

Original research

Whole genome and exome sequencing identify NDUFV2 mutations as a new cause of progressive cavitating leukoencephalopathy

Zhimei Liu ⁽¹⁾, ¹ Li Zhang, ^{2,3} Changhong Ren, ¹ Manting Xu ⁽¹⁾, ¹ Shufang Li, ¹ Rui Ban, ¹ Ye Wu, ⁴ Ling Chen, ⁵ Suzhen Sun, ⁵ Matthias Elstner, ⁶ Masaru Shimura, ⁷ Minako Ogawa-Tominaga 💿 ,⁷ Kei Murayama,⁷ Tieliu Shi,^{2,3} Holger Prokisch,^{1,8,9} Fang Fang

ABSTRACT

Background Progressive cavitating leukoencephalopathy (PCL) is thought to result from mutations in nuclear genes affecting mitochondrial function and energy metabolism. To date, mutations in two subunits of complex I, NDUFS1 and NDUFV1, have been reported to be related to PCL.

Methods Patients underwent clinical examinations, brain MRI, skin biopsy and muscle biopsy. Whole-genome or whole-exome sequencing was performed on the index patients from two unrelated families with PCL. The effects of the mutations were examined through complementation of the NDUFV2 mutation by cDNA expression.

Results The common clinical features of the patients in this study were recurring episodes of acute or subacute developmental regression that appeared in the first years of life, followed by gradual remissions and prolonged periods of stability. MRI showed leukoencephalopathy with multiple cavities. Three novel NDUFV2 missense mutations were identified in these families. Complex I deficiency was confirmed in affected individuals' fibroblasts and a muscle biopsy. Functional and structural analyses revealed that these mutations affect the structural stability and function of the NDUFV2 protein, indicating that defective NDUFV2 function is responsible for the phenotypes in these individuals. **Conclusions** Here, we report the clinical presentations, neuroimaging and molecular and functional analyses of novel mutations in *NDUFV2* in two sibling pairs of two Chinese families presenting with PCL. We hereby expand the knowledge on the clinical phenotypes associated with mutations in NDUFV2 and the genotypes causative for PCL.

Leukoencephalopathy is seen in disorders of toxic,

acquired vascular or infectious origin as well as in

inherited disorders. Inherited leukoencephalopathies

are a large group of heterogeneous disorders that result

in white matter abnormalities.^{1 2} Cavitating leukoen-

cephalopathy associated with mitochondrial complex I

deficiency was first described in 1999,³ while progres-

sive cavitating leukoencephalopathy (PCL) was first

proposed in 2005 and is thought to result from muta-

tions in nuclear genes affecting mitochondrial func-

tion and energy metabolism.⁴ In 2011, Ferreira and

colleagues first reported that the NDUFS1 gene was

related to PCL and associated with respiratory chain

complex I (Co I) deficiency. The study concluded that

the clinical and neuroradiological entity of PCL is a syndrome strongly related to mitochondrial respiratory chain enzyme deficiency.⁵ Subsequently, NDUFV1 and other genes involved in mitochondrial function and energy metabolism were also reported to be related to PCL.⁶⁻¹⁰ Both NDUFS1 and NDUFV1 encode mitochondrial Co I subunits.

Co I (NADH: quinone oxidoreductase) is by far the largest and most complicated enzyme of the respiratory chain and is composed of more than 40 subunits, 14 central subunits and up to 32 accessory subunits, 7 of which are encoded by mitochondrial DNA and the remaining by nuclear DNA (nDNA).5611 Co I can be subdivided into three functional modules: the electron input (N) module that oxidises NADH, the P module that pumps protons across the membrane and the electron output (Q) module that reduces quinone. The N module consists of the subunits encoded by the nDNA genes NDUFV1, NDUFV2 and NDUFS1.611

NDUFV2 is a highly conserved core subunit of Co I and functions in the transfer of electrons from NADH to the respiratory chain. To date, only six likely pathogenic mutations in NDUFV2 have been reported, of which two mutations (c.427C>T, c.580G>A) result in Leigh syndrome,¹² two mutations (c.120+5 120+8delGTAA, c.669 670insG) lead to hypertrophic cardiomyopathy and encephalopathy^{13–17} and Leigh syndrome,¹⁷ while the other two mutations (c.86C>T, c.626A>G) are likely related to Parkinson's disease.¹⁸¹⁹ To date, no leukoencephalopathy phenotype has been reported.

Here, we report the clinical findings, neuroimaging and molecular and functional analyses of novel mutations in NDUFV2 in four Chinese patients (two sets of siblings) presenting with PCL. Thus, we expand the knowledge on the clinical phenotypes of NDUFV2 and the genotypes of PCL.

METHODS

Participants

Sibling pairs from two families (F-1 II-1 and II-2; F-2 II-1 and II-2) participated in this study, and their pedigrees are shown in figure 1. The families were recruited by the Department of Neurology, Beijing Children's Hospital, Capital Medical University and by the Department of Pediatrics, Peking University First Hospital (China).

For numbered affiliations see end of article. Correspondence to Professor Fang Fang,

Additional material is

published online only. To view,

please visit the journal online

(http://dx.doi.org/10.1136/

imedgenet-2020-107383).

Department of Neurology, Beijing Children's Hospital, Capital Medical University, National Center for Children's Health, Beijing, China; 13910150389@163.com Professor Tieliu Shi; tieliushi@yahoo.com Professor Holger Prokisch; Prokisch@helmholtz-muenchen. de

ZL and LZ contributed equally.

Received 9 August 2020 Revised 21 December 2020 Accepted 10 January 2021

INTRODUCTION

Check for updates

© Author(s) (or their employer(s)) 2021. Re-use permitted under CC BY-NC. No commercial re-use. See rights and permissions. Published by BMJ.

