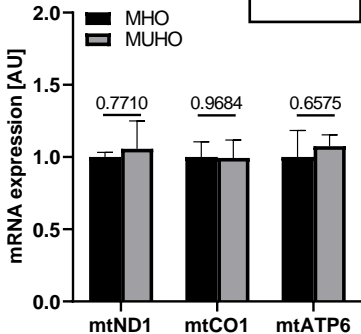


fig. S3



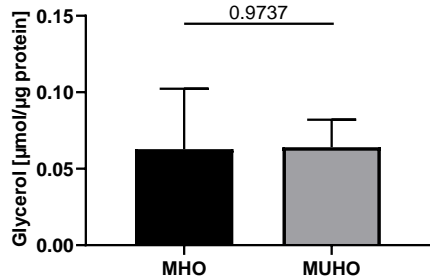
A**B**

fig. S4

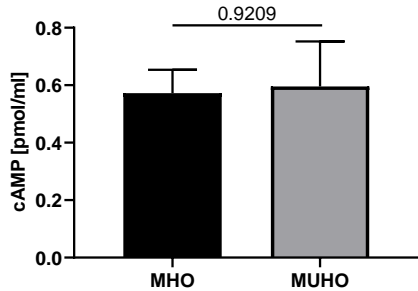
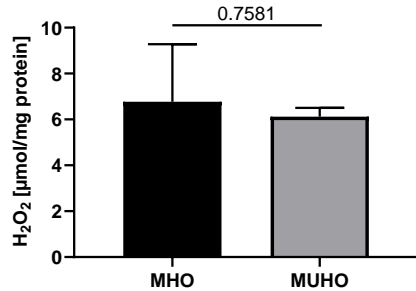
**C**

