



Supporting Information

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Genetic Manipulation of Mammalian Cells in Microphysiological Hydrogels

*Anna C. Jäkel, Dong-Jiunn Jeffery Truong and Friedrich C. Simmel**

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Supporting Information

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Doxycycline Titration

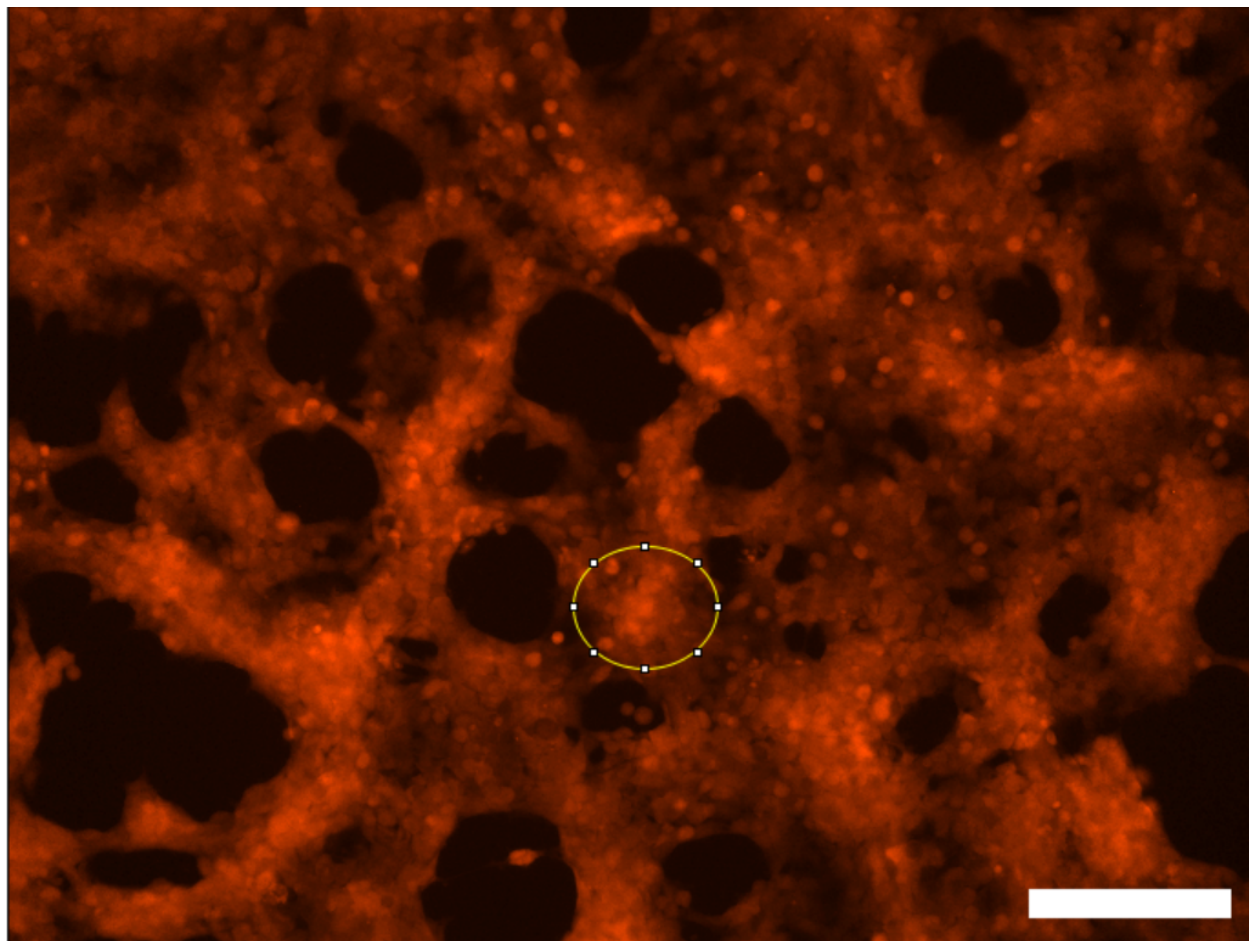
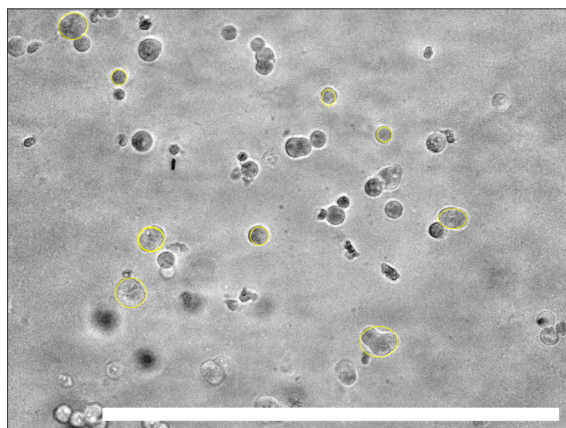
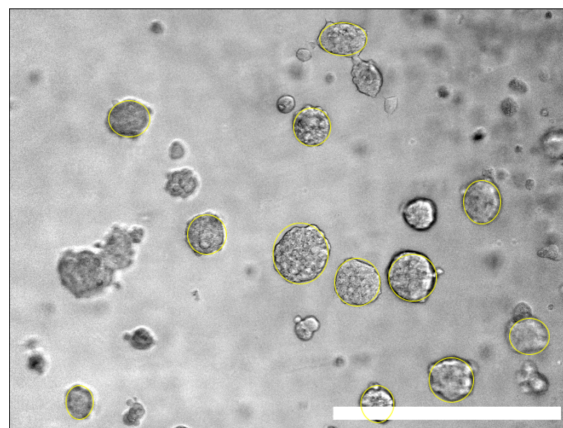


Figure S1: A region of interest (ROI) was selected for each sample that was 100% confluent. The following parameters were measured for over a period of 60 h: area, mean intensity, standard deviation, min and max intensity. Measurements were taken every 2 h for the first 24 h, then after 24 h, and then after 12 h. Scalebar: 100 μm

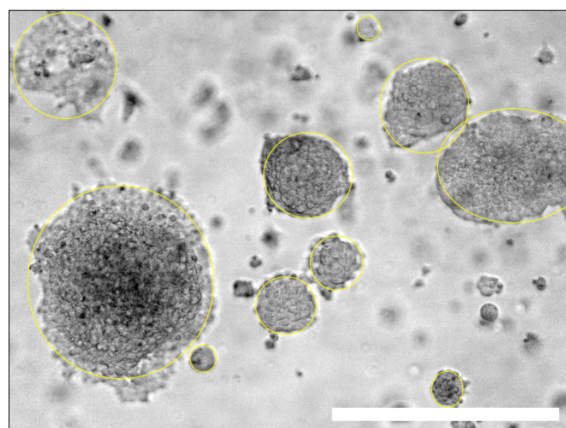
Cell growth in GelMa



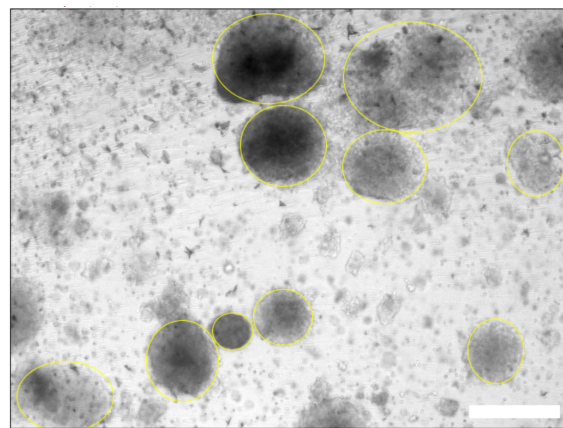
(a) Day 1 after seeding cells in GelMa.



(b) Day 5 after seeding cells in GelMa.

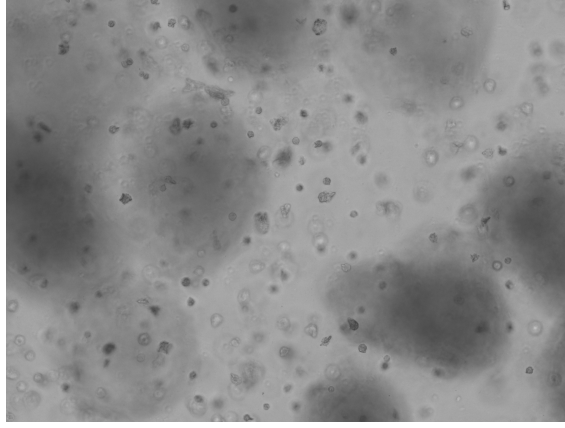


(c) Day 8 after seeding cells in GelMa.

