# Feasibility of reducing differences in estimated doses in nuclear medicine between a patientspecific and a reference phantom

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### Abstract

The feasibility of reducing the differences between patient-specific internal doses and doses estimated using reference phantoms was evaluated. Relatively simple adjustments to a polygon-surface ICRP adult male reference phantom were applied to fit selected individual dimensions using the software Rhinoceros<sup>®</sup>4.0. We tested this approach on two patient-specific phantoms: the biggest and the smallest phantoms from the Helmholtz Zentrum München library. These phantoms have unrelated anatomy and large differences in body-mass-index. Three models approximating each patient's anatomy were considered: the voxel and the polygon-surface ICRP adult male reference phantoms and the adjusted polygon-surface reference phantom. The Specific Absorbed Fractions (SAFs) for internal photon and electron sources were calculated with the Monte Carlo code EGSnrc. Employing the timeintegrated activity coefficients of a radiopharmaceutical (S)-4-(3-18F-fluoropropyl)-L-glutamic acid and the calculated SAFs, organ absorbed-dose coefficients were computed following the formalism promulgated by the Committee on Medical Internal Radiation Dose. We compared the absorbed-dose coefficients between each patient-specific phantom and other models considered with emphasis on the cross-fire component. The corresponding differences for most organs were notably lower for the adjusted reference models compared to the case when reference models were employed. Overall, the proposed approach provided reliable dose estimates for both tested patient-specific models despite the pronounced differences in their anatomy. To capture the full range of inter-individual anatomic variability more patient-specific phantoms are required. The results of this test study suggest a feasibility of estimating patient-specific doses within a relative uncertainty of 25% or less using adjusted reference models, when only simple phantom scaling is applied.

Key words: nuclear medicine, human computational phantom, personalised internal dose, cross-fire

### Introduction

Radiation-based modalities are used extensively in diagnostic and therapeutic medical applications. Their usage requires the estimation of radiation doses to the patient. International Commission on Radiological Protection (ICRP) Publication 110 [1] describes the reference voxel computational phantoms that are widely used in dosimetric calculations. Xu [2] stated that for medical dose tracking individualised dosimetry is needed. This is in contrast to the "Reference Man" methodology promulgated by ICRP and used in prospective radiation protection. Bolch et al. [3] mentioned the limited use of reference phantoms in medical imaging and therapy applications, because anatomical variations among individuals can notably contribute to the uncertainty of the estimated organ doses. These variations are disregarded if reference phantoms are employed. Thus, the application of individualised phantoms has the potential to improve the accuracy of medical dosimetric calculations. This is especially important for medical dose estimation in therapeutic applications. The accuracy level of dosimetry depends on the application and on the available resources. The accurate segmentation of a complete patient-specific phantom in nuclear medicine is often difficult, due to the very time-consuming manual work necessary. Another shortcoming of the segmentation of individual phantoms is the need of individual tomographic images of sufficient resolution and body coverage, which are usually not available in routine nuclear medicine procedures.

Up to now, many groups created families of human computational phantoms via segmentation from available individual tomographic data or via scaling BREP (Boundary REPresentation) phantoms in terms of body height and weight [4-19]. Total body height, weight or body mass index (BMI) are often used as parameters for matching a specific individual with a library of available phantoms for the purpose of dosimetric calculations. Nonetheless, this usually does not reflect the organ topology of the investigated individual and could cause large differences between the real patient-specific doses and those obtained for the available computational phantom. Zhang et al. [20] investigated the differences in organ doses, effective doses, and risk indices in computed tomography (CT) examinations for adults, caused by the employment of four types of computational phantoms in the dosimetry. This included the extended cardiac-torso (XCAT) family [21], the reference voxel phantoms described in ICRP Publication 110 [1] and two stylized mathematical phantoms. The study of Zhang et al. [20] revealed considerable differences in organ doses from CT even in case of closely matched weight, height and organ masses of the investigated phantoms. Other authors also quantified and showed cases with notable differences in individual organ dose conversion coefficients for external irradiation [4, 6]

and Specific Absorbed Fractions (SAFs) or organ absorbed-dose coefficients for internal irradiation [22-24], caused by the variations in organ masses and locations in different computational phantoms.

This work focuses on nuclear medicine diagnostic investigations. In this case, organs and tissues of a patient receive low to moderate doses due to the accumulation of an administered radiopharmaceutical in various body organs and tissues, so-called source regions. There are two components of an organ absorbed-dose: (1) the dose from the self-absorption (for source regions only), (2) the dose from the cross-fire from other source regions [25, 26]. The self-absorption dose is the dose deposited in a tissue from radiation contained in the tissue itself. The dose from the cross-fire denotes a dose deposited in a tissue from radiation coming from the radionuclides contained in another tissue. The relative values of the two dose components depend upon the organ, the emitted radiation type and its energy. The positron radiation emitted in positron emission tomography (PET) is absorbed in short distances from the emission point. Thus, the self-absorption component is the dominant dose component for source regions. A numerical example of this is given in the Results. Organ selfabsorption is dominated by organ mass, as demonstrated in Pamphlet 11 of the Committee on Medical Internal Radiation Dose (MIRD) [27], which provides a guidance on how patient-specific scaling of reference radionuclide SAF values is to be done. This scaling requires only the mass of the target (= source) region and is, hence, otherwise independent of the organ topology of an individual. The crossfire (except the cross-fire from blood and total body), on the contrary, is influenced by the organ topology. Thus, this study focuses on the anatomy-dependent cross-fire components of organ absorbed doses. We evaluate whether the accuracy of internal dose estimations can be efficiently improved with less effort than would be required were we to segment a patient-specific model.

Whalen et al. [28] demonstrated the feasibility of reducing the uncertainties in organ volumes to 21–29 % for all organs, except spleen, by using the trunk height as a parameter to match a patient with a library of computational phantoms. Whalen et al. [28] also showed that the uncertainties in organ volumes can be further reduced, if ventral cavity volumes are matched. Little work has been done to investigate the possible reduction in the uncertainty of estimated dose that is achievable by adjusting reference phantoms to match the individual patient.

The objective of this work is to check the feasibility of reducing the uncertainties in the cross-fire component of organ internal doses by adjusting a reference phantom to selected external dimensions of individuals. Since it is beyond the scope of this study to evaluate the differences of the cross-fire SAFs for all potential source/target region combinations, as an example, we focused on the biokinetic

behaviour of a novel radiopharmaceutical (S)-4-(3-<sup>18</sup>F-fluoropropyl)-L-glutamic acid (<sup>18</sup>F-FSPG), for which comprehensive datasets were available.

