Genetic studies of leptin concentrations implicate leptin in the regulation of early

adiposity

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Abstract

Leptin influences food intake by informing the brain about the status of body fat stores. Rare *LEP* mutations associated with congenital leptin deficiency cause severe earlyonset obesity that can be mitigated by administering leptin. However, the role of genetic regulation of leptin in polygenic obesity remains poorly understood. We performed an exome-based analysis in up to 57,232 individuals of diverse ancestries to identify genetic variants that influence adiposity-adjusted leptin concentrations. We identify five novel variants, including four missense variants, in *LEP, ZNF800, KLHL31*, and *ACTL9*, and one intergenic variant near *KLF14.* The missense variant Val94Met (rs17151919) in *LEP* was common in individuals of African ancestry only and its association with lower leptin concentrations was specific to this ancestry (P=2x10-16, n=3,901). Using *in vitro* analyses, we show that the Met94 allele decreases leptin secretion. We also show that the Met94 allele is associated with higher BMI in young African-ancestry children but not in adults, suggesting leptin regulates early adiposity.

Introduction

Leptin is an adipocyte-derived hormone that helps maintain homeostatic control of fat tissue mass by signaling the status of body energy stores to the appetite-regulating circuits of the brain [1]. Rare homozygous mutations in the leptin (*LEP*) gene can cause complete leptin deficiency that results in hyperphagia and severe early-onset obesity, which can be treated effectively by exogenous leptin administration [2, 3]. Mice and patients heterozygous for these mutations show partial leptin deficiency and increased body weight [4-6].

In the general population, leptin concentrations correlate closely with body fat mass. However, there is wide inter-individual variability; about 10-20% of obese individuals have leptin concentrations that are similar to those observed in non-obese individuals, which is in part due to genetic differences [7, 8]. Twin and family studies suggest that 30-50% of variation in leptin at any given level of adiposity and across different ethnic groups is explained by genetic differences [8]. The implications of this variability for body weight regulation remain poorly understood.

Identification of genetic variants associated with circulating leptin may shed new light on the role of variability in leptin levels in the general population. In a recent genome-wide association study (GWAS) of leptin concentrations, we identified four loci associated with leptin concentrations independent of body mass index (BMI) [9]. The variant most strongly associated with leptin concentrations was rs10487505, located 21 kb upstream from *LEP,* in a region shown to harbor a long non-coding RNA (EST EL947753) that influences the transcriptional control of leptin expression [10]. The leptin-decreasing allele of

rs10487505 was nominally associated with ~0.03 kg/m² higher BMI in adults and 1.05fold increased risk of early-onset obesity [9]. More recently, the association of the leptindecreasing allele of rs10487505 with higher adult BMI, body fat percentage, and risk of extreme obesity was replicated in the UK Biobank [10]. The most pronounced association, however, was observed for body size at 10 years of age; carriers of the leptin-decreasing allele reported being "plumper" at age 10 compared to peers" more frequently than carriers of the allele associated with higher leptin concentration. The association between rs10487505 and childhood body size was recently replicated in 14,521 Norwegian children, and the peak effect of rs10487505 on BMI was observed in 1.5-year-old children [11].

In the present study, we sought to elucidate the genetic basis of leptin concentrations through screening genetic variants with an exome-targeted array in up to 57,232 individuals of European, African, East Asian or Hispanic ancestry. We confirm five previously established and identify five novel variants associated with leptin concentrations, including four missense variants in *LEP, ZNF800, KLHL31*, and *ACTL9*, and one intergenic variant near *KLF14.* The novel *LEP* variant, Val94Met (rs17151919), is associated with leptin concentrations in adults of African ancestry only. The leptinlowering Met94 allele of the rs17151919 variant is associated with higher BMI in young children, but shows a weak or no association with BMI in adulthood, suggesting leptin regulates early adiposity.

Research Design and Methods

Study design

We performed an exome-based association study using data from 35 cohorts comprising up to 57,232 adults (≥18 years) of whom 50,321 were of European descent, 4,387 of African descent, 2,036 of East Asian descent, and 488 of Hispanic descent. We carried out additional analyses in men and women separately. All analyses were performed in models combining studies of all ancestries and in European ancestry cohorts only, for both additive and recessive genetic models. All participating institutions and coordinating centers approved the project. Informed consent was obtained from all study participants. We have reported the study-specific design, sample quality control, and descriptive statistics in **Tables S1-S2**.

Outcome traits

The participating studies acquired residuals for leptin concentrations (in ng/mL) using linear regression, adjusting for age, genome-wide principal components, and any studyspecific covariates (e.g. study center). The residuals were calculated with and without adjustment for BMI. Studies with unrelated individuals acquired the residuals in men and women separately, whereas family-based studies additionally acquired sex-combined residuals adjusting for sex as a covariate. Case-control studies acquired the residuals in cases and controls separately. Finally, we rank-transformed the residuals using inverse normal transformation to follow a distribution with a mean of 0 and a standard deviation of 1.

Genotyping

All participating studies performed genotyping using the Illumina HumanExome BeadChip. The genotype calling was performed using the designated manufacturer's software, followed by zCall. Study-specific quality control measures were implemented before the association analyses to remove poorly genotyped variants (**Table S3**).

Study-level association analyses

Associations of the exome-wide variants with the residuals of leptin concentrations were examined using linear mixed models implemented in either RAREMETALWORKER [12] or RVTEST [13] (**Table S3**). The model accounted for potential cryptic relatedness by incorporating a kinship matrix. We performed the single variant association analyses using both additive and recessive genotypic models. We also calculated covariance matrices capturing LD relationships between markers within 1 Mb for use in gene-level meta-analyses.