fig. S5

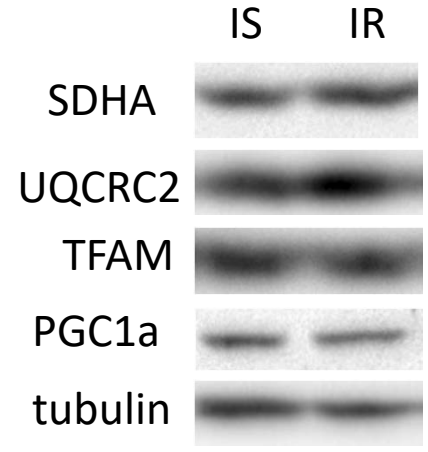
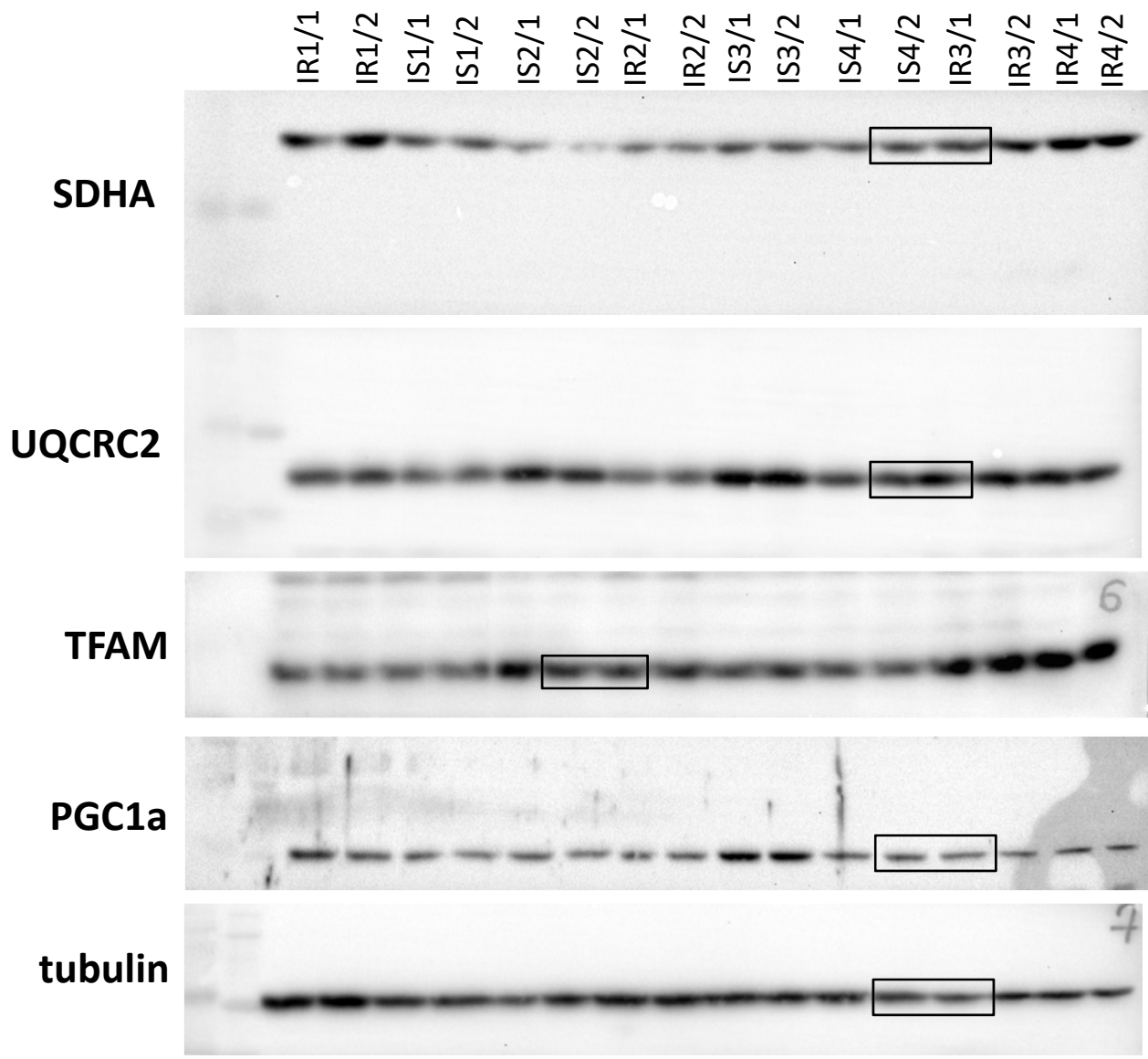
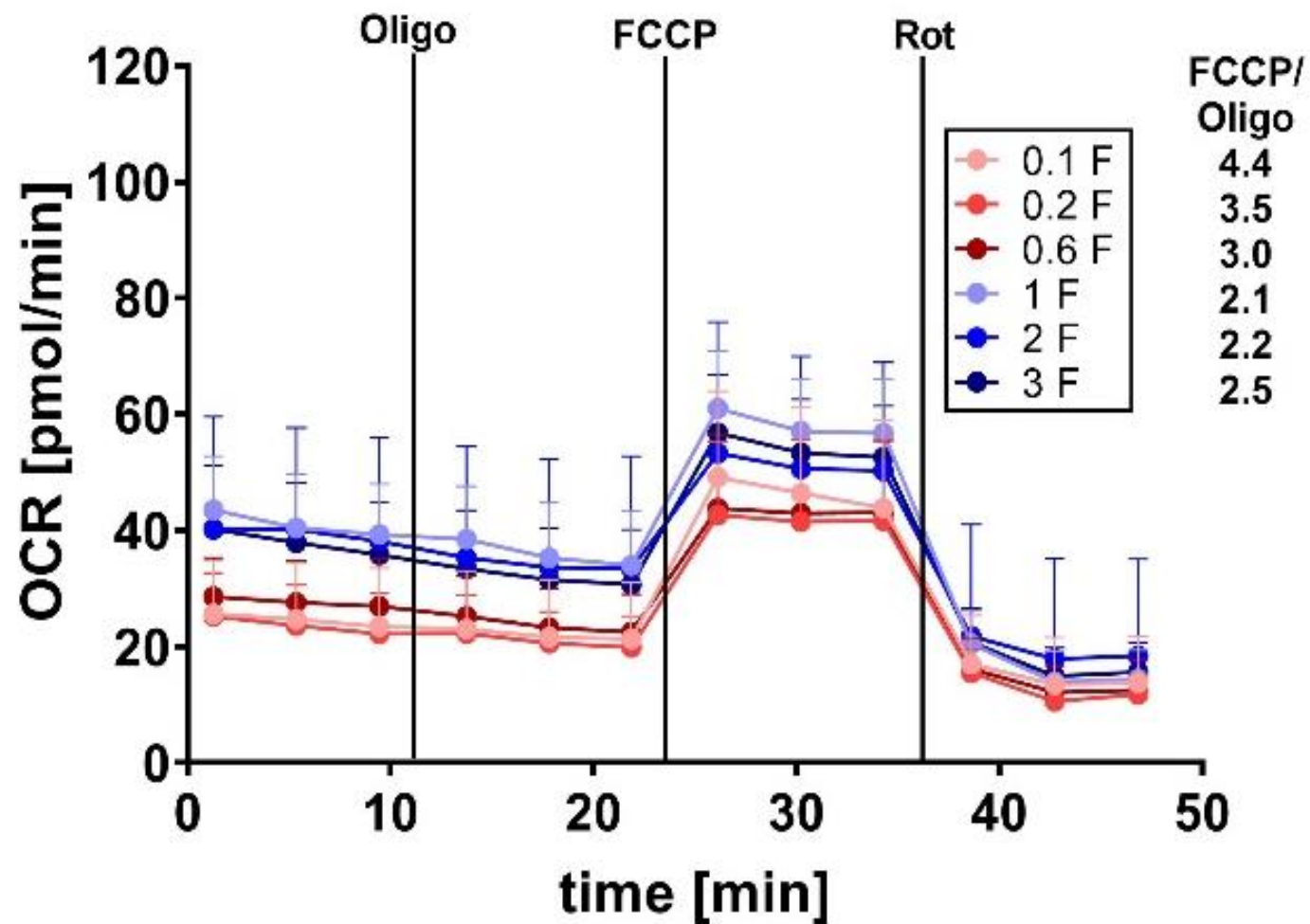


