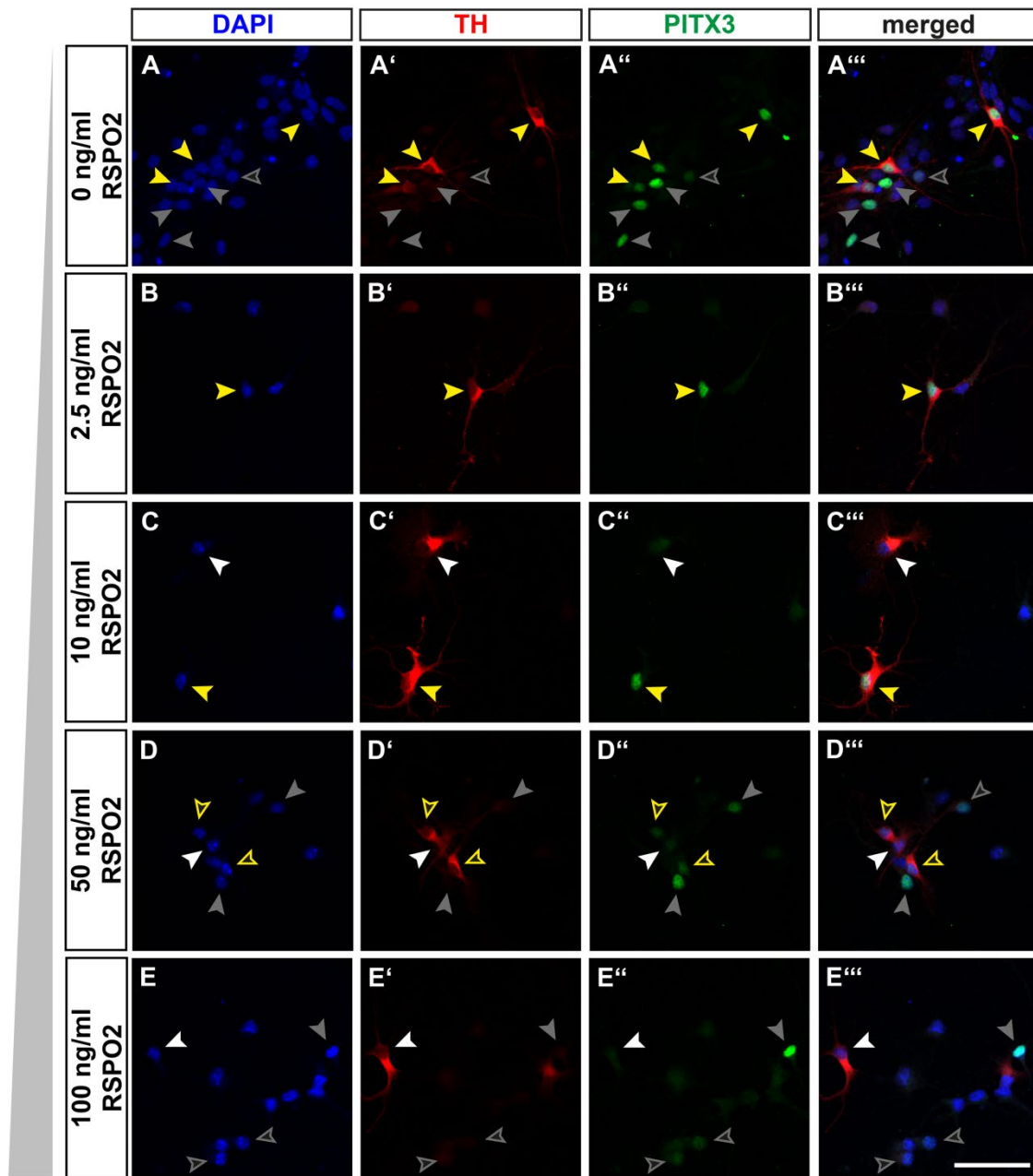




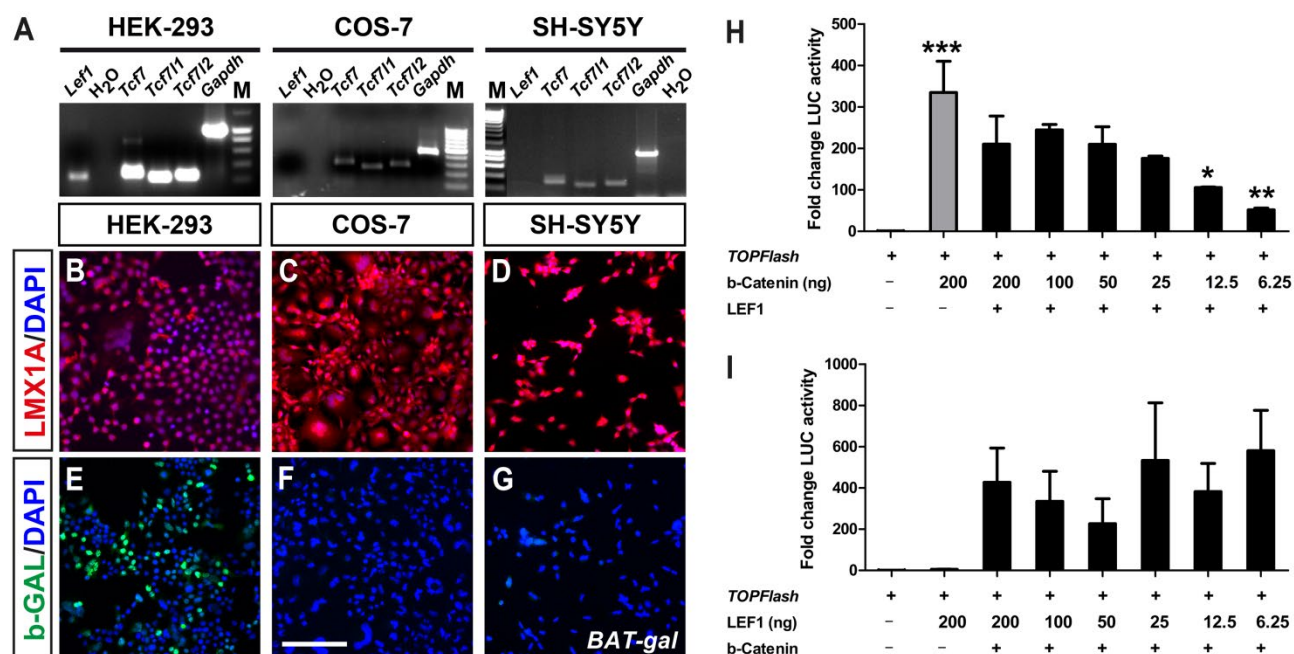
Supplementary Material

1 Supplementary Figures and Tables

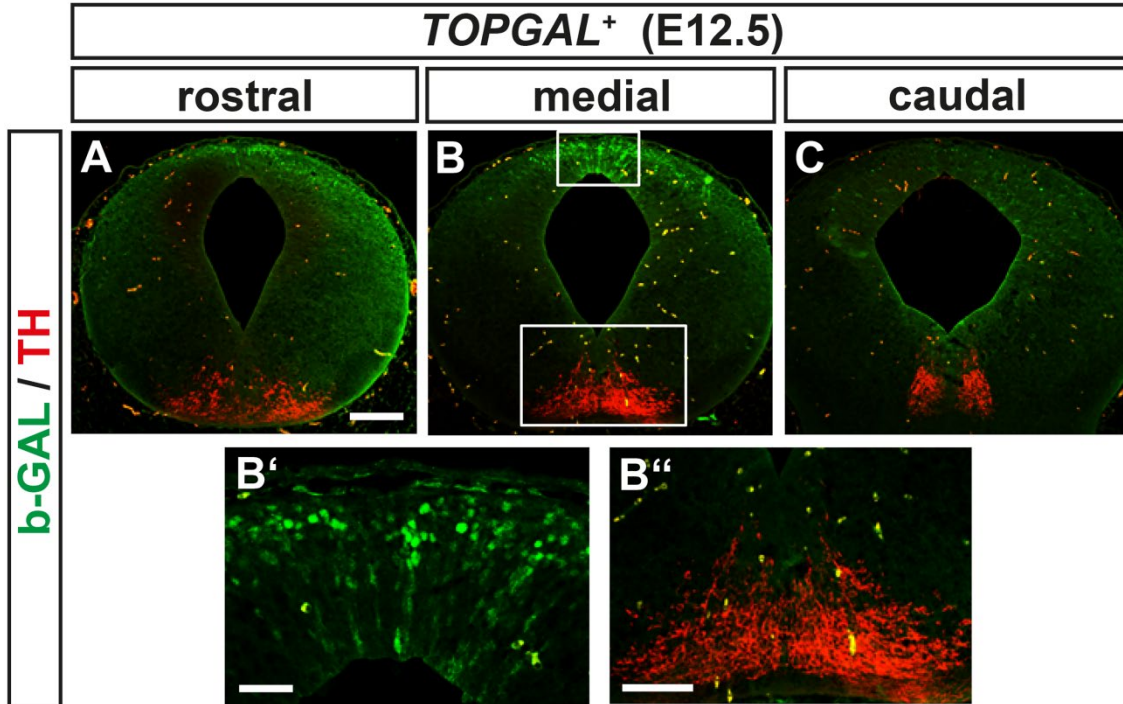
1.1 Supplementary Figures



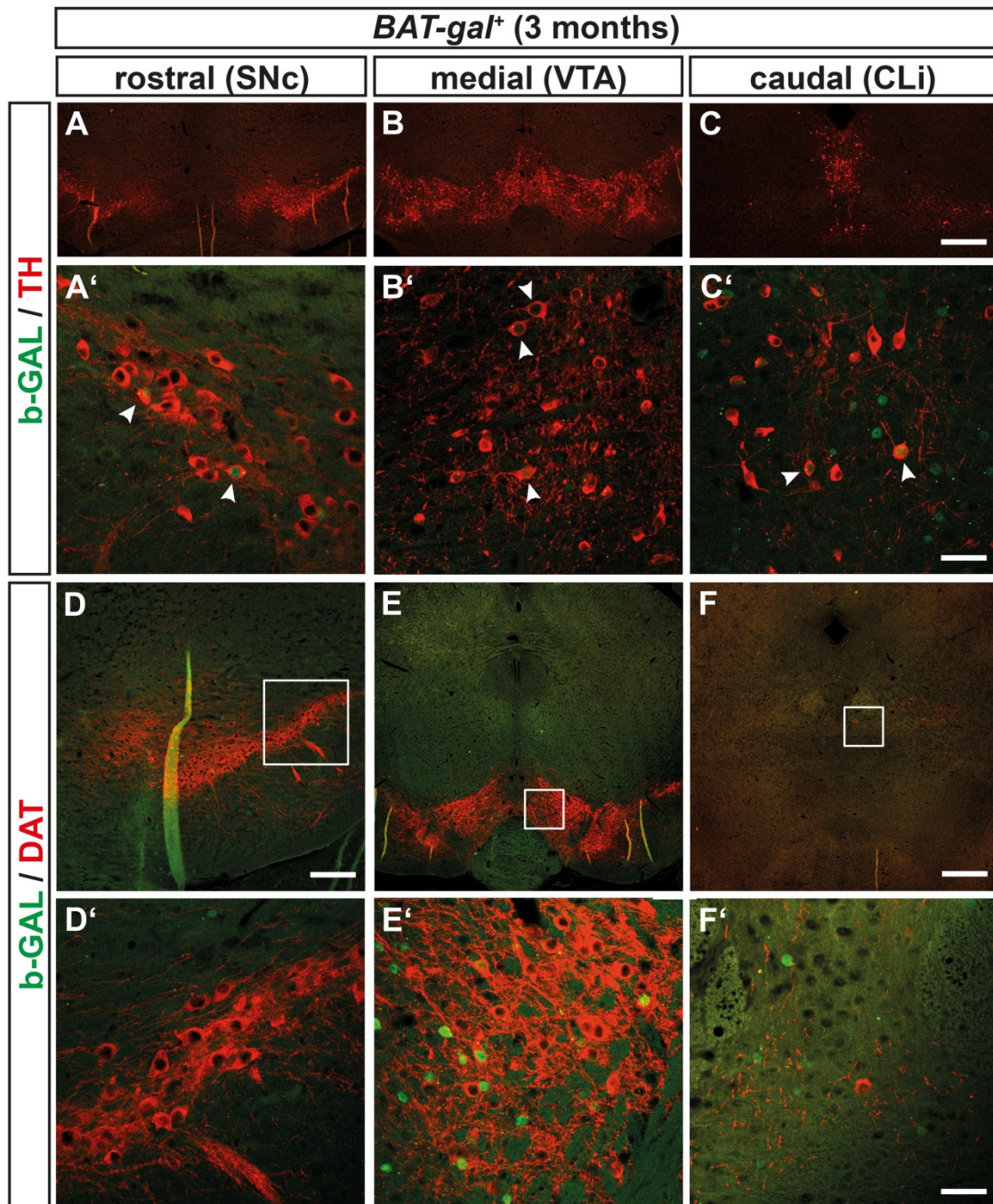
Supplementary Figure 1: RSPO2-mediated activation of WNT1/b-catenin signaling inhibits the differentiation of PITX3⁺ mdDA neurons *in vitro*. (A-E''') Representative confocal close-up views of VM primary cells isolated from E11.5 wild-type (*CD-1*) mouse embryos and treated for 7 d (10 DIV) with increasing concentrations of RSPO2 protein. Cells were immunostained for TH (red; A'-E'), PITX3 (green; A''-E''), and counterstained with DAPI (blue; A-E). The rightmost panel (A'''-E''') depicts the merged images. Yellow arrowheads in (A-D''') point at PITX3⁺/TH⁺ double-positive cells; white arrowheads in (C-E''') point at PITX3⁻/TH⁺ single-positive cells; gray arrowheads in (A-A'''; D-E''') point at PITX3⁺/TH⁻ single-positive cells; and open arrowheads in (A-A'''; D-E''') point at PITX3⁺ cells with very low immunocytochemical signal intensities. Scale bar: 50 μ m (E''').



