#### ARTICLE



# Role of human plasma metabolites in prediabetes and type 2 diabetes from the IMI-DIRECT study

Sapna Sharma<sup>1</sup> · Qiuling Dong<sup>1,2</sup> · Mark Haid<sup>3</sup> · Jonathan Adam<sup>1,4</sup> · Roberto Bizzotto<sup>5</sup> · Juan J. Fernandez-Tajes<sup>6</sup> · Angus G. Jones<sup>7</sup> · Andrea Tura<sup>5</sup> · Anna Artati<sup>3</sup> · Cornelia Prehn<sup>3</sup> · Gabi Kastenmüller<sup>8</sup> · Robert W. Koivula<sup>9</sup> · Paul W. Franks<sup>10</sup> · Mark Walker<sup>11</sup> · Ian M. Forgie<sup>12</sup> · Giuseppe Giordano<sup>13</sup> · Imre Pavo<sup>14</sup> · Hartmut Ruetten<sup>15</sup> · Manolis Dermitzakis<sup>16,17,18</sup> · Mark I. McCarthy<sup>6</sup> · Oluf Pedersen<sup>19,20</sup> · Jochen M. Schwenk<sup>21</sup> · Konstantinos D. Tsirigos<sup>22</sup> · Federico De Masi<sup>22</sup> · Soren Brunak<sup>20,23</sup> · Ana Viñuela<sup>24</sup> · Andrea Mari<sup>5</sup> · Timothy J. McDonald<sup>25</sup> · Tarja Kokkola<sup>26</sup> · Jerzy Adamski<sup>27,28,29</sup> · Ewan R. Pearson<sup>12</sup> · Harald Grallert<sup>1,4</sup>

Received: 28 February 2024 / Accepted: 29 July 2024 © The Author(s) 2024

# Abstract

**Aims/hypothesis** Type 2 diabetes is a chronic condition that is caused by hyperglycaemia. Our aim was to characterise the metabolomics to find their association with the glycaemic spectrum and find a causal relationship between metabolites and type 2 diabetes.

Methods As part of the Innovative Medicines Initiative - Diabetes Research on Patient Stratification (IMI-DIRECT) consortium, 3000 plasma samples were measured with the Biocrates Absolute IDQ p150 Kit and Metabolon analytics. A total of 911 metabolites (132 targeted metabolomics, 779 untargeted metabolomics) passed the quality control. Multivariable linear and logistic regression analysis estimates were calculated from the concentration/peak areas of each metabolite as an explanatory variable and the glycaemic status as a dependent variable. This analysis was adjusted for age, sex, BMI, study centre in the basic model, and additionally for alcohol, smoking, BP, fasting HDL-cholesterol and fasting triacylglycerol in the full model. Statistical significance was Bonferroni corrected throughout. Beyond associations, we investigated the mediation effect and causal effects for which causal mediation test and two-sample Mendelian randomisation (2SMR) methods were used, respectively. **Results** In the targeted metabolomics, we observed four (15), 34 (99) and 50 (108) metabolites (number of metabolites observed in untargeted metabolomics appear in parentheses) that were significantly different when comparing normal glucose regulation vs impaired glucose regulation/prediabetes, normal glucose regulation vs type 2 diabetes, and impaired glucose regulation vs type 2 diabetes, respectively. Significant metabolites were mainly branched-chain amino acids (BCAAs), with some derivatised BCAAs, lipids, xenobiotics and a few unknowns. Metabolites such as lysophosphatidylcholine a C17:0, sum of hexoses, amino acids from BCAA metabolism (including leucine, isoleucine, valine, N-lactoylvaline, N-lactoylleucine and formiminoglutamate) and lactate, as well as an unknown metabolite (X-24295), were associated with HbA<sub>1c</sub> progression rate and were significant mediators of type 2 diabetes from baseline to 18 and 48 months of follow-up. 2SMR was used to estimate the causal effect of an exposure on an outcome using summary statistics from UK Biobank genome-wide association studies. We found that type 2 diabetes had a causal effect on the levels of three metabolites (hexose, glutamate and caproate [fatty acid (FA) 6:0]), whereas lipids such as specific phosphatidylcholines (PCs) (namely PC aa C36:2, PC aa C36:5, PC ae C36:3 and PC ae C34:3) as well as the two n-3 fatty acids stearidonate (18:4n3) and docosapentaenoate (22:5n3) potentially had a causal role in the development of type 2 diabetes.

**Conclusions/interpretation** Our findings identify known BCAAs and lipids, along with novel *N*-lactoyl-amino acid metabolites, significantly associated with prediabetes and diabetes, that mediate the effect of diabetes from baseline to follow-up (18 and 48 months). Causal inference using genetic variants shows the role of lipid metabolism and n-3 fatty acids as being causal for metabolite-to-type 2 diabetes whereas the sum of hexoses is causal for type 2 diabetes-to-metabolite. Identified metabolite markers are useful for stratifying individuals based on their risk progression and should enable targeted interventions.

Sapna Sharma, Qiuling Dong and Mark Haid contributed equally to this study.

Extended author information available on the last page of the article

# Research in context

#### What is already known about this subject?

- Type 2 diabetes is a chronic condition that is characterised by hyperglycaemia
- The primary objectives of the DIRECT consortium include the identification of potential biomarkers that can assist in categorising individuals based on their glycaemic deterioration

#### What is the key question?

• What is the correlation between metabolomics and the glycaemic spectrum, and is there a causal relationship between any metabolites and type 2 diabetes?

#### What are the new findings?

- Novel metabolites such as picolinoylglycine, N-lactoylvaline, N-lactoylleucine and formiminoglutamate, as well as
  lactate and an unknown metabolite (X-24295, from untargeted metabolomics), were associated with prediabetes,
  type 2 diabetes and HbA<sub>1c</sub> progression rate and were significant mediators of type 2 diabetes from baseline to 18
  and 48 months of follow-up
- Causal inference using genetic variants shows that the *n*-3 fatty acids stearidonate (18:4n3) and docosapentaenoate (22:5n3) have a causal role in type 2 diabetes

#### How might this impact on clinical practice in the foreseeable future?

• Identified metabolite markers may be useful for stratifying individuals based on their risk of progression and may enable targeted interventions

**Keywords** Causality · Glycaemic traits · HbA<sub>1c</sub> · IMI-DIRECT · Mediation · Metabolomics · *N*-lactoylaminoacids · Patient stratification · Targeted metabolomics · Type 2 diabetes · Untargeted metabolomics

#### Abbreviations

2SMR	Two-sample MR
GWAS	Genome-wide association study
H1	Hexoses
IGR	Impaired glucose regulation
IGT	Impaired glucose tolerance
IMI-DIRECT	Innovative Medicines Initiative - Diabetes
	Research on Patient Stratification
Lac-Phe	N-lactoyl-phenylalanine
LysoPC	Lysophosphatidylcholine
MOVE	Multi-omics variational autoencoders
MR	Mendelian randomisation
NA	Unidentified metabolite
NGR	Normal glucose regulation
PC	Phosphatidylcholine

# Introduction

Type 2 diabetes is a complex and common metabolic disorder, resulting from the body's ineffective use of insulin. It can be characterised by hyperglycaemia (high blood sugar) due to impaired insulin secretion and insulin resistance, with most affected people being overweight or obese [1]. Impaired glucose tolerance (IGT) and impaired fasting glucose, together known as impaired glucose regulation (IGR) or prediabetes, characterise an intermediate condition before converging towards diabetes. Recent studies show that a complex interplay of genetic susceptibility, environmental factors, lifestyle (including diet, physical activity, smoking and alcohol consumption), clinical heterogeneity, drugs and gut microbiome orchestrates the development of type 2 diabetes [2]. Over time, individuals with type 2 diabetes are more likely to have a higher risk for heart attacks, strokes [3], neuropathy (nerve damage), retinopathy (causing blindness) and kidney failure as well as several infectious diseases including COVID-19, reducing life quality and causing social burden [4, 5].