Quality control of study-level association results

We applied the EasyQC package in R to association summary statistics from each participating study to identify cohort-specific QC issues. This included (i) identifying issues with calculation of leptin residuals and transformation of the residuals, (ii) identifying strand issues by comparing allele frequencies against reference alleles from the 1000 Genomes Project phase 1, and (iii) identifying issues arising from population stratification.

Single variant meta-analyses

The meta-analyses of summary statistics from the participating studies were carried out using RAREMETAL [14] by two different analysts in parallel. We excluded all variants with a call rate <98%, Hardy Weinberg equilibrium *P*-value <1x10-6, or an allele frequency that strongly deviated from the 1000 Genomes reference frequency (>0.60 for all-ancestry analyses and >0.30 for ancestry-specific analyses). To identify the leptin-associated variants, we used the array-wide Bonferroni-corrected threshold of *P*<2x10-7 for ~250,000 variants in the single variant analyses.

Gene-based meta-analyses

We performed gene-based analyses using the sequence kernel association test [15] (SKAT) and variable threshold [16] (VT) methods in RAREMETAL. The analyses were performed with two different sets of criteria (broad and strict) to select predicted damaging rare and low-frequency variants with MAF<5% annotated using five prediction algorithms: PolyPhen-2, HumDiv, HumVar, LRT, MutationTaster, and SIFT. The broad gene-based tests included nonsense, stop-loss, splice-site, and missense variants that were annotated as damaging by at least one of the five algorithms whereas the strict tests only included variants predicted as damaging by all of the five algorithms. The statistical significance for the gene-based tests was set at a Bonferroni-corrected threshold of *P*<2.5x10-6 for 20,000 genes.

Age-stratified BMI analyses of variants in and near *LEP*

To study the influence of age on the association of the Val94Met variant in *LEP* and the rs10487505 variant near *LEP* with childhood BMI, we performed age-stratified analyses

in children with African and European ancestry from the Center for Applied Genomics at Children's Hospital of Philadelphia (CHOP) cohort recruited from 2006 to present [17]. The participants had multiple BMI measurements at different ages and analyses were performed with measurements in 1-year age bins. The number of BMI measurements in each age bin is shown in **Tables S9** and **S10**. Statistical significance was defined as P<0.05. The Val94Met and rs10487505 variants were genotyped using the Illumina Infinium II HumanHap550 and Human610 BeadChip and imputed to the HRC r1.1 reference panel using the Sanger Imputation Server. All participants were biologically unrelated, aged between 2 and 18 years, and between -3 and +3 standard deviations of CDC-corrected BMI. The study was approved by the Institutional Review Board of the Children's Hospital of Philadelphia. Parental informed consent was given for each study participant.

Additionally, we used information on comparative body size at age 10 (data field 1687) for 452,264 individuals of European ancestry and 8,154 individuals of African ancestry from the UK Biobank. The participants were asked to choose one of the three categories of "about average", "thinner", or "plumper" to describe their body size compared to average when they were 10 years old.

Pathway enrichment analyses

We utilized the EC-DEPICT [18, 19] gene set enrichment analysis method to evaluate nonsynonymous index variants (strongest nonsynonymous variant within ±1 Mb boundary) with *P*<5x10-4 for association with either i) leptin unadjusted for BMI, or ii) leptin adjusted for BMI. EC-DEPICT's primary innovation is the use of "reconstituted" gene sets,

which consist of gene sets downloaded from several databases that have been extended based on publicly available large-scale co-expression data [18]. Two analyses were performed: (i) all coding variants (N=93 loci for leptin unadjusted for BMI and N=91 loci for leptin adjusted for BMI) and (ii) coding variants with MAF<5% only (N=77 loci for leptin unadjusted for BMI and N=65 loci for leptin adjusted for BMI).

We also utilized PASCAL [20] to study the enrichment of exome-wide association results in gene sets and pathways using two estimation approaches: MAX and SUM. The MAX estimation is more powerful for single variant-driven associations whereas the SUM estimation is more powerful when multiple variants are driving the signal [20]. We used reconstituted gene sets from DEPICT and the reference data from UK10K [TwinsUK [21] and ALSPAC [22]] to estimate LD. The PASCAL analyses were performed for all exomechip variants ($N_{all}=265,780$ for leptin adjusted for BMI, $N_{all}=265,780$ for leptin unadjusted). and for coding variants only (N_{coding} =176,035 for leptin adjusted for BMI, N_{coding} =180,864 for leptin unadjusted). No allele frequency or *P*-value thresholds were used to select variants for the PASCAL analyses. The pathway scoring method used by PASCAL combines individual gene scores without the need for a tuneable threshold parameter to determine inclusion of genes in the enrichment analysis [20].

Leptin adjusted for BMI is correlated with body fat free mass (correlation with fat-free mass index in the Fenland cohort = -0.39). The initial pathway analyses for leptin adjusted for BMI using EC-DEPICT and PASCAL suggested enrichment of skeletal-muscle related pathways. To make sure that the gene set enrichment results were not due to correlation between leptin adjusted for BMI and fat-free mass index, we corrected the effect sizes

using the following equation [23]: Beta_{corrected} = beta_{leptin} – (beta_{FFMI} x r_{FFMIvs} LEPTIN), where $r_{FFMIVs,LEPTIN}$ = -0.39 (Pearson correlation coefficient in the Fenland Study). The beta_{FFMI}coefficients were extracted from an ongoing exome-wide association study of fat-free mass index in ~500,000 individuals.