To cite: Liu Z, Zhang L, Ren C, et al. J Med Genet Epub ahead of print: [please include Day Month Year]. doi:10.1136/ jmedgenet-2020-107383

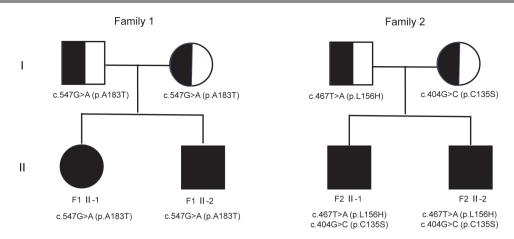


Figure 1 Pedigree of the families. Black/white represents the healthy parents with heterozygous variant in *NDUFV2*, while the black symbols depict the affected patients with homozygous or compound heterozygous variant in *NDUFV2*.

Next-generation sequencing

Targeted next-generation sequencing was performed for each patient, which contained 115 genes involved in vanishing white matter disease, Krabbe disease, adrenoleukodystrophy, metachromatic leukodystrophy, Alexander disease and so on. No suspected disease-causing mutations were found. Therefore, we further applied whole-genome sequencing (WGS) or wholeexome sequencing (WES) to the individuals' DNA extracted from peripheral blood. Family 1 (including the two sibling patients and their parents) was investigated by WGS, and Family 2 was investigated by WES. WES coding regions were enriched using the Agilent SureSelect Human All Exon V6 Kit followed by sequencing as 150 bp paired-end runs on an Illumina HiSeq2000. WGS sequencing libraries were prepared using Illumina TruSeq Nano Kits and sequenced on Illumina HiSeq X sequencers via 2×150 bp reads. Reads were aligned to the human reference genome (UCSC Genome Browser build hg19, https://genome. ucsc.edu/) using Burrows-Wheeler Aligner²⁰ (BWA, http://biobwa.sourceforge.net/). ANNOVAR²¹ (http://annovar.openbioinformatics.org/) software was used for variant filtering and annotation. Rare variants were subsequently filtered by a minor allele frequency (MAF) lower than 0.1% in 1000 Genomes Project²² (http://www.internationalgenome.org/), Exome Aggregation Consortium²³ (ExAC, http://exac.broadinstitute.org/) and gnomAD²⁴ (https://gnomad.broadinstitute.org/) databases. The missense variants in highly conserved amino acid residues predicted by PolyPhen-2²⁵ (http://genetics.bwh.harvard.edu/ pph2/), MutationTaster²⁶ (http://www.mutationtaster.org/), (https://sift.bii.a-star.edu.sg/) and M-CAP²⁸ (http:// SIFT²⁷ bejerano.stanford.edu/mcap/) tools, and those causing protein truncation or frameshift were considered as pathogenic. Furthermore, the potential pathogenic variants should also be shared by the two patients of the same family. Variants were interpreted according to the guidelines published by ACMG using the HGVS (https://varnomen.hgvs.org/) nomenclature.²⁹ Sanger sequencing was used to confirm the identified mutations.

Muscle histopathology study

Muscle biopsy was carried out for F1 II-1. Muscle samples used for histopathological study were frozen after collection in liquid N2-cooled isopentane and stored at -80° C until use. The samples were assessed for H&E, modified Gomori trichrome, succinate dehydrogenase (SDH), NADH-tetrazolium reductase (NADH-TR), cytochrome oxidase (COX), glycogen and lipid staining.³⁰

Mitochondrial respiratory chain (MRC) enzyme assay

Skin biopsy was performed for II-1 and II-2 of Family 1. The skeletal muscle used for the MRC enzyme assay was stored at -80° C before use. Activities of MRC complexes I, II, II+III, III, IV and the mitochondrial marker enzyme citrate synthase (CS) were assayed in the crude post-600 g supernatant of the muscle sample and in isolated mitochondria from skin fibroblasts using spectrophotometric assays as described previously.^{12 31 32} The enzyme activities of each complex are presented as the percentage of the normal control mean relative to appropriate reference enzyme activities, such as CS or MRC complex II. Enzyme activity was defined as being decreased at a level of <40% in a cell line or <30% in tissue.³³

Oxygen consumption rate (OCR) measurement

OCR was measured in fibroblasts from the two patients (F1 II-1 and II-2) with a XF96 Extracellular Flux Analyzer (Seahorse Bioscience, Billerica, Massachusetts, USA). Samples were prepared as previously described.^{12 34} In each run, we measured two controls along with the patient samples. The control cell lines used were derived from fHDF (Toyobo Biotech Support Department, Japan). Each control and patient fibroblast cell line was seeded in at least 14 wells of two XF96 cell culture microplates (Seahorse Bioscience) at a density of 20000 cells/well in 80 µL of growth medium and incubated at 5% CO₂ and 37°C overnight. The following day, the growth medium was replaced with 160 µL of 25 mM glucose medium or 10 mM galactose medium, respectively, and the cell culture microplate was placed into a CO₂-free incubator at 37°C for 60 min before measurement. After measurement of the basal OCR, 10 µM oligomycin, 4 µM carbonyl cyanide 4-(trifluoromethoxy) phenylhydrazone (FCCP) and 20 µM rotenone were added sequentially, and the OCR was recorded after each addition.³⁴ The maximum respiration rate (MRR) corresponds to the OCR after FCCP injection minus the rotenone-insensitive OCR. MRR was expressed as a percentage relative to the average of controls, in which a reduction to <71.6% was considered to represent a significant decline.¹²

Complementation of NDUFV2 mutation by cDNA expression

The fibroblasts of F1 II-1 and II-2 were obtained by skin biopsy. The complementation assay was based on the widely used ViraPower HiPerform Lentiviral Expression System. The wild-type (wt) cDNAs from *NDUFV2* were cloned into the pLenti6.3

