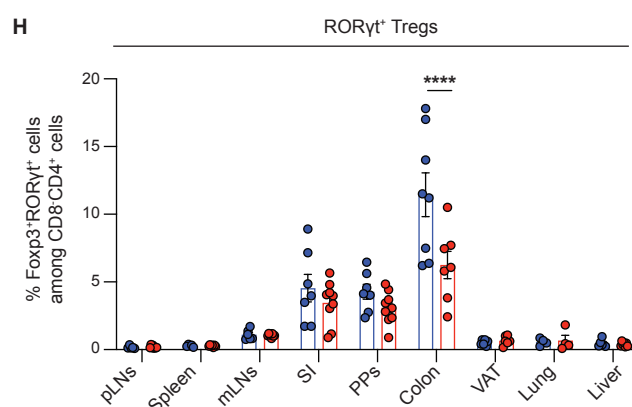
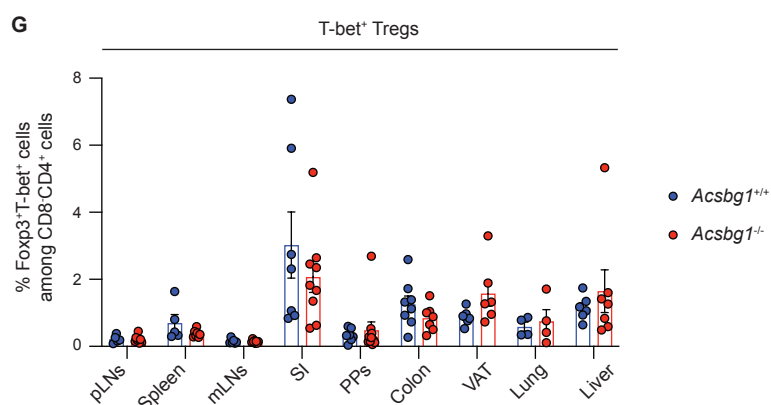
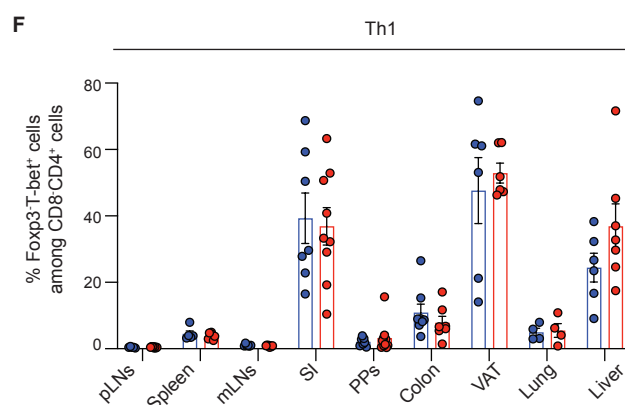
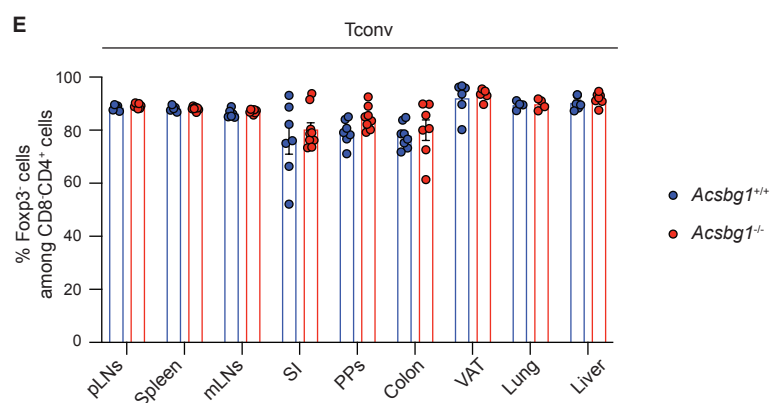