Collider bias

Given that we adjusted leptin concentrations for BMI in our exome-based analyses and leptin and BMI are strongly correlated (r~0.5-0.8) [9], we tested all exome-based significant loci for evidence of collider bias [23-25]. For each index we extracted the association results from our BMI-unadjusted leptin analyses and from the largest published exome-wide analysis for BMI [19]. We corrected BMI-adjusted associations for potential bias due to phenotypic correlation between leptin concentrations and BMI, and compared the strength and significance of association with leptin concentrations unadjusted for BMI, leptin adjusted for BMI, and association with BMI (**Table S6**).

eQTL colocalization analyses

The *cis*-expression quantitative trait locus (*cis*-eQTL) analyses were carried out by using abdominal subcutaneous adipose tissue from 770 participants of the METSIM (Metabolic Syndrome in Men) study who all were Finnish men from Kuopio, Finland [26]. The eQTL mapping in 770 METSIM individuals was performed by EPACTS implementing a linear mixed model to account for the population structure among the samples. The eQTLs were defined as *cis* (local) if the peak association was within 1 Mb on either side of the exon boundaries of the gene. We also identified variants most strongly associated with

genes/transcripts from the index variant ("eSNP"). We used METSIM LD (based on $n=770$, HRC imputation) to assess LD r^2 between the index variant and the lead eSNP. If the pairwise LD was $r^2 > 0.80$, we performed a reciprocal conditional analysis. We tested association between the lead SNP and transcript level when the lead eSNP was included in the model, and *vice versa*.

Expression of the potential causal genes in preadipocytes and mature adipocytes

We compared the expression of the candidate causal genes in the novel leptin-associated loci, including *ZNF800, KLF14, KLHL31*, *ACTL9*, *CNTD1* and *DNAJC18* in preadipocytes and mature adipocytes, two major constituent cell types of adipose tissue. Human preadipocytes isolated from adipose tissue were induced to undergo adipocyte differentiation *in vitro* [27]. RNA samples were obtained from preadipocytes and lipidladen mature adipocytes at post-differentiation day 12.

Impact of Val94Met variant in *LEP* **on leptin protein stability**

We used UCSF Chimera 1.13.1 to model the 3D protein structure and valine-tomethionine substitution in the leptin protein [28]. The Rotamers tool and the Dunbrack Rotamer Libary were used to view and evaluate amino acid sidechain rotamers. The displayed orientation of methionine was chosen based on the clashes and contacts observed in the protein and hydrogen bonds [29]. To predict protein stability, we used SDM [30], iStable [31], Cupsat [32] and iMutant 2.0 [33]. All analyses applied the 3D structure for leptin (ID 1AX8) from the RSCCP Protein Data Bank as the reference data set.

Effect of the Val94Met variant in *LEP* **on leptin protein stability and secretion rate**

We tested the effect of the Val94Met variant on leptin protein stability and secretion rate in HEK293 cells *in vitro*. Human leptin cDNA clone was obtained from Open Biosystems Inc (Huntsville, AL) and subcloned into pcDNA3.1 vector. The original cDNA clone encodes the Val94 variant. The 94Met variant was created using Quikchange II sitedirected Mutagenesis kit (Agilent, Santa Clara, CA), with the Val94 plasmid as template and the following mutagenesis primers (forward: 5'-atgccttccagaaacatgatccaaa tatccaac-3', reverse: 5'-gttggatatttggatcatgtttctggaaggcat-3'). Plasmids carrying Val94 or 94Met cDNA (0.05 μg) were introduced into HEK293 cells (0.65 million cells/well in 12-well plate) using Lipofectamine 2000 as previously described [34]. To measure intracellular leptin protein turnover and secretion rates, cells were treated with protein synthesis inhibitor cycloheximide (CHX, 20 μg/ml) in fresh media for 0.5 and 1 hr at 72 hr post-transfection. Cells incubated with fresh media for 1 hr without CHX were used as untreated controls. Conditioned media were saved for leptin assay, and cell lysate were prepared using NP-40 lysis buffer (50 mM Tris-HCl, pH 8.0, 150 mM NaCl, 1 mM DTT, 1 mM EDTA, 0.5% NP-40, 10% glycerol, and 1x Roche protein inhibitor mixture). Leptin concentrations in cell lysates and the amount of leptin in conditioned media were determined using a human leptin ELISA kit (R&D Systems, Minneapolis, MN). Little or no cell debris was observed in the conditioned media after centrifugation, suggesting little or no cell breakage during the incubation. The experiments were carried in duplicates or triplicates and repeated four times.

Results

Five novel genetic variants show association with leptin concentrations independent of adiposity

To identify genetic variants associated with leptin concentrations, we tested the associations of 246,328 single nucleotide variants (SNVs), genotyped on an exometargeted genotyping array, with leptin concentrations in up to 57,232 individuals of European (n=50,321), African (n=4,387), East Asian (n=2,036) or Hispanic ancestry (n=488) from 35 studies (**Tables S1-S3**). The exome-array provides a detailed coverage of gene-coding regions and includes tags for variants identified in previously published GWASs for human complex traits. Given the strong correlation between leptin and BMI (r~0.5-0.8) [9], we examined associations with leptin concentrations with and without adjustment for BMI. Additional analyses were performed in men (n=23,862) and women (n=32,940) separately. All the analyses were performed in all ancestries combined and in European-ancestry individuals only.

We confirmed five previously established [9] and identified five novel variants associated with leptin concentrations. The novel associations include four missense variants, in *LEP, ZNF800*, *KLHL31*, and *ACTL9*, and one intergenic variant near *KLF14* (**Table 1, Table S4**). The associations at already established loci include intergenic variants near *LEP* and *CCNL1*, a missense variant in *GCKR*, and intronic variants in *COBLL1* and *FTO* (**Table 1, Table S4**). To detect additional independent signals at the 10 leptin-associated loci, we performed conditional analyses, but no further signals were identified.

The association between rs1121980 near *FTO* and leptin concentrations became nonsignificant after adjustment for BMI ($P_{\text{unadi}}=8x10^{-17}$; $P_{\text{adiBM}}=0.45$). The effects of all other known and novel loci were independent of BMI (**Table S5**). We tested whether the adjustment for BMI, a strongly correlated covariate [23], may have introduced collider bias, but found no evidence of such bias (**Table S6**).