### Methods and materials

#### Voxel and polygon-surface based ICRP adult male reference computational phantoms

ICRP Publication 110 [1] gives the detailed description of the voxel ICRP adult reference computational phantoms, their applications and possible limitations. These models are the official computational models of reference adults, adopted by ICRP [1]. Voxel-based computational phantoms are limited in their flexibility though [2, 3]. Such phantoms can be modified only by increasing or decreasing the dimensions of each voxel [29] or adding or removing of voxel layers in an organ. Selective adjustment or scaling of various body parts of a voxel phantom by different factors cannot be easily done. Thus, as a base model in the current work we used the polygon-surface ICRP male reference phantom [30]. It was developed by converting the voxel ICRP male reference phantom to a polygon-surface format. As a BREP model, the polygon-surface phantom can be relatively easily modified and deformed [2, 3]. Note that the polygon-surface ICRP male reference phantom used here and described by Yeom et al. [30] is not the final version, since it is currently under development [31, 32]. Analogous to the abbreviation for the voxel ICRP adult male reference computational phantom used by e.g. Schlattl et al. [33] and utilised in this work (RCP-AM), we denote the polygon-surface ICRP adult male reference computational phantom

### Target patient-specific voxel models and adjusted characteristics

Two voxel models from the Helmholtz Zentrum München (HMGU) model library were considered to be fictitious individual target patients in this work. We refer to these models as to patient-specific phantoms. The proposed approach was to apply relatively simple adjustments to P-RCP-AM and, thus, to make P-RCP-AM more specific to the two selected individual target patients. The goodness of this approach was subsequently evaluated depending on how close the organ doses for the adjusted reference phantoms are to the real patient-specific doses. To test the approach on phantoms segmented from individuals with potentially high differences in anatomy, the biggest available model Visible Human [6] and the smallest available model Irene [5, 6] (see Figure 1) were selected as fictitious patients. The BMI of Visible Human and Irene is equal to 31.9 kg/m<sup>2</sup> and 19.2 kg/m<sup>2</sup>, respectively. Despite P-RCP-AM being male, the female model Irene was chosen since no male model with similarly small BMI is available in the HMGU model library. Thus the doses for genderspecific organs could not be compared between Irene and P-RCP-AM adjusted to Irene. The comparison of the doses for other target regions appears to be appropriate though.



**Figure 1.** Voxel phantoms Visible Human (left) and Irene (right) [33], denoted here as Pat1 and Pat2, respectively.

The Visible Human model used here was segmented from the CT images obtained from the National Library of Medicine's Visible Human Project. This phantom is a partial-body phantom covering the head to mid-thigh. It is described in detail by Zankl et al. [6]. The Visible Human phantom has been constructed from an individual who had only one testicle. This circumstance resulted in a testes mass that is lower than the reference value (25.5 g compared to 35 g). However, for the radiopharmaceutical under consideration the testes are not a source region. Hence, the testes dose is only due to cross-fire irradiation. Consequently, this lower target region mass does not have an impact. Another voxel phantom of the same individual described by Xu et al. [34] exists, segmented from whole body colour photographs. Irene has been constructed from the CT images of a patient, covering the whole body [5, 6].

For the sake of simplicity and to emphasize that Visible Human and Irene represent fictitious individual patients in this work, we denote them as Pat1 and Pat2, respectively. The following dimensions of P-RCP-AM [30] were adjusted to the corresponding individual characteristics of Pat1 and Pat2: maximum distance between the iliac crest and the clavicles, total height, front-to-back distance and width of the rib cage (both are the corresponding maximum dimensions on the level of the xiphoid process). Sitting height or trunk height along with total standing height were previously considered by various authors to create a family of human computational phantoms or to match a patient with a library of phantoms [8, 10-12, 15, 17, 18, 28]. The distance between the iliac crest and the clavicles was selected as an analogue to the trunk height. Whalen et al. [28] matched ventral cavity

volumes. To scale this body part, we used the front-to-back distance and width of the rib cage, since these dimensions are relatively easy to measure. The dimensions of the rib cage correspond to those of the skeleton, not the external body dimensions. It is expected that the dimensions of the volume spanned by the rib cage correlate with the relative locations of the organs inside the rib cage better than those of the external body, especially for overweight individuals with a thick layer of subcutaneous fat. The front-to-back distance and the width of the rib cage (skeleton) can be estimated by subtraction of the approximate thickness of the subcutaneous fat from the corresponding external body dimensions. Alternatively, the rib cage dimensions can be determined easily from e.g. a CT scan which usually accompanies the actual PET diagnosis. The two new phantoms, obtained by matching P-RCP-AM to Pat1 and Pat2, are indicated as Pat1M and Pat2M, respectively. Table 1 summarises all abbreviations used for naming the phantoms in this work. The selected set of adjustments was restricted to individual characteristics that are relatively easy to measure in actual patients. The head of the phantoms was not scaled in any dimension. The expected variability in the size of a head should not be high. In this study, as in a test study, only the dimensions with high expected variability are considered. Since the female individual Pat2 is represented by the male model Pat2M with only limited set of adjustments made, waist to hip ratio is not matched, bone size and muscle distribution are different. Additionally, gender-specific organs as uterus, gonads or prostate are excluded from the analysis.

All adjustments were made in the 3D modelling software Rhinoceros<sup>®</sup> 4.0 (McNeel North America, Seattle, WA), which provides powerful tools for creating and editing non-uniform rational B-spline curves, surfaces and polygon meshes. The adjustments were applied in three steps, which are schematically shown in Figure 2. At the first step the distance between the iliac crest and the clavicles and the total height were matched. This was achieved by scaling the rib cage, the spine, the clavicles, the sternum, the scapulae and all the organs inside the rib cage on the level of thoracic and lumbar spine, including the small intestine and the colon, in superior-inferior directions by a factor needed to match the distance between the iliac crest and the clavicles. For convenience, prior to the first scaling step, the pelvis along with the legs, the bladder, the prostate, the testes and the inguinal lymphatic nodes were moved in inferior direction, as shown in Figure 2. After matching the distance between the iliac crest and the clavicles the phantom was re-unified. If needed, the first scaling step was followed by a scaling of the legs in inferior direction to match the total height. At the second step all the aforementioned organs, except the arms, were scaled together with the rib cage in anterior-posterior direction to fit the outer front-to-back distance of the rib cages of Pat1 and Pat2. At the third step the

scaling in lateral dimension was applied to match the outer width of the rib cages of the two patients considered. The arms were not scaled, but moved laterally to fit the scaled thorax. The scaling was done with the Scale 1D command. Organ intersections were then found with the MeshIntersect command (Rhinoceros<sup>®</sup> 4.0) and eliminated by applying some minor adjustments of the intersecting organs with the Smooth command and/or Mesh Edit and Mesh Repair Tools of Rhinoceros<sup>®</sup> 4.0. To scale and repair the skin mesh, the Scale 1D command as well as the Mesh Edit and Mesh Repair Tools along with the Smooth command were employed. No further adjustments of individual organ positions and sizes of Pat1M and Pat2M to match the corresponding values of Pat1 and Pat2 were applied.