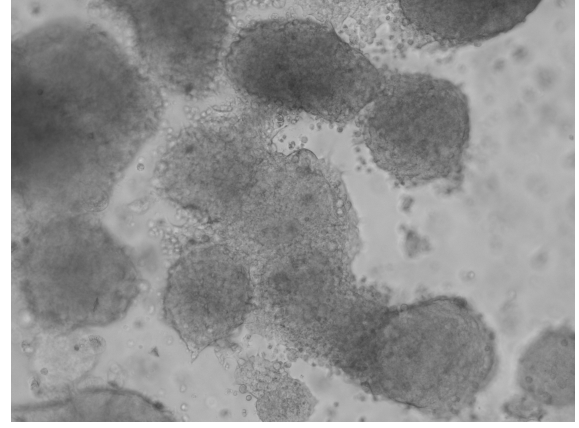


(d) Day 16 after seeding cells in GelMa.

Figure S2: For each sample 10 clusters were selected to measure cluster size. The feret radius was determined with Fiji and the mean was calculated for each sample. Scalebar: 500 μm

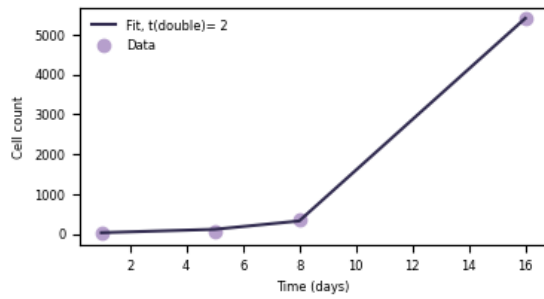


(a)

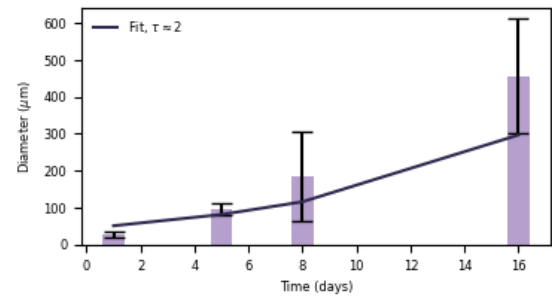


(b)

Figure S3: Cells were cultured for 8 days in $\approx 500\mu\text{m}$ thick GelMA. Shown are two images focusing on a lower and higher layer within the gel. a) The cells in lower layers (more than $200\mu\text{m}$ away from the surface) were not sufficiently supplied with DMEM and did not grow further. b) The cells in the upper layers, down to $\approx 200\mu\text{m}$ depths, did grow into large cell clusters.



(a) Number of cells calculated in one cell cluster over time.



(b) Cell cluster diameter over time. Fitting the data results in a doubling time of ≈ 2 days.

Figure S4: The cell count N_c was calculated assuming a spherical volume of each cluster. And the doubling time was calculated using the formula $N_c(t) = N_0 \times 2^{t/t_D}$.

Transfection

Transfection of mCherry - FACS

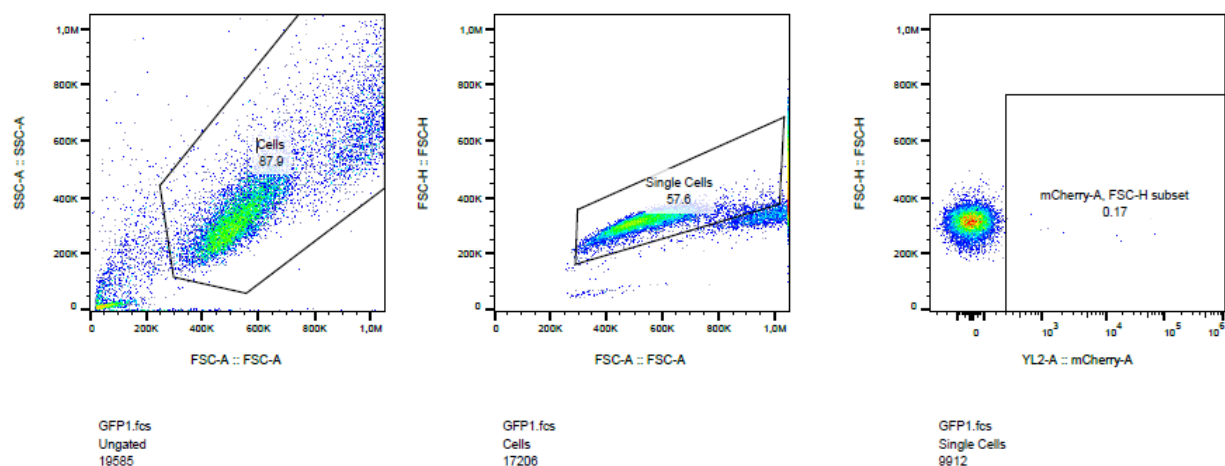


Figure S5: Control measurement of HEK293T cells.

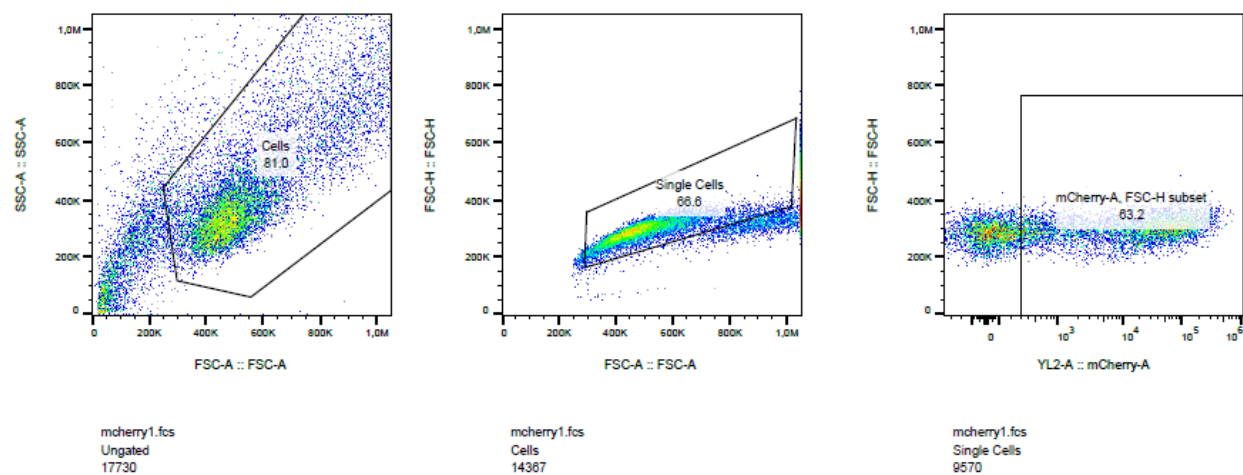


Figure S6: HEK293T were transfected using jetOptimus transfection reagent with plasmid DNA encoding for the fluorescent protein mCherry.

Rheometer Measurements

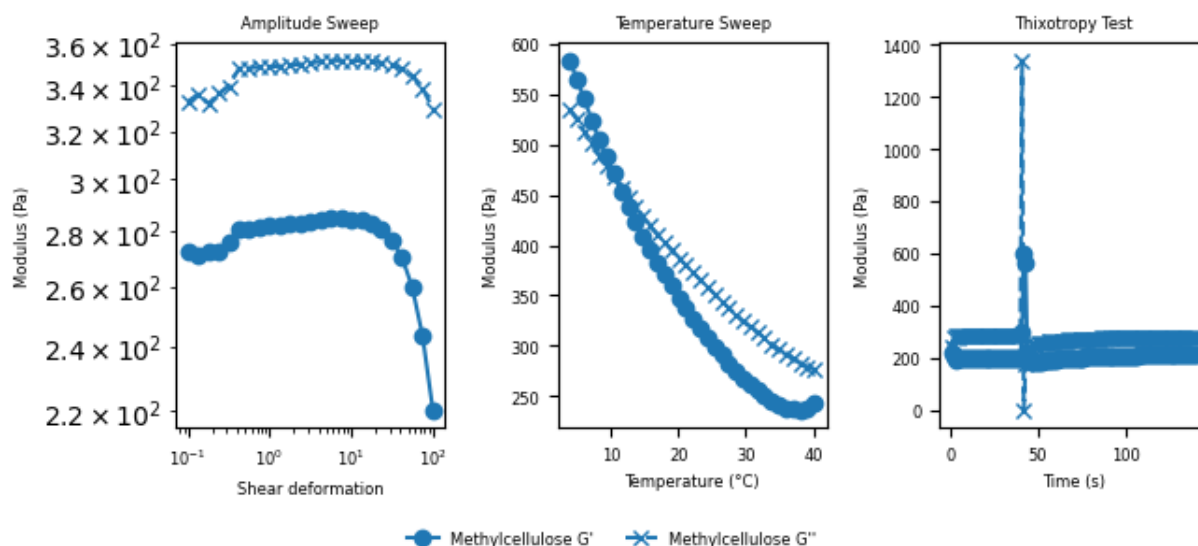


Figure S7: Methylcellulose (5 % v/w) was measured as benchmark. The amplitude sweep provides insight into the viscoelastic properties of methylcellulose by evaluating its storage modulus (G') and loss modulus (G'') across increasing strain amplitudes. In the linear viscoelastic region (LVR), G'' remains higher than G' , indicating that the material can be described as a viscoelastic liquid. As strain increases beyond the critical strain, G' decreases, and the limit of the LVE-region is reached. This suggests that methylcellulose maintains its gel-like properties under low deformation but undergoes yielding at higher strains. The temperature-dependent rheological behavior of methylcellulose is characterized by a decrease in G' at higher temperatures, indicative of thermogelling properties. Initially, at lower temperatures, the material behaves more elastic with $G' > G''$. As temperature rises, the viscous proportion predominates, reflected in a crossover point where G' surpasses G'' . This transition temperature is a critical parameter for applications requiring thermal responsiveness. The thixotropy test assesses the recovery of methylcellulose after shear-induced breakdown. Upon applying high shear, both G' and viscosity drop significantly, reflecting structural disruption. When shear is reduced, a gradual recovery of G' is observed, indicating partial structural reformation. However, if full recovery is not achieved within the measured timeframe, this suggests that methylcellulose exhibits a degree of irreversible structural breakdown or slow rebuilding dynamics. This behavior is relevant for applications where shear-induced fluidization and recovery kinetics are important.