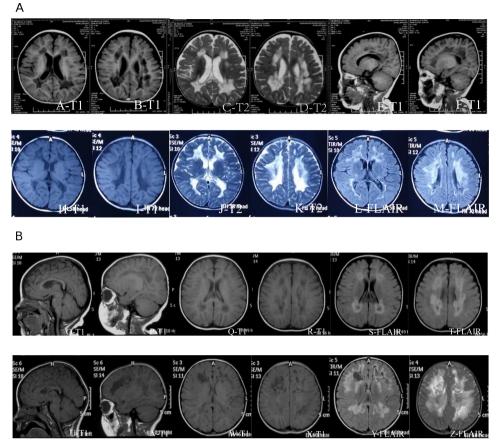


Figure 2 Brain MRI of F1 II-1 (A) at the age of 16 months (A–F) and 6 years and 3 months (H–M), and F1 II-2 (B) at the age of 12 months (O–T) and 4 years and 8 months (U–Z). A, B, H, I, Q, R, W and X—axial T1-weighted images; C, D, J and K—axial T2-weighted images; E, F, O, P, U and V—sagittal T1-weighted images and L, M, S, T, Y and Z—axial T2-FLAIR images. MRI of F1 II-1 showed long T1 and T2 signals with some cystic changes in the corpus callosum and cerebral hemispheric white matter, which were relatively stable up to 6 years and 3 months of age. Earlier MRI of F1 II-2 showed long T1 and T2 signals with some cystic changes in cerebral hemispheric white matter, mainly involving periventricular white matter and centrum semiovale, with sparing of the corpus callosum, while the later MRI showed worsening and widening of cystic lesions involving the white matter and the corpus callosum.

vector, and virions were obtained as previously described.³⁴ Patient and control fibroblasts from healthy individuals were infected with viral supernatant and selected on exposure to 5 μ g/mL blasticidin. The OCR of infected fibroblasts was measured, and the rescue effect was analysed.

RESULTS

Patients' history and clinical presentations Family 1

F1 II-1, the first child of healthy, non-consanguineous parents from different countries, had normal prenatal and perinatal histories, with a birth weight of 3.15 kg. Her parents cannot accurately describe her developmental milestones, but she could sit, stand alone and speak 'baba and mama' when she was 1 year old. At the age of 14 months, she suddenly showed signs of developmental regression after a fever. Gradually, she was not able to stand or sit alone, nor she could speak; she also showed difficulties opening her mouth and eating. MRI showed a characteristic pattern of cystic changes in the corpus callosum and deep periventricular white matter (figure 2A-F). After IVIG, ribavirin and oxiracetam treatment, her clinical symptoms gradually improved. At the age of 2 years, she could again walk alone. MRI seemed relatively stable up to 6 years and 3 months old (figure 2H-M). When examined at 7 years and 2 months old, her neurological condition was stable, with slight infantile behaviour. Although her speech and gait were normal,

Liu Z, et al. J Med Genet 2021;**0**:1–7. doi:10.1136/jmedgenet-2020-107383

her school examination scores were poor. Physical examination revealed that her height and weight were both below the third centile (17 kg, 112 cm), intermittent external strabismus could be observed in her left eye and bilateral knee tendon reflexes were active. After genetic diagnosis, she was treated with thiamine (150 mg/day), riboflavin (150 mg/day), coenzyme Q10 (150 mg/day), L-carnitine (1000 mg/day) and vitamins C (100 mg/day) and E (100 mg/day) for only 2–3 months. Up to the age of 7 years and 8 months, she walked, ran and jumped normally and had relatively normal cognitive abilities.

F1 II-2, the second child of Family 1, was born after an uneventful perinatal period. He started to present mild developmental delay at the age of 12 months when he was able to sit alone but could not climb or stand. After that, he suddenly displayed developmental regression with no obvious predisposing causes. MRI showed long T1 and T2 signals with some cystic changes in cerebral hemispheric white matter, mainly involving periventricular white matter and centrum semiovale, with sparing of the corpus callosum (figure 2O-T). The symptoms then abated, and he remained stable for some time. At the age of 20 months, he could walk alone. However, when he was 4 years and 8 months old, he had episodes of neurological deterioration, which were characterised by irritability, salivation, dysarthria, dysphagia, dystonia, developmental regression, strabismus and pyramidal tract impairment. During the most serious episode, he could only speak word by word with an unclear voice, and he could

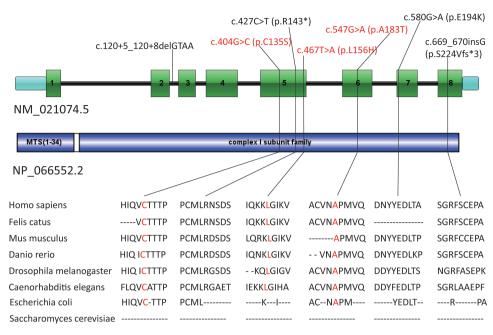


Figure 3 NDUFV2 gene structure shows the localisation of the reported and investigated (red) gene variations. Conservation of the affected amino acid residues is presented in the alignment of homologs across different species.