**Figure 2.** Three main steps of the adjustments of P-RCP-AM to fit selected dimensions of Pat1. The adjustments applied to P-RCP-AM to fit Pat2 were similar to those of Pat1 and thus are not shown here.

Description		Used abbreviation		
Voxel ICRP adult male reference computational phantom [1]	RCP-AM			
Polygon-surface ICRP adult male reference computational phantom	P-RCP-AM			
Visible Human [6]	(fictitious patient 1)	Pat1		
Irene [5, 6]	(fictitious patient 2)	Pat2		
Phantom obtained via adjustments of polygon-surface ICRP adult m computational phantom to fit selected characteristics of Visible Hum	Pat1M			
Phantom obtained via adjustments of polygon-surface ICRP adult m computational phantom to fit selected characteristics of Irene	Pat2M			

Table 1. Abbreviations used for indicating the computational models in this work

### Monte Carlo calculations

The Monte Carlo code EGSnrc [35] with a user code developed at the HMGU was employed in this work. The SAFs for homogeneous volume sources of monoenergetic photons and electrons were calculated in the geometry of the six computational phantoms. The EGSnrc system has been previously tested against other Monte Carlo codes [1, 36-38]. The user code has been previously utilised and benchmarked in various works to compute dose conversion coefficients for public, occupational and medical exposures. The detailed description of the user code can be found elsewhere [7, 26, 33]. A total of 18 electron energies, ranging from 10 keV to 1 MeV, and one photon energy of 511 keV were considered. The simulated energies were selected to cover the beta spectra of <sup>11</sup>C- and <sup>18</sup>F-labeled compounds, which are commonly used in PET, and the energy of annihilation photons. The electron energies were in accordance to ICRP Publication 133 [26] with an additional energy of 250 keV. For photon internal sources and electron internal sources in the energy range of 10 keV -600 keV, 40 million histories were simulated per source region and energy. For electron energies of 800 keV and higher the number of simulated histories was reduced to 10 million for the sake of computational time. Photon histories were terminated at 2 keV. Electron histories with kinetic energies less than 50 keV were followed down to 2 keV. All other electron histories were terminated at 20 keV. The source regions considered included total blood, heart wall, heart contents, kidneys, liver, pancreas, salivary glands, spleen, stomach wall, total body, thyroid and urinary bladder content. The source region "total blood" comprised blood vessels, if present in the phantom, and regional blood volumes (RBVs) of organs and tissues. Target regions included all organs and tissues segmented in the phantoms.

Since the EGSnrc user code utilises voxel geometry, all polygon-surface phantoms considered had to be voxelised. Using the software Binvox (Patrick Min, <u>http://www.patrickmin.com/binvox/</u>) [39] all polygon-surface organs have been transferred into individual organ voxel models with 1 × 1 × 1 mm<sup>3</sup> voxel size. They were then combined to complete phantoms using an in-house IDL-based (IDL<sup>®</sup>, Exelis Visual Information Solutions, Inc., version 8.2.1) program. For simplicity, here and in the following we use the same abbreviations for the polygon-surface models and their voxelised versions, since only the latter were used for the Monte Carlo calculations. Media elemental compositions and densities from ICRP Publication 110 [1] were used in P-RCP-AM, Pat1M and Pat2M. For each target phantom, Pat1 and Pat2, three models approximating it were considered: RCP-AM, P-RCP-AM and Pat1M for Pat1, RCP-AM, P-RCP-AM and Pat2M for Pat2 (see Table 1). Table 2 summarises selected properties of the employed phantoms.

It was previously shown that blood activity has a substantial effect on absorbed doses for highly vascular organs, which are not source regions [40]. As mentioned above, total blood as source region consists of blood vessels and RBVs. The total blood in Pat1 and Pat2 originally did not comprise RBVs. To include RBVs into the total blood, mass fractions of blood in different organs and tissues are required. Nadler's formula [41] was applied to calculate the total blood volumes for Pat1 and Pat2. Total blood masses were computed with the blood density from ICRU Report 46 [42] and ICRP Publication 110 [1] (1.06 g/cm<sup>3</sup>). Using the resulting total blood masses, reference RBVs from ICRP Publication 89 [43] and the organ masses of Pat1 and Pat2, mass fractions of blood were calculated for organs and tissues of the aforementioned phantoms. These fractions were subsequently considered in the Monte Carlo calculations.

Tuble II beletica properties of constanted computational phantoms									
Phantom	Height, [cm]	Weight, [kg]	Number of segmented regions	Voxel dimensions, [mm <sup>3</sup> ]					
Pat1	180.0 (125.0) <sup>a</sup>	103.2 (89.1) <sup>a</sup>	131	0.9  imes 0.9  imes 5.0					
Pat2	163.0	51.1	62	$1.9 \times 1.9 \times 5.0$					
RCP-AM	176.0	73.0	138	$2.1\times2.1\times8.0$					
P-RCP-AM	176.4	73.6	111	$1.0\times1.0\times1.0$					
Pat1M	180.2	95.6	111	$1.0\times1.0\times1.0$					
Pat2M	162.4	51.1	111	$1.0\times1.0\times1.0$					

Table 2. Selected properties of considered computational phantoms

<sup>a</sup>Pat1 is a partial-body phantom. The values in parentheses correspond to the available part of the phantom, without parentheses to the actual height and weight of the individual

#### Mass-correction of SAFs derived for whole-body anatomical models

The MIRD formalism [44] assumes reference mass values for the organs (source and target) and the total body. MIRD Pamphlet 11 [27] provides guidance on how patient-specific scaling of reference radionuclide S values (and eventually SAF values) is to be done for the electron and photon component of the emission spectrum. Based on this guidance and results of extensive analysis of the mass dependences of photon [24] and electron [45] SAFs, the SAFs derived for RCP-AM, P-RCP-AM, Pat1M and Pat2M were mass-corrected. This was done to transfer the simulated values to the investigated fictitious patients (Pat1 and Pat2), who have organ masses differing from those of RCP-AM, P-RCP-AM, Pat1M and Pat2M. The mass-dependences of the SAFs for whole-body phantoms, demonstrated by Petoussi-Henss et al. [24] and Zankl et al. [45], include self-irradiation, i.e. source equal to target for both photons and electrons and the case of electron total-body source with the target being a part of it.

If the source is equal to the target, the SAFs for electrons are inversely proportional to the mass of the target (=source) and the SAFs for photons are proportional to the  $(mass)^{-\frac{2}{3}}$  of the target (=source). The corresponding mass-corrections of SAFs were applied in this case to account for the differences in the masses of target=source between Pat1 and Pat2 and the models approximating them.