The strongest variant associated with leptin concentrations was rs791600, an intergenic variant near the *LEP* gene. The rs791600 variant is in linkage disequilibrium (LD) (EUR r^2 =0.70) with the rs10487505 variant identified in our previously published GWAS [9], which is not included in the exome array and was therefore not available for analyses in the present study. In the prior GWAS study, the rs10487505 variant showed a more significant association with BMI-adjusted leptin concentrations (beta=0.034 per allele, P=2.7x10⁻¹¹, n=29,252) than rs791600 (beta=0.029 per allele, P=3.0x10⁻⁹, n=31,800) and thus is still considered the lead variant at this locus (**Figure S1**).

Nine of the 10 identified loci showed an association with leptin concentrations in all ancestries combined and in European ancestry only analyses. However, the novel *LEP* variant Val94Met (rs17151919) only showed a significant association in all ancestries combined (P=2x10-16) and not in European-ancestry individuals alone (P=0.47, **Table S7**). In further ancestry-stratified analyses, we observed that the Met94-coding allele is common in populations of African ancestry (MAF=8%), less common in those with Hispanic ancestry (MAF=2%), very rare in those with European ancestry (MAF=0.02%) and monomorphic in people with East Asian ancestry [35]. In individuals of African descent, each Met94-coding allele was associated with 0.34 standard deviations (SD)

lower leptin concentrations (*P*=2x10-16, n=3,901) (**Figure S2**). The direction of effect was consistent in individuals with Hispanic (-0.21 SD effect per allele, *P*=0.29, n=488) and European ancestry (-0.19 SD effect per allele, *P*=0.47, n=44,401), but did not reach statistical significance, most likely because very few carriers were available ($N_{HIS}=24$, NEUR=15) (**Table S7**).

Gene-based analysis identifies two novel genes with sex-specific effect on leptin

In addition to single variant-based association tests, we performed gene-based tests using rare and low-frequency coding variants in aggregate [15, 16] (**Methods**). We identified two genes associated with leptin concentrations. *CNTD1* showed association with leptin concentrations unadjusted for BMI in men (*P*=1×10−7) but not in women (*P*=0.27) (**Table 2, Table S8**). The association in men was driven by five coding variants and was strongly attenuated by adjusting for BMI (*P*=0.007), suggesting that the association of *CNTD1* with leptin concentrations may be due to a link between *CNTD1* and adiposity, although no such connection has been previously reported. The *CNTD1* gene encodes cyclin N-terminal domain-containing 1, which is critical for meiotic crossover maturation and deselection of excess pre-crossover sites.

Another gene, *DNAJC18*, showed association with BMI-adjusted leptin concentrations in women (*P*=6×10⁻⁸), but not men (P=0.02). The association in women was driven by two coding variants (**Table 2, Table S8**). *DNAJC18* is part of the Dnaj heat shock protein family. However, no function has yet been described to C18 subfamily.

LEP **Val94Met regulates leptin secretion and early adiposity**

The Val94Met (rs17151919) *LEP* variant was associated with BMI-adjusted leptin concentrations in individuals of African ancestry. A previous study in 2,129 African Americans in the CARDIA study (not included in the present meta-analyses) reported a significant association between the leptin-decreasing Met94 allele of the Var94Met (rs17151919) variant in *LEP* and up to 1.12 kg/m² higher BMI in adulthood (P=0.018) [36]. However, results from two larger studies of BMI by the African Ancestry Anthropometry Genetics Consortium (n=42,752; *P*=0.88) [37] and the African ancestry population of the UK Biobank study (N=7,820, *P*=0.17), did not replicate the association. Nevertheless, among the African ancestry population in the UK Biobank, carriers of the leptindecreasing Met94 allele reported more often that, at age 10, they were "plumper" (compared to peers) (OR=1.11, *P*=0.04), suggesting that the effect of this variant may be age-dependent. To study the influence of age, we performed age-stratified analyses in up to 2,726 children with African-ancestry from the Center for Applied Genomics at Children's Hospital of Philadelphia (CHOP) cohort [17]. Comparing the effect sizes across different age points revealed that each leptin-decreasing Met94 allele was associated with 0.12-0.20 units higher BMI z-score between the ages 3 and 7 (P<0.05). The most pronounced effect was reached at age 6 years (**Figure 1, Table S9**), and no association with BMI was observed after age 8 years (betas -0.04 to 0.05) (**Figure 1, Table S9**), suggesting that the BMI-increasing effect of the Met94 allele wanes shortly before puberty. The rs10487575 variant near *LEP* showed a similar trajectory of association with childhood BMI as the Val94Met variant but the effect sizes were much more modest (**Figure 1, Table S10**), consistent with the five-fold smaller effect of rs10487505 on leptin concentrations compared to Val94Met in adults (**Table 1**).

The Val94Met variant is located at position 94 in the 167 amino acid leptin precursor protein and results in a valine to methionine change at position 73 of the mature protein (**Figure 2A**). Position 73 is situated at the leptin protein surface and is not believed to be involved in binding of leptin to its receptor. Nevertheless, structural prediction tools [30- 33] suggested that the substitution of valine with methionine at this position is likely to lead to reduced stability of the mature leptin protein (**Figure 2A, Figures S3-4, Table S11**). This is consistent with our observation that the methionine-coding allele is associated with lower leptin concentrations.