Metabolomics profiles involve a set of low-molecularweight biochemicals (metabolites) that includes sugars, amino acids, organic acids, nucleotides, lipids, xenobiotics and other compound classes. Identifying biochemical changes occurring between prediabetes and diabetes improves risk prediction for better-targeted prevention [6, 7]. In addition, genetic composition can be used to make predictions regarding disease susceptibility. Genome-wide association studies (GWAS) show that more than 400 loci influence the risk of type 2 diabetes [8] and that 900 genetic variants have been associated with BMI [9]. Therefore, linking metabolites with genetics gives access to genetics' influence on the metabolic compositions [10–13], providing comprehensive molecular understanding of the disease.

In the Innovative Medicines Initiative - Diabetes Research on Patient Stratification (IMI-DIRECT), we characterised 132 metabolites from targeted measurements and 779 metabolites from untargeted measurements profiled in 3000 individuals at baseline. The study population was stratified by following ADA 2011 glycaemic categories as follows: 23.89% (n=692) had normal glucose regulation (NGR) with fasting glucose 5.23 (SD=0.39) mmol/l; 48.91% (n=1418) had IGR with fasting glucose 5.90 (SD=0.51) mmol/l; and 27.2% (*n*=890) had type 2 diabetes with fasting glucose 7.15 (SD=1.39) mmol/l [14]. For the integration of non-omics data such as health status, lifestyle and medication with metabolomics, advanced statistical techniques were applied to analyse the data (see Methods). Beyond multivariate and association analyses we performed causal mediation analysis to evaluate potential causal roles of mediators on outcome [15, 16]. A study on drug-omics associations in type 2 diabetes [17] used an unsupervised deep learning framework of multi-omics variational autoencoders (MOVE) to extract significant drug response patterns from 789 individuals newly diagnosed with type 2 diabetes in the IMI-DIRECT cohort. We integrated the polypharmacy effect on metabolomics knowledge from MOVE and compared with our molecular findings in this study.

Our aims in this study were as follows: (1) to characterise 911 small molecular (132 targeted, 779 untargeted metabolomics analysis approach) features associated with prediabetes/ IGR and type 2 diabetes; (2) to identify baseline metabolites associated with progression rate estimated from cross-sectional data; (3) to investigate potential mediation effects of metabolites from baseline glycaemic status to follow-up using mediation analysis; and (4) to identify causal relationships between metabolites and type 2 diabetes using genetics drivers using two-sample Mendelian randomisation (2SMR) tests.

# Methods

# **DIRECT cohort**

The Diabetes Research on Patient Stratification (DIRECT) cohort encompasses 24,682 European participants at varying risk of glycaemic deterioration, identified and enrolled into a prospective cohort (study 1) of prediabetes (n=2235) and type 2 diabetes (n=830). Using ADA 2011 glycaemic categories in study 1, 33% (n=692) of cohort 1 (prediabetes risk) had NGR, 67% (n=1418) had IGR and 108 were excluded. In study 2, 789 samples were included and 41 samples were excluded. From study 1, 101 excluded samples entered study 2 (n=890). The ratio of self-reported sex varied in each study.

Detailed characteristics on inclusion and exclusion criteria, along with the protocol timeline for visits and tests for both studies, have been described elsewhere [14, 18]. In summary, venous blood fasting samples were obtained, followed by performance of DNA extractions and additional biochemical analyses. Metabolomics measurements for distinct samples at the baseline is considered in this study.

#### Targeted metabolomics (Absolute/DQ p150 Kit)

Blood samples in the study were analysed with the Absolute*IDQ* p150 Kit (BIOCRATES Life Sciences, Innsbruck, Austria) (see electronic supplementary material [ESM] Methods for details) [19]. After data export, lower and upper outliers were defined as samples with >33% of metabolite concentrations below 25% quantile ( $\pm$ 1.5 × IQR). Metabolite traits with too many zero-concentration samples and unidentified metabolites (NAs, >50%) were excluded (none). The CV was calculated in reference samples for each metabolite over all plates. Metabolite traits with CV>0.25 were excluded. After quality control, 132 metabolites were included in this study (ESM Table 1). Metabolite concentrations were log<sub>e</sub>-transformed and scaled (mean=0, SD=1) to ensure comparability between the metabolites.

#### Untargeted metabolomics (Metabolon platform)

Untargeted LC/MS-based techniques covers a broad spectrum of metabolites, in contrast to the targeted techniques wherein metabolites are limited to a predefined set of molecules. For details on sample preparation, measurement and identification of metabolites, see ESM Methods. Incomplete databases and the presence of unknown or novel metabolites have been reported with an asterisk (\*) against the metabolite name. The measured volume of the datasets contained 12% missing values. We screened for outlier remover (see ESM Fig. 1 for an example), which added 4% more missing values onto existing missing values (ESM Table 2). Peaks were quantified using AUC. For studies spanning multiple days, a data normalisation step was performed to correct variation resulting from instrument inter-day tuning differences. Essentially, each compound was corrected in run-day blocks by registering the medians to equal one and normalising each data point proportionately (termed the 'block correction'; ESM Fig. 2). Principal component analysis was performed on the metabolite dataset and checked for technical effects such as centre and sex (see ESM Fig. 3). The data missing pattern was tested using logistic regression considering missing as 0 and non-missing as 1; there was no significant association between missing and regressors indicating the missing-atrandom pattern. The K-nearest neighbour (KNN)-based

imputation method was applied using K=10 as suggested and optimised from German Cohort KORA F4 [20].

### Statistics

Multivariable logistic regression and linear regression Identifying metabolites specifically associated with the presence of IGR and type 2 diabetes, we ran the logistic regression with adjustment for age, sex, BMI and centre as the basic model, and adjusted additionally for alcohol consumption, smoking, BP, fasting HDL-cholesterol and fasting triacylglycerol as the full model. The concentration of each metabolite was log<sub>a</sub>-transformed and scaled to have a mean of zero and an SD of 1. Each metabolite was taken as exposure and a binary NGR-IGR, NGR-type 2 diabetes (NGR-T2D) or IGR-type 2 diabetes (IGR-T2D) variable as an outcome. The OR of outcomes was calculated using the  $\beta$  coefficient from logistic regression, where OR>1 indicates higher odds of outcome and OR<0 shows lower odds of outcome. To account for multiple testing, the *p* values from regression analyses were adjusted for multiple testing using the Bonferroni correction  $(p_{fdr}$  values). To stratify sex-dependent metabolites, men and women were separated to test the associations by performing the logistic regression full models.

For incidents of IGR and type 2 diabetes analysis, a binary NGR-IGR, NGR-T2D or IGR-T2D variable at follow-up times of 18 months and 48 months was taken as the outcome; transformed metabolites and the same risk factors in the full model were taken as exposure and covariates, respectively. The same *p* correction method was adopted.

The linear regression model was used to explore the association between  $HbA_{1c}$  progression rate and metabolites at the baseline.  $HbA_{1c}$  progression rate was computed with a conditional linear mixed effect model and adjusted for changes in BMI and diabetes medications [21]. Each transformed metabolite was taken as the independent variable and  $HbA_{1c}$  concentration as the dependent variable, with adjustment for age and sex. Bonferroni correction was performed for *p* correction.

Mediation analysis Mediation analysis followed the basic steps suggested by Baron and Kenny [22], and the significance of the mediation effect was tested with a non-parametric causal mediation analysis [22, 23]. Each identified metabolite was taken as a mediator, glycaemic category status at the baseline as the independent variable and glycaemic category at the follow-up (18 months and 48 months) as the dependent variable. R package 'mediation (4.5.0)' was used to calculate the *p* value and proportion of the mediation effect by bootstrapping with 1000 resamples.

**Mendelian randomisation** We used 2SMR approaches from the MRInstruments (0.3.2) and TwoSampleMR library

(v0.5.6) to check causal inference [24]. The 2SMR technique enables the establishment of a causal relationship between two observational studies (ESM Fig. 4), solely relying on summary statistics obtained from GWAS [24, 25]. To evaluate the influence of type 2 diabetes on metabolite levels, we conducted a 2SMR examination. Type 2 diabetes instruments were obtained from the genome-wide genotyping study [26] and the corresponding SNP estimates on metabolites were extracted from the metabolite-GWAS [10, 27]. Prior to performing Mendelian randomisation (MR) analysis, exposure and outcome data were harmonised by aligning the SNPs on the same effect allele. We employed the inverse-variance weighting [10, 26, 27] to estimate the causal effect.