neither hold his head up nor sit. MRI showed diffuse cystic changes involving the cerebral hemispheric white matter and corpus callosum (figure 2U-Z), which was worse than before. Cerebral spinal fluid (CSF) lactate was 2.22 mmol/L (normal <2.8 mmol/L), and CSF cell count, protein, glucose, chloride and cultures were normal. Urine organic acid and plasma amino acid results were normal. After oxiracetam, thiamine and mecobalamin treatment, his movement and language gradually improved. When examined at 5 years and 8 months old, his condition was stable, with a slightly unstable gait and low intelligence but normal speech. Physical examination revealed external strabismus in his left eye as well as a spastic gait, accompanied by hypertonia, bilateral active knee tendon reflexes and positive Babinski signs. His weight was at the 10th centile (17kg), and his height was between the 10th and 25th centiles (112 cm). After the genetic diagnosis was established, he received thiamine (150 mg/day), riboflavin (150 mg/day), coenzyme Q10 (150 mg/ day), L-carnitine (1000 mg/day) and vitamins C (100 mg/day)

| Table 1 Mitochondrial respiratory chain assay from fibroblasts and muscle | | | | | | | | | |
|---|-------------------------------|---------|------------------------------|---------|---------------------------------|---------|--|--|--|
| MRC from fibroblasts | Per cent of control (%) | | Absolute values (mU/U CS) | | Reference range | | | | |
| | F1 II-1 | F1 II-2 | F1 II-1 | F1 II-2 | (Mean±SD) | Range | | | |
| 1 | 34 | 20 | 147 | 88 | (433±85) | 237–618 | | | |
| II | 89 | 82 | 487 | 447 | (545±97) | 364–826 | | | |
| + | 62 | 72 | 327 | 381 | (526±133) | 298–795 | | | |
| III | 123 | 112 | 116 | 105 | (94±27) | 32–143 | | | |
| IV | 77 | 98 | 15 | 19 | (19±5) | 13–35 | | | |
| MRC from muscle of F1 II-1 | Per cent of control (%) | | Absolute values (mU/U CS) | | Reference range (Mean±SD) | Range | | | |
| 1 | 24 | | 82 | | (346±38) | 279–405 | | | |
| II | 91 | | 285 | | (313±55) | 209–425 | | | |
| + | 103 | | 308 | | (299±98) | 131–488 | | | |
| 111 | 108 | | 186 | | (172±71) | 53–296 | | | |
| IV | 171 | | 52 | | (31±13) | 20–62 | | | |
| MRC, mitochondrial respiratory chain. | | | | | | | | | |

and E (100 mg/day) for 3 months. Up to the age of 6 years and 2 months, he walked, ran and jumped with a slightly abnormal gait but had relatively preserved cognitive abilities.

Family 2

F2 II-1, the first child born of healthy, non-consanguineous parents, was born after an uneventful perinatal period. Before 4 months of age, his developmental milestones were normal, but after that, his development was arrested. When he was admitted to the hospital at the age of 11 months, he had developmental delay but relatively preserved cognitive function, which showed that he could turn over and understand simple instructions but could not sit nor stand alone. Physical examination indicated hypertonia of both lower extremities, bilateral active knee tendon reflexes and positive Babinski signs. Serum lactate, urine organic acid and plasma amino acid levels were normal. MRI showed a characteristic pattern of cystic changes in the deep periventricular white matter. He was clinically diagnosed with leukoencephalopathy and treated with coenzyme Q10 (30 mg/ day), L-carnitine (500 mg/day), thiamine (20 mg/day), riboflavin (10 mg/day), vitamin B6 (20 mg/day) and mecobalamin (0.5 mg/ day) for only 1 year. During the clinical course, he presented mild development regression after an infection but gradually returned to baseline with the recovery from the infection. Followed up to 8 years and 6 months old, he could turn over and sit alone but could not stand or walk, yet his cognitive function was relatively preserved.

F2 II-2, the second child of Family 2, had normal prenatal and perinatal histories and normal developmental milestones before the age of 1. He had gradual developmental regression after vaccine inoculation at the age of 1 year. When he was admitted to the hospital at the age of 13 months, he could not turn over nor stand, but his cognitive function was relatively preserved. Physical examination showed hypertonia of both lower extremities, bilateral active knee tendon reflexes and positive Babinski signs. Serum lactate, urine organic acid and plasma amino acid levels were normal. MRI showed a characteristic pattern of cystic changes in the deep periventricular white matter. He

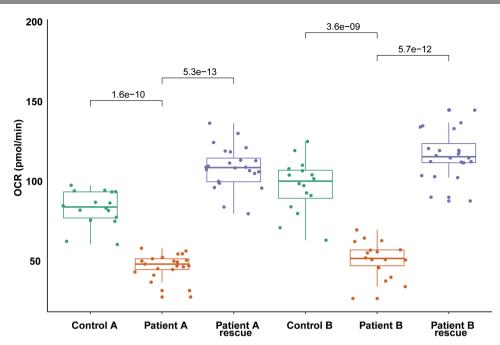


Figure 4 Complementation of *NDUFV2* variants by cDNA expression. Individual OCR expressed as pmol O_2 /min/cell in cell lines of the two patients of family 1 with *NDUFV2* mutations compared with controls and complemented patient cell lines. Significant rescue effect (p < 0.01) can be found. OCR, oxygen consumption rate.

was clinically diagnosed with leukoencephalopathy and treated like his brother with coenzyme Q10 (30 mg/day), L-carnitine (500 mg/day), thiamine (20 mg/day), riboflavin (10 mg/day), vitamin B6 (20 mg/day) and mecobalamin (0.5 mg/day). Up to the age of 4 years and 11 months, he walked, ran and jumped with a slightly unstable gait but had relatively preserved cognitive abilities.

Next-generation sequencing (WGS and WES)

We selected the potential causative variants by excluding those silent (noncoding or synonymous), common (MAF >0.1%) or not following autosomal-recessive inheritance. Those variants have been provided in the online supplemental tables 1 and 2. Furthermore, the missense variants in highly conserved amino acid residues predicted by PolyPhen-2, MutationTaster, SIFT and M-CAP tools, and those causing protein truncation or frameshift were considered as pathogenic. Ultimately, only NDUFV2 has been identified as the pathogenic gene in the two families.