To study the impact of the Val94Met variant on the intracellular turnover of the leptin protein and its secretion rate, we performed *in vitro* experiments in HEK293 cells. Leptin secretion rate – calculated as the amount of leptin secreted in 1 hour normalized to the respective cellular leptin content – was 20.4% lower in Met94 than in Val94 cells (*P*=0.0007 by repeated measures 1-way ANOVA) 72 hours post-transfection (**Figure 2B**). Leptin secretion rates between 48-72 hours post-transfection and during a 1-hour treatment with cycloheximide were 11.8% (*P*=0.0005) and 17.9% (*P*=0.0002) lower, respectively, in Met94 compared to Val94 (**Figure S5**). Notably, no difference was found in the intracellular turnover rate of leptin between Val94 and Met94 cells during a 0.5 or 1-hour incubation with cycloheximide to impair protein synthesis (**Figure 2C**). The unchanged turnover rate incorporates protein secretion and degradation, suggesting that decreased leptin secretion rate was likely associated with increased intracellular leptin degradation in Met94 cells. Overall, these *in vitro* experiments suggest that methionine substitution in position 73 of the mature leptin protein decreases the rate of leptin

secretion from the cells, which may contribute to the association of the Met94 allele with lower leptin concentrations.

ZNF800 **locus regulates adipose gene expression and body composition**

The Pro103Ser (rs62621812) variant in *ZNF800* changes the amino acid sequence of CH2 zinc finger protein, a putative transcription factor [38]. We found that the Ser103 allele (frequency=2.8%) is associated with lower BMI-adjusted leptin concentrations (*P*=2.0x10-12). As shown before [26], Pro103Ser is the lead variant associated with expression of *ZNF800* in subcutaneous adipose tissue in the Finnish METSIM Study (*P*=2.4x10-16); the Ser103 allele is associated with higher *ZNF800* expression levels (**Table S12**). *ZNF800* is a master regulator in subcutaneous adipose tissue, as the Pro103Ser variant has also been associated with adipose tissue expression of nine other genes [26]. In the eQTL data, the leptin-decreasing Ser103 allele was not significantly associated with the expression of *LEP* (*P*=0.20), located 866 kb downstream, and the observed direction of the effect on *LEP* expression was opposite to that observed for leptin concentrations (beta=0.14 SD/allele vs. beta= -0.13 SD/allele, respectively), suggesting that the leptin-lowering effect of the Ser103 allele on leptin concentrations is unlikely to be mediated by direct transcriptional regulation of *LEP*.

In the UK Biobank study, we found that each leptin-decreasing Ser103 allele is associated with 0.14 kg/m² higher BMI (P=8.1x10⁻⁶). However, there was no association between Ser103 allele and body fat percentage (-0.045% per allele, *P*=0.25), indicating that the variant impacts BMI primarily by increasing fat free body mass. Indeed, the leptindecreasing Ser103 allele was associated with a 0.33 kg higher fat free mass (*P*=4.6x10-

²⁰) and only 0.13 kg higher fat mass (*P*=0.023). The Ser103 allele is associated with higher expression of the *ZNF800* gene in the tibial nerve (GTEx v8, P=1.4x10-6, n=532) that innervates the muscles of the leg, and has been previously identified for association with increased appendicular lean mass [39]. There was no association between the Ser103 allele and self-reported body size at age 10 (P=0.75).

The *KLF14* **locus regulates adipogenesis and fat distribution**

The rs972283 variant (MAF_{EUR}=48%), associated with leptin concentrations, is located 51 kb upstream from *KLF14* and 2.5 Mb downstream from *LEP*, and is in near-perfect LD with previously reported GWAS variants for type 2 diabetes [40], insulin-resistance [41], HDL cholesterol [42], and body fat distribution [43]. As reported earlier [26], rs972283 is associated with *KLF14* expression in subcutaneous adipose tissue (**Table S12**). As *KLF14* is a master regulator in adipose tissue, rs972283 is also associated with the expression of multiple other genes in *trans* [44]. No significant association was observed between rs972283 and *LEP* expression in the METSIM eQTL study [44], suggesting that *KLF14* may not regulate leptin production at the transcriptional level, at least not in men. Lower expression of *KLF14* has been implicated in impaired adipogenesis due to defective adipocyte glucose uptake in women, characterized by the presence of fewer but larger adipocytes and a shift in fat distribution from gynoid stores to abdominal tissues [44]. However, while the effects of *KLF14* on adipogenesis and adipose redistribution have been found to be specific to women, there was no difference in the association of rs972283 with leptin levels between men and women (**Table S4**).

Interestingly, the carriers of the rs972283-G allele reported more frequently being plumper $(P=2.8x10^{-5})$ and shorter (P=0.014) than average at age 10 in the UK Biobank than noncarriers, whereas the same allele was associated with a lower BMI (P=6.8x10-9) and increased height (P=0.010) in adults, suggesting that the effect of the rs972283 variant on body size may change during life course. In previous GWAS of adults, the rs972283- G allele has been identified to be associated with higher risk of type 2 diabetes [40] and insulin resistance [41], and lower hip circumference (adjusted for BMI) [43] and HDL cholesterol [42]. In the UK Biobank study, the rs972283-G allele was associated with lower body fat percentage in adults (P=5.9x10-22).

The *KLHL31* **locus is implicated in adipogenesis in adult females**

The Val156Ile (rs3799260) variant (MAF_{EUR}=18%) in *KLHL31*, associated with leptin concentrations in female-only analyses, changes the amino acid sequence of the kelchlike family member 31 protein. *KLHL31* suppresses Wnt-β-catenin signaling that is involved in promoting adipocyte differentiation and suppressing oxidative metabolism in adipocytes. The Val156Ile variant is predicted to be benign/tolerated by SIFT/Polyphen [45, 46]. Previous genetic associations have identified a variant in low LD (rs7739232; EUR r^2 =0.27) to be associated with BMI-adjusted hip circumference, also specific to women [43]. The rs7739232 variant was not included in the exome-array and was thus not analyzed in the present study. Our *in vitro* experiments showed that *KLHL31* is only expressed in mature adipocytes, but not in preadipocytes (**Figure S6**), suggesting that the gene is developmentally regulated.