# Results

#### **Study populations**

After stringent quality control (see ESM Methods), we identified 132 (ESM Table 1) and 779 (ESM Table 2) metabolites from targeted and untargeted metabolomics measurements, respectively, that were profiled for 3000 samples (ESM Table 3) [28]. Baseline characteristics (Table 1) revealed that there were significant differences in BMI, fasting variables and health status observed between NGR, IGR and type 2 diabetes groups. No significant differences in age and smoking status were observed between these three groups. In addition, the study was conducted across seven countries; type 2 diabetes participants were recruited in all centres while participants with NGR or IGR were only recruited in the Amsterdam, Copenhagen, Kuopio and Lund centres.

# Metabolites associated with prediabetes and diabetes from targeted metabolomics measurements

A multivariable logistic regression model was used with known diabetes-related variables as covariates to identify significant metabolites. Study centre, sex, age and BMI were covariates in the basic model while the additional variables systolic BP, fasting HDL-cholesterol, fasting triacylglycerol, smoking status, alcohol status and health status were added in the full model. Based on the full model, four metabolites differed significantly between the NGR and IGR groups (Fig. 1a). Of these, hexoses (H1) showed the strongest association (OR 1.81 [95% CI 1.59, 2.06],  $p_{fdr}$ =3.97×10<sup>-17</sup>) and served as a positive control throughout our analysis. Thirty-four and 50 metabolites differed significantly between NGR and IGR vs type 2 diabetes, respectively (Fig. 1b,c). As a general pattern, phosphatidylcholines (PCs) and lysophosphatidylcholine (lysoPC) were negatively associated with progression to type 2 diabetes, while branched-chain and aromatic amino acids as

 
 Table 1
 Baseline characteristics
 of the DIRECT participants based on their glycaemic category

Characteristic	NGR	IGR	T2D	p value
Sample size	692	1418	890	
Male sex	519 (75.0)	1074 (75.7)	525 (59.0)	< 0.001
Centre				< 0.001
Amsterdam	167 (24.1)	300 (21.2)	183 (20.6)	
Copenhagen	54 (7.8)	223 (15.7)	97 (10.9)	
Dundee	0	0	164 (18.4)	
Exeter	0	0	142 (16.0)	
Kuopio	407 (58.8)	820 (57.8)	34 (3.8)	
Lund	64 (9.2)	75 (5.3)	104 (11.7)	
Newcastle	0	0	166 (18.7)	
Age, years	62.15±6.43	62.08±6.19	61.99 <u>+</u> 7.96	0.894
BMI, kg/m <sup>2</sup>	27.15±3.65	28.33±4.06	30.59±4.92	< 0.001
Systolic BP, mmHg	128.48±15.21	131.62±15.20	132.02±15.78	< 0.001
Diastolic BP, mmHg	79.18 <u>+</u> 8.73	81.20±8.97	76.48 <u>+</u> 9.88	< 0.001
Fasting glucose, mmol/l	5.23±0.39	5.90±0.51	7.13±1.39	< 0.001
Fasting HDL-cholesterol, mmol/l	1.37±0.35	1.30±0.36	1.18±0.38	< 0.001
Fasting LDL-cholesterol, mmol/l	3.21±0.90	3.19 <u>+</u> 0.95	$2.43 \pm 1.00$	< 0.001
Fasting TG, mmol/l	$1.22 \pm 0.53$	1.44 <u>+</u> 0.66	1.56±0.88	< 0.001
Fasting cholesterol, mmol/l	5.14 <u>±</u> 0.97	$5.15 \pm 1.01$	$4.33 \pm 1.17$	< 0.001
Fasting HbA <sub>1c</sub> , mmol/mol	35.34±2.22	37.86±2.88	45.86±5.94	< 0.001
Fasting HbA <sub>1c</sub> , %	$5.38 \pm 0.20$	$5.61 \pm 0.26$	$6.35 \pm 0.54$	< 0.001
Fasting insulin, pmol/l	$50.84 \pm 30.90$	$72.42 \pm 50.22$	96.56±72.69	< 0.001
Smoking status				0.717
Current smoker	93 (13.4)	215 (15.2)	117 (13.2)	
Ex-smoker	326 (47.1)	681 (48.0)	445 (50.1)	
Never	272 (39.3)	520 (36.7)	326 (36.7)	
Not Known	1 (0.1)	2 (0.1)	1 (0.1)	
Alcohol consumption status				0.004
Never	96 (13.9)	166 (11.7)	140 (15.7)	
Occasionally	134 (19.4)	282 (19.9)	214 (24.1)	
Regularly	462 (66.8)	968 (68.3)	534 (60.1)	
Not known	0	2 (0.1)	1 (0.1)	
Health status				< 0.001
Poor	1 (0.1)	10 (0.7)	28 (3.1)	
Fair	49 (7.1)	74 (5.2)	34 (3.8)	
Good	331 (47.8)	744 (52.5)	428 (48.1)	
Very good	213 (30.8)	396 (27.9)	239 (26.9)	
Excellent	49 (7.1)	74 (5.2)	34 (3.8)	
Not known	4 (0.6)	11 (0.8)	19 (2.1)	

Quantitative variables are expressed as mean  $\pm$  SD; categorical variables are expressed as n (%)

The significant difference of population characteristics between the individuals with IGR/type 2 diabetes and the normal participants (NGR) was calculated. Test statistics for categorical variables were calculated via the  $\chi^2$  test and Student's *t* test for continuous variables

T2D, type 2 diabetes; TG, triacylglycerol

well as valeryl/glutaryl-related acylcarnitines were positively associated with type 2 diabetes.

H1 (OR 9.67 [95% CI 6.54, 14.32],  $p_{fdr}$ =1.13×10<sup>-27</sup>) also had the strongest associations in NGR-T2D while C5-M-DC (OR=5.31 [95% CI 4.16, 6.77],  $p_{fdr}$ =1.07×10<sup>-38</sup>) had the strongest association in IGR-T2D. Three metabolites (H1, lysoPC a C17:0, lysoPC a C18:0) were significantly different in all comparisons (NGR-IGR, NGR-T2D and IGR-T2D), suggesting their important roles in diabetes indication and severity. Detailed statistics for the basic model and full model are shown in ESM Tables 3-8. As there were many more male participants than female participants



and the metabolites as independent variables, adjusted for study centre, sex, age, BMI, BP, fasting HDL-cholesterol, fasting triacylglycerol, smoking status, alcohol status and health status. The *x*-axis shows OR (95% CI) and the *y*-axis shows each significant metabolite; metabolite classes are represented by different colours. SM, sphingomyelin

enrolled in the study, a sensitivity analysis stratified by sex was conducted, and is reported in ESM Results, ESM Tables 9–14 and ESM Fig. 5.

# Metabolites associated with prediabetes and diabetes from untargeted metabolomics measurements

Fifteen metabolites were significantly changed between NGR and IGR based on the logistic regression analyses in the full model (Fig. 2a). Fructosyl lysine had the highest statistically significant association with progression to IGR (OR 1.53 [95% CI 1.37, 1.71],  $p_{fdr}$ =8.64×10<sup>-12</sup>). Similarly, 99 and 108 metabolites differed significantly between NGR or IGR and type 2 diabetes, respectively (Fig. 2b,c). As a general pattern, lipids were negatively associated and amino acids were positively associated with progression to type 2 diabetes. 1-(1-Enyl-palmitoyl)-2-oleoyl-GPC (P-16:0\_18:1)\* (OR 0.23 [95% CI 0.17, 0.31],  $p_{fdr}$ =3.48×10<sup>-18</sup>) had the strongest association for the NGR-T2D comparison, while cysteine-S-sulphate (OR 3.25 [95% CI 2.55, 4.15],  $p_{fdr}$ =3.11×10<sup>-18</sup>) was significantly associated in the IGR-T2D comparison. Seven metabolites (fructosyl lysine, glutamate, 1-stearoyl-GPC (18:0), N-lactoylphenylalanine, N-lactoylvaline, picolinoyl glycine, mannonate) appeared significant in all comparison groups, suggesting their important roles as diabetes risk indicators. Detailed statistics are presented in ESM Tables 15-20. A sex-based sensitivity analysis of metabolomics data from the untargeted measurements is reported in ESM Results, ESM Table 21-26, ESM Fig. 6.