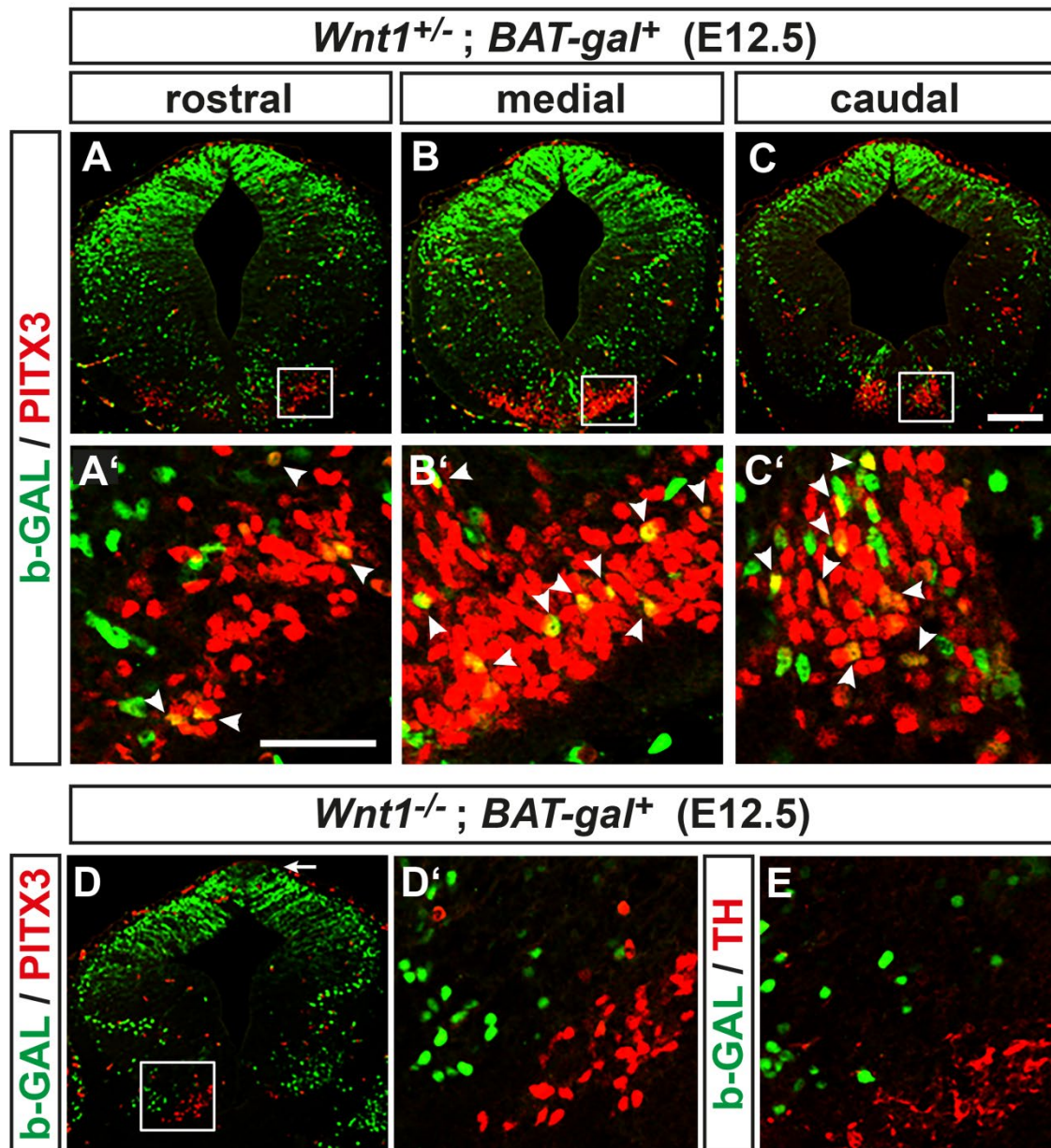
Supplementary Figure 2: Dose-dependent activation of LEF1/TCF-mediated WNT/b-catenin signaling in WNT/b-catenin-responding HEK-293 cells. (A) RT-PCR analyses of total RNA isolated from HEK-293, COS-7 and SH-SY5Y cells for the detection of *LEF1/Lef1*, *TCF7/Tcf7*, *TCF7L1/Tcf7l1* and *TCF7L2/Tcf7l2* transcription. H₂O, negative PCR control; *GAPDH/Gapdh* (Glyceraldehyde-3-phosphate dehydrogenase), positive PCR control; M, DNA ladder. (B-G) Representative confocal overviews of HEK-293 (B,E), COS-7 (C,F) and SH-SY5Y (D,G) cells, immunostained for LMX1A (red; B-D) or b-GAL after transfection of the *BAT-gal* reporter construct (green; E-G) and counterstained with DAPI (blue; B-G). (H,I) Fold change of luciferase (LUC) activity in HEK-293 cells (relative to only *TOPFlash*-transfected cells, set as 1) after transfection with *TOPFlash* reporter and decreasing amounts of *S33Y-b-catenin* (H) or *LEF1* cDNA (I) alone or together with *LEF1* (H) or *S33Y-b-catenin* (I) plasmids, respectively (n = 3 independent experiments; statistical testing for significance was done between the only *TOPFlash*-transfected cells and the *TOPFlash* + *S33Y-b-catenin* or *LEF1*-transfected cells (grey bars), and between the latter and cells transfected with *TOPFlash* reporter and decreasing amounts of *S33Y-b-catenin* or *LEF1* plasmids in the presence of *LEF1* or *S33Y-b-catenin* (black bars), respectively, using one-way ANOVA followed by Bonferroni's multiple comparisons *post hoc* tests; F(7,15) = 7.1, P = 0.0008). Single asterisk, P<0.05; double asterisks, P<0.005; triple asterisks, P<0.0001. Scale bar: 100 μm (F).