The WGS performed in the affected individuals and their parents of Family 1 revealed a homozygous missense variant c.547G>A (p.A183T) in *NDUFV2* (NM_021074.5) in the affected individuals and the heterozygous missense variant in

| Table 2Clinical manifestation summary of the four patients fromtwo Chinese families | | | | | | | | |
|---|---------|---------|---------|---------|--|--|--|--|
| Clinical manifestations | F1 II-1 | F1 II-2 | F2 II-1 | F2 II-2 | | | | |
| Developmental regression (HP:0002376) | + | + | + | + | | | | |
| Strabismus (HP:0000486) | + | + | - | - | | | | |
| Lower limb hypertonia (HP:0006895) | - | + | + | + | | | | |
| Lower limb hyperreflexia (HP:0002395) | + | + | + | + | | | | |
| Short stature (HP:0004322) | + | - | NA | NA | | | | |
| Failure to thrive (HP:0001508) | + | + | NA | NA | | | | |
| Deep cerebral white matter hyperdensities (HP:0030892) | + | + | + | + | | | | |

+, present; -, absent; NA, not available

NDUFV2 in their parents encoding a mitochondrial Co I subunit (online supplemental table 1). This variant is a novel mutation that has not been reported before. The WES performed in the affected individuals of Family 2 revealed compound heterozygous missense variants c.467T>A (p.L156H) and c.404G>C (p.C135S) in the same gene, NDUFV2 (online supplemental table 2). The three variants are all absent in the 1000 Genomes Project, Exome Aggregation Consortium (ExAC) and gnomAD databases and are all predicted as deleterious by a group of tools, including SIFT, Polyphen-2, MutationTaster and M-CAP. Sequence alignments of the NDUFV2 proteins from bacteria to humans showed that all the affected amino acids are highly conserved (figure 3).

Muscle histopathology study and MRC enzyme assay

There were no abnormalities detected using H&E, Gomori trichrome, SDH, NADH-TR, COX, NSE, glycogen and lipid staining, and no ragged red, SDH-rich, ragged blue or COX-negative fibres were detected. Low activity of Co I was found in both affected individuals' fibroblasts and in the muscle of II-1 of Family 1. Co I activity in F1 II-2 was lower than that in F1 II-1 (table 1).

OCR and complementation studies in fibroblasts

In addition to measuring the OCR in cells grown in glucose medium, we measured the OCR in cells grown in galactose medium, a condition that forces cells to depend on mitochondrial respiration rather than glycolysis for ATP production. A significant decline of MRR with glucose and galactose medium was found in both siblings of Family 1 compared with the control. The OCR of F1 II-1 was 64% and 53% relative to the average of controls with glucose and galactose medium, respectively, while the OCR of F1 II-2 was 56% and 43%, respectively. Next, we took advantage of the cellular phenotype in a complementation experiment. The OCR of both Family 1 patients increased to normal values after transduction with a recombinant lentiviral

construct expressing the wt cDNA of *NDUFV2* (figure 4), which validates the pathogenic role of the novel *NDUFV2* variant in defective mitochondrial respiration.

DISCUSSION

We report sibling pairs from two Chinese families presenting a similar clinical course of recurring episodes of acute or subacute developmental regression that appeared in the first year of life (table 2). This decline was followed by gradual remissions and prolonged periods of stability. These patients had similar MRI findings that showed leukoencephalopathy with multiple cavities. Gene sequencing revealed that all four patients carried pathogenic novel variants in the *NDUFV2* gene.

Since the first disease gene, NDUFS1, was reported to be related to PCL in 2011,⁵ NDUFV1 and other genes related to mitochondrial function and energy metabolism have been reported, while mutations in NDUFS1 and NDUFV1 seem to be more common.⁶⁻⁸ NDUFV1, NDUFV2 and NDUFS1-encoding subunits compose the N module of Co I, which oxidises NADH.^{6 11} With a similar function, NDUFV2 was supposed to be another causal gene of PCL. NDUFV2 is highly expressed in the brain, mainly in neurons, astrocytes and oligodendrocytes, so a NDUFV2 mutation can induce dysfunction of neurons, astrocytes and oligodendrocytes, causing the corresponding clinical syndromes. To date, six mutations have been described in NDUFV2, most of which result in Leigh syndrome, but none has been reported to be associated with PCL. In this study, we identified three novel NDUFV2 mutations in patients with PCL. There are several lines of evidence suggesting the pathogenicity of the following NM 021074.5 mutations: c.547G>A (p.A183T), c.467T>A (p.L156H) and c.404G>C (p.C135S) in NDUFV2.²⁹ First, homozygous and compound heterozygous mutations occurred in sibling pairs of two families with similar clinical and neuroimaging phenotypes, segregating from the two heterozygous healthy parents. Second, the three mutations were all absent from controls in the 1000 Genomes Project, ExAC and gnomAD databases, were all in highly conserved amino acid residues and were all predicted as deleterious by a group of tools, including SIFT, Polyphen-2, MutationTaster and M-CAP. Third, the MRC enzyme assay showed Co I deficiency in the fibroblasts of both affected individuals and in the muscle of the elder sister of Family 1. Finally, the two patients of Family 1 showed a significant decrease in MRR compared with the control, and a significant rescue effect could be shown by functional complementation of patient-derived fibroblast cell lines, which is an established method to validate the disease-causing variant in various genes.34

In the present patients, CNS involvement is the main clinical presentation. Common clinical features include developmental regression, followed by gradual remissions and prolonged periods of stability. Brain MRI revealed abnormalities in the white matter of all four patients. Our results suggest that leuko-encephalopathy in these patients is caused by *NDUFV2* mutations. The clinical features described in these patients differ from the previously described *NDUFV2*-associated neurological diseases.