In the UK Biobank, similar to the variants in and near *LEP*, the carriers of the leptindecreasing Ile156 allele reported more often being plumper than average at age 10 $(P=5.6x10^{-6})$, but there was a weaker association with higher BMI (P=0.045) in adulthood.

In men, the *ACTL9* **locus may regulate leptin concentrations in a cell nonautonomous fashion**

Homozygosity for the minor allele of the Ser37Phe (rs2340550) variant in *ACTL9* was associated with leptin concentrations in men only in a recessive genetic model. While the Ser37Phe variant is predicted to be benign/tolerated by SIFT/Polyphen, another missense variant, Ala51Val (rs10410943), in high LD (EUR $r^2=0.99$) is predicted to be deleterious/probably damaging and could be the causal variant at the locus. The Ser37Phe variant is also in high LD (r^2 >0.8) with several nearby non-coding variants (**Figure S7**). However, none of these overlaps with regulatory elements in adipocytes. The expression of *ACTL9* is restricted to the testis and it is therefore likely to act in an adipocyte non-autonomous fashion to influence leptin concentrations. Actin proteins have cytoskeletal functions and have also been implicated in signaling and nuclear activities.

Gene-set analyses implicate adipocyte-related pathways

We performed gene-set enrichment analyses using EC-DEPICT [18, 19, 47] and PASCAL [20] to identify biological processes and candidate pathways enriched for loci associated with leptin unadjusted or adjusted for BMI. Among coding variants associated with BMIunadjusted leptin concentrations, PASCAL identified significant enrichment of the geneset for "positive regulation of reproductive success" ($P_{empirical}$ =1.6x10⁻⁵) (**Table S13**),

consistent with the crucial permissive role of leptin in the integrity of the gonadal axis [48]. Among coding variants associated with leptin adjusted for BMI, we found enrichment of the immune-related TRIM39 protein-protein interaction subnetwork [49, 50] (*P*empirical=8.4x10-6) (**Table S14**). No gene sets were found to be significantly enriched in PASCAL analyses where all exome-wide variants (coding and non-coding) for leptin adjusted for BMI were included, nor in the EC-DEPICT analyses.

Discussion

We identified 10 genetic variants associated with leptin concentrations and two genebased associations using an exome-based genotyping array in up to 57,232 individuals with varying ancestries. The two independent variants most strongly associated with leptin concentrations were located in and near the *LEP* gene. The African ancestry-driven variant within *LEP*, Val94Met (rs17151919), was found to decrease leptin secretion in HEK293 cells whereas rs10487505 located near *LEP* overlaps a lncRNA that regulates *LEP* expression [10]. Both variants showed significant association with increased adiposity in children, whereas only a nominal or no association was observed in adults.

Previous analyses have shown that the leptin-lowering allele of rs10487505 is only weakly associated with higher BMI in adulthood but shows a pronounced association with BMI in early childhood [10, 11]. Similarly, we showed that the *LEP* Met94 allele, associated with lower leptin concentration, is associated with early childhood BMI. Our results suggest that leptin has an impact specifically on early adiposity, encouraging further studies to uncover the molecular mechanisms that underlie this age-dependent relationship between leptin and BMI.

The Val94Met and rs10487505 variants in and near *LEP* are likely to influence leptin concentrations by different molecular mechanisms. The novel African ancestry-driven variant Val94Met may affect circulating levels of leptin by reducing leptin secretion. The rs10487505 variant is associated with leptin mRNA levels in adipose tissue. Located upstream of *LEP* within a lncRNA (EL947753), we hypothesize that this variant interacts with enhancer regions to regulate the expression of *LEP*. Defects in *LEP* regulation in

mice lead to a relative hypoleptinemic form of obesity that is responsive to leptin administration [10].

We identified four new loci associated with leptin concentrations, located in or near the *ZNF800*, *KLF14*, *KLHL31* and *ACTL9* genes. Two additional genes, *CNTD1* and *DNAJC18* were identified in gene-based analyses. The *ZNF800* and *KLF14* genes are master *trans*-regulators of adipose tissue gene expression [26] and located in the proximity of the *LEP* gene (866 kb and 2.5 Mb away, respectively). The variants in *ZNF800* and near *KLF14* were not associated with *LEP* mRNA levels, however, suggesting that they may be involved in translational or post-translational rather than transcriptional regulation of leptin production. *KLHL31* has been shown to promote adipocyte differentiation and suppress oxidative metabolism in adipocytes, whereas *ACTL9* is not expressed in adipocytes and could affect circulating levels by a non-cell autonomous mechanism. The *KLHL31* and *ACTL9* loci, and the *CNTD1* and *DNAJC18* genes, were only identified in sex-specific models and narrowly passed the array-wide significance threshold. Further validation of the association of these loci with leptin concentrations is warranted.

In summary, we identified a new genetic association of an African ancestry-specific missense variant rs17151919 in *LEP* with leptin concentrations and replicated the association of the rs10487505 variant near *LEP*. The pronounced association of these variants with BMI in early childhood implicates genetic regulation of *LEP* in early growth and suggests that young children may be particularly sensitive to the metabolic/behavioral effects of leptin. We also identified novel loci at *ZNF800*, *KLF14*, *KLHL31*, *ACTL9*, *CNTD1*

and *DNAJC18* associated with leptin concentrations, providing additional insights into leptin physiology.

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consultation.pdf?phpMyAdmin=trmKQlYdjjnQIgJ%2CfAzikMhEnx6.

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Guarantor Statement

HY and TOK are the guarantor of this work and, as such, had full access to all the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis.

