IPF: Let's Keep the Focus on the A(ge)TII cell

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Chronic lung diseases (CLDs) such as idiopathic pulmonary fibrosis (IPF) are the third leading cause of death worldwide [1], and lung transplantation remains the only curative therapy to date. In the case of IPF, aging has emerged as the single most significant risk factor. With the worldwide aging population, its incidence is expected to rise significantly [2]. On a cellular level, several hallmarks have been identified as drivers of aging [3, 4], and IPF exhibits strong aging hallmarks, leading to severely compromised regeneration of the diseased lung [2]. Among these hallmarks are cellular senescence and metabolic reprogramming, which have been described as significant cellular changes in IPF, especially in ATII cells [2, 5]. Transient development of fibrosis characterizes the bleomycin mouse model of pulmonary fibrosis. However, when bleomycin is instilled into aged animals, the fibrosis is persistent, further highlighting the contribution of lung aging to the development of non-resolving fibrosis [6, 7]. The increase in cellular senescence leads to a senescence-associated secretory phenotype (SASP), culminating in a proinflammatory and pro-fibrotic milieu in the aged lung. At the same time, geroprotective factors such as Klotho or SIRT1 are downregulated. The main drivers of cellular aging in ATII cells are incompletely understood and deciphering the upstream regulators will be a significant step in identifying therapeutic targets to reduce or reverse this process.

Here, the authors describe a role of a cardinal nicotinamide adenine dinucleotide (NAD) hydrolase, CD38, in promoting mitochondrial dysfunction and cellular senescence of ATII cells leading to pulmonary fibrosis [8]. The authors use mouse models, cell lines, and primary mouse cells to show that CD38, a NAD consuming enzyme, increases fibrosis. At the same time, in both IPF and experimental fibrosis, there is a decrease in NAD levels in ATII cells. Changes in NAD levels are accompanied by a differential regulation of cellular energetics and mitochondrial function that have been well described in the literature [9]. Similarly, the authors confirm the existence of senescent ATII cells in experimental fibrosis and explanted lungs from IPF patients [10]. It remains unclear what drives CD38 overexpression in aged lungs, and future studies are

needed to define the mechanisms in the aging human lung. In this paper, the authors demonstrate that overexpression of CD38 in ATII cell lines leads to decreased NAD levels, mitochondrial dysfunction, and increased cellular senescence. Notably, CD38 downregulation or inhibition blunts in vitro, bleomycin-induced cellular senescence, and mitochondrial dysfunction, suggesting CD38 as a promising pharmacological target. By decreasing NAD levels, CD38 overexpression inhibits major NAD consuming enzymes like Sirtuins and PARPs, likely to contribute to the observed mitochondrial and senescence effects.

It has been identified in other models of fibrosis that in vivo CD38 inhibition effectively decreases fibrosis in aged mice, improving mitochondrial function and decreasing the cellular senescence [11]. In the present manuscript, similarly, CD38 knockout mice are protected from the development of pulmonary fibrosis. Even more importantly, bleomycin-induced fibrosis can be reversed with a molecule inhibition of CD38.

This study builds upon a growing body of literature identifying cellular aging hallmarks in ATII cells of IPF patients [2, 9, 10, 12]. Targeting specific hallmarks such as cellular senescence with the help of senolytics showed promising preclinical results, and first in human trials demonstrated initial tolerability in IPF patients [10, 12-14]. This further highlights the applicability of therapies targeting cellular aging mechanisms in IPF. The finding that CD38 inhibition does not only decrease the level of cellular senescence but also reduces oxidative stress and reverts mitochondrial dysfunction is intriguing and argues that CD38 is a master regulator of multiple aging-related mechanisms. This suggests that aging processes are highly interlinked and supports the "Unitary theory of targeting fundamental aging mechanisms," which suggests that targeting one aging mechanism might positively impact one or all other aging mechanisms [15]. Interestingly, therapies clearing senescent cells can be administered in an intermittent manner suggesting that potential off-target effects will be reduced. It will be of specific interest to

Page 4 of 8

understand the role of CD38 in other age-related lung diseases that have been linked to ATII senescence, such as COPD.

Although these studies demonstrate that pharmacological inhibition of CD38 has both preventive and therapeutic benefits in relevant mouse models of pulmonary fibrosis, several unknowns need to be considered when moving towards the clinical application of CD38 inhibition as a novel IPF therapy. The authors show an effect of CD38 inhibition one week after Bleomycin administration; it remains unknown if CD38 inhibition might be able to revert established fibrosis, 14 to 21 days after fibrosis, as seen in IPF patients. Furthermore, the study presents limited human data, and a human preclinical model of IPF such as PCLS is needed to confirm suitability for human disease. Again, not only ATII cells express CD38 In the fibrotic lung. Other epithelial cells, such as basal and basaloid cells, show detectable CD38 expression, and future studies are needed to address the relationship between CD38 and premature aging in other cell types. Similarly, other senescent cells, such as fibroblasts, have been implicated in the pathogenesis of IPF [7, 16], do not seem to overexpress CD38, and the effect of CD38 inhibition on these cell populations remains elusive.