Supplementary Figure 3: The *TOPGAL* mouse is not a reporter of WNT/b-catenin signaling in the mouse VM. (A-C) Representative overviews (A-C) and close-up views of the DM (B') and VM (B'') (corresponding to the boxed areas in B) on coronal sections (dorsal top) at different rostrocaudal levels of the midbrain from a *TOPGAL* embryo at E12.5, immunostained with antibodies against b-GAL (green) and TH (red); overlapping expression domains appear in yellow. Scale bars: 200 μ m (A); 100 μ m (B''); 50 μ m (B').



Supplementary Figure 4: mdDA neurons in the adult mouse brain are mostly non-WNT/b-catenin-responding. (A-F') Representative overviews (A-C,D-F) and close-up views of the SNc, VTA and CLi (A'-C'; D'-F', corresponding to the boxed areas in D-F, respectively) on coronal sections (dorsal top) at different rostrocaudal levels of the midbrain from 3 months-old *BAT-gal*⁺ mice, immunostained with antibodies against b-GAL (green) and TH (red; A-C') or DAT (red; D-F'); overlapping expression domains appear in yellow. White arrowheads in (A'-C') point at b-GAL⁺/TH⁺ double-positive cells. Abbreviations: CLi, caudal linear nucleus of the raphe; SNc, Substantia nigra pars compacta; VTA, ventral tegmental area. Scale bars: 250 μ m (C,F); 100 μ m (D); 50 μ m (C',F').



Supplementary Figure 5: *Wnt1* is the main activating WNT-ligand in the mouse FP. (A-E) Representative overviews (A-D) and close-up views of the VM (A'-D',E; corresponding to the boxed areas in A-D, respectively) on coronal sections (dorsal top) at different rostrocaudal levels of the midbrain from *Wnt1*^{+/-}; *BAT-gal*⁺ (A-C') and *Wnt1*^{-/-}; *BAT-gal*⁺ (D-E) mouse embryos at E12.5, immunostained with antibodies against b-GAL (green) and PITX3 (red; A-D') or TH (red; E); overlapping expression domains appear in yellow. White arrowheads in (A'-C') point at b-GAL⁺/PITX3⁺ double-positive cells. White arrow in (D) points at the b-GAL-negative roof plate. Scale bars: 200 μm (C); 50 μm (A').

1.2 Supplementary Tables