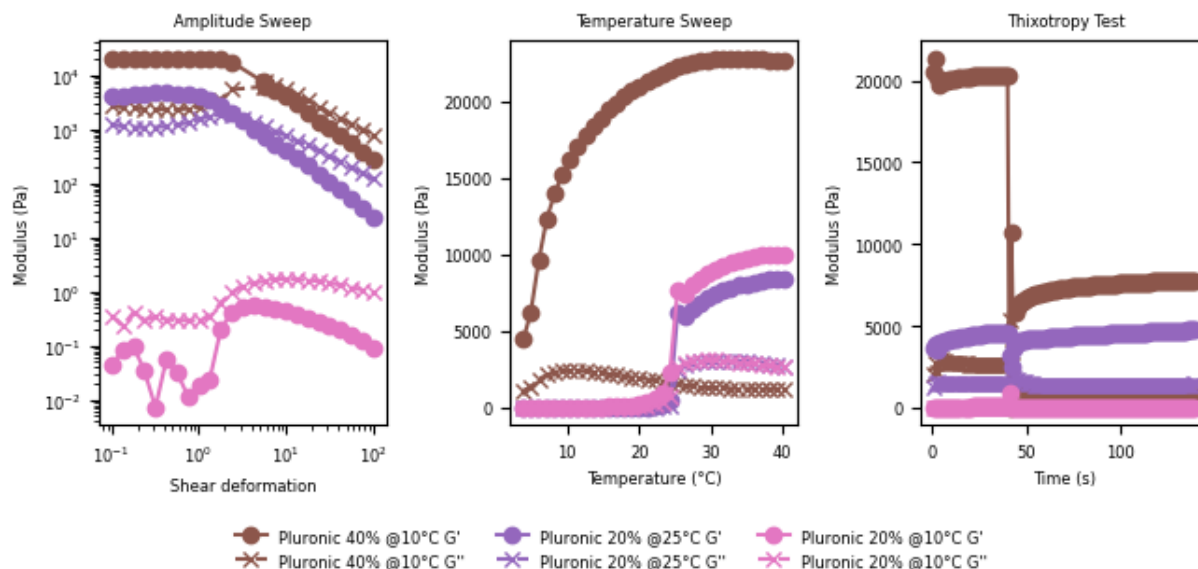


Figure S8: Pluronic is printable at 20 % and 40 % at room temperature for ~ 15 min when stored before at -4°C . An amplitude sweep of pluronic hydrogels at different concentrations and temperatures are shown. Compared to methylcellulose (Figure S7), the pluronic curves exhibit a less steep decline in the storage modulus (G') and loss modulus (G'') with increasing strain, indicating a more gradual structural breakdown. Additionally, pluronic 20 % at 10°C shows significantly lower G' values compared to pluronic 20 % at room temperature or pluronic 40 %, suggesting a temperature-dependent shift in mechanical strength. This behavior reflects the thermoresponsive nature of pluronic, where gel stiffness varies with both concentration and temperature. Interestingly pluronic with a concentration of 40 % at 10°C behaves similar as pluronic with a concentration of 20 % at 25°C . The temperature sweep of pluronic hydrogels at different concentrations shows that in contrast to methylcellulose, which shows a more linear increase in modulus with temperature, pluronic exhibits a non-monotonic "hill-shaped" curve. The storage modulus (G') initially increases with rising temperature, reaches a peak, and then declines at higher temperatures. This trend suggests a gelation process followed by structural weakening, likely due to micellar rearrangement or phase separation. Additionally, the difference between storage (G') and loss modulus (G'') is more pronounced compared to methylcellulose, highlighting distinct viscoelastic behavior and phase transitions in pluronic hydrogels. The thixotropy test shows that pluronic at 40 % does not recover its structure and viscosity as the 20 % pluronic does.

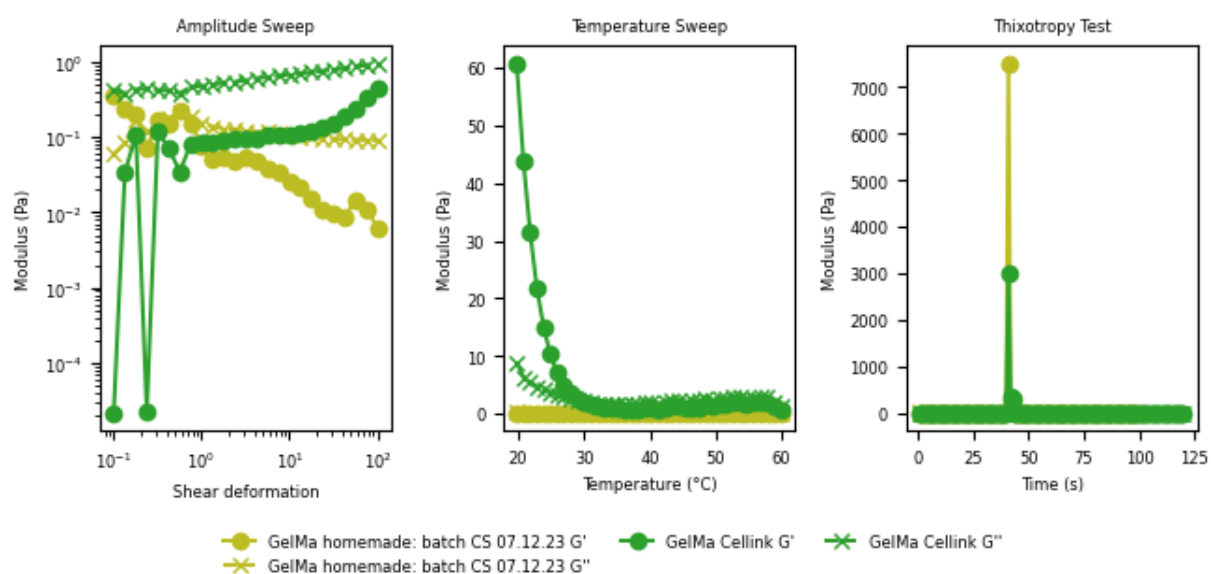


Figure S9: Rheological analysis of GelMa purchased from Cellink and our homemade GelMa. Amplitude-, temperature sweep, and thixotropy test. The temperature sweep shows similar behaviour of the hydrogels above 30 $^{\circ}\text{C}$. The thixotropy test shows that both hydrogels recover its structure and viscosity after deformation.

Cell viability in different gels

Cell line test

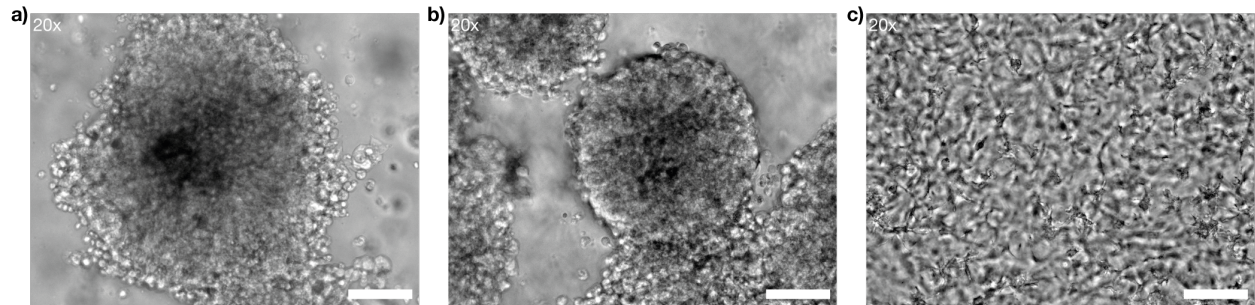


Figure S10: Different celltypes grown in GelMa for 16 days: a) HEK293T (ATCC, CRL-3216™) b) NIH-3T3 (ATCC, CRL-1658™) c) hMSC (The cells were a kind gift from the Clausen-Schaumann lab). Scalebar: 100 μ m.