Author affiliations

¹Department of Neurology, Beijing Children's Hospital, Capital Medical University, National Center for Children's Health, Beijing, China

²Center for Bioinformatics and Computational Biology, Institute of Biomedical Sciences, School of Life Sciences, East China Normal University, Shanghai, China ³Key Laboratory of Advanced Theory and Application in Statistics and Data Science -MOE, School of Statistics, East China Normal University, Shanghai, China ⁴Department of Pediatrics, Peking University First Hospital, Beijing, China ⁵Department of Neurology, Children's Hospital of Hebei Province, Hebei Medical University, Shijiazhuang, China

⁶Department of Neurology, Technical University Munich, Munich, Germany ⁷Center for Medical Genetics, Department of Metabolism, Chiba Children's Hospital, Chiba, Japan

⁸Institute of Human Genetics, Technical University Munich, Munich, Germany ⁹Institute of Neurogenomics, Helmholtz Zentrum Munich, Neuherberg, Germany

Acknowledgements The authors would like to thank the families for their participation in this study. We thank AJE Team (www.aje.com) for editing a draft of this manuscript.

Contributors ZL collected and analysed the data and was responsible for drafting the manuscript. LZ analysed the genetic data and was responsible for drafting the manuscript. CR, MX, SL, RB, YW, LC and SS contributed to phenotyping and to acquiring and analysing the clinical and histopathological data. ME and HP conducted functional experiments, collected and analysed the functional data and contributed to the manuscript. ZL, MS, MO-T and KM conducted the enzyme assay and OCR experiments and collected and analysed these data. TS, HP and FF were responsible for conceiving, designing and supervising the study and for writing the manuscript. All authors revised the manuscript for intellectual content.

Funding This study was funded by the Capital Health Development Research Foundation Project (2018-2-2096), the National Natural Science Foundation of China (81541115) and the Henan Provincial Key Laboratory of Children's Genetics and Metabolic Diseases Foundation (SS201904).

Competing interests None declared.

Patient consent for publication Parental/guardian consent obtained.

Ethics approval The Ethics Committee of each hospital approved the study, and prior informed consent was obtained from the patients' parents.

Provenance and peer review Not commissioned; externally peer reviewed.

Data availability statement Data are available on reasonable request. The data that support the findings of this study are available from the corresponding author on reasonable request.

Supplemental material This content has been supplied by the author(s). It has not been vetted by BMJ Publishing Group Limited (BMJ) and may not have been peer-reviewed. Any opinions or recommendations discussed are solely those of the author(s) and are not endorsed by BMJ. BMJ disclaims all liability and responsibility arising from any reliance placed on the content. Where the content includes any translated material, BMJ does not warrant the accuracy and reliability of the translations (including but not limited to local regulations, clinical guidelines, terminology, drug names and drug dosages), and is not responsible for any error and/or omissions arising from translation and adaptation or otherwise.

Open access This is an open access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited, appropriate credit is given, any changes made indicated, and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/.

ORCID iDs

Zhimei Liu http://orcid.org/0000-0003-0393-7598 Manting Xu http://orcid.org/0000-0002-3155-4810 Minako Ogawa-Tominaga http://orcid.org/0000-0003-1449-5916

REFERENCES

- 1 Parikh S, Bernard G, Leventer RJ, van der Knaap MS, van Hove J, Pizzino A, McNeill NH, Helman G, Simons C, Schmidt JL, Rizzo WB, Patterson MC, Taft RJ, Vanderver A, GLIA Consortium. A clinical approach to the diagnosis of patients with leukodystrophies and genetic leukoencephelopathies. *Mol Genet Metab* 2015;114:501–15.
- 2 Vanderver A, Prust M, Tonduti D, Mochel F, Hussey HM, Helman G, Garbern J, Eichler F, Labauge P, Aubourg P, Rodriguez D, Patterson MC, Van Hove JLK, Schmidt J, Wolf NI, Boespflug-Tanguy O, Schiffmann R, van der Knaap MS, GLIA Consortium. Case definition and classification of leukodystrophies and leukoencephalopathies. *Mol Genet Metab* 2015;114:494–500.
- 3 Schuelke M, Smeitink J, Mariman E, Loeffen J, Plecko B, Trijbels F, Stöckler-Ipsiroglu S, van den Heuvel L. Mutant NDUFV1 subunit of mitochondrial complex I causes leukodystrophy and myoclonic epilepsy. *Nat Genet* 1999;21:260–1.
- 4 Naidu S, Bibat G, Lin D, Burger P, Barker P, Rosemberg S, Braverman N, Arroyo H, Dowling M, Hamosh A, Kimonis V, Blank C, Fiumara A, Facchini S, Singhal B, Moser H, Kelley R, DiMauro S. Progressive cavitating leukoencephalopathy: a novel childhood disease. *Ann Neurol* 2005;58:929–38.
- 5 Ferreira M, Torraco A, Rizza T, Fattori F, Meschini MC, Castana C, Go NE, Nargang FE, Duarte M, Piemonte F, Dionisi-Vici C, Videira A, Vilarinho L, Santorelli FM, Carrozzo

R, Bertini E. Progressive cavitating leukoencephalopathy associated with respiratory chain complex I deficiency and a novel mutation in NDUFS1. *Neurogenetics* 2011;12:9–17.