Supplementary Table 1: Primer and PCR conditions used for genotyping, ISH probe cloning, ChIP-PCR, site-directed mutagenesis of LEF1/TCF BSs/WREs and RT-PCR; and siRNAs

Gene (application)	Forward primer (5' → 3') Reverse primer (5' → 3') or mutagenic primer or siRNA (5' → 3')	Product length (bp)	T _m (°C)	Cycles
<i>LacZ</i> (<i>BAT-gal</i> genotyping)	GGTGGCGCTGGATGGTAA CGCCATTTGACCACTACC	613	60	30
<i>LacZ</i> (<i>TOPGAL</i> genotyping)	CGTGGCCTGATTCATTCC ATCCTCTGCATGGTCAGGTC	315	60	30
<i>Lef1 wt</i> (<i>Lef1</i> ^{+/+} genotyping)	CCGTTTCAGTGGCACGCCCTCTCC TGTCTCTCTTTCCGTGCTAGTTC	71	58	38
<i>Lef1 ko</i> (<i>Lef1</i> ^{-/-} genotyping)	CCGTTTCAGTGGCACGCCCTCTCC CATGGCGATGCCTGCTTGC	321	58	38
<i>Apcddl</i> (ISH probe)	TCTACCGGCCGTCCAGTTAC TATCTGAGGATGCGCCAATG	814	60	34
<i>Cck</i> (ISH probe)	TGGACCCCAGCCATAGAA GGAAACACTGCCTTCCGA	275	60	34
<i>Fgf14</i> (ISH probe)	GCAACCTGGTGGATATCTTCTC GAGTTTAGCTGGTTTGTCAGG	830	63	35
<i>Pbx1</i> (ISH probe)	CATGCGACTGGACAACATGC CATGGGCTGACACATTGGTG	644	63	35
<i>Rspo2</i> (ISH probe)	CTCATGGCGTTCTCAGCATC GCCTTGCTTTGGATGTTTCC	941	63	35
<i>Smarca1</i> (ISH probe)	CGGAATGCTCCCCAGTTTAG CGCACACTGACAGCAAACAC	878	60	34
<i>Sulf1</i> (ISH probe)	CTCTTCACTCCAGCCACACG ACCAACATTGTGAGCCATGC	807	60	34
<i>Sulf2</i> (ISH probe)	CTGGGACAGCTATGGGAAGG GGGGGCAGGAACACTGTAAG	852	60	34
<i>Tcf7</i> (ISH probe)	CTCCTTCCCCACAGAACTGC GGGTGCACACTGGGTTTAGG	866	64	35
<i>Tcf7l1</i> (ISH probe)	GCAAATCCAAGAGGCAGAGG TCTACAGTGGTAAGGCACTGCTG	613	65	35
<i>Pitx3</i> (LEF1 ChIP Primer 1)	CCTGTCCCTTGCAAACACTT CCCCTCCTTCCATCCTATC	362 (-1025 to -664)	60	32
<i>Pitx3</i> (LEF1 ChIP Primer 2)	GGAAAGGAGGGGTGGTCTTT TTGCCATCCACTGAGCAAC	412 (-675 to -264)	60	32
<i>Pitx3</i> (LEF1 ChIP Primer 3)	AAGGCCTCAATTAGCCACAG GCTAGCGGAGGAGAGAGTGA	388 (-406 to -19)	60	32
<i>Pitx3</i> (LEF1 ChIP Primer 4)	TGGCACTGCACTGTGAGACT CCCCTAGATTTCAAGGTGCTC	478 (-57 to +421)	60	32
<i>Pitx3</i> (LEF1 ChIP Primer 5)	CCTTTACCAGAGAGCACCTTG ACATTCCGTTCACTACTGCT	474 (+391 to +864)	60	32
<i>Pitx3</i> (LEF1 ChIP Primer 6)	CAGACGATGTGTCCAGCAGT TTCGAGTTTGTGAGGGGACT	396 (+831 to +1226)	60	32