Compatibility with glue

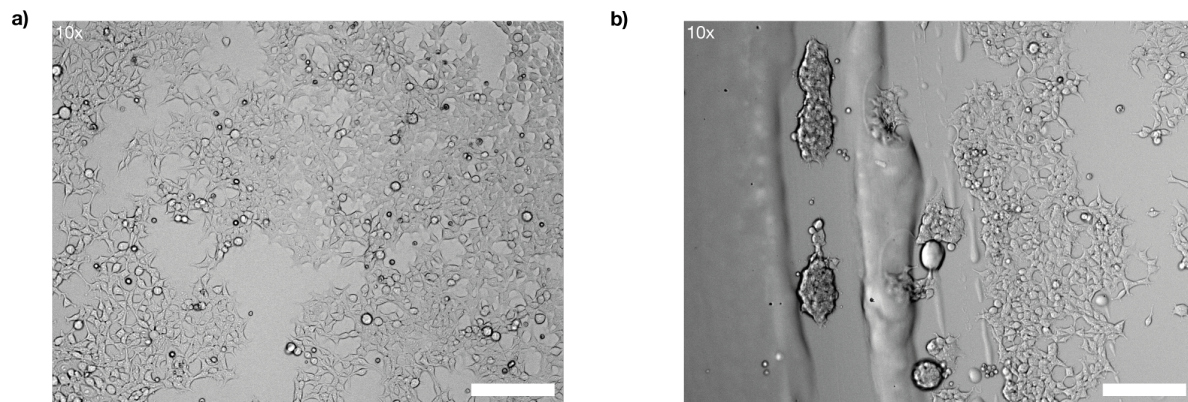


Figure S11: a) Cell growth in a well plate next to a glue drop. b) Cell growth inside the gel drop. Cells do grow in clusters similar to cells growing in GelMa. Scalebar: 200 μ m.

Test of nozzle precision

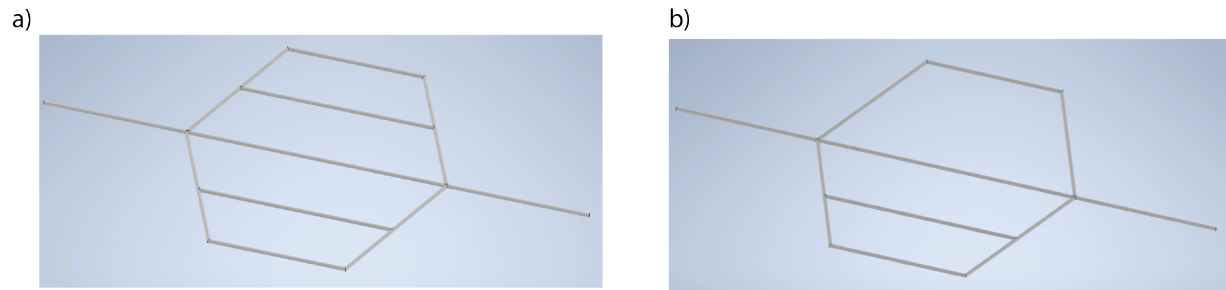


Figure S12: Printed vascular structures. a) Even distribution network. b) Uneven distribution network.

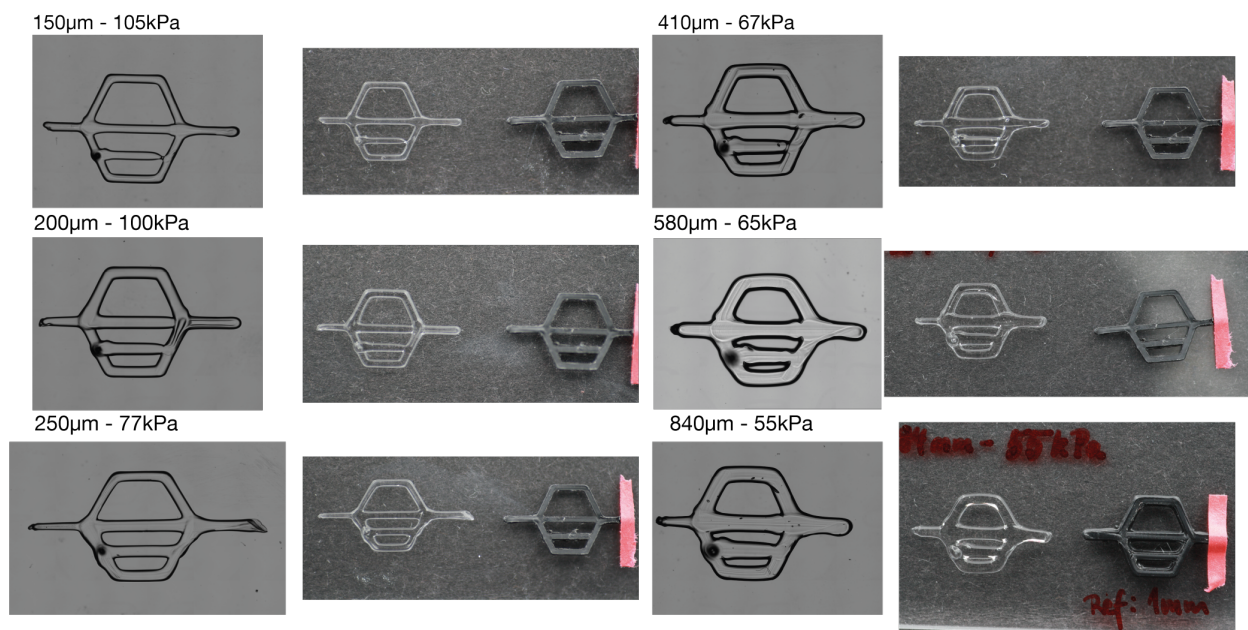


Figure S13: Channel structures printed with a BIOX2 printer (Cellink). Different needle sizes were tested and images are presented of the structure for each needle with the optimal pressure value. Right: microscope image, left: photograph of the structure next to a PLA printed construct which was printed with a Bambulab X1E printer. The channel thickness was 1 mm to compare resolution.

Fluorescein Channel Experiments

To fabricate vasculature-like structures, a 40 % (w/v) Pluronic F-127 solution was prepared by dissolving pluronic powder (Sigma-Aldrich) in double-distilled water (ddH₂O) at 4 °C overnight, with periodic vortexing to ensure complete dissolution. This temperature-responsive bioink served as a sacrificial support material, facilitating the creation of vascular channels within the GelMa matrix.

A BIOX 3D bioprinter (Cellink) equipped with a 200 µm nozzle was used to print the vascular structures. The pluronic solution, stored at 4 °C, was printed at room temperature, maintaining its optimal viscosity (cf. Figure S8). To prevent premature gelation, printing was performed within 15 minutes of removing the solution from cold storage. If the solution warmed beyond a critical threshold, re-cooling was necessary before continuing the process. Printing parameters were optimized, including a pressure of 100 kPa, a printing speed of 5 mm s⁻¹, and a preflow adjustment of -50 ms to ensure controlled extrusion.

The bioprinting workflow for generating vascular-like channels is illustrated in Figure S14. First, Pluronic F-127 (40 % w/v) was printed as a sacrificial ink to define the channel structure within GelMa. Figure S14a presents a photograph of a pluronic-printed channel stained in red, to highlight the channel structures within the hydrogel. The complete bioprinting process (Figure S14b) involved printing pluronic structures, overlaying the construct with GelMa, and crosslinking the hydrogel using 405 nm UV light for 30 seconds. Following crosslinking, the constructs were cooled to 4 °C for 5 minutes, liquefying the pluronic and enabling its removal, thereby creating perfusable channels.

To validate the functionality of these channels, Figure S14c shows a brightfield image of a pluronic structure before GelMa casting, demonstrating the precision and integrity of the printed channels. Additionally, Figure S14d presents fluorescence imaging of fluorescein diffusion from the channels into the hydrogel, confirming molecular transport into the surrounding matrix. This approach establishes a controlled environment for studying molecular diffusion and cellular responses within engineered hydrogel constructs.