### Metabolites associated with HbA<sub>1c</sub> progression rate

HbA<sub>1c</sub> progression rate was computed with a conditional linear mixed effect model and adjusted for changes in BMI and diabetes medications [21]. In multivariable linear regression analysis, lysoPC a C17:0 (β -0.0535 [95% CI -0.08, -0.0269],  $p_{fdr}$ =0.0109), glycine (Gly) ( $\beta$ -0.0509 [95% CI -0.0782, -0.0236],  $p_{fdr}=0.0347$ ) and H1 ( $\beta$  0.0481 [95% CI 0.0218, 0.0745],  $p_{fdr}$ =0.0452) were significantly correlated with HbA1c progression rate and all were related to glycaemic-deterioration traits as well. In untargeted metabolomic profiling, 20 metabolites were significantly related to HbA<sub>1c</sub> progression rate, with pyruvate ( $\beta$  0.0877 [95% CI 0.0609, 0.114],  $p_{fdr}$ =1.28×10<sup>-7</sup>) showing the strongest association. Besides pyruvate, N-lactoylleucine, lactate, N-lactoylphenylalanine, X-15245, N-lactoylisoleucine, N-lactoylvaline, 1-(1-enyl-palmitoyl)-2-oleoyl-GPC (P-16:0/18:1)\*, cortolone glucuronide, X-24295, formiminoglutamate and N-lactoyltyrosine were also significantly associated with glycaemic categories.

Tables 2 and 3 show the metabolites with significant associations, while the complete results are reported in ESM Tables 27–28.

# Metabolite association with incident diabetes (IGR/ type 2 diabetes)

Several metabolites were identified to be significantly associated with HbA<sub>1c</sub> progression rate as well as glycaemic category: three targeted metabolites (lysoPC a C17:0; glycine, H1); and 12 untargeted metabolites (pyruvate, N-lactoylleucine, lactate, N-lactoylphenylalanine, X-15245, N-lactoylisoleucine, N-lactoylvaline, 1-[1-enyl-palmitoyl[-2-oleoyl-GPC\* [PC(P-16:0/18:1)], cortolone glucuronide, X-24295, formiminoglutamate, N-lactoyltyrosine). Next, we investigated their predictive value for IGR and type 2 diabetes by including baseline metabolite concentrations and incident IGT or type 2 diabetes in follow-up timelines in multivariable logistic regression. As shown in Table 4, lysoPC a C17:0 concentration at baseline was observed to significantly differ in 244 incident IGR individuals compared with 398 NGR control individuals after 18 months. The sum of H1 at baseline concentrations showed significant differences between incident IGR (at 48 month follow-up) and NGR or incident type 2 diabetes and IGR at both the 18 month and the 48 month follow-up.

In untargeted metabolomic profiling, lactate and X-24295 baseline concentrations were significantly correlated with IGR or type 2 diabetes incidence at the 18 month and 48 month follow-up (Table 5). Formiminoglutamate, *N*-lactoylleucine and *N*-lactoylvaline significantly differed in 244 incident IGT individuals compared with 398 NGT control individuals after 18 months. We did not find any significant metabolites from untargeted measurements to predict the incidence of IGR from NGR at 48 months.

#### **Mediation analysis**

Causal mediation analysis was employed to explore the potential mediation effects of the identified metabolites from baseline glycaemic status to follow-up. Consistent with incidence results, lysoPC a C17:0 showed strong significance (proportion of mediation by 13%, mediation effect p=0.034, Fig. 3a), indicating that this metabolite partially mediated the glycaemic deterioration from NGR to IGR at 18 months. The positive control H1 exhibited significant mediation effects in all groups (between 6% and 9%) as it is mainly represented by blood glucose.

*N*-Lactoylvaline (proportion of mediation 24%, mediation effect  $p < 2 \times 10^{-16}$ ), lactate (proportion of mediation 22%, mediation effect p=0.002), *N*-lactoylleucine (proportion of mediation 20%, mediation effect p=0.006), formimino-glutamate (proportion of mediation 11%, mediation effect



Fig. 2 Flag plots representing the results of the multivariable logistic regression models for NGR vs IGR (a), NGR vs type 2 diabetes (b) and IGR vs type 2 diabetes (c) as dependent variables and the metabolites as independent variables, adjusted for study centre, sex, age, BMI, BP, fasting HDL-cholesterol, fasting triacylglycerol, smoking status, alcohol status and health status. The x-axis shows OR (95% CI) and the y-axis shows each significant metabolite; metabolite classes are represented by different colours. Asterisks (\*) indicate the presence of unknown or novel metabolites

**Table 2** Metabolites from targeted measurements significantly associated with HbA<sub>1c</sub> progression rate from a linear regression model

	-	-	
Metabolite	β (95% CI)	p value	$p_{fdr}$ value
LysoPC a C17:0	-0.053 (-0.080, -0.027)	8.25×10 <sup>-5</sup>	0.011
Gly	-0.051 (-0.078, -0.024)	2.63×10 <sup>-4</sup>	0.0345
H1	0.048 (0.022, 0.075)	$3.42 \times 10^{-4}$	0.045

The dependent variable is the  $HbA_{1c}$  progression rate while the independent variable is the  $log_e$ -transformed and standardised baseline concentration of a given metabolite, adjusted by age and sex

The  $p_{fdr}$  values represent the adjusted p value for multiple testing by Bonferroni correction

p=0.034) and X-24295 (proportion of mediation 11%, mediation effect p=0.042) were all observed to show significant mediation effects from baseline NGR to IGR at 18 months' follow-up (Fig. 3b). Furthermore, formiminoglutamate (proportion of mediation 23%, mediation effect p=0.006) showed a significant mediation effect from NGR to IGR at 48 months. These results suggest that these metabolites own a significant mediation effect on glycaemic deterioration.

#### MR

The availability of genetic data on type 2 diabetes makes the use of MR particularly compelling. To assess bidirectional causal relationships between type 2 diabetes and metabolites (Fig. 4), we employed 2SMR tests. After multiple testing correction only the concentration of the sum of H1 was determined by type 2 diabetes (p < 0.05/117 = 0.00042). For untargeted metabolites we found instruments for only 19% of the metabolites (i.e. 151 out of 779). For example, instruments are from genes TCF7L2, IGF2BP2, NOTCH2, CDKAL1, PABPC4, FTO and JAZF1, known to be associated with diabetes and that have been further significantly associated with the metabolites. Following multiple testing correction, it suggests that the change in an amino acid (glutamate) and a lipid (caproate, FA C6:0) was caused by change in type 2 diabetes status (p < 0.05/151 = 0.000331). However, metabolites that are causal for type 2 diabetes (meaning that the change in metabolite caused change in the disease status) included several phosphatidylcholines, namely PC aa C36:2, PC aa C36:5, PC ae C36:3 and PC ae C34:3, from the targeted metabolomics dataset. From the untargeted metabolomics