Gene (application)	Forward primer (5' → 3') Reverse primer (5' → 3') or mutagenic primer or siRNA (5' → 3')	Product length (bp)	T _m (°C)	Cycles
<i>Pitx3</i> WRE 1 (site-directed mutagenesis) *	tgagaggccgagtgctcacacAGCGAaagcacacaaggccagacagc wt: tgagaggccgagtgctcacac cttgaagcacaca aggccagacagc	#	#	#
<i>Pitx3</i> WRE 2 (site-directed mutagenesis) *	aagatgctgctgataccaAGCGAatttctctgttccaaatcctgc wt: aagatgctgctgatacca cttgaatttctctgttccaaatcctgc	#	#	#
<i>Pitx3</i> WRE 3 (site-directed mutagenesis) *	tgggtctcagctccacaAGCGActgcccgtcgtttgccc wt: tgggtctcagctccaca cttgcgcccgtcgtttgccc	#	#	#
<i>Lef1</i> (RT-PCR)	AAAGAAATGAGGGCGAATGTCGTA GCTGTCATTCTGGGACCTGTACCT	257	59	30
<i>Tcf7</i> (RT-PCR)	CCCCTCAATGCTTTTCATGCTTTAC CGAATGCATTTCTTTTTCCTCCTG	283	59	30
<i>Tcf7l1</i> (RT-PCR)	TACCCCTTCCTGATGATTCCAGAC GGAGAAGTGGTCGTTGCTGTAGGT	288	59	30
<i>TCF7L2</i> (RT-PCR)	GCATCAGGACTCCAAAAGGAAGA TTCCCATAGTTATCCCGTGCAGAC	270	59	30
<i>GAPDH</i> (RT-PCR)	TGAAGGTCGGAGTCAACGGATTTGGT CATGTGGGCCATGAGGTCCACCAC	983	59	30
<i>LMX1A</i> (RT-PCR)	TGTCTGCGAGGGCTGTCAGC CAGCAGCAGAAGCAGCTCAGG	270	60	35
<i>Gapdh</i> (RT-PCR)	ACCACAGTCCATGCCATCAC TCCACCACCCTGTTGCTGTA	450	60	35
<i>LMX1A</i> siRNA #144	GCUUGAUGCACUUAAGUUAdTdT (sense) dTdTTCGAACUACGUGAAUUCAAU (antisense)	—	—	—
nt siRNA (control)	ON-TARGETplus Non-targeting siRNA #1, Cat. No. D-001810-01-05	—	—	—

Footnotes: Oligonucleotides were purchased from Metabion International AG and biomers.net GmbH, Germany. Targeting and non-targeting siRNAs were purchased from Fermentas GmbH (Thermo Fisher Scientific), Germany.

* Mutagenized nucleotides in CAPITALS, *WRE* sequence in **bold**, and predicted LEF1/TCF BS is underlined.

According to the instructions of the manufacturer.