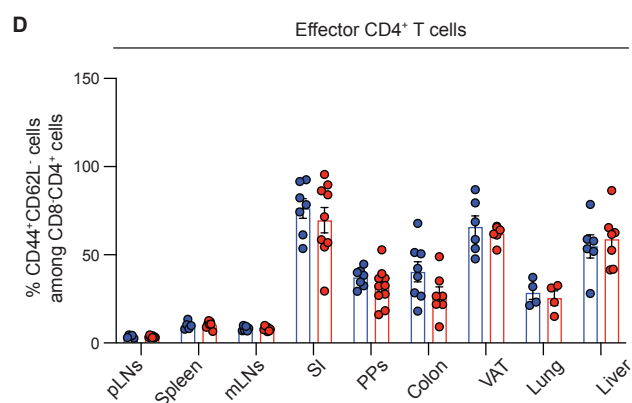
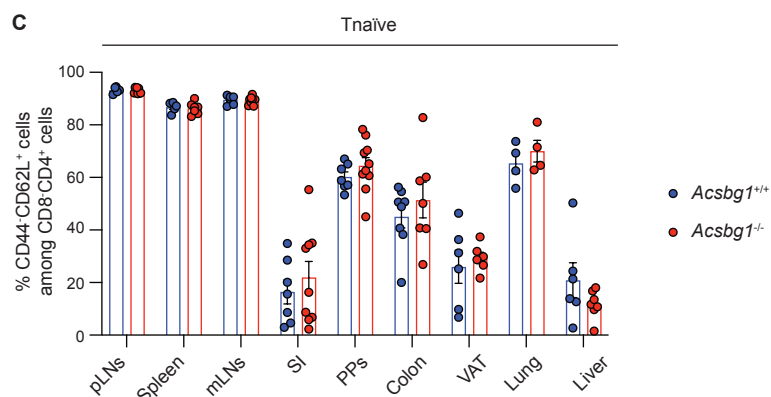
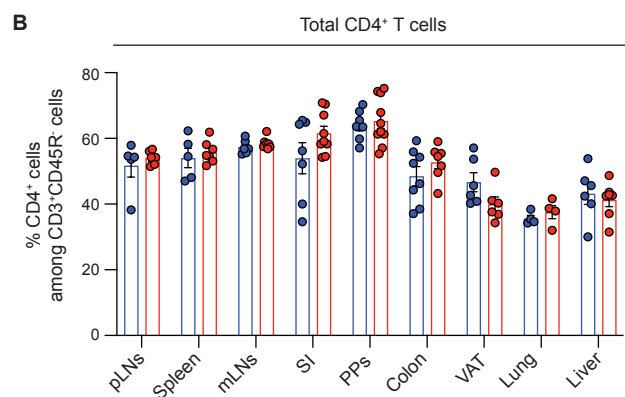
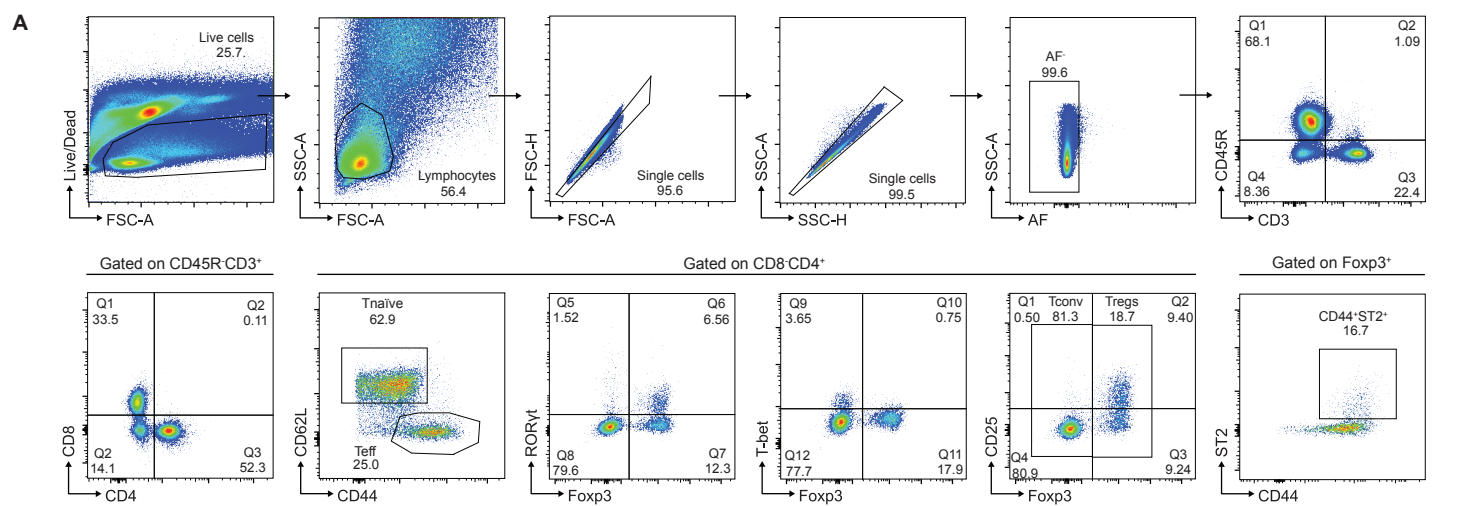