Metabolite	β (95% CI)	p value	$p_{fdr}$ value
Pyruvate	0.087 (0.060, 0.114)	$1.65 \times 10^{-10}$	$1.28 \times 10^{-7}$
N-Lactoylleucine	0.082 (0.056, 0.109)	$8.43 \times 10^{-10}$	$6.57 \times 10^{-7}$
Lactate	0.075 (0.049, 0.102)	$3.30 \times 10^{-8}$	$2.57 \times 10^{-5}$
N-Lactoylphenylalanine	0.074 (0.048, 0.100)	$3.66 \times 10^{-8}$	$2.85 \times 10^{-5}$
X-15245	0.074 (0.047, 0.100)	$6.24 \times 10^{-8}$	$4.86 \times 10^{-5}$
N-Lactoylisoleucine	0.068 (0.042, 0.095)	$3.11 \times 10^{-7}$	$2.42 \times 10^{-4}$
N-Lactoylvaline	0.067 (0.041, 0.094)	$5.69 \times 10^{-7}$	$4.43 \times 10^{-4}$
X-11444	0.068 (0.041, 0.094)	$6.22 \times 10^{-7}$	$4.84 \times 10^{-4}$
Orotidine	0.065 (0.038, 0.091)	$1.74 \times 10^{-6}$	$1.35 \times 10^{-3}$
Metabolonic lactone sulphate	0.063 (0.036, 0.089)	$2.9 \times 10^{-6}$	$2.28 \times 10^{-3}$
3,4-Dihydroxybutyrate	0.060 (0.033, 0.087)	$1.11 \times 10^{-5}$	$8.64 \times 10^{-3}$
N4-Acetylcytidine	0.059 (0.033, 0.085)	$1.16 \times 10^{-5}$	$9.06 \times 10^{-3}$
X-24337	0.058 (0.032, 0.085)	$1.47 \times 10^{-5}$	0.011
1-(1-Enyl-palmitoyl)-2-oleoyl-GPC(P-16:0/18:1)*	-0.058 (-0.084, -0.032)	$1.49 \times 10^{-5}$	0.016
X-25828	-0.058 (-0.085, -0.032)	$1.50 \times 10^{-5}$	0.017
Cortolone glucuronide	0.058 (0.032, 0.085)	$1.73 \times 10^{-5}$	0.013
X-24295	0.057 (0.031, 0.084)	$1.77 \times 10^{-5}$	0.014
Formiminoglutamate	0.059 (0.032, 0.088)	$2.75 \times 10^{-5}$	0.021
1-Palmitoyl-2-oleoyl-GPE (16:0/18:1)	0.056 (0.029, 0.082)	$3.59 \times 10^{-5}$	0.028
N-Lactoyltyrosine	0.055 (0.029, 0.082)	$3.98 \times 10^{-5}$	0.031

The dependent variable is the HbA<sub>1c</sub> progression rate while the independent variable is the  $\log_e$ -transformed and standardised baseline concentration of a given metabolite, adjusted by age and sex. The  $p_{fdr}$  are adjusted p for multiple testing by Bonferroni correction

Table 3Metabolites fromuntargeted metabolomicsmeasurements significantlyassociated with HbA1cprogression rate from a linearregression model

 Table 4
 Metabolites from targeted measurements that were significantly associated with incidence of IGR and type 2 diabetes in different pairwise comparisons

Comparison	OR (95% CI)	p value
18 months		
398 NGR vs 244 IGR		
lysoPC a C17:0	-0.246 (-0.452, -0.043)	0.018
897 IGR vs 71 T2D		
H1	0.545 (0.164, 0.945)	0.006
48 months		
244 NGR vs 295 IGR		
H1	0.433 (0.189, 0.690)	$7x10^{-3}$
821 IGR vs 128 T2D		
H1	0.347 (0.064, 0.642)	0.018

Baseline metabolites were taken as the independent variables with glycaemic category in different timelines (18 months and 48 months) as the dependent variables, adjusted by study centre, sex, age, BMI, BP, fasting HDL-cholesterol, fasting triacylglycerol, smoking status, alcohol status and health status

ORs and p values were calculated from the logistic regression model T2D, type 2 diabetes

 Table 5
 Metabolites from untargeted measurements that were significantly associated with incidence of IGR and type 2 diabetes in different pairwise comparisons

Comparison	OR (95% CI)	p value
18 months		
398 NGR vs 244 IGR		
Formiminoglutamate	0.369 (0.157, 0.588)	$7.7 \times 10^{-4}$
Lactate	0.373 (0.143, 0.557)	0.002
N-Lactoylleucine	0.294 (0.079, 0.514)	0.008
N-Lactoylvaline	0.248 (0.039, 0.460)	0.021
X-24295	0.225 (0.022, 0.432)	0.031
897 IGR vs 71 T2D		
X-24295	0.474 (0.162, 0.801)	$3.6 \times 10^{-3}$
Lactate	0.409 (0.077, 0.747)	$1.6 \times 10^{-2}$
48 months		
821 IGR vs 128 T2D		
X-24295	0.474 (0.162, 0.801)	$3.6 \times 10^{-3}$
Lactate	0.409 (0.077, 0.747)	$1.6 \times 10^{-2}$

Baseline metabolites were taken as the independent variables with glycaemic category in different timelines (18 months and 48 months) as the dependent variables, adjusted by study centre, sex, age, BMI, BP, fasting HDL-cholesterol, fasting triacylglycerol, smoking status, alcohol status and health status

ORs and *p* values were calculated from the logistic regression model T2D, type 2 diabetes

dataset, two *n*-3 fatty acids, namely stearidonate (18:4n3) and docosapentaenoate (n3 DPA; 22:5n3), were identified to be causal for type 2 diabetes. Detailed statistics of our MR analysis are presented in ESM Tables 29–32.

# Discussion

In this study, we used untargeted metabolomics to provide semi-quantitative global screening of metabolites in the development of a disease whereas targeted metabolomics was used to quantify a pre-selected subset of metabolites with absolute concentrations. However, the overlap between the two metabolomic techniques was limited to a few amino acids and lipids. In the current study we report 19 metabolites (three from targeted and 14 from global profiling, plus one common lysoPC a C18:0 / 1-stearoyl-GPC [18:0]) that were significantly associated with prediabetes in the DIRECT cohort. The advantages of global profiling become evident as it allows for the identification of a broader spectrum of metabolites. Few notable examples are given here. First, picolinoylglycine (HMDB0059766), which is potentially a phase II product of picolinic acid, a degradation product of tryptophan [29] and glycine [30], and shows potential as a novel marker for glycaemic deterioration. Prediabetes is often associated with dyslipidaemia, marked by an imbalanced lipid profile compared with individuals with NGR [24]. Second, N-lactoyl amino acids are not infrequently observed in metabolomic datasets. In fact it has come to light that N-lactoyl amino acids were misidentified in some metabolomic studies and were erroneously reported as 1-carboxyethyl amino acids. In particular, N-lactoyl-phenylalanine (Lac-Phe) is known to act as an appetite suppressant when given to obese mice [31]. However, in humans Lac-Phe concentrations were observed to rise following vigorous exercise [32]. In fact, the most recent study shows that Lac-Phe facilitates the impact of metformin on both food intake and body weight [33, 34]. It seems that the exact role of Lac-Phe in the human body and pathways downstream, such as energy metabolism, insulin signalling, exercise-induced pathways, are unclear and needs further research.

We are aware of several limitations to our study. Although metabolomics screening showcases numerous valuable attributes in health science, challenges inherent to this approach continue to exist, especially in the accurate identification of metabolites which is crucial for the biological interpretation and validation of metabolomics



**Fig.3** Schematic overview of mediation analysis with lysoPC a C17:0 and hexoses (**a**) or *N*-lactoylvaline, lactate, *N*-lactoylleucine, formiminoglutamate and X-24295 (**b**) as mediators. Numbers above

the red arrows indicate the percentage and significance of mediation effects. T2D, type 2 diabetes

data [35]. Variability in sample collection, preparation and analytical techniques can impact the reproducibility and comparability of results across different studies. Standardisation efforts are ongoing but may not fully address all sources of variation. The identification of metabolites, especially in untargeted metabolomics, can be challenging. Incomplete databases and the presence of unknown or novel metabolites have been reported with a metabolite name with an asterisk (\*) sign. However, ongoing advancements in technology, methodology and standardisation efforts aim to enhance the robustness and applicability of metabolomics studies [35]. The current study is predominantly based on White male participants from the Kuopio region of Europe, and for this reason an additional sex-based sensitivity analysis has been performed and reported separately (ESM Results 1 and 2). Challenges in MR studies include limited statistical power, potential reverse causation, confounding and pleiotropy [36]. Caution is advised in interpreting causality inference,

considering the various limitations mentioned in the methods, and precautionary measures were taken by using valid MR instruments and reporting Bonferroni significance.