- 6 Björkman K, Sofou K, Darin N, Holme E, Kollberg G, Asin-Cayuela J, Holmberg Dahle KM, Oldfors A, Moslemi A-R, Tulinius M. Broad phenotypic variability in patients with complex I deficiency due to mutations in NDUFS1 and NDUFV1. *Mitochondrion* 2015;21:33–40.
- 7 Ortega-Recalde O, Fonseca DJ, Patiño LC, Atuesta JJ, Rivera-Nieto C, Restrepo CM, Mateus HE, van der Knaap MS, Laissue P. A novel familial case of diffuse leukodystrophy related to NDUFV1 compound heterozygous mutations. *Mitochondrion* 2013;13:749–54.
- 8 Dallabona C, Abbink TEM, Carrozzo R, Torraco A, Legati A, van Berkel CGM, Niceta M, Langella T, Verrigni D, Rizza T, Diodato D, Piemonte F, Lamantea E, Fang M, Zhang J, Martinelli D, Bevivino E, Dionisi-Vici C, Vanderver A, Philip SG, Kurian MA, Verma IC, Bijarnia-Mahay S, Jacinto S, Furtado F, Accorsi P, Ardissone A, Moroni I, Ferrero I, Tartaglia M, Goffrini P, Ghezzi D, van der Knaap MS, Bertini E. LYRM7 mutations cause a multifocal cavitating leukoencephalopathy with distinct MRI appearance. *Brain* 2016;139:782–94.
- 9 Melchionda L, Haack TB, Hardy S, Abbink TEM, Fernandez-Vizarra E, Lamantea E, Marchet S, Morandi L, Moggio M, Carrozzo R, Torraco A, Diodato D, Strom TM, Meitinger T, Tekturk P, Yapici Z, Al-Murshedi F, Stevens R, Rodenburg RJ, Lamperti C, Ardissone A, Moroni I, Uziel G, Prokisch H, Taylor RW, Bertini E, van der Knaap MS, Ghezzi D, Zeviani M. Mutations in APOPT1, encoding a mitochondrial protein, cause cavitating leukoencephalopathy with cytochrome c oxidase deficiency. *Am J Hum Genet* 2014;95:315–25.
- 10 Ishiyama A, Sakai C, Matsushima Y, Noguchi S, Mitsuhashi S, Endo Y, Hayashi YK, Saito Y, Nakagawa E, Komaki H, Sugai K, Sasaki M, Sato N, Nonaka I, Goto Y-I, Nishino I. *IBA57* mutations abrogate iron-sulfur cluster assembly leading to cavitating leukoencephalopathy. *Neurol Genet* 2017;3:e184.
- 11 Brandt U. Energy converting NADH:quinone oxidoreductase (complex I). *Annu Rev Biochem* 2006;75:69–92.
- 12 Ogawa E, Shimura M, Fushimi T, Tajika M, Ichimoto K, Matsunaga A, Tsuruoka T, Ishige M, Fuchigami T, Yamazaki T, Mori M, Kohda M, Kishita Y, Okazaki Y, Takahashi S, Ohtake A, Murayama K. Clinical validity of biochemical and molecular analysis in diagnosing Leigh syndrome: a study of 106 Japanese patients. *J Inherit Metab Dis* 2017;40:685–93.
- 13 Koene S, Rodenburg RJ, van der Knaap MS, Willemsen MAAP, Sperl W, Laugel V, Ostergaard E, Tarnopolsky M, Martin MA, Nesbitt V, Fletcher J, Edvardson S, Procaccio V, Slama A, van den Heuvel LPWJ, Smeitink JAM. Natural disease course and genotype-phenotype correlations in complex I deficiency caused by nuclear gene defects: what we learned from 130 cases. *J Inherit Metab Dis* 2012;35:737–47.
- 14 Bénit P, Beugnot R, Chretien D, Giurgea I, De Lonlay-Debeney P, Issartel J-P, Corral-Debrinski M, Kerscher S, Rustin P, Rötig A, Munnich A. Mutant NDUFV2 subunit of mitochondrial complex I causes early onset hypertrophic cardiomyopathy and encephalopathy. *Hum Mutat* 2003;21:582–6.
- 15 Liu H-Y, Liao P-C, Chuang K-T, Kao M-C. Mitochondrial targeting of human NADH dehydrogenase (ubiquinone) flavoprotein 2 (NDUFV2) and its association with earlyonset hypertrophic cardiomyopathy and encephalopathy. J Biomed Sci 2011;18:29.
- 16 Pagniez-Mammeri H, Lombes A, Brivet M, Ogier-de Baulny H, Landrieu P, Legrand A, Slama A. Rapid screening for nuclear genes mutations in isolated respiratory chain complex I defects. *Mol Genet Metab* 2009;96:196–200.
- 17 Cameron JM, MacKay N, Feigenbaum A, Tarnopolsky M, Blaser S, Robinson BH, Schulze A. Exome sequencing identifies complex I NDUFV2 mutations as a novel cause of Leigh syndrome. *Eur J Paediatr Neurol* 2015;19:525–32.
- 18 Hattori N, Yoshino H, Tanaka M, Suzuki H, Mizuno Y. Genotype in the 24-kDa subunit gene (NDUFV2) of mitochondrial complex I and susceptibility to Parkinson disease. *Genomics* 1998;49:52–8.
- 19 Nishioka K, Vilariño-Güell C, Cobb SA, Kachergus JM, Ross OA, Hentati E, Hentati F, Farrer MJ. Genetic variation of the mitochondrial complex I subunit NDUFV2 and Parkinson's disease. *Parkinsonism Relat Disord* 2010;16:686–7.