Before the SAFs for electrons were explicitly derived from particle transport calculations, an approximation given by ICRP [46] was commonly used. According to it, the SAFs for all internal organs as targets and a total body source are approximately equal to the inverse of the total body mass [46]. The validity of this approximation has also been demonstrated by Zankl et al. [45]. The SAFs derived for a total body source of electrons and internal organs as targets were corrected in this way to account for differences in total body masses of RCP-AM, P-RCP-AM, Pat1M (Pat2M) relative to Pat1 (Pat2).

#### Correction of SAFs evaluated with a partial-body anatomical model

As mentioned above, the Visible Human phantom (Pat1) is a partial-body phantom, from head to mid-thigh. Thus the SAFs simulated for this model had to be adjusted to correspond to a whole-body model. Several cases had to be considered. If the source and the target are both entirely inside the partial-body model, the SAFs of the whole-body model would not differ from those of the partial-body model. Hence, the same mass-correction as for the whole-body models was applied in this case. The second case is for total body as a source and a target being entirely inside the partial-body model. As mentioned above, the electron SAFs in this case are approximately equal to the inverse of the total body mass [45, 46]. Hence, the SAFs evaluated with a phantom comprising of only a portion of the body are overestimated and should be corrected, such that they correspond to the inverse mass of the total body of the whole person, rather than to the mass of the phantom only. The same is applied here for photons. If the source is another extended region ( $\neq$  total body) that is partly outside the partialbody phantom, e.g. total blood or muscles, and the target is entirely inside the phantom, the SAFs are inversely proportional to the total mass of the source region. Thus, the SAFs simulated for partial-body phantoms should be corrected in this case to account for the missing mass of the extended source region. If the source is entirely inside the phantom and the target is an extended region that is only partly inside the available part of the phantom, e.g. total body, skin and muscles, the reciprocity principle, introduced by Cristy and Eckerman [47] for photons and confirmed for electrons by Zankl et

al. [45], suggests that the situation is the same as when the target is entirely inside the partial-body phantom and the source is not. Hence, the SAFs are inversely proportional to the total mass of the target and the SAFs evaluated for partial-body phantoms should be corrected for the missing target mass. If both source and target extend beyond the phantom, electron and photon SAFs are inversely proportional to the highest mass (source or target). Thus, the SAFs simulated for a partial-body phantom had to be corrected for the missing mass of a heavier region (source or target). All the aforementioned cases of mass-correction of the SAFs were implemented in the developed in-house program for the internal organ absorbed-dose calculations and applied in this work.

### Activity data and internal organ dose calculation

For internal dosimetry in this work we employed five sets of time-integrated activity coefficients derived in a previous study [40] for a novel radiopharmaceutical for PET/CT imaging <sup>18</sup>F-FSPG. The case of blood being a distinct source region was considered [40].

We calculated absorbed-dose coefficients five times (i.e. once for each set of activity data) for each of the models Pat1, RCP-AM, P-RCP-AM and Pat1M with the SAFs corrected to correspond to the masses of Pat1 (see sections 2.3.1 and 2.3.2). For each set of activity data the dose differences between RCP-AM, P-RCP-AM and Pat1M relative to Pat1 were computed. Average, maximum and minimum values of these differences are reported. The same procedure was done for the models Pat2, RCP-AM, P-RCP-AM and Pat2M with SAFs scaled to the masses of Pat2. The study [40] was the source of the activity data only. No attempt was made to consider the individual anatomy of the healthy volunteers from the previous study [40] or to track their doses.

An in-house MATLAB-based (MATLAB, version 8.0.0.783 (R2012b), The MathWorks, Inc., Natick Massachusetts, 2012) program was used to calculate organ absorbed-dose coefficients, according to the MIRD system [44]. <sup>18</sup>F is a  $\beta^+$  emitter ( $E_{mean}=249.8$  keV,  $E_{max}=633.5$  keV). The detailed beta spectrum of <sup>18</sup>F from ICRP Publication 107 [48] and a single photon energy at 511 keV (annihilation photons) were utilised for the dosimetric calculations. The SAFs corresponding to those intermediate electron energies which were not explicitly considered in Monte Carlo calculations were derived via a logarithmic interpolation of the simulated data. The curves describing the dependence of the explicitly calculated SAFs upon the electron energy are smooth and, hence, the interpolation was appropriate. The SAFs for bladder contents as source and bladder wall as target depend upon the volume of the bladder contents and the thickness of the bladder wall [49, 50]. The SAFs simulated for the computational phantoms in this work do not reflect these dependences, though, and correspond to fixed volumes of the bladder contents, which amount to 39.3 and 24.3 cm<sup>3</sup> for Pat1 and Pat2, respectively. Since these volumes are substantially smaller than the reference volume of the bladder contents (192.3 cm<sup>3</sup>) [1] and no scaling was done to the bladder, we did not use the SAF values obtained from the phantoms, but exploited those calculated by Andersson et al. [51].

In Pat1 breast was not segmented. Hence for breast in Pat1 the total breast of the RCP-AM was used. The heart wall of Pat1 and Pat2 was surrogated by heart (wall and content). Pat2 is a computational phantom representing a female individual whereas P-RCP-AM and, consequently, Pat2M represent male individuals. Thus, the doses for gonads, uterus or prostate cannot be compared. Because P-RCP-AM is still under development [30-32], some complex structures, e.g. muscles, are not present in P-RCP-AM, Pat1M and Pat2M. Thus subcutaneous fat and skeletal muscle were considered together as only one tissue (residual tissue). The skeletal muscle in P-RCP-AM, Pat1M and Pat2M was surrogated by residual tissue in the dose computation. The elemental composition and the density of the residual tissue were adjusted to correspond to a mixture of muscles and adipose (66 % and 34 %, respectively, for the parenchyma mass fractions).

### Results

### Anatomical models Pat1M and Pat2M

A representative subset of four slices of the voxelised P-RCP-AM is presented in Figure 3. The grey values on the images correspond to the organ labels of the phantom P-RCP-AM. As can be seen from Figure 3, the contours of the organs of P-RCP-AM are smooth and the voxel size of  $1 \times 1 \times 1$  mm<sup>3</sup> is sufficient for realistic representations of most of the organs.



**Figure 3.** A subset of four slices of P-RCP-AM. The voxel size is  $1 \times 1 \times 1$  mm<sup>3</sup>. Grey values correspond to the organ labels.

Figure 4 shows the polygon-surface model P-RCP-AM along with Pat1M and Pat2M that were obtained via the adjustment of P-RCP-AM to some individual characteristics of Pat1 and Pat2. The adjusted dimensions are summarised in Table 3. It can be seen from Figure 4 that the adjusted phantoms Pat1M and Pat2M retain their anatomic realism.



**Figure 4.** P-RCP-AM with the selected dimensions and the result of its adjustment to Pat1 and Pat2 (from left to right).