A drug-metabolomics associations study [17] was examined to determine whether or not metabolites linked to type 2 diabetes from the DIRECT study were also associated with a particular drug. Looking at our results and those of Allesøe et al [17], we found that 44% (15 out of 34) of targeted metabolites and 3% (three out of 99) of non-targeted metabolites that were significantly associated with type 2 diabetes also showed a significant association with at least one of the 20 drugs. This suggests that metabolites linked to type 2 diabetes may be confounded by polypharmacy.

However, metabolite association with incident prediabetes or diabetes (IGR-T2D) showed that lysoPC a C17:0 could predict the risk of developing IGR at 18 months and 48 months. It has already been shown that lysoPCs differ significantly between individuals with incident IGT or type 2 diabetes and individuals with NGR in the KORA study



Fig. 4 Forest plot representing causal estimates of type 2 diabetes on targeted and untargeted metabolites in the two-sample MR test. T2D, type 2 diabetes

[37]. LysoPC a C17:0 was negatively associated with diabetes, a finding that was confirmed in several studies [38, 39]. The aforementioned drug–metabolomics association study [17] showed that lysoPC a 17:0 was not associated with the drugs. However, the origin of odd-chain fatty acids (mainly C15:0 and C17:0) remains elusive. Jenkins et al [40] investigated the origin of circulating odd-chain fatty acids (C17:0, C15:0) through a combination of animal and human studies to determine possible contributions of fatty acids from the gut-microbiota, diet and novel endogenous biosynthesis [41]. The findings suggested that C15:0 was linked to dietary intake, while C17:0 was predominantly biosynthesised, indicating independent origins and non-homologous roles in disease causation.

Causal mediation analysis indicated that plasma lactate strongly mediates the effects of identified metabolites in the transition from baseline glycaemic status to follow-up [42]. In a longitudinal study of Swedish men, elevated serum lactate was independently linked to a higher incidence of type 2 diabetes, irrespective of obesity measures [43]. Formiminoglutamate was confirmed to be associated with a higher risk of incident type 2 diabetes in older Puerto Ricans [44]. N-lactoylleucine and N-lactoylvaline, derivatives of leucine and valine, respectively, are ubiquitous pseudodipeptides of lactic acid and amino acids that are formed by reverse proteolysis [32] and are correlated with underivatised amino acids in human plasma. The Microbiome and Insulin Longitudinal Evaluation Study (MILES) [45] investigated the association between ABO haplotypes and insulin-related characteristics, and explored possible pathways that could mediate these

associations. The study showed that the A1 haplotype potentially enhances favourable insulin sensitivity in non-Hispanic White individuals, with lactate likely influencing this mechanism, while gut bacteria are not believed to be a contributing factor.

In MR, causality signifies that modifying exposure leads to a predictable change in the outcome. Our 2SMR analysis suggests that the metabolites causal for type 2 diabetes are PC aa C36:2, PC aa C36:5, PC ae C34:3 and PC ae C36:3 and all these metabolites are significantly associated with drug-metabolomics. However, from untargeted metabolomics two n-3 fatty acids, namely stearidonate (18:4n3) and docosapentaenoate DPA 22:5n3), are not further associated with drugs. In 2012, Banz et al [46] explored the therapeutic implications of stearidonate acid in preventing or managing type 2 diabetes. The Fatty Acids and Outcomes Research Consortium (FORCE) [47] found that higher circulating biomarkers of seafood-derived n-3 fatty acids were associated with lower type 2 diabetes risk. On the contrary, branchedchain amino acids [48] and sphingomyelin [15] have been shown to have a causal role in type 2 diabetes development, a correlation not observed in the DIRECT study.

# Conclusions

Our study demonstrates that alteration in blood plasma metabolites is associated with glycaemic deterioration. The progression from prediabetes to diabetes is mediated by novel metabolites such as picolinoylglycine and *N*-lactoyl-amino acids, as demonstrated by evidence from the DIRECT study. *N*-lactoyl-amino acids are known to be exercise-induced metabolites that suppress food intake and influence glucose homeostasis. Additional functional research and quantification are needed to advance the identification of early metabolic biomarkers such as *N*-lactoyl-amino acids, which have the potential to forecast the onset of type 2 diabetes. Collectively, these findings direct attention towards novel metabolic signatures associated with glycaemic deterioration.

Supplementary Information The online version contains peer-reviewed but unedited supplementary material available at https://doi.org/10. 1007/s00125-024-06282-6.

Acknowledgements We extend our gratitude to the IMI-DIRECT study participants who willingly participated in phenotyping as well as to the clinical and technical staff across European study centres for their contributions to participant recruitment and clinical assessment. This publication's development has been supported by the Innovative Medicines Initiative Joint Undertaking under grant agreement 115317 (DIRECT), with resources derived from the European Union's Seventh Framework Programme (FP7/2007–2013) and EFPIA companies' in-kind contributions. Special thanks go to the study centre team, L. M. 't Hart, F. Rutters, J. Vangipurapu and T. H. Hansen, for providing internal review on this manuscript.

Data availability Access to the molecular and clinical raw data, as well as the processed data, is restricted. This is in accordance with the informed consent provided by study participants, the various national ethical approvals obtained for the study, and compliance with the European General Data Protection Regulation (GDPR). Individual-level clinical and molecular data cannot be transferred from the centralised IMI-DIRECT repository. Requests for access will receive guidance on accessing data through the DIRECT secure analysis platform after submitting an appropriate application. The IMI-DIRECT data access policy and additional information about the IMI-DIRECT research consortium's initiatives and activities can be found at https://directdiabetes.org.

Code used for MR in the study is included as ESM.

**Funding** Open Access funding enabled and organized by Projekt DEAL. We would like to thank Helmholtz Munich, German Diabetes Center (DZD) for their support in current research and China Research Council (CRC) funding for a PhD student hosted by Helmholtz Munich.

Authors' relationships and activities The authors declare that there are no relationships or activities that might bias, or be perceived to bias, their work.

**Contribution statement** SS, QD, MH, JA and HG conceptualised the analysis plan. RWK, PWF, MW, IF, GG, IP, HR, MD, MM, OP, JS, KT, FDM, SB, AV, AM, TM, TK, JA, EP and HG were involved in conception and design of the DIRECT study. SS, QD, MH, JA, GK, AA, CP, RB, JFT, AJ and AT were involved in the data acquisition, pre-processing and interpretation of data. SS organised inclusion of outlined sections and, along with QD, wrote the original draft of the manuscript. All authors contributed to drafting the article or critically revising it for significant intellectual content and have provided approval to the final version to be published. SS and HG are the guarantors of this work and, as such, had full access to all the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

# References

- Zimmet PZ (2017) Diabetes and its drivers: the largest epidemic in human history? Clin Diabetes Endocrinol 3:1. https://doi.org/ 10.1186/s40842-016-0039-3
- Wesolowska-Andersen A, Brorsson CA, Bizzotto R et al (2022) Four groups of type 2 diabetes contribute to the etiological and clinical heterogeneity in newly diagnosed individuals: an IMI DIRECT study. Cell Rep Med 3(1):100477. https://doi.org/10. 1016/j.xcrm.2021.100477
- Emerging Risk Factors Collaboration, Sarwar N, Gao P et al (2010) Diabetes mellitus, fasting blood glucose concentration, and risk of vascular disease: a collaborative meta-analysis of 102 prospective studies. Lancet 375(9733):2215–2222. https:// doi.org/10.1016/S0140-6736(10)60484-9
- Emerging Risk Factors Collaboration, Di Angelantonio E, Kaptoge S et al (2015) Association of cardiometabolic multimorbidity with mortality. JAMA 314(1):52–60. https://doi.org/10. 1001/jama.2015.7008
- Rao KondapallySeshasai S, Kaptoge S, Thompson A et al (2011) Diabetes mellitus, fasting glucose, and risk of causespecific death. N Engl J Med 364(9):829–841. https://doi.org/ 10.1056/NEJMoa1008862
- Wurtz P, Kangas AJ, Soininen P, Lawlor DA, Davey Smith G, Ala-Korpela M (2017) Quantitative serum nuclear magnetic resonance metabolomics in large-scale epidemiology: a primer on -Omic technologies. Am J Epidemiol 186(9):1084–1096. https://doi.org/10.1093/aje/kwx016
- Guasch-Ferre M, Hruby A, Toledo E et al (2016) Metabolomics in prediabetes and diabetes: a systematic review and meta-analysis. Diabetes Care 39(5):833–846. https://doi.org/10.2337/dc15-2251
- Cai L, Wheeler E, Kerrison ND et al (2020) Genome-wide association analysis of type 2 diabetes in the EPIC-InterAct study. Sci Data 7(1):393. https://doi.org/10.1038/s41597-020-00716-7
- Yengo L, Sidorenko J, Kemper KE et al (2018) Meta-analysis of genome-wide association studies for height and body mass index in approximately 700000 individuals of European ancestry. Hum Mol Genet 27(20):3641–3649. https://doi.org/10.1093/hmg/ddy271
- Shin SY, Fauman EB, Petersen AK et al (2014) An atlas of genetic influences on human blood metabolites. Nat Genet 46(6):543–550. https://doi.org/10.1038/ng.2982
- Gieger C, Geistlinger L, Altmaier E et al (2008) Genetics meets metabolomics: a genome-wide association study of metabolite profiles in human serum. PLoS Genet 4(11):e1000282. https:// doi.org/10.1371/journal.pgen.1000282
- Lanznaster D, Veyrat-Durebex C, Vourc'h P, Andres CR, Blasco H, Corcia P (2020) Metabolomics: a tool to understand the impact

