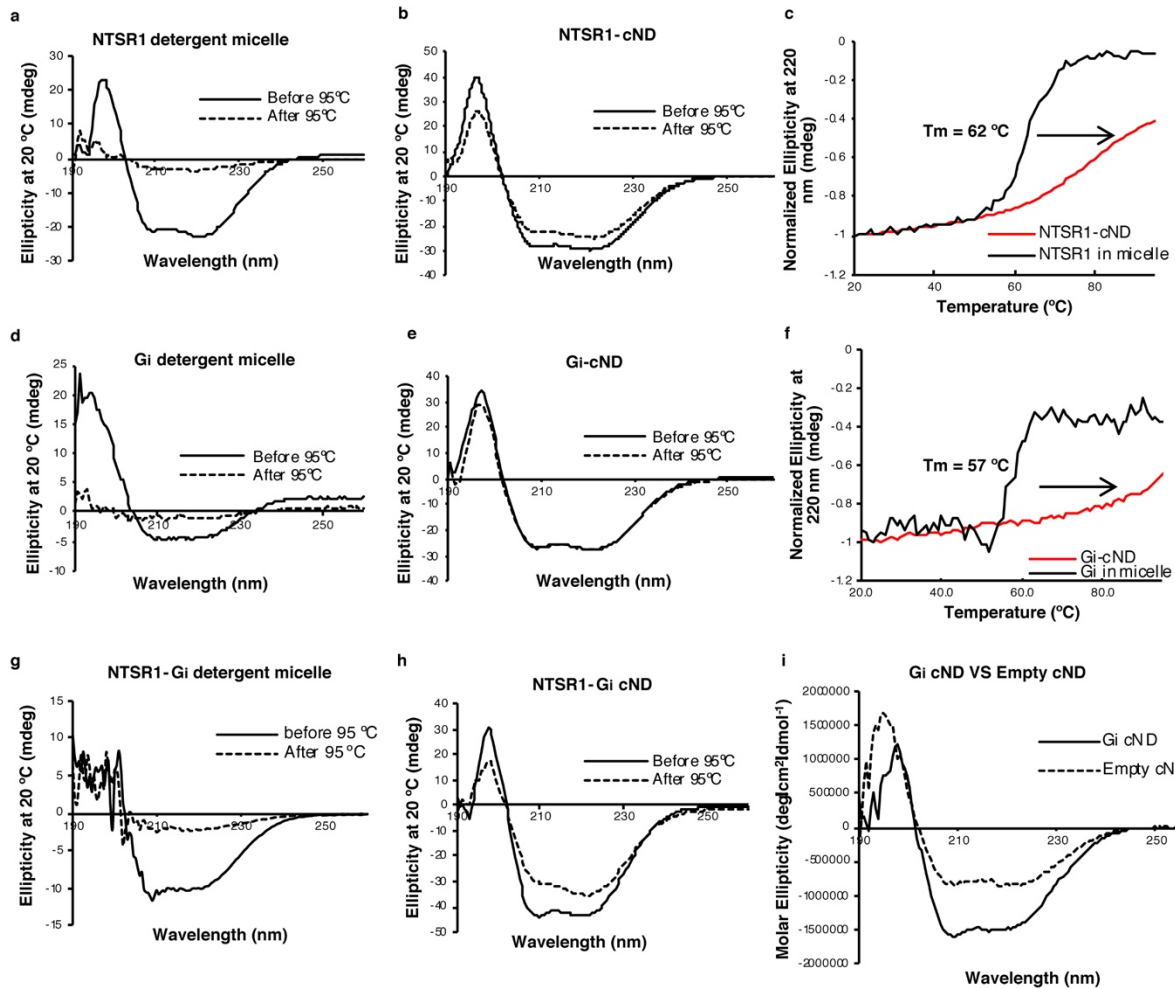

Supplementary information

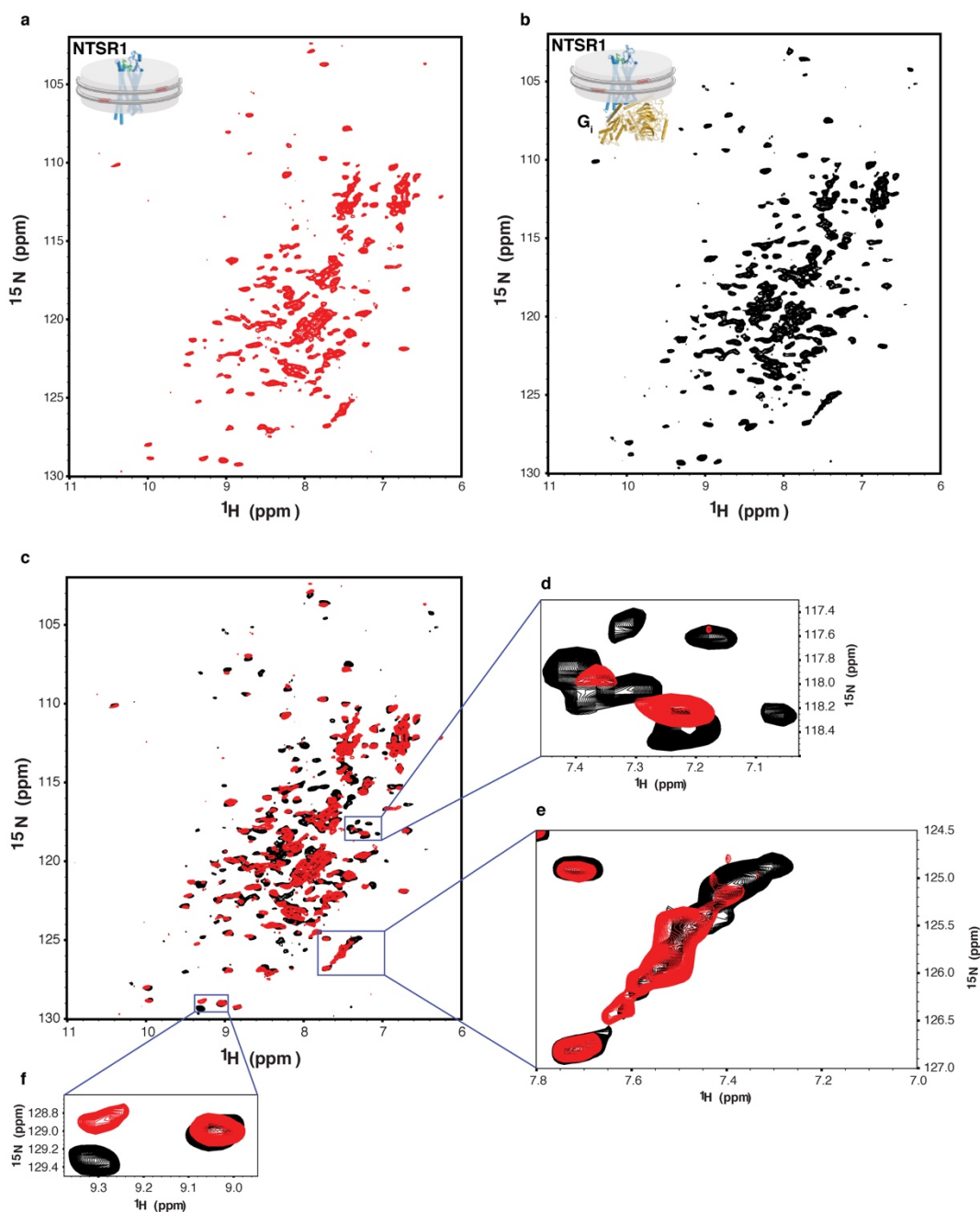
Cryo-EM structure of an activated GPCR–G protein complex in lipid nanodiscs

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Supplementary Fig. 1 | Thermostability enhancement of NTSR1, G_i , and NTSR1- G_i complexes by incorporation into cNDs. **a-b**, Circular Dichroism (CD) spectra at 20 °C before (solid line) and after (dashed line) treatment at 95 °C of **(a)** NTSR1 in DH₇PC detergent micelles; **(b)** NTSR1 in cNDs. **c**, Temperature-dependent CD signals of NTSR1 in detergent micelles (black) and cNDs (red) at 220 nm. **d-e**, CD spectra at 20 °C before (solid line) and after (dashed line) treatment at 95 °C of **(d)** G_i in DH₇PC detergent micelles; **(e)** G_i in cNDs. G_i was reconstituted into cNDs by incubation with POPC/POPG lipid, cNW9, and cholate, followed by detergent removal and size-exclusion chromatography. **f**, Temperature-dependent CD signals of G_i in detergent micelles (black) and cNDs (red) at 220 nm. The melting temperature (T_m) of cNDs is 93 °C (data

not shown) and therefore does not affect transitions before this temperature. **g-h**, CD spectra at 20 °C before (solid line) and after (dashed line) treatment at 95 °C of **(g)** NTSR1-G_i in LMNG/GDN/CHS detergent micelles; **(h)** NTSR1-G_i in cNDs. **i**, CD spectra of 2 µM G_i-cND (solid line) and 2 µM empty cND (dashed line), showing nearly 50% signal contribution from G_i. NTSR1 and G_i account for at least 50% of CD signals even in the presence of cNDs. NTSR1 in detergent micelles irreversibly unfolds during temperature increase