Adjusted characteristic	Pat1	Pat1M	Difference, %	Pat2	Pat2M	Difference, %
Distance between the iliac crest and clavicles	46.0	46.2	0.4%	40.0	39.7	-0.8%
Front-to-back distance of the rib cage	23.6	23.7	0.3%	15.0	15.1	0.5%
Width of the rib cage	32.7	32.7	0.1%	23.6	23.6	0.2%
Height	180.0	180.2	0.1%	163.0	162.4	-0.4%

**Table 3.** Selected dimensions of Pat1 and Pat2, [cm] and the corresponding matched dimensions of Pat1M and Pat2M, [cm]

The differences are given in % of the original dimensions of Pat1 and Pat2

#### Calculated organ absorbed-dose coefficients

The statistical uncertainties of the Monte Carlo calculations performed notably depend on the absolute values of the absorbed fractions (AF). For the target regions located distantly from the source only a small fraction of energy released from the source can reach the target. This results in low AF and higher statistical uncertainties. For the calculations done here, the coefficients of variance for  $AF \ge 10^{-3}$  were below 0.5 %, in most cases even below 0.25 %. For the values  $10^{-4} \le AF < 10^{-3}$ , the coefficients of variance were below 1.5 %, for  $10^{-5} \le AF < 10^{-4}$  the coefficients of variance amount to 2.5 % at most, and for  $10^{-6} \le AF < 10^{-5}$  they were below 10 %.

The five sets of time-integrated activity coefficients derived in [40] and the simulated SAFs for the six computational phantoms were used for dose computation. We assessed absorbed-dose coefficients 40 times in total: five times for each of the models Pat1, RCP-AM, P-RCP-AM and Pat1M with SAFs scaled to the masses of Pat1 and five times for each of the models Pat2, RCP-AM, P-RCP-AM and Pat2M with SAFs scaled to the masses of Pat2.

To distinguish the impact of the individual organ topology on the computed organ absorbed-dose coefficients, the self-absorption and the cross-fire components of the respective organ absorbed-dose coefficients are given separately in the following.

For each set of activity data the dose differences of RCP-AM, P-RCP-AM and Pat1M relative to Pat1 and of RCP-AM, P-RCP-AM and Pat2M relative to Pat2 were computed. We calculated average, minimum and maximum values of these relative differences for the cross-fire components of the organ dose coefficients. They are summarised in Table 4 and Table 5.

#### Self-absorption component of absorbed-dose coefficients

The source regions fixating the radiopharmaceutical <sup>18</sup>F-FSPG are: stomach wall, liver, thyroid, salivary glands, heart wall, kidneys, urinary bladder-contents, pancreas, spleen and blood. Due to the accumulation of activity in these regions and relatively low energies of the emitted positrons in case of

 $^{18}$ F ( $E_{max}(\beta^+) = 633.5$  keV), the doses received by the source regions are predominantly due to selfabsorption. The calculated absorbed-dose coefficients due to self-absorption along with the relative deviation from the respective values calculated for Pat1 and Pat2 and the absolute contributions of self-absorption to the total organ absorbed-dose coefficients are presented in Figure 5. Note that the absorbed-dose coefficients from Figure 5 are computed with one set of activity data. The corresponding figures obtained using the other four sets of activity data are similar and, hence, are not presented here.

Except for heart wall and stomach wall, no substantial differences in the self-absorption dose coefficients are observed. The contributions of self-absorption to total absorbed-dose coefficients are shown in Figure 5. Notably, the contribution is particularly high for kidneys and pancreas.



**Figure 5.** Calculated self-absorption components of absorbed-dose coefficients, differences relative to the corresponding values for Pat1 (left) and Pat2 (right) and absolute contributions of the self-absorption to the total absorbed-dose coefficients. The total height of the bars on the upper row corresponds to the total organ absorbed-dose coefficients, the filled areas to the self-absorption components. For these calculations, the SAFs estimated with RCP-AM, P-RCP-AM and Pat1M (Pat2M) for organs where source is equal to target were adjusted according to the masses of the target patient Pat1 (Pat2).

### Cross-fire component of absorbed-dose coefficients

The subset of the calculated absorbed-dose coefficients from cross-fire along with the relative deviations from those calculated for Pat1 and Pat2 employing one set of activity data are presented in Figure 6 and Figure 7, respectively. The corresponding figures for the other four sets of activity data

are similar to those of Figure 6 and Figure 7 and, therefore, are not presented here. The same set of activity data is used for Figure 5, Figure 6 and Figure 7.

Table 4 and Table 5 show the relative differences in the calculated absorbed-dose coefficients from cross-fire for Pat1 and Pat2, respectively, and the models approximating Pat1 and Pat2. The values given in Table 4 and Table 5 are computed as average, minimum and maximum within dose differences calculated with five sets of activity data. For the majority of organs of Pat1M (Table 4) and Pat2M (Table 5) the relative deviations in cross-fire absorbed-dose coefficients from those calculated for Pat1 and Pat2 are lower than the respective dose differences between the target phantoms Pat1 and Pat2 and the reference phantoms (RCP-AM, P-RCP-AM).

**Table 4.** Relative differences<sup>a</sup> in the organ absorbed-dose coefficients from cross-fire<sup>b</sup>, calculated for three models approximating Pat1, compared to the corresponding doses of Pat1 (five sets of activity data were used).