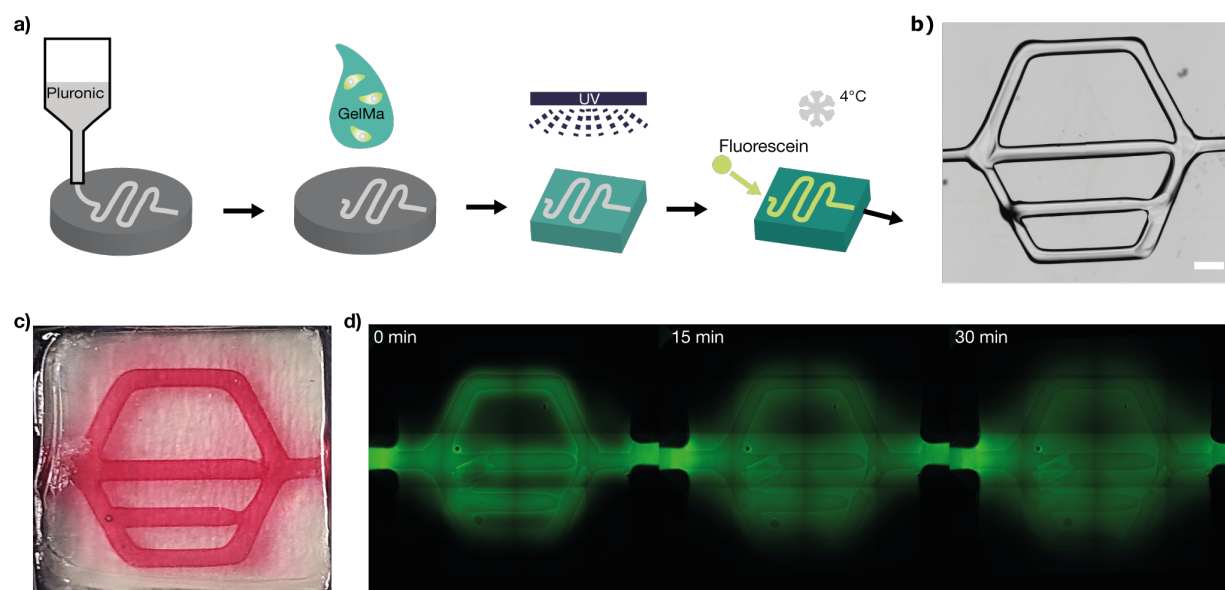


Figure S14: Construction of vascular-like structures. a) Schematic representation of the method for preparing vascular structures using a commercial 3D bioprinter. Pluronic F-127 is printed, followed by casting GelMa with or without cells on top. The GelMa is crosslinked using 405 nm UV light for 30 seconds. Subsequently, the construct is cooled to 4 °C for 5 minutes, liquefying the pluronic to enable channel perfusion with a material of interest. b) Brightfield image of the printed channel before hydrogel application. Scale bar: 1000 m. c) Photograph of a constructed channel filled with red food dye to visualize perfusion within the GelMa matrix. d) Channels perfused with fluorescein to observe diffusion into the surrounding hydrogel over time. e) Diffusion of fluorescein from a single channel. Scale bar: 1000 m. f) Quantitative analysis of fluorescein diffusion.

Bioreactor Setup

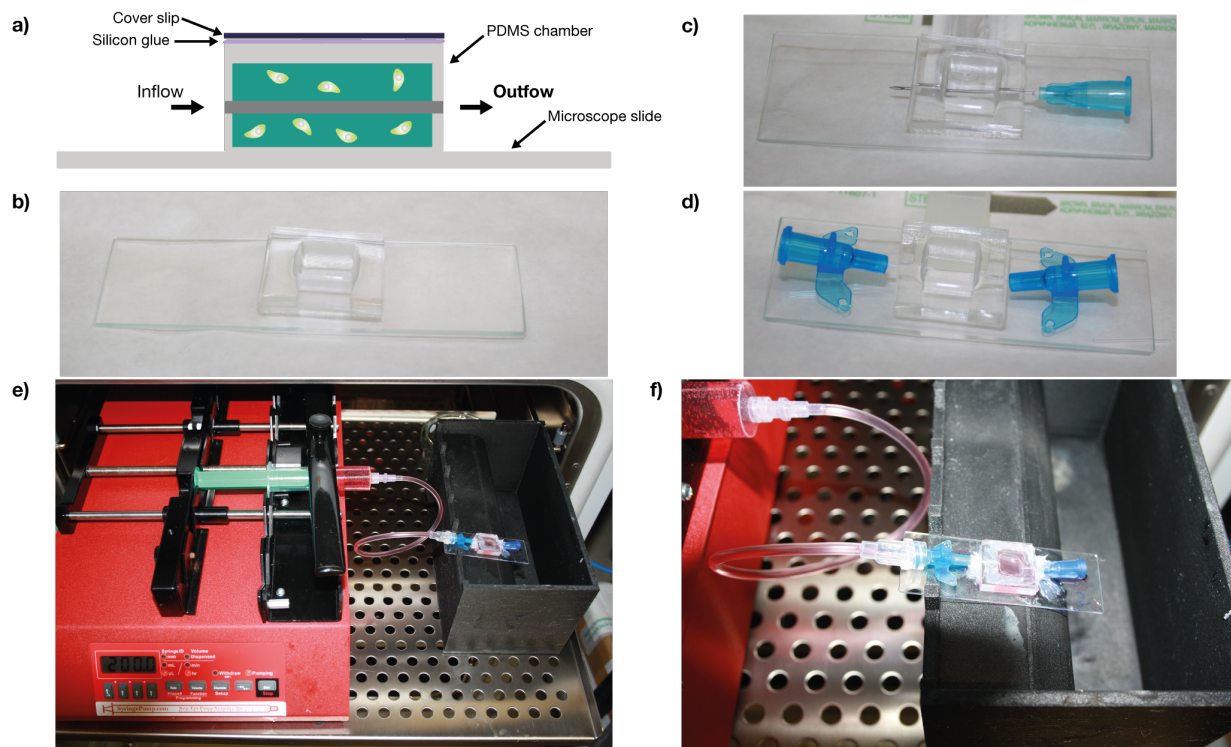


Figure S15: a) Schematic of the bioreactor. b) PDMA chamber bonded to a microscope slide with O₂ plasma. c) A needle (ID 0.6 mm) was punched through PDMS for casting gel on top for the creation of a channel. d) In- and outlet connection were achieved with catheters (22 G). e) Photograph of the whole setup. A syringe pump is used to be able to run 6 bioreactors in parallel. A 3D printed autoclavable PAHT chamber is used as collection tank for material flushed through the bioreactors. f) Zoom in on the bioreactor itself.

Plasmids

The full sequence of the plasmid used for transfection studies of mCherry is provided below

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CS3_mCh_3xSV40_AnnaLiedlMA
4488 bp

Key Features:

- CMV enhancer** (grey arc, top right)
- CMV promoter** (grey arc, top right)
- J23119 promoter** (grey arc, top right)
- mCherry** (red arc, top right)
- SV40 DTS** (blue arcs, bottom right)
- SV40 poly(A)** (blue arc, bottom right)
- SV40 signal** (blue arc, bottom right)
- SV40 copy** (blue arc, bottom right)
- ori** (yellow arc, top left)
- AmpR** (green arc, bottom left)
- IRES** (green arc, bottom left)
- Part of M13 ori** (green arc, bottom left)
- SV40 promoter** (grey arc, bottom left)
- SV40 DTS** (blue arcs, bottom left)
- SV40 poly(A)** (blue arc, bottom left)
- SV40 signal** (blue arc, bottom left)
- SV40 copy** (blue arc, bottom left)

Restriction Enzyme Sites (Clockwise from top):

- BciVI
- HincII
- SpeI
- CspCI
- Eco53kI
- SacI
- BsaI
- PmeI, +3
- AleI
- BstEII
- AhdI
- StuI
- +1
- +2
- +4
- AlwNI
- Bpu10I
- KasI, +4
- AleI
- NmeAIII, +6
- PmeI
- BclI
- BbsI
- EagI
- NotI, PaeR7I, PspXI, TliI, XhoI
- BclI
- Bpu10I
- CspCI
- SspI
- BspHI
- BciVI
- SspI
- AgI
- AclI
- AvaII
- +1
- BsaI
- BsrDI
- AhdI
- BspHI
- AlwNI

S16

The full sequence of the plasmid that was used to create the stable cell line HEK293T that expresses mScarlet-I upon doxycycline induction is provided below