Conflict of Interest statement

This work was conducted prior to M.E.G's current affiliation with the National Heart, Lung, and Blood Institute, and, as such, the views expressed in this article do not represent the views of the NHLBI, NIH, or other government entity. D.M.-K. is a part-time clinical research consultant for Metabolon, Inc. M.A.N's participation is supported by a consulting contract between Data Tecnica International and the National Institute on Aging, National Institutes of Health. V.S. has served in advisory boards for Novo Nordisk and Sanofi and received honoraria from these companies. He also has ongoing research collaboration with Bayer Ltd (all unrelated to the present study). B.M.P. serves on the Steering Committee of the Yale Open Data Access Project funded by Johnson & Johnson. J.R.K. reports stock ownership in Bristol Myers Squibb, Johnson & Johnson, Merck, and Pfizer.

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Tables

Table 1. Leptin-associated loci identified in exome-based association analyses

The chromosomal positions are based on hg19.

Chr, chromosome; EA, Effect allele; OA, Other allele; EAF, Effect allele frequency; LeptinAdjBMI, leptin adjusted for body mass index
Table 2. Leptin-associated genes identified by gene-based exome-wide association analyses

The chromosomal positions are based on hg19.

Chr, chromosome; LeptinAdjBMI, leptin adjusted for body mass index

Figure 1. Association of the leptin-decreasing alleles of the *LEP* **Val94Met (rs17151919) variant (on the left) and the rs10487505 variant near** *LEP* **(on the right) with BMI standard deviation score (SDS) in the CHOP cohort.** The analyses for the Val94Met variant were performed in up to 2,726 African ancestry participants and the analyses for the rs10487505 variant in up to 3,681 African and European ancestry participants of the CHOP cohort. The y-axis reports the effect of each leptin-decreasing allele on BMI at each age year. The error bars indicate 1 standard error of the mean (SEM).

Figure 2. Impact of Val94Met transversion at LEP rs17151919 on leptin secretion rate in HEK293 cells. The rs17151919 variant changes valine to methionine in position 73 of the mature leptin protein. A) The 3D illustration of leptin structure derived from RSCCB Protein Data Bank and modified with UCSF Chimera1.13.1. The prediction of protein stability is derived from the SDM2 server [30]. B) Leptin secretion rates for Val94 and Met94 expressed as the amount of leptin secreted in ng during a 1 hr incubation (72-73 hr post-transfection) (LEPs/hr) normalized by the respective cellular leptin content (LEPc) in untreated control cells at the end of incubation. Individual data points from four separate experiments (each with 2-3 technical replicates) are plotted. The normality of data distribution was examined using D'Agostino & Pearson normality test (p=0.65 and 0.54 for LEPV94 and LEPM94, respectively) and repeated measures oneway ANOVA was performed to assess the difference in secretion rate between the genotypes. Mean ± SD and AVOVA results (F and p values) are reported in the table below the graph. C) Intracellular leptin turnover rates for Val94 and Met94 alleles, obtained by measuring the relative cellular leptin contents in the untreated control cells

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(defined as 1 for the respective *LEP* variant) and in samples treated with the protein synthesis inhibitor cycloheximide (CHX, 20 μ g/ml) for 0.5 and 1.0 hour. Mean \pm SD at each time point from four separate experiments (each with 2-3 technical replicates) are plotted. Paired t-test was used to assess the genotype effect on the fractions of cellular LEP remained after 0.5 hr and 1 hr of CHX treatment (p values are reported in the table below the graph). The average hourly turnover rates for Val94 and Met94 were 61±2%, and 60±3%, respectively, calculated by subtracting the percent cellular LEP remained after one hour of CHX treatment from those of the respective untreated controls (defined as 100%).

Figure 1

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LEPV94

LEPM94

 $F(1,8)$

p

 1.62 ± 0.18

1.29±0.09

28.82

0.0007

Met9

Table S1. Study design, number of individuals and sample quality control for ExomeChip study cohorts

* Call rate to exclude individuals for whom genotyping success rate is less than a certain percentage (to exclude 'bad' samples/DNA)

**Exome-chip samples from this study

Table S2. Study-specific descriptive statistics of ExomeChip cohorts.

* only report descrptives for the individuals included in each of the analyses

CHS NOTE: For age and BMI, I included all individuals who are included in one or more of the analyses

CHS NOTE: For leptin and adiponectin, I included all individuals in the biggest analysis (not adjusted for fat percentage or BMI)

Table S3. Information on genotyping methods, quality control of SNPs, imputation, and statistical analysis for ExomeChip study cohorts

* Call rate to exclude SNPs for which less than a certain percentage of individuals were successfully genotyped (i.e. to exclude 'bad' SNPs)

** Includes monomorphic SNPs

Table S4. Single-variant results in all statistical models for the leptin-associated loci

The chromosomal positions are based on hg19.

Chr, chromosome; EA, Effect allele; OA, Other allele; EAF, Effect allele frequency; LeptinAdjBMI, leptin adjusted for body mass index

Table S6. Examination of collider bias with BMI among the exome-array significant loci associated with leptin adjusted for BMI

xL, Effect size for leptin

pL, P value for leptin

xLadjB, Effect size for leptin adjusted for BMI

pLadjB, P value for leptin adjusted for BMI

xLadjBa, Corrected effect size for leptin adjusted for BMI

xB, Effect size for BMI

pB, P value for BMI

* The *ACTL9*, *KLHL31*, and *LEP* rs17151919 loci reached array-wide significance (P<2x10-7) in meta-analyses of European-ancestry men (recessive model), all-ancestry women, and Africanancestry men and women combined, respectively. The results shown are from meta-analyses of European ancestry individuals only.