	RCP-AM				P-RCP-AM				Pat1M			
	Mean	Min	Max	N	lean	Min	Max		Mean	Min	Max	
Active bone marrow	38.1%	32.7%	46.3%	35	5.2%	30.2%	43.3%		14.5%	11.4%	19.2%	
Colon wall	8.6%	5.8%	10.3%	11	1.0%	7.2%	13.1%		-17.2%	-18.4%	-16.4%	
Lungs	20.8%	20.1%	21.6%	23	3.7%	22.2%	25.3%		-3.1%	-4.6%	-1.0%	
Stomach wall <sup>c</sup>	51.4%	49.6%	54.3%	54	4.7%	52.3%	58.2%		12.4%	9.4%	14.5%	
Breast, total	0.0%	0.0%	0.0%	4	.5%	4.0%	5.5%		-16.4%	-18.1%	-14.1%	
Gonads	-20.6%	-26.5%	-18.3%	-20	0.8%	-26.3%	-18.5%		-28.4%	-34.1%	-26.2%	
Urinary bladder wall	0.6%	0.3%	0.7%	0	.5%	0.3%	0.7%		0.2%	0.1%	0.3%	
Oesophagus	36.0%	30.1%	44.0%	32	2.7%	26.4%	40.4%		8.5%	3.8%	12.2%	
Liver <sup>c</sup>	40.9%	39.8%	41.8%	44	1.3%	43.0%	45.7%		2.2%	0.8%	4.6%	
Thyroid <sup>c</sup>	50.7%	46.8%	57.1%	51	1.5%	47.3%	58.5%		18.9%	16.8%	21.4%	
Brain	29.4%	29.1%	30.3%	31	1.5%	30.4%	34.2%		11.7%	10.3%	12.4%	
Salivary glands <sup>c</sup>	15.7%	15.5%	16.1%	20	).7%	19.8%	22.5%		-9.2%	-10.6%	-8.2%	
Skin, total	12.7%	9.5%	14.6%	11	1.9%	9.1%	13.3%		-5.7%	-10.9%	-2.8%	
Adrenals	38.7%	31.5%	46.9%	40	0.1%	32.7%	48.4%		3.3%	-1.9%	11.4%	
Extrathoracic airways (ET)	19.7%	19.2%	20.7%	25	5.1%	24.0%	27.9%		3.2%	0.9%	4.5%	
Gall bladder wall	80.6%	67.8%	88.7%	75	5.5%	64.4%	83.7%		29.4%	23.2%	35.6%	
Heart <sup>c</sup>	42.3%	38.2%	49.7%	41	1.1%	37.1%	49.2%		8.9%	5.2%	15.0%	
Kidneys <sup>c</sup>	37.7%	36.4%	40.0%	41	1.9%	39.8%	45.0%		4.2%	2.3%	5.9%	
Pancreas <sup>c</sup>	32.8%	29.7%	37.2%	32	2.8%	30.4%	36.8%		-6.5%	-8.7%	-2.9%	
Prostate	-10.2%	-11.1%	-9.8%	-12	2.8%	-13.6%	-12.4%		-15.3%	-16.2%	-14.9%	
Small intestine wall	57.9%	47.7%	65.8%	45	5.3%	34.4%	54.2%		14.0%	5.5%	22.9%	
Spleen <sup>c</sup>	39.9%	30.9%	47.3%	43	3.2%	33.0%	51.7%		6.6%	-0.6%	12.2%	
Thymus	34.6%	32.5%	38.2%	33	3.4%	31.2%	35.6%		10.9%	9.6%	12.4%	
Spinal cord	34.5%	33.4%	36.6%	36	5.8%	35.3%	39.7%		14.0%	13.5%	15.0%	
Skeleton, total	-3.3%	-4.4%	-2.6%	-3	8.6%	-4.5%	-3.0%		-11.7%	-13.3%	-10.9%	
Muscle tissue	23.1%	22.4%	24.8%	31	1.7%	29.0%	35.4%		18.5%	16.8%	20.6%	
Eyes	18.7%	15.9%	20.1%	27	7.2%	25.0%	28.5%		4.9%	-1.9%	8.4%	
Eye lenses	10.3%	7.8%	11.7%	14	4.4%	8.4%	17.6%		-9.9%	-17.6%	-5.9%	

<sup>a</sup>The differences are expressed relative to the absorbed-dose coefficients from cross-fire for Pat1, since these are the patient-specific ones.

<sup>b</sup>Total absorbed-dose coefficients for source organs include in addition the contribution from the self-absorption, which is dominated by the organ mass and is otherwise independent of the anatomy of an individual. Here only the cross-fire components of the absorbed-dose coefficients are given, since only they are affected by the individual organ topology.

<sup>c</sup>These are the source organs for <sup>18</sup>F-FSPG.

For  $AF \ge 10^{-4}$  the coefficients of variance were 1.5 % at most, for  $10^{-5} \le AF \le 10^{-4}$  the coefficients of variance were below 2.5 %. More information on statistical uncertainties of performed Monte Carlo calculations is given in the text.



**Figure 6.** Calculated cross-fire components of the organ absorbed-dose coefficients for Pat1 and different models approximating it. The dose differences in the upper panel are given relative to Pat1 (see also footnote b of Table 4).

The computed relative differences are similar for all activity datasets. No differences in absorbeddose coefficients for urinary bladder wall were observed, since the same SAFs for bladder contents as source and bladder wall as target derived by Andersson et al. [51] were employed. The breast of RCP-AM was used as a surrogate for the breast of Pat1. Hence, no significant differences exist between Pat1 and RCP-AM, while the differences between Pat1 and Pat1M are below 18.1 %. For Pat1 the relative differences in cross-fire absorbed-dose coefficients of liver, extrathoracic airways, kidneys, pancreas and eyes decreased from about 30.0–40.0 % for the reference phantoms RCP-AM and P-RCP-AM to less than 10.0 % for the matched phantom Pat1M. For oesophagus, brain, adrenals, heart wall, spleen, thymus and spinal cord the cross-fire dose differences between Pat1 and the reference phantoms amount to approximately 40.0 %, while the corresponding differences between Pat1 and Pat1M are below 15.0 %. For Pat1 the differences in absorbed-dose coefficients of gall bladder wall amount to 80.6 % and 75.5 % on average for RCP-AM and P-RCP-AM, respectively. The corresponding differences relative to Pat1M are below 30.0 % on average. For active bone marrow, thyroid and salivary glands the calculated relative dose differences between Pat1M and Pat1 are only half of those between reference phantoms and Pat1. For lungs the computed dose differences decreased on average from 21–24 % between Pat1 and reference phantoms to -3 % between Pat1 and Pat1M. The differences in the calculated cross-fire absorbed-dose coefficients for stomach wall and small intestine wall amount, respectively, to 12.4 % and 14.3 % on average between Pat1 and Pat1M. The corresponding differences between Pat1 and both reference phantoms are within 54.7 % and 57.9 % on average. We observed better agreement between Pat1 and the reference phantoms (RCP-AM and P-RCP-AM) in the absorbed-dose coefficients for gonads, prostate, skeleton and colon wall. Absorbed-dose coefficients for all organs, except gonads and gall bladder wall are consistent between Pat1 and Pat1M within 25.0 % (Table 4).

**Table 5.** Analogue to Table 4 the relative differences<sup>a</sup> in the organ absorbed-dose coefficients from cross-fire, for three models approximating Pat2, compared to corresponding doses of Pat2 (five sets of activity data were used).