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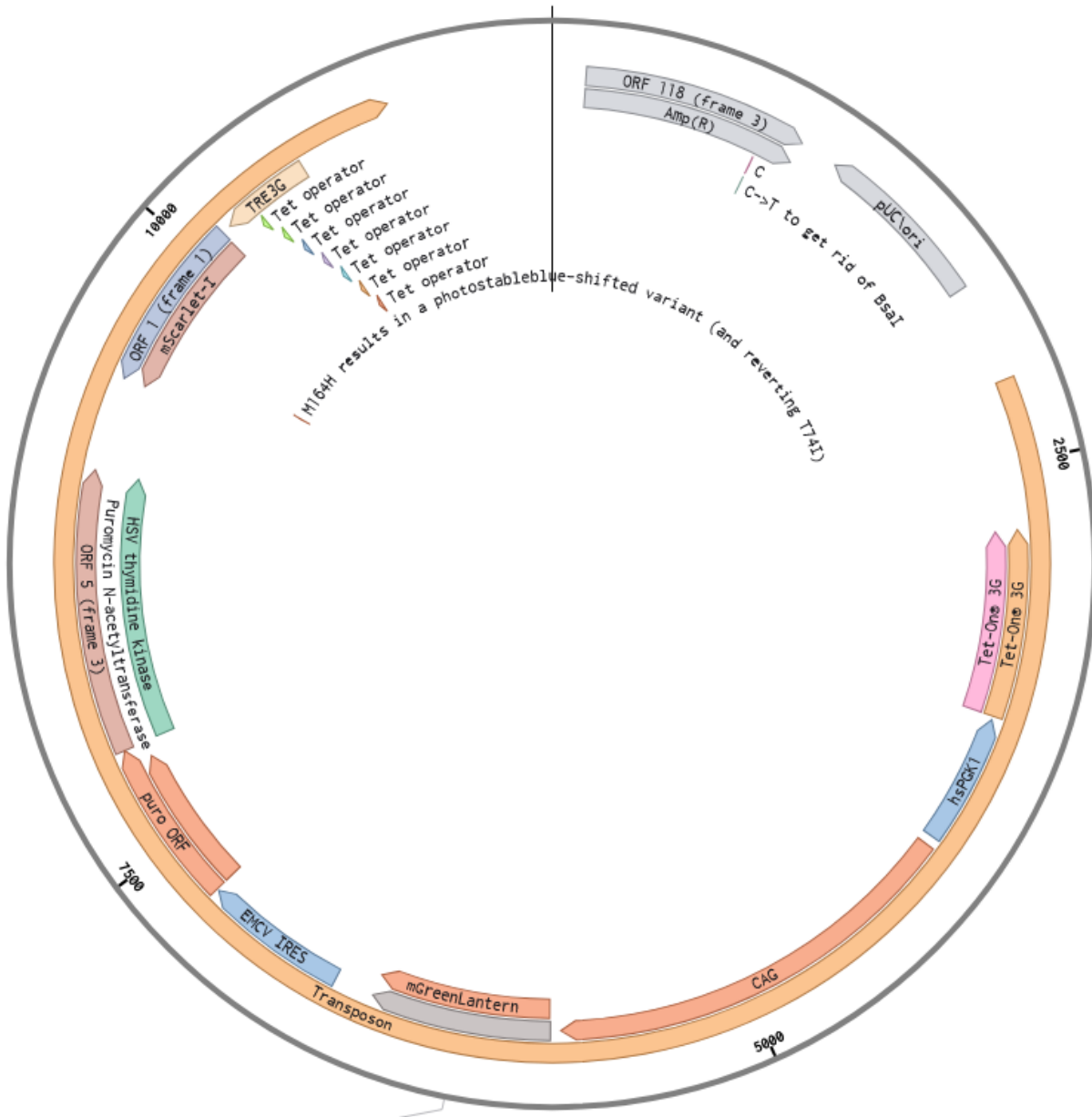


Figure S17: Transposon used for engineering the stable cell line HEK293T which expresses mScarlet-I upon doxycycline addition.

The full sequence of the plasmid encoding the transposase is provided below

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The full sequence of the plasmid encoding the pegRNA is provided below

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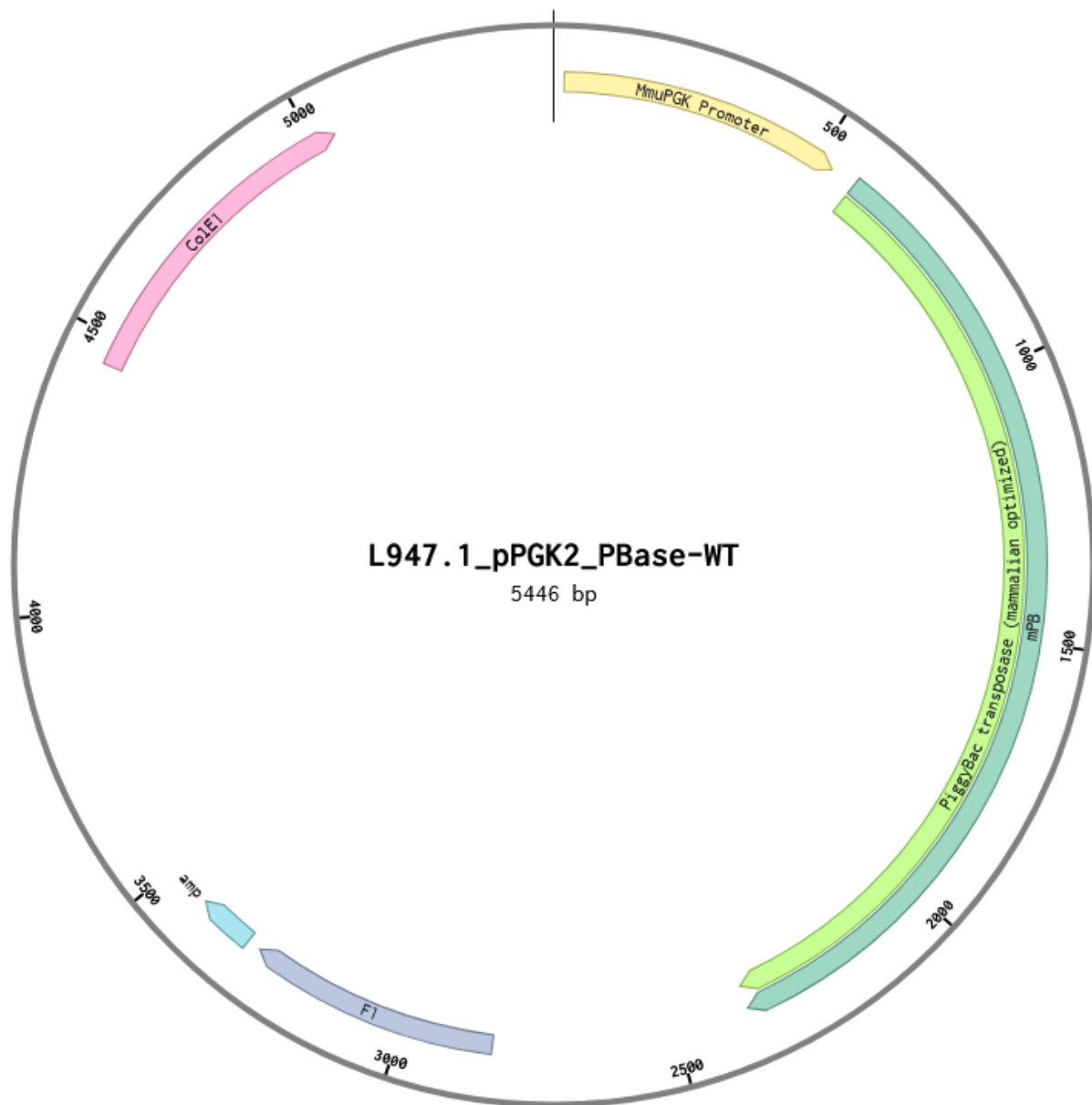


Figure S18: Transposase used for engineering the stable cell line HEK293T which expresses mScarlet-I upon doxycycline addition.

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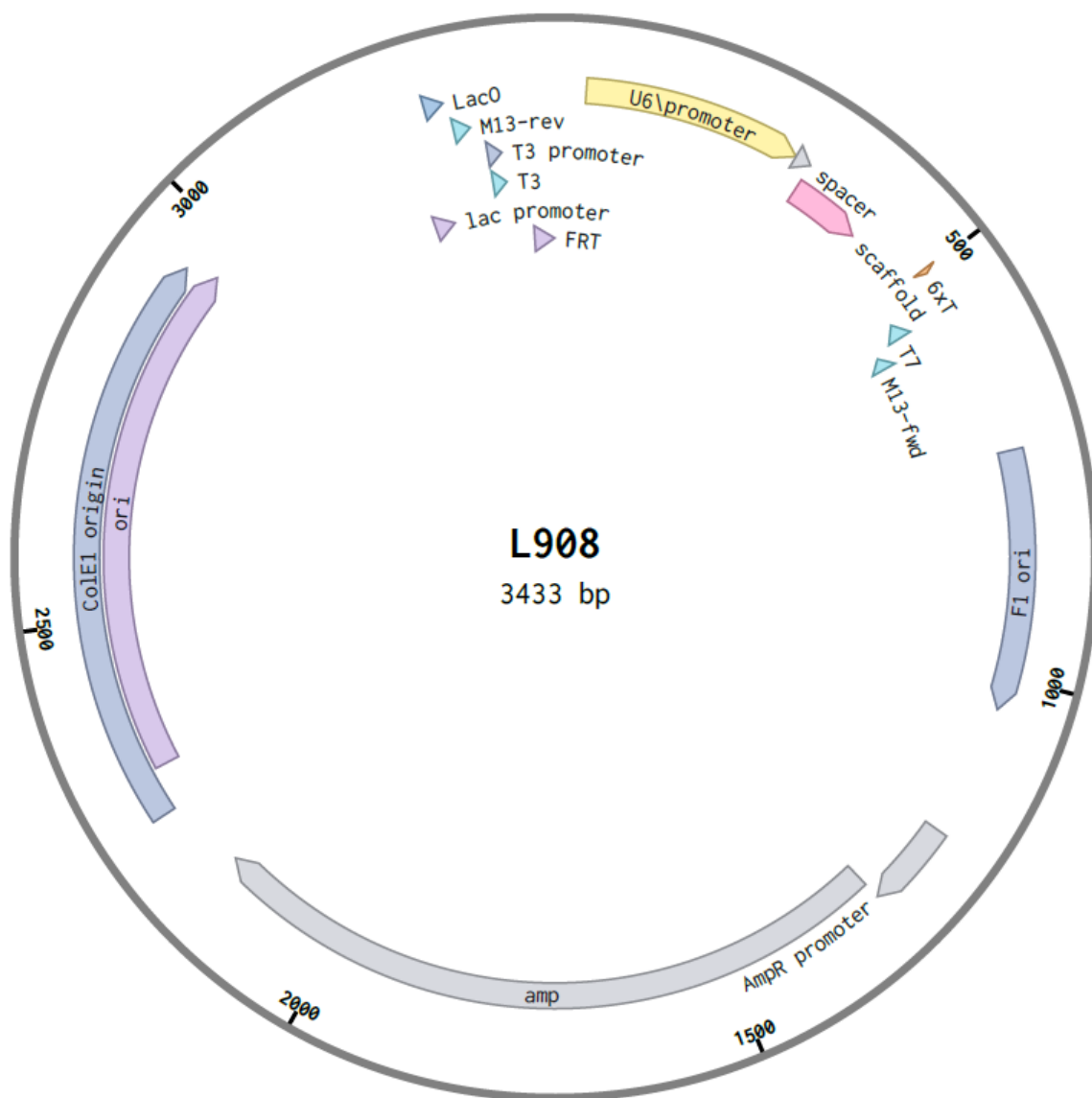