with a T_m of 62 °C. In contrast, NTSR1-cND changes structure around 80 °C and does not lose much secondary structure after decreasing temperature to 20 °C. Similar observations were made for G_i, where the protein irreversibly and completely unfolds with T_m of 57 °C in detergent micelles but displays no clear transition temperature in cNDs. For the NTSR1-G_i complex in cND, only mild unfolding was observed around 82 °C. These observations indicate that lipid bilayers improve the stability of NTSR1, G_i and NTSR1-G_i complexes relative to detergent micelles.



Supplementary Fig. 2 | Characterization of the interaction between NTS-NTSR1 and G_i in cNDs by two-dimensional 1H , ^{15}N -TROSY HSQC NMR spectroscopy. a-b, NMR spectrum of ^{15}N -labeled NTS-NTSR1 in cNDs in the (a) absence and (b) presence of G_i . **c**, Overlay of (a) (red) onto (b) (black) showing structural and dynamical changes of NTS-NTSR1 upon binding to G_i in cNDs. **d**, A region showing conformational stabilization of NTSR1. More peaks are observed in

the presence of G_i , suggesting that NTSR1 is highly dynamic in the absence of G_i and resonances are averaged out among a wide range of conformers resulting in low signal-to-noise ratio and even disappeared peaks. Upon interaction with G_i , NTSR1 is stabilized into fewer conformers and becomes less dynamic, which leads to better signal-to-noise ratio and more resonances being observed. **e**, A region showing dynamically slow-exchange shift of NTSR1 upon interaction with G_i . **f**, A region showing chemical shift perturbation of NTSR1, suggesting conformational change of NTSR1 upon binding to G_i in cNDs.