	RCP-AM			P-RCP-AM				Pat2M			
	Mean	Min	Max	 Mean	Min	Max		Mean	Min	Max	
Active bone marrow	0.7%	-2.1%	6.4%	-1.3%	-4.3%	4.3%		17.9%	14.1%	26.3%	
Colon wall	-15.5%	-17.4%	-13.8%	-13.9%	-16.4%	-12.0%		10.2%	5.9%	12.4%	
Lungs	-28.0%	-33.5%	-25.1%	-26.5%	-31.5%	-23.9%		-7.2%	-11.0%	-4.7%	
Stomach wall <sup>b</sup>	-29.8%	-31.6%	-24.7%	-28.1%	-29.9%	-22.9%		-6.7%	-8.9%	-0.8%	
Breast, total	-7.0%	-9.7%	-3.2%	-3.5%	-6.7%	1.4%		19.2%	13.2%	28.0%	
Urinary bladder wall	-0.9%	-1.1%	-0.5%	-1.0%	-1.2%	-0.5%		-0.4%	-0.5%	-0.2%	
Oesophagus	-4.7%	-6.5%	-2.8%	-6.9%	-8.3%	-5.4%		17.1%	14.0%	19.0%	
Liver <sup>b</sup>	-34.4%	-35.2%	-34.0%	-32.8%	-33.4%	-32.4%		-10.0%	-10.8%	-9.4%	
Thyroid <sup>b</sup>	-7.5%	-11.9%	-5.4%	-7.0%	-11.1%	-5.1%		17.5%	14.5%	18.9%	
Brain	-13.4%	-14.4%	-13.0%	-12.4%	-12.4%	-12.3%		6.7%	5.7%	9.2%	
Salivary glands <sup>b</sup>	-42.1%	-42.6%	-41.6%	-39.7%	-40.9%	-38.1%		-16.3%	-17.9%	-13.9%	
Skin, total	-15.5%	-16.9%	-14.5%	-16.3%	-17.4%	-15.6%		-0.9%	-1.6%	0.3%	
Adrenals	-25.4%	-29.4%	-19.8%	-24.7%	-28.7%	-19.0%		-2.8%	-8.0%	3.6%	
Extrathoracic airways (ET)	-25.5%	-29.9%	-23.3%	-22.7%	-26.3%	-21.0%		-4.8%	-6.2%	-4.1%	
Gall bladder wall	-8.2%	-13.1%	-3.6%	-11.0%	-15.2%	-7.5%		19.4%	9.9%	23.8%	
Heart <sup>b</sup>	-5.1%	-13.5%	-0.3%	-5.8%	-14.7%	-0.5%		18.5%	8.3%	24.5%	
Kidneys <sup>b</sup>	-38.0%	-38.9%	-35.8%	-36.0%	-36.8%	-33.7%		-14.6%	-16.0%	-11.3%	
Pancreas <sup>b</sup>	-38.1%	-38.7%	-37.1%	-38.1%	-38.9%	-37.3%		-16.9%	-18.2%	-15.8%	
Small intestine wall	-29.4%	-33.8%	-24.5%	-34.8%	-38.2%	-30.9%		-18.1%	-22.9%	-13.6%	
Spleen <sup>b</sup>	-21.9%	-26.3%	-18.0%	-20.1%	-25.1%	-16.0%		3.3%	-4.0%	7.8%	
Thymus	-26.9%	-29.5%	-23.1%	-27.5%	-29.9%	-23.8%		-12.7%	-16.3%	-8.3%	
Spinal cord	-25.3%	-28.4%	-22.2%	-24.1%	-26.9%	-21.3%		-7.9%	-9.4%	-5.4%	
Skeleton, total	-8.9%	-10.9%	-6.5%	-9.2%	-11.2%	-6.7%		7.3%	4.6%	12.0%	
Muscle tissue	-24.7%	-27.7%	-22.2%	-20.2%	-22.3%	-18.8%		-4.2%	-5.0%	-3.5%	
Eyes	-27.1%	-32.2%	-24.4%	-22.7%	-27.6%	-20.2%		-2.6%	-5.6%	-1.0%	
Eye lenses	-28.4%	-33.7%	-25.6%	-26.2%	-33.3%	-22.4%		-12.4%	-17.6%	-9.5%	

<sup>a</sup>The differences are expressed relative to the absorbed-dose coefficients from cross-fire for Pat2, since these are the patient-specific ones.

<sup>b</sup>These are the source organs for <sup>18</sup>F-FSPG.



**Figure 7.** Calculated cross-fire components of the organ absorbed-dose coefficients for Pat2 and different models approximating it. The dose differences in the upper panel are given relative to Pat2 (see also footnote b of Table 4).

For Pat2 the differences in the cross-fire absorbed-dose coefficients for kidneys, pancreas, brain, small intestine, thymus, and eye lenses relative to Pat2M are approximately lower by a factor of two compared to those relative to the reference phantoms. As a general finding for all sets of activity data, the differences in the absorbed-dose coefficients for lungs, stomach wall (cross-fire), adrenals, extrathoracic airways (ET) and spinal cord decreased from approximately 30.0 % between RCP-AM, P-RCP-AM and Pat2 to less than 10.0 % between Pat2M and Pat2. The absorbed-dose coefficients for eyes and muscles in Pat2M are consistent with the corresponding values in Pat2 within 5.6 %. For liver and salivary glands the calculated relative cross-fire dose differences between Pat2M and Pat2 are only one third of those between reference phantoms and Pat2, for skin and eyes only one tenth. For the cross-fire absorbed-dose coefficients for active bone marrow, breast, oesophagus, thyroid, gall-bladder wall and heart wall better agreement was observed between Pat2 and reference phantoms,

though. Absorbed-dose coefficients from cross-fire for all organs and tissues of Pat2M considered, except the active bone marrow and breast, are consistent with the ones of Pat2 within 25.0 % (Table 5). Despite the gross gender-specific differences between Pat2 and Pat2M, the difference in the organ absorbed-dose coefficients from cross-fire were still held to <25.0 % for most organs. The gender-specific organs cannot be compared in this case.

### Discussion

The voxelised versions  $(1 \times 1 \times 1 \text{ mm}^3 \text{ voxel size})$  of the polygon-surface models were employed for the Monte Carlo calculations in this work. This potentially introduces the limitation of the voxelised models to realistically represent very small (< 1 mm in at least one dimension) anatomical structures. Nonetheless, the voxel size used is notably smaller than in the models segmented directly from the tomographic images (see e.g. Table 2) and it was sufficient for retaining the anatomical realism of most of the organs. The voxelisation can be theoretically done with even finer voxel sizes if accurate dosimetry is needed in specific cases, e.g. for ET, oral mucosa, eye lenses, basal cells of the epidermis etc. The limiting factors in this case are the available memory to store the phantom and the increasing calculation time of the Monte Carlo simulations when decreasing the voxel size.

Self-absorption is the main contribution to the organ absorbed-dose coefficients for most source organs, especially kidneys, pancreas, salivary glands, liver and thyroid. Figure 5 shows that the differences in the self-absorption are mostly eliminated by applying a mass-correction to the SAFs reflecting the masses of a target patient. Higher differences in the self-absorption dose coefficients between the target patient-specific models Pat1 and Pat2 and the phantoms approximating them were observed for heart wall (16–18 %) and stomach wall (4–8 %). The mass-scaling of the SAFs applied might have limitations for walled organs. Additionally, Pat1 and Pat2 do not have heart wall segmented separately, but only the whole heart instead. This could be another reason for the differences observed for the heart wall, since the self-absorption in the whole heart is expected to be higher than that in the heart wall only. For some source organs, e.g. stomach wall and spleen, the absolute contributions to the absorbed-dose coefficients from the cross-fire were also substantial due to high activities of some neighbouring source organs.