Figure S19: Plasmid used for prime editing experiments. pegRNA is expressed which leads the prime editor.

The full sequence of the plasmid encoding the pegRNA for RNA production is provided below

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The full sequence of the plasmid encoding the prime editor is provided below

gacattgattattgactagttattaatagtaatcaattacggggtcattagttcatagcccatatatggagttccgcgttacataacttacggtaaatggccgc

The full sequence of the plasmid encoding the prime editor for mRNA production is provided below

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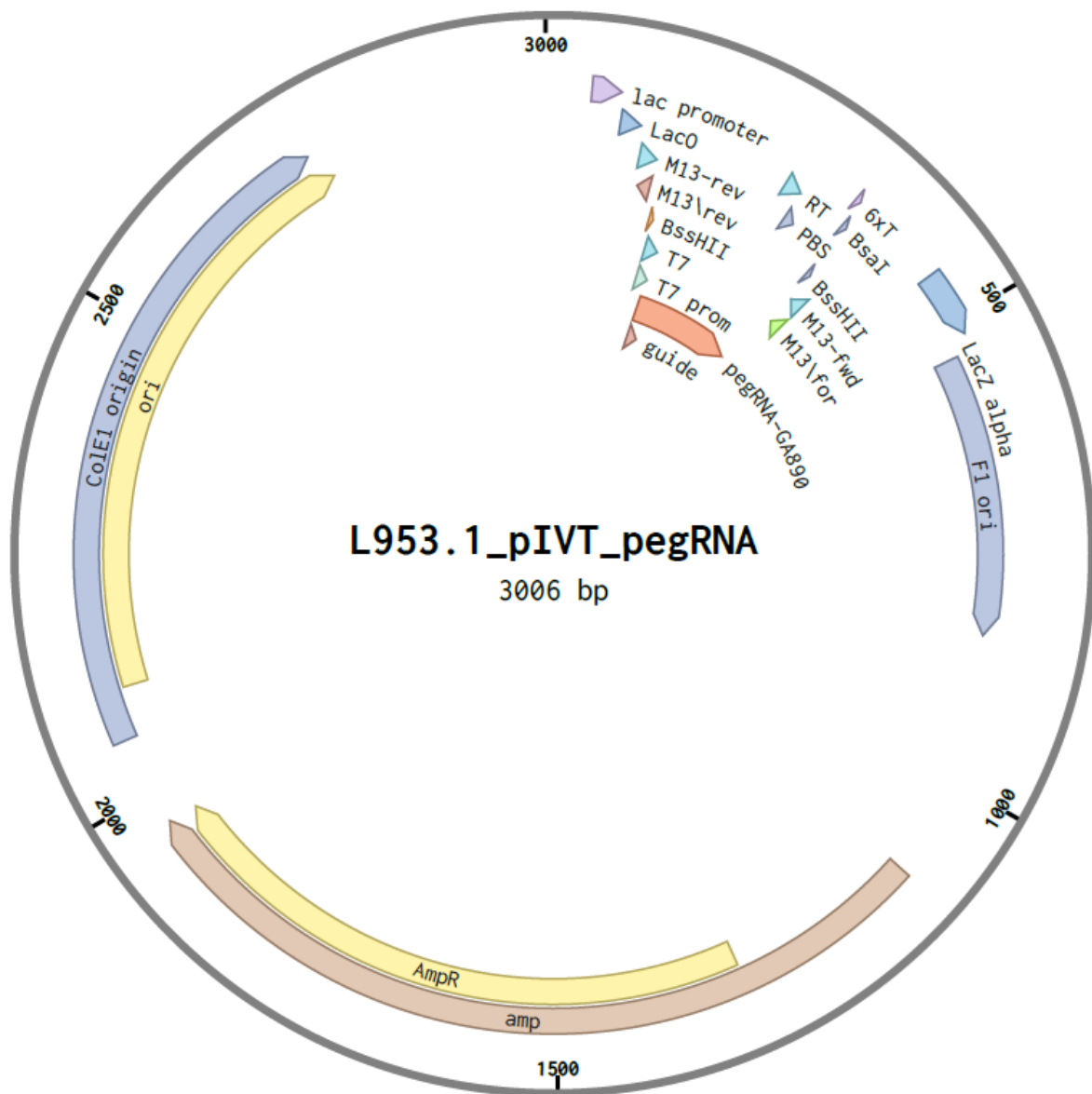


Figure S20: Plasmid used for prime editing experiments. Plasmids used to produce pegRNA via IVT.

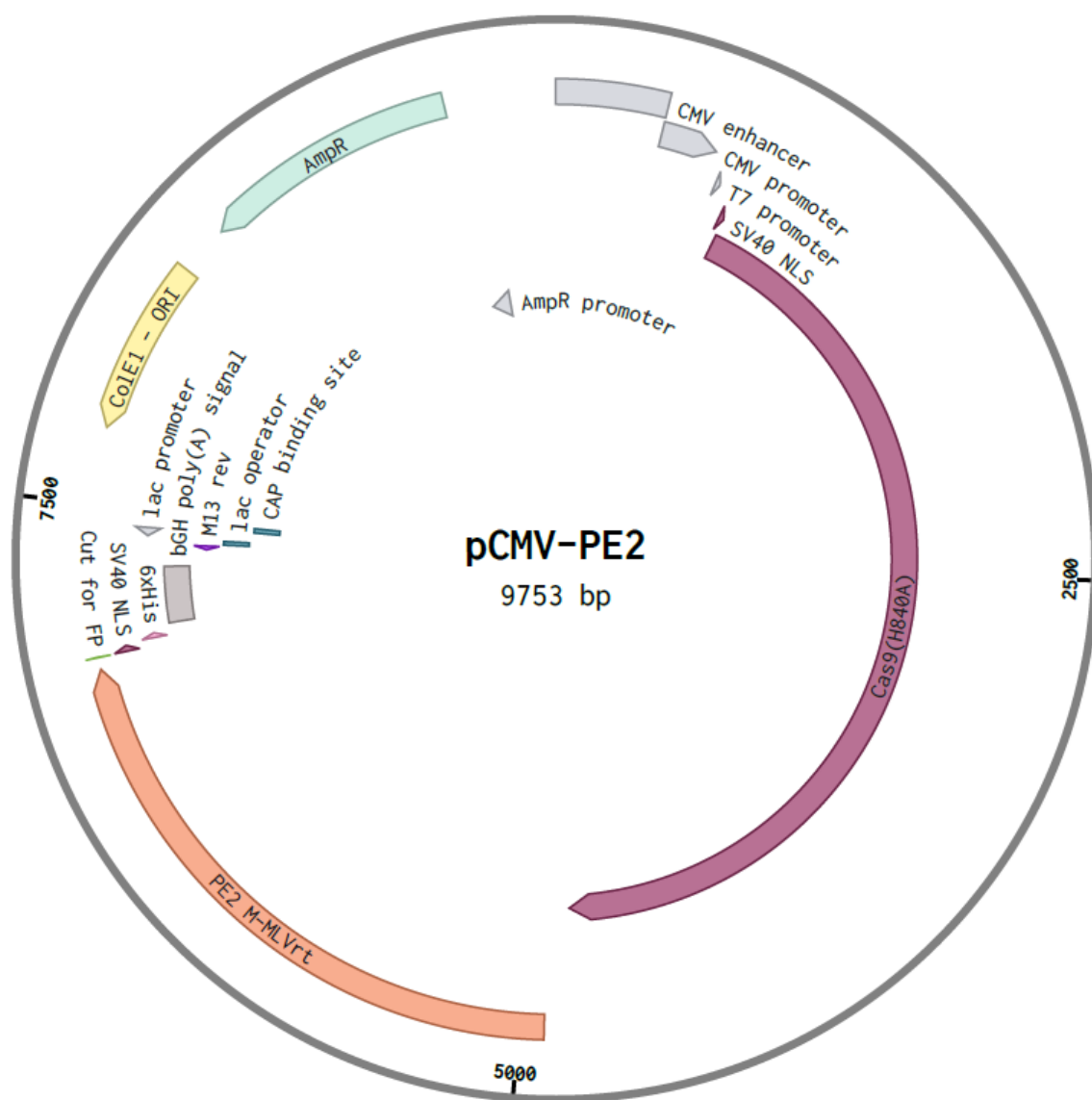


Figure S21: Plasmid used for prime editing experiments. The prime editor is expressed.