Table S7. Ancestry-specific results for the Val94Met (rs17151919) missense variant in *LEP*

EA, effect allele; OA, other allele; EAF, effect allele frequency

Table S8. Gene-based results in all statistical models for leptin-associated genes

Table S9. Association of the leptin-decreasing Met94 allele of *LEP* **Val94Met (rs1715919) with BMI z-score in African-ancestry children from the CHOP cohort.**

Table S10. Association of the leptin-decreasing C allele of rs10487505 near *LEP* **with BMI z-score in a metaanalysis of African-ancestry and European ancestry children from the CHOP cohort.**

Table S11. Predicted change in leptin protein stability upon the Val94Met change (Val73Met in the mature leptin protein) in the amino acid sequence

Table S12. Colocalization of METSIM subcutaneous adipose tissue eQTLs at GWAS loci for leptin

LD r2 calculated using 770 METSIM samples (Finnish males) included in eQTL data

A1 (column E) is the leptin raising allele from the Exome Chip analysis. A1 is also the effect allele for the effect sizes listed in columns H and J. Allele 1 in column O is the effect allele for the effect in columns O/Q. FDR<1% (P < 2.37 x 10-4)

Table S13. PASCAL gene set enrichment analysis results for leptin unadjusted for BMI using coding variants only.

(A) Leptin not adjusted for BMI, European, additive model, sex-combined analysis. Coding variants included. SUM method used (Bonferroni correction for 1000 gene sets and 2 traits: P<2.5E-05 for both chi2Pvalue and empPvalue)

(B) Leptin not adjusted for BMI, European, additive model, sex-combined analysis. Coding variants included. MAX method used (Bonferroni correction for 1000 gene sets and 2 traits: P<2.5E-05 for both chi2Pvalue and empPvalue)

Table S14. PASCAL gene set enrichment analysis for leptin adjusted for BMI using coding variants only.

SUPPLEMENTARY INFORMATION

Yaghootkar H, Zhang Y, Spracklen CN, Karaderi T, Huang LO, Bradfield J, et al. Genetic studies of leptin concentrations implicate leptin in the regulation of early adiposity

Figure S1. Association of rs10487505 and rs791600 variants near *LEP* with leptin concentrations adjusted for BMI in a genome-wide association study of up to 32,161 individuals of European ancestry (Kilpeläinen et al., 2016).

Figure S2. Meta-analysis of the association of the Met94 allele of rs17151919 with leptin concentrations adjusted for BMI in cohorts of African ancestry.

Figure S3: Surface region of the leptin protein with the Val94Met position (Val73Met in the mature protein) highlighted.

Figure S4: Leptin structure and the predicted impact of mutagenesis in position 73 from valine to methionine. The Rotamer list on the left shows sidechain torsions (Chi 1 and 2), with the probability and number of interatomic clashes, i.e. unfavourable interactions where atoms are too close together. On the right, the lower picture shows all possibilities for sidechain torsions when methionine is substituted with valine, whereas the upper picture displays the substitution with the highest probability (marked with red square in the Rotamer list).

Fig S5. Impact of Val94Met transversion at *LEP* **rs17151919 on leptin secretion rate in HEK293 cells in different conditions.** A) Leptin secretion rates for Val94 and Met94 during a 24-hr incubation period (48-72 hr post-transfaction), expressed as the amount of leptin secreted in ng per hour over 24 hrs (LEPs/hr) normalized by the respective cellular leptin content (LEPc, ng) at the end of incubation. B) Leptin secretion rates for Val94 and Met94 during a 1-hour incubation (72-73 hr post-transfection) in the presence of cycloheximide (CHX, 20 μg/ml) expressed as the amount of leptin secreted in ng during the 1-hour incubation (LEPs/hr), normalized by the respective cellular leptin content (LEPc, ng). Individual data points from four separate experiments (each with 2-3 technical replicates) are plotted. All data passed D'Agostino & Pearson normality test and repeated measures one-way ANOVA was performed to assess the difference in secretion rate between the genotypes. Mean ± SD and AVOVA results (F and p values) are reported in the table below each graph. C-D. The amounts of leptin secreted (LEPs) during a 1 hr incubation (72-73 hr posttransfection) in untreated control cells (C), and the corresponding cellular leptin content (LEPc) at the end of the

incubation (D). Leptin secretion rates shown in Fig 2B were ratios of the amounts of leptin secreted (LEPs) over the corresponding cellular leptin contents (LEPc) shown here.

Figure S6. Expression of leptin modifiers in human preadipocytes and mature adipocytes. De-identified human subcutaneous adipose stromal cells were generously provided by the Boston NORC and were cultured and differentiated as previously described (Lee and Fried, 2014). Preadipocytes and *in vitro*-differentiated adipocytes from two females and two males were studied. Lipid-laden cells were assayed between 10-14 days after initial treatment with differentiation factors. Transcript levels were determined by RT-qPCR, normalized to the geometric mean of *RPLP0* and *PPIA*, and expressed relative to levels in preadipocytes. Two-way repeated measures ANOVA with posthoc Sidak's multiple comparison tests were performed *: p<0.05, **: p<0.01, ****: p<0.0001, ns (no statistical difference) are indicated, comparing the transcript levels between preadipocytes and mature adipocytes. There was an interaction

between sex and differentiation stage for ZNF800 (p<0.01). No *ACTL9* transcript was detected (nd: none detected). Genotypes of the individuals were marked as R-reference allele and A-alternative allele.

Figure S7. Linkage disequilibrium between the Ser37Phe (rs2340550) variant in *ACTL9* and variants within ±500 kb in the 1000 Genomes European ancestry reference panel. The numbering refers to Regulome DB score of the variants (www.regulomedb.org). Non-coding variants are marked in orange color and coding variants in red. The plot was produced using LDlink (https://ldlink.nci.nih.gov).

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