This does not diminish enthusiasm for targeting cellular aging in ATII cells of IPF patients but highlights the need to characterize cell-type-specific susceptibility to specific therapeutic strategies further. Additionally, it underlines the potential market and promise of combination therapies affecting different aging hallmarks in multiple cellular compartments.

In summary, this exciting study identifies CD38 as a potential regulator of aging hallmarks in fibrotic ATII cells and suggests CD38 inhibition as a possible novel therapeutic strategy to target age-associated IPF.

References

1. Lozano R, Naghavi M, Foreman K, Lim S, Shibuya K, Aboyans V, Abraham J, Adair T, Aggarwal R, Ahn SY, Alvarado M, Anderson HR, Anderson LM, Andrews KG, Atkinson C, Baddour LM, Barker-Collo S, Bartels DH, Bell ML, Benjamin EJ, Bennett D, Bhalla K, Bikbov B, Bin Abdulhak A, Birbeck G, Blyth F, Bolliger I, Boufous S, Bucello C, Burch M, Burney P, Carapetis J, Chen H, Chou D, Chugh SS, Coffeng LE, Colan SD, Colguhoun S, Colson KE, Condon J, Connor MD, Cooper LT, Corriere M, Cortinovis M, de Vaccaro KC, Couser W, Cowie BC, Criqui MH, Cross M, Dabhadkar KC, Dahodwala N, De Leo D, Degenhardt L, Delossantos A, Denenberg J, Des Jarlais DC, Dharmaratne SD, Dorsey ER, Driscoll T, Duber H, Ebel B, Erwin PJ, Espindola P, Ezzati M, Feigin V, Flaxman AD, Forouzanfar MH, Fowkes FG, Franklin R, Fransen M, Freeman MK, Gabriel SE, Gakidou E, Gaspari F, Gillum RF, Gonzalez-Medina D, Halasa YA, Haring D, Harrison JE, Havmoeller R, Hay RJ, Hoen B, Hotez PJ, Hoy D, Jacobsen KH, James SL, Jasrasaria R, Jayaraman S, Johns N, Karthikeyan G, Kassebaum N, Keren A, Khoo JP, Knowlton LM, Kobusingye O, Koranteng A, Krishnamurthi R, Lipnick M, Lipshultz SE, Ohno SL, Mabweijano J, MacIntyre MF, Mallinger L, March L, Marks GB, Marks R, Matsumori A, Matzopoulos R, Mayosi BM, McAnulty JH, McDermott MM, McGrath J, Mensah GA, Merriman TR, Michaud C, Miller M, Miller TR, Mock C, Mocumbi AO, Mokdad AA, Moran A, Mulholland K, Nair MN, Naldi L, Narayan KM, Nasseri K, Norman P, O'Donnell M, Omer SB, Ortblad K, Osborne R, Ozgediz D, Pahari B, Pandian JD, Rivero AP, Padilla RP, Perez-Ruiz F, Perico N, Phillips D, Pierce K, Pope CA, 3rd, Porrini E, Pourmalek F, Raju M, Ranganathan D, Rehm JT, Rein DB, Remuzzi G, Rivara FP, Roberts T, De Leon FR, Rosenfeld LC, Rushton L, Sacco RL, Salomon JA, Sampson U, Sanman E, Schwebel DC, Segui-Gomez M, Shepard DS, Singh D, Singleton J, Sliwa K, Smith E, Steer A, Taylor JA, Thomas B, Tleyjeh IM, Towbin JA, Truelsen T, Undurraga EA, Venketasubramanian N, Vijayakumar L, Vos T, Wagner GR, Wang M, Wang W, Watt K, Weinstock MA, Weintraub R, Wilkinson JD, Woolf AD, Wulf S, Yeh PH, Yip P, Zabetian A, Zheng

ZJ, Lopez AD, Murray CJ, AlMazroa MA, Memish ZA. Global and regional mortality from 235 causes of death for 20 age groups in 1990 and 2010: a systematic analysis for the Global Burden of Disease Study 2010. *Lancet* 2012: 380(9859): 2095-2128.

2. Meiners S, Eickelberg O, Konigshoff M. Hallmarks of the ageing lung. *The European respiratory journal* 2015: 45(3): 807-827.

3. Lopez-Otin C, Blasco MA, Partridge L, Serrano M, Kroemer G. The hallmarks of aging. *Cell* 2013: 153(6): 1194-1217.

4. Pun FW, Leung GHD, Leung HW, Liu BHM, Long X, Ozerov IV, Wang J, Ren F, Aliper A, Izumchenko E, Moskalev A, de Magalhaes JP, Zhavoronkov A. Hallmarks of aging-based dualpurpose disease and age-associated targets predicted using PandaOmics AI-powered discovery engine. *Aging* 2022: 14(6): 2475-2506.