High contribution to the absorbed-dose coefficients by the cross-fire from blood was observed for some organs, e.g. lungs, oesophagus, extrathoracic airways, thymus, eyes and eye lenses. The differences in the cross-fire from blood cannot be reduced by phantom adjustments. The cross-fire contribution from blood depends on how blood is attributed to different body organs and tissues in the Monte Carlo calculations. Since individual blood volumes and individual blood contributions in different organs and tissues are usually not available, the cross-fire from blood is a source of uncertainty of the absorbed-dose estimation and requires further careful considerations that go beyond the scope of this work.

We showed the feasibility of reducing the differences between the patient-specific cross-fire and the cross-fire estimated for adjusted reference models for most organs. Nonetheless, the results might

differ for other radiopharmaceuticals, which have other source regions than <sup>18</sup>F-FSPG. Comprehensive conclusions could be drawn if we compared the SAFs evaluated for all source-target combinations of the phantoms and different energies of the emitted  $\beta$ -particles (not only those corresponding to the  $\beta$ spectrum of  ${}^{18}$ F). However, such a comparison is practically very difficult because there are too many source-target combinations. Additionally, it is not generally obvious how the differences in the SAFs are propagated to the differences in the resulting organ absorbed-dose coefficients, since this also depends on the absolute values of SAFs and the absolute values of time-integrated activity coefficients. A considerable methodological limitation of our work is the usage of only two target patient-specific phantoms. We selected Pat1 and Pat2 as the biggest and the smallest models in the HMGU library, respectively. They represent individuals with a pronounced difference in the BMI (31.8 kg/m<sup>2</sup> and 19.2 kg/m<sup>2</sup> for Pat1 and Pat2, respectively). Pat1 and Pat2 also have unique and unrelated organ topology. Relatively simple adjustments without matching the locations and masses of individual organs to those of Pat1 and Pat2 were applied in this work. Consequently, the cross-fire dose coefficients between the two fictitious target patients considered and the adjusted polygonsurface reference phantom P-RCP-AM were consistent for most organs within 25 %. Employing reference phantoms without any adjustment resulted in estimations of patient cross-fire dose coefficients within approximately 50 % relative uncertainty for the majority of target regions. However, the achieved improvement in the accuracy of organ dose coefficients might not apply to other individuals. To capture the whole range of inter-individual anatomic variability, more patientspecific phantoms would be required. The female fictitious target patient Pat2 was represented by male anatomy of the adjusted P-RCP-AM. Thus gender-specific organs of Pat2 were not considered. This is a limitation of the presented work. The results of our test study showed the feasibility of reducing the uncertainties in the cross-fire absorbed-dose coefficients for both patient-specific models employed, despite them being segmented from individuals with unrelated anatomy and pronounced difference in the BMI.

For a convenient application of the proposed method the dimensions to be adjusted should be relatively easy to obtain for individual patients. The set of dimensions adjusted in this work was selected accordingly. For constructing Pat1M and Pat2M individual organ masses and positions were not adjusted to match the corresponding values of the patient-specific phantoms Pat1 and Pat2. The employed mass-correction of the SAFs is phantom-independent and requires only the masses of the target=source regions. The clinical quantification of the absolute activities in the source regions – which in any case needs to be determined to yield accurate personalised self-absorption doses –

implicitly includes the estimation of the source region masses as part of its procedure. The absolute activities are calculated via the multiplication of the activity concentrations, determined from the PET images, and the source region volumes, obtained from the registered CT images.

Patient-specific dosimetry potentially provides more accurate dose values compared to our proposed approach. The former can be implemented in two ways: a direct dosimetry based on patient CT images or a conversion of patient tomographic images into a voxelized patient-specific phantom, followed by the corresponding dosimetry. However, both patient-specific options require the contouring of all regions of interest. This is often difficult in clinical conditions, especially with a lack of automated segmentation tools. Additionally, patient-specific dosimetry requires individual CT or magnetic resonance images of sufficiently large body coverage and adequate resolution to realistically represent even small organs and tissues. Such tomographic data are not always available. Although patient-specific dosimetry on patient tomographic images is considered the most accurate dosimetry option, the abovementioned factors often limit its application in medical practice. The proposed approach can be useful in those scenarios where the patient-specific dosimetry is not feasible. Our study showed the potential to improve the dosimetry in nuclear medicine by adjusting P-RCP-AM to the dimensions of a patient. This approach requires less effort than would be needed were we to segment a patient-specific model. An alternative approach could be to select the phantom, which agrees best with the specific patient under consideration, from the already existing large libraries [8, 9, 11, 12, 14, 15, 17-19]. However, since most of these libraries were constructed based on variations in height and weight as primary parameters, the approach chosen here can help adjusting the selected phantom to the dimensions of the specific patient even better, without too much effort. The dimensions adjusted in our study include front-to back distance and width of the rib cage, height, distance between the iliac crest and clavicles. These dimensions could be also considered in selecting the phantom that best matches to the individual patient. This might further reduce the differences in the estimated doses.

### Conclusions

To evaluate the feasibility of reducing the uncertainty in the estimated doses in nuclear medicine caused by anatomical difference between the patient-specific and the reference phantom, we adjusted the polygon-surface ICRP adult male reference phantom to a set of external dimensions of two investigated individuals. They were represented by the voxel phantoms Visible Human (=Pat1) and Irene (=Pat2). Using the two resulting phantoms from the adjustment and five sets of activity data of a radiopharmaceutical, designated for PET/CT imaging, we studied which agreement in the computed doses can be achieved. The results of our pilot study showed the feasibility of reducing the uncertainty in the cross-fire component of the estimated dose coefficients by a relatively simple phantom adjustment; that is, < 25 % of relative uncertainty in dose coefficients from cross-fire for most organs and tissues. Overall, the proposed approach provided reliable dose estimates for both tested patients despite the pronounced differences in their anatomy. To capture the full range of inter-individual variability more patients would be required. We believe the proposed approach could be a good tradeoff balancing the effort to be expended and the accuracy to be achieved. It has the potential to be directly applicable in medical practice, in contrast to the less viable segmentation of a patient-specific phantom. The level of personalisation of an employed anatomical model should depend on the application and on the available resources. Our results suggest that the proposed approach could be reliable enough for most situations involving low to moderate doses (i.e. outside the therapy realm). Better agreement in cross-fire could also be achieved by using a phantom selected from a collection of diversified models which best matches the external dimensions of the patient.

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## **Conflict of interest statement**

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