of genetic mutations in amyotrophic lateral sclerosis. Genes (Basel) 11(5):537. https://doi.org/10.3390/genes11050537

- Suhre K, Raffler J, Kastenmuller G (2016) Biochemical insights from population studies with genetics and metabolomics. Arch Biochem Biophys 589:168–176. https://doi.org/10.1016/j.abb.2015.09.023
- Koivula RW, Forgie IM, Kurbasic A et al (2019) Discovery of biomarkers for glycaemic deterioration before and after the onset of type 2 diabetes: descriptive characteristics of the epidemiological studies within the IMI DIRECT Consortium. Diabetologia 62(9):1601–1615. https://doi.org/10.1007/s00125-019-4906-1
- Zhang Z, Zheng C, Kim C, Van Poucke S, Lin S, Lan P (2016) Causal mediation analysis in the context of clinical research. Ann Transl Med 4(21):425. https://doi.org/10.21037/atm.2016.11.11
- Dong Q, Sidra S, Gieger C et al (2023) Metabolic signatures elucidate the effect of body mass index on type 2 diabetes. Metabolites 13(2):227. https://doi.org/10.3390/metabo13020227
- Allesøe RL, Lundgaard AT, Hernandez Medina R et al (2023) Discovery of drug-omics associations in type 2 diabetes with generative deep-learning models. Nat Biotechnol 41(3):399–408. https:// doi.org/10.1038/s41587-022-01520-x
- Koivula RW, Heggie A, Barnett A et al (2014) Discovery of biomarkers for glycaemic deterioration before and after the onset of type 2 diabetes: rationale and design of the epidemiological studies within the IMI DIRECT Consortium. Diabetologia 57(6):1132–1142. https://doi.org/10.1007/s00125-014-3216-x
- Römisch-Margl W, Prehn C, Bogumil R, Röhring C, Suhre K, Adamski J (2012) Procedure for tissue sample preparation and metabolite extraction for high-throughput targeted metabolomics. Metabolomics. 8:133–142. https://doi.org/10.1007/s11306-011-0293-4
- Do KT, Wahl S, Raffler J et al (2018) Characterization of missing values in untargeted MS-based metabolomics data and evaluation of missing data handling strategies. Metabolomics 14(10):128. https://doi.org/10.1007/s11306-018-1420-2
- Bizzotto R, Jennison C, Jones AG et al (2021) Processes underlying glycemic deterioration in type 2 diabetes: an IMI DIRECT study. Diabetes Care 44(2):511–518. https://doi.org/10.2337/dc20-1567
- Baron RM, Kenny DA (1986) The moderator-mediator variable distinction in social psychological research: conceptual, strategic, and statistical considerations. J Pers Soc Psychol 51(6):1173– 1182. https://doi.org/10.1037/0022-3514.51.6.1173
- Tingley D, Yamamoto T, Hirose K, Keele L, Imai K (2014) mediation: R package for causal mediation analysis. J Stat Soft 59(5):1–38. https://doi.org/10.18637/jss.v059.i05
- 24. Hemani G, Zheng J, Elsworth B et al (2018) The MR-Base platform supports systematic causal inference across the human phenome. Elife 7. https://doi.org/10.7554/eLife.34408
- Zaghlool SB, Sharma S, Molnar M et al (2021) Revealing the role of the human blood plasma proteome in obesity using genetic drivers. Nat Commun 12(1):1279. https://doi.org/10.1038/ s41467-021-21542-4
- 26. Xue A, Wu Y, Zhu Z et al (2018) Genome-wide association analyses identify 143 risk variants and putative regulatory mechanisms for type 2 diabetes. Nat Commun 9(1):2941. https://doi.org/10. 1038/s41467-018-04951-w
- Draisma HHM, Pool R, Kobl M et al (2015) Genome-wide association study identifies novel genetic variants contributing to variation in blood metabolite levels. Nat Commun 6:7208. https://doi. org/10.1038/ncomms8208
- Brown AA, Fernandez-Tajes JJ, Hong MG et al (2023) Genetic analysis of blood molecular phenotypes reveals common properties in the regulatory networks affecting complex traits. Nat Commun 14(1):5062. https://doi.org/10.1038/s41467-023-40569-3
- 29. Hu Q, Jin L, Zeng J et al (2020) Tryptophan metabolite-regulated Treg responses contribute to attenuation of airway inflammation during specific immunotherapy in a mouse asthma model. Hum

Vaccin Immunother 16(8):1891–1899. https://doi.org/10.1080/ 21645515.2019.1698900

- 30. Choi JY, Kim SH, Kim JE et al (2019) Four amino acids as serum biomarkers for anti-asthma effects in the ovalbumininduced asthma mouse model treated with extract of Asparagus cochinchinensis. Lab Anim Res 35:32. https://doi.org/10.1186/ s42826-019-0033-x
- Li VL, He Y, Contrepois K et al (2022) An exercise-inducible metabolite that suppresses feeding and obesity. Nature 606(7915):785–790. https://doi.org/10.1038/s41586-022-04828-5
- 32. Jansen RS, Addie R, Merkx R et al (2015) N-lactoyl-amino acids are ubiquitous metabolites that originate from CNDP2-mediated reverse proteolysis of lactate and amino acids. Proc Natl Acad Sci U S A 112(21):6601–6606. https://doi.org/10.1073/pnas.1424638112
- Xiao S, Li VL, Lyu X et al (2024) Lac-Phe mediates the effects of metformin on food intake and body weight. Nat Metab 6(4):659– 669. https://doi.org/10.1038/s42255-024-00999-9
- Scott B, Day EA, O'Brien KL et al (2024) Metformin and feeding increase levels of the appetite-suppressing metabolite Lac-Phe in humans. Nat Metab 6(4):651–658. https://doi.org/10.1038/ s42255-024-01018-7
- 35. Kirwan JA, Gika H, Beger RD et al (2022) Quality assurance and quality control reporting in untargeted metabolic phenotyping: mQACC recommendations for analytical quality management. Metabolomics 18(9):70. https://doi.org/10.1007/s11306-022-01926-3
- Davey Smith G, Hemani G (2014) Mendelian randomization: genetic anchors for causal inference in epidemiological studies. Hum Mol Genet 23(R1):R89-98. https://doi.org/10.1093/hmg/ddu328
- 37. Wang-Sattler R, Yu Z, Herder C et al (2012) Novel biomarkers for pre-diabetes identified by metabolomics. Mol Syst Biol 8:615. https://doi.org/10.1038/msb.2012.43
- Floegel A, Stefan N, Yu Z et al (2013) Identification of serum metabolites associated with risk of type 2 diabetes using a targeted metabolomic approach. Diabetes 62(2):639–648. https://doi.org/ 10.2337/db12-0495
- Lee H-S, Xu T, Lee Y et al (2016) Identification of putative biomarkers for type 2 diabetes using metabolomics in the Korea Association REsource (KARE) cohort. Metabolomics 12(12):178. https://doi.org/10.1007/s11306-016-1103-9
- Jenkins BJ, Seyssel K, Chiu S et al (2017) Odd chain fatty acids; new insights of the relationship between the gut microbiota, dietary intake, biosynthesis and glucose intolerance. Sci Rep 7:44845. https://doi.org/10.1038/srep44845
- Weitkunat K, Schumann S, Nickel D et al (2017) Odd-chain fatty acids as a biomarker for dietary fiber intake: a novel pathway for endogenous production from propionate. Am J Clin Nutr 105(6):1544–1551. https://doi.org/10.3945/ajcn.117.152702
- Crawford SO, Hoogeveen RC, Brancati FL et al (2010) Association of blood lactate with type 2 diabetes: the Atherosclerosis Risk in Communities Carotid MRI Study. Int J Epidemiol 39(6):1647– 1655. https://doi.org/10.1093/ije/dyq126
- 43. Ohlson LO, Larsson B, Bjorntorp P et al (1988) Risk factors for type 2 (non-insulin-dependent) diabetes mellitus. Thirteen and one-half years of follow-up of the participants in a study of Swedish men born in 1913. Diabetologia 31(11):798–805. https://doi. org/10.1007/BF00277480
- Rivas-Tumanyan S, Pacheco LS, Haslam DE et al (2022) Novel plasma metabolomic markers associated with diabetes progression in older puerto ricans. Metabolites 12(6):513. https://doi.org/10. 3390/metabo12060513
- 45. Li-Gao R, Grubbs K, Bertoni AG et al (2022) The roles of gut microbiome and plasma metabolites in the associations between ABO blood groups and insulin homeostasis: the Microbiome and Insulin Longitudinal Evaluation Study (MILES). Metabolites 12(9):787. https://doi.org/10.3390/metabo12090787