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The full sequence of the plasmid encoding the prime editor and an additional mScarlet-I for observation of prime editor expression is provided below

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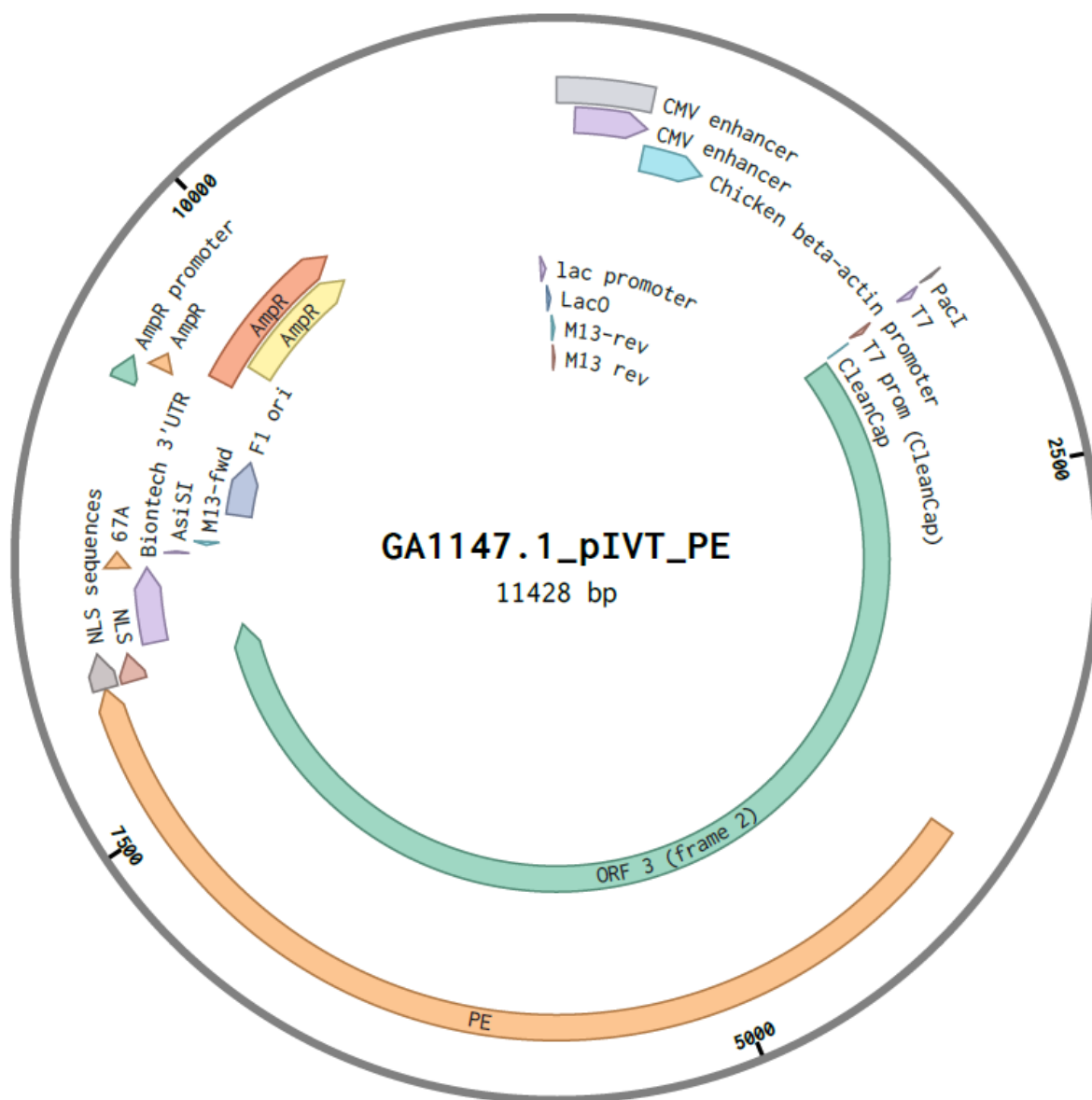


Figure S22: Plasmid used for prime editing experiments. The prime editor and mScarlet-I are expressed to visualize expression of the prime editor.

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The full sequence of the plasmid encoding the prime editor and an additional mScarlet-I for observation of prime editor expression for mRNA production is provided below

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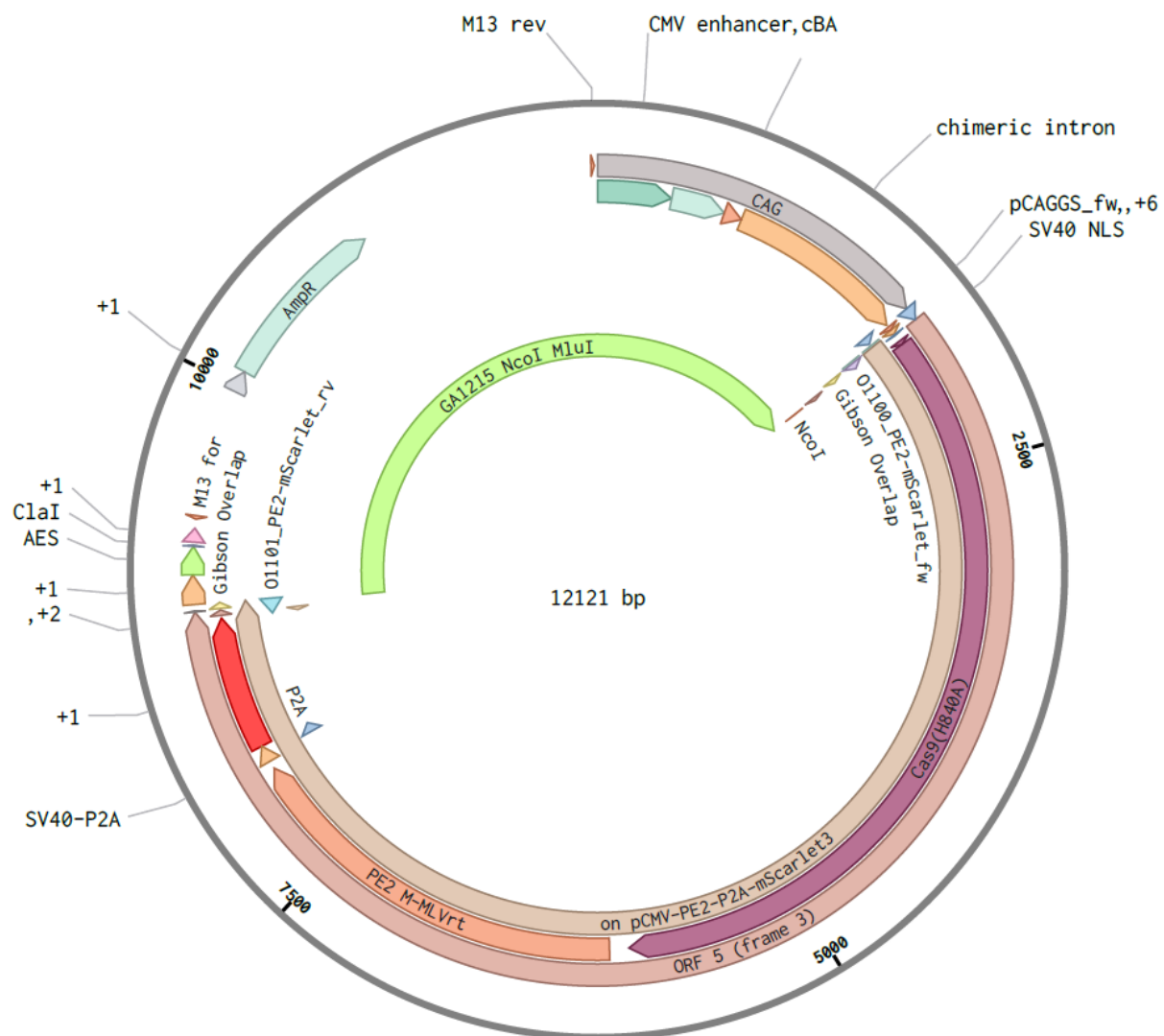


Figure S24: Plasmid used for prime editing experiments. Plasmid used to produce RNA for the prime editor and mScarlet-I for visualization via IVT.

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