fig. S6

FCCP titration



gene	sequence 5' - 3'
ACACA up	CATTCCGAGCAAGGGATAAG
ACACA do	CAGAGCAGGCTCCAGATGA
ACACB do	TGCGGAACATCTCATAGGC
ACACB up	TCGGGTCATCGAGAAGGT
ATP5B do	ACCAGAATTTCTGCTCAACA
ATP5B up	TGATGAAAGAGGTCCCATCAA
ATP5J do	AAGCTCCCTCTCCAGCTCTT
ATP5J up	CAGAAACTCTTTGTGGACAAGATTAG
COX4I1 do	CAGACAGGTGCTTGACATGG
COX4I1 up	TCCACCTCTGTGTGTGTACGA
COX5B do	TGTATGGGTCCAGTCCCTTCT
COX5B up	TCCATGGCATCTGGAGGT
CPT1b do	GAAGGAGGAACCCACTGTTG
CPT1b up	CCTGGAAGAAACGCCTGAT
ESRRA do (ERRa)	TCAACCACCAGCAGATGAGA
ESRRA up (ERRa)	GTGGGCGGCAGAAGTACA
LipE do	TTCGTTCCCCTGTTGAGC
LipE up	CGCTGGAGGAGTGCTTCTT
NDUFS1 do	AAGAAGATAATTGGAACGCAAATC
NDUFS1 up	TCTGACACCTTATGCACTGAAGA
NDUFS3 do	AACTCATTGAAGCAGGACACCT
NDUFS3 up	AGCAGCTCTCAGCTTTTGGGA
NFE2L2 up (NRF2)	GAGACAGGTGAATTTCTCCCAAT
NFE2L2 do (NRF2)	TTTGGGAATGTGGGCAAC
NRF1 do	TGGAGGGTGAGATACAGAGGA
NRF1 up	TATCCGGAAGAGGCAACAAA
PGC1a do	ATAAATCACACGGCGCTCTT
PGC1a up	TGAGAGGGCCAAGCAAAG
PNPLA2 do	CCCTGCTTGACATCTCTC
PNPLA2 up	CTCCACCAACATCCACGAG
PPARa do	GTCCCCGCAGATTCTACATT
PPARa up	GAAGCTGTCCTGGCTCAGAT
SDHA do	CCAGCGTTTGGTTTAATTGG
SDHA up	GGACCTGGTTGTCTTTGGTC
SDHC do	CTGCACTCAAAGCAATACCAGT
SDHC up	TCCTCTGTCTCCCCACATTAC
TFAM do	GAATCAGGAAGTTCCTCCA
TFAM up	GAACAACCTACCCATATTTAAAGCTCA
UQCR10 do	CTCCAGCTCTGGGCAAAC
UQCR10 up	GGGGTGACAGTGGAGTAGAGAA
UQCRC2 do	TGCACTTGTGAAATGGTTCTG
UQCRC2 up	TGTCCTGACTATAGGATTGGAAAA
mtND1 up	AACCTCTCCACCCTTATCACAA
mtND1 do	TCATATTATGGCCAAGGGTCA
mtCO1 up	CCCTCCCTTAGCAGGGAAC

mtCO1 do	TGAAATTGATGGCCCCTAAG
mtATP6 up	GCCGCAGTACTGATCATTCTATT
mtATP6 do	GGGTGGTGATTAGTCGGTTG

Table S1: Primer sequences used for quantitative real-time PCR.