- 20 Li H. Exploring single-sample SNP and indel calling with whole-genome de novo assembly. *Bioinformatics* 2012;28:1838–44.
- 21 Wang K, Li M, Hakonarson H. ANNOVAR: functional annotation of genetic variants from high-throughput sequencing data. *Nucleic Acids Res* 2010;38:e164.
- 22 1000 Genomes Project Consortium, Abecasis GR, Auton A, Brooks LD, DePristo MA, Durbin RM, Handsaker RE, Kang HM, Marth GT, McVean GA. An integrated map of genetic variation from 1,092 human genomes. *Nature* 2012;491:56–65.
- 23 Lek M, Karczewski KJ, Minikel EV, Samocha KE, Banks E, Fennell T, O'Donnell-Luria AH, Ware JS, Hill AJ, Cummings BB, Tukiainen T, Birnbaum DP, Kosmicki JA, Duncan LE, Estrada K, Zhao F, Zou J, Pierce-Hoffman E, Berghout J, Cooper DN, Deflaux N, DePristo M, Do R, Flannick J, Fromer M, Gauthier L, Goldstein J, Gupta N, Howrigan D, Kiezun A, Kurki MI, Moonshine AL, Natarajan P, Orozco L, Peloso GM, Poplin R, Rivas MA, Ruano-Rubio V, Rose SA, Ruderfer DM, Shakir K, Stenson PD, Stevens C, Thomas BP, Tiao G, Tusie-Luna MT, Weisburd B, Won H-H, Yu D, Altshuler DM, Ardissino D, Boehnke M, Danesh J, Donnelly S, Elosua R, Florez JC, Gabriel SB, Getz G, Glatt SJ, Hultman CM, Kathiresan S, Laakso M, McCarroll S, McCarthy MI, McGovern D, McPherson R, Neale BM, Palotie A, Purcell SM, Saleheen D, Scharf JM, Sklar P, Sullivan PF, Tuomilehto J, Tsuang MT, Watkins HC, Wilson JG, Daly MJ, MacArthur DG, Exome Aggregation Consortium. Analysis of protein-coding genetic variation in 60,706 humans. *Nature* 2016;536:285–91.
- 24 Karczewski KJ, Francioli LC, Tiao G, Cummings BB, Alföldi J, Wang Q, Collins RL, Laricchia KM, Ganna A, Birnbaum DP, Gauthier LD, Brand H, Solomonson M, Watts NA, Rhodes D, Singer-Berk M, England EM, Seaby EG, Kosmicki JA, Walters RK, Tashman K, Farjoun Y, Banks E, Poterba T, Wang A, Seed C, Whiffin N, Chong JX, Samocha KE, Pierce-Hoffman E, Zappala Z, O'Donnell-Luria AH, Minikel EV, Weisburd B, Lek M, Ware JS, Vittal C, Armean IM, Bergelson L, Cibulskis K, Connolly KM, Covarrubias M, Donnelly S, Ferriera S, Gabriel S, Gentry J, Gupta N, Jeandet T, Kaplan D, Llanwarne C, Munshi R, Novod S, Petrillo N, Roazen D, Ruano-Rubio V, Saltzman A, Schleicher M, Soto J, Tibbetts K, Tolonen C, Wade G, Talkowski ME, Neale BM, Daly MJ, MacArthur DG, Genome Aggregation Database Consortium. The mutational constraint spectrum quantified from variation in 141,456 humans. *Nature* 2020;581:434–43.
- 25 Adzhubei I, Jordan DM, Sunyaev SR. Predicting functional effect of human missense mutations using PolyPhen-2. *Curr Protoc Hum Genet* 2013; Chapter 7:t7–20.
- 26 Schwarz JM, Rödelsperger C, Schuelke M, Seelow D. MutationTaster evaluates disease-causing potential of sequence alterations. *Nat Methods* 2010;7:575–6.
- 27 Ng PC, Henikoff S. SIFT: predicting amino acid changes that affect protein function. *Nucleic Acids Res* 2003;31:3812–4.
- 28 Jagadeesh KA, Wenger AM, Berger MJ, Guturu H, Stenson PD, Cooper DN, Bernstein JA, Bejerano G. M-CAP eliminates a majority of variants of uncertain significance in clinical exomes at high sensitivity. *Nat Genet* 2016;48:1581–6.
- 29 Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, Grody WW, Hegde M, Lyon E, Spector E, Voelkerding K, Rehm HL, ACMG Laboratory Quality Assurance Committee. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of medical genetics and genomics and the association for molecular pathology. *Genet Med* 2015;17:405–23.
- 30 Parikh S, Goldstein A, Koenig MK, Scaglia F, Enns GM, Saneto R, Anselm J, Cohen BH, Falk MJ, Greene C, Gropman AL, Haas R, Hirano M, Morgan P, Sims K, Tarnopolsky M, Van Hove JLK, Wolfe L, DiMauro S. Diagnosis and management of mitochondrial disease: a consensus statement from the mitochondrial medicine Society. *Genet Med* 2015;17:689–701.
- 31 Kirby DM, Crawford M, Cleary MA, Dahl HH, Dennett X, Thorburn DR. Respiratory chain complex I deficiency: an underdiagnosed energy generation disorder. *Neurology* 1999;52:1255–64.
- 32 Frazier AE, Thorburn DR. Biochemical analyses of the electron transport chain complexes by spectrophotometry. *Methods Mol Biol* 2012;837:49–62.
- 33 Bernier FP, Boneh A, Dennett X, Chow CW, Cleary MA, Thorburn DR. Diagnostic criteria for respiratory chain disorders in adults and children. *Neurology* 2002;59:1406–11.
- 34 Kremer LS, Prokisch H. Identification of disease-causing mutations by functional complementation of patient-derived fibroblast cell lines. *Methods Mol Biol* 2017;1567:391–406.

J Med Genet: first published as 10.1136/jmedgenet-2020-107383 on 2 April 2021. Downloaded from http://jmg.bmj.com/ on May 26, 2021 at Helmholtz Zentrum Muenchen Zentralbibliothek Protected by copyright.