- Banz WJ, Davis JE, Clough RW, Cheatwood JL (2012) Stearidonic acid: is there a role in the prevention and management of type 2 diabetes mellitus? J Nutr 142(3):635S-640S. https://doi. org/10.3945/jn.111.146829
- 47. Qian F, Ardisson Korat AV, Imamura F et al (2021) n-3 fatty acid biomarkers and incident type 2 diabetes: an individual participantlevel pooling project of 20 prospective cohort studies. Diabetes Care 44(5):1133–1142. https://doi.org/10.2337/dc20-2426

# **Authors and Affiliations**

 Lotta LA, Scott RA, Sharp SJ et al (2016) Genetic predisposition to an impaired metabolism of the branched-chain amino acids and risk of type 2 diabetes: a mendelian randomisation analysis. PLoS Med 13(11):e1002179. https://doi.org/10.1371/journal.pmed.1002179

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Sapna Sharma<sup>1</sup> · Qiuling Dong<sup>1,2</sup> · Mark Haid<sup>3</sup> · Jonathan Adam<sup>1,4</sup> · Roberto Bizzotto<sup>5</sup> · Juan J. Fernandez-Tajes<sup>6</sup> · Angus G. Jones<sup>7</sup> · Andrea Tura<sup>5</sup> · Anna Artati<sup>3</sup> · Cornelia Prehn<sup>3</sup> · Gabi Kastenmüller<sup>8</sup> · Robert W. Koivula<sup>9</sup> · Paul W. Franks<sup>10</sup> · Mark Walker<sup>11</sup> · Ian M. Forgie<sup>12</sup> · Giuseppe Giordano<sup>13</sup> · Imre Pavo<sup>14</sup> · Hartmut Ruetten<sup>15</sup> · Manolis Dermitzakis<sup>16,17,18</sup> · Mark I. McCarthy<sup>6</sup> · Oluf Pedersen<sup>19,20</sup> · Jochen M. Schwenk<sup>21</sup> · Konstantinos D. Tsirigos<sup>22</sup> · Federico De Masi<sup>22</sup> · Soren Brunak<sup>20,23</sup> · Ana Viñuela<sup>24</sup> · Andrea Mari<sup>5</sup> · Timothy J. McDonald<sup>25</sup> · Tarja Kokkola<sup>26</sup> · Jerzy Adamski<sup>27,28,29</sup> · Ewan R. Pearson<sup>12</sup> · Harald Grallert<sup>1,4</sup>

- Sapna Sharma sapna.sharma@helmholtz-munich.de
- Harald Grallert harald.grallert@helmholtz-munich.de
- <sup>1</sup> Research Unit of Molecular Epidemiology, Institute of Epidemiology, German Research Center for Environmental Health, Helmholtz Zentrum München, Neuherberg, Germany
- <sup>2</sup> Faculty of Medicine, Ludwig-Maximilians-University München, Munich, Germany
- <sup>3</sup> Metabolomics and Proteomics Core, German Research Center for Environmental Health, Helmholtz Zentrum München, Neuherberg, Germany
- <sup>4</sup> German Center for Diabetes Research (DZD), München Neuherberg, Germany
- <sup>5</sup> Institute of Neuroscience, National Research Council, Padova, Italy
- <sup>6</sup> Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford, UK
- <sup>7</sup> Department of Clinical and Biomedical Sciences, University of Exeter College of Medicine & Health, Exeter, UK
- <sup>8</sup> Institute of Computational Biology, Helmholtz Zentrum München, Munich, Germany
- <sup>9</sup> Oxford Centre for Diabetes, Endocrinology and Metabolism, University of Oxford, Oxford, UK
- <sup>10</sup> Department of Clinical Science, Genetic and Molecular Epidemiology, Lund University Diabetes Centre, Malmö, Sweden
- <sup>11</sup> Translational and Clinical Research Institute, Faculty of Medical Sciences, University of Newcastle, Newcastle upon Tyne, UK
- <sup>12</sup> Population Health and Genomics, Ninewells Hospital and Medical School, University of Dundee, Dundee, UK
- <sup>13</sup> Department of Clinical Science, Genetic and Molecular Epidemiology, Lund University Diabetes Centre, Malmö, Sweden

- <sup>14</sup> Eli Lilly Regional Operations GmbH, Vienna, Austria
- <sup>15</sup> Sanofi Partnering, Sanofi-Aventis Deutschland GmbH, Frankfurt am Main, Germany
- <sup>16</sup> Department of Genetic Medicine and Development, University of Geneva Medical School, Geneva, Switzerland
- <sup>17</sup> Institute for Genetics and Genomics in Geneva (iGE3), University of Geneva, Geneva, Switzerland
- <sup>18</sup> Swiss Institute of Bioinformatics, Geneva, Switzerland
- <sup>19</sup> Center for Clinical Metabolic Research, Herlev and Gentofte University Hospital, Copenhagen, Denmark
- <sup>20</sup> Novo Nordisk Foundation Center for Basic Metabolic Research, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark
- <sup>21</sup> Science for Life Laboratory, School of Biotechnology, KTH
   Royal Institute of Technology, Solna, Sweden
- <sup>22</sup> Department of Health Technology, Technical University of Denmark, Kongens Lyngby, Denmark
- <sup>23</sup> Department of Health Technology, Technical University of Denmark, Kongens Lyngby, Denmark
- <sup>24</sup> Biosciences Institute, Faculty of Medical Sciences, University of Newcastle, Newcastle upon Tyne, UK
- <sup>25</sup> Blood Sciences, Royal Devon and Exeter NHS Foundation Trust, Exeter, UK
- <sup>26</sup> Internal Medicine, Institute of Clinical Medicine, University of Eastern Finland, Kuopio, Finland
- <sup>27</sup> Department of Biochemistry, Yong Loo Lin School of Medicine, National University of Singapore, Singapore, Singapore
- <sup>28</sup> Institute of Experimental Genetics, German Research Center for Environmental Health, Helmholtz Zentrum München, Neuherberg, Germany
- <sup>29</sup> Institute of Biochemistry, Faculty of Medicine, University of Ljubljana, Ljubljana, Slovenia