5. Barnes PJ, Baker J, Donnelly LE. Cellular Senescence as a Mechanism and Target in Chronic Lung Diseases. *American journal of respiratory and critical care medicine* 2019: 200(5): 556-564.

6. Hecker L, Logsdon NJ, Kurundkar D, Kurundkar A, Bernard K, Hock T, Meldrum E, Sanders YY, Thannickal VJ. Reversal of persistent fibrosis in aging by targeting Nox4-Nrf2 redox imbalance. *Science translational medicine* 2014: 6(231): 231ra247.

7. Kato K, Logsdon NJ, Shin YJ, Palumbo S, Knox A, Irish JD, Rounseville SP, Rummel SR, Mohamed M, Ahmad K, Trinh JM, Kurundkar D, Knox KS, Thannickal VJ, Hecker L. Impaired Myofibroblast Dedifferentiation Contributes to Nonresolving Fibrosis in Aging. *American journal of respiratory cell and molecular biology* 2020: 62(5): 633-644.

8. Cui H, Xie N, Banerjee S, Dey T, Liu RM, Antony VB, Sanders YY, Adams TS, Gomez JL, Thannickal VJ, Kaminski N, Liu G. CD38 Mediates Lung Fibrosis by Promoting Alveolar Epithelial Cell Aging. *Am J Respir Crit Care Med* [online ahead of print] 06 June 2022; https://www.atsjournals.org/doi/abs/10.1164/rccm.202109-2151OC.

9. Bueno M, Lai YC, Romero Y, Brands J, St Croix CM, Kamga C, Corey C, Herazo-Maya JD, Sembrat J, Lee JS, Duncan SR, Rojas M, Shiva S, Chu CT, Mora AL. PINK1 deficiency impairs mitochondrial homeostasis and promotes lung fibrosis. *The Journal of clinical investigation* 2015: 125(2): 521-538.

10. Lehmann M, Korfei M, Mutze K, Klee S, Skronska-Wasek W, Alsafadi HN, Ota C, Costa R, Schiller HB, Lindner M, Wagner DE, Gunther A, Konigshoff M. Senolytic drugs target alveolar epithelial cell function and attenuate experimental lung fibrosis ex vivo. *The European respiratory journal* 2017: 50(2).

11. Shi B, Wang W, Korman B, Kai L, Wang Q, Wei J, Bale S, Marangoni RG, Bhattacharyya S, Miller S, Xu D, Akbarpour M, Cheresh P, Proccissi D, Gursel D, Espindola-Netto JM, Chini CCS, de Oliveira GC, Gudjonsson JE, Chini EN, Varga J. Targeting CD38-dependent NAD(+) metabolism to mitigate multiple organ fibrosis. *iScience* 2021: 24(1): 101902.

12. Schafer MJ, White TA, Iijima K, Haak AJ, Ligresti G, Atkinson EJ, Oberg AL, Birch J, Salmonowicz H, Zhu Y, Mazula DL, Brooks RW, Fuhrmann-Stroissnigg H, Pirtskhalava T, Prakash YS, Tchkonia T, Robbins PD, Aubry MC, Passos JF, Kirkland JL, Tschumperlin DJ, Kita H, LeBrasseur NK. Cellular senescence mediates fibrotic pulmonary disease. *Nature communications* 2017: 8: 14532.

13. Justice JN, Nambiar AM, Tchkonia T, LeBrasseur NK, Pascual R, Hashmi SK, Prata L, Masternak MM, Kritchevsky SB, Musi N, Kirkland JL. Senolytics in idiopathic pulmonary fibrosis: Results from a first-in-human, open-label, pilot study. *EBioMedicine* 2019: 40: 554-563.

14. Zhu Y, Prata L, Gerdes EOW, Netto JME, Pirtskhalava T, Giorgadze N, Tripathi U, Inman CL, Johnson KO, Xue A, Palmer AK, Chen T, Schaefer K, Justice JN, Nambiar AM, Musi N,

Kritchevsky SB, Chen J, Khosla S, Jurk D, Schafer MJ, Tchkonia T, Kirkland JL. Orally-active, clinically-translatable senolytics restore alpha-Klotho in mice and humans. *EBioMedicine* 2022: 77: 103912.

15. Tchkonia T, Palmer AK, Kirkland JL. New Horizons: Novel Approaches to Enhance Healthspan Through Targeting Cellular Senescence and Related Aging Mechanisms. *J Clin Endocrinol Metab* 2021: 106(3): e1481-e1487.

16. Alvarez D, Cardenes N, Sellares J, Bueno M, Corey C, Hanumanthu VS, Peng Y, D'Cunha H, Sembrat J, Nouraie M, Shanker S, Caufield C, Shiva S, Armanios M, Mora AL, Rojas M. IPF lung fibroblasts have a senescent phenotype. *American journal of physiology Lung cellular and molecular physiology* 2017: 313(6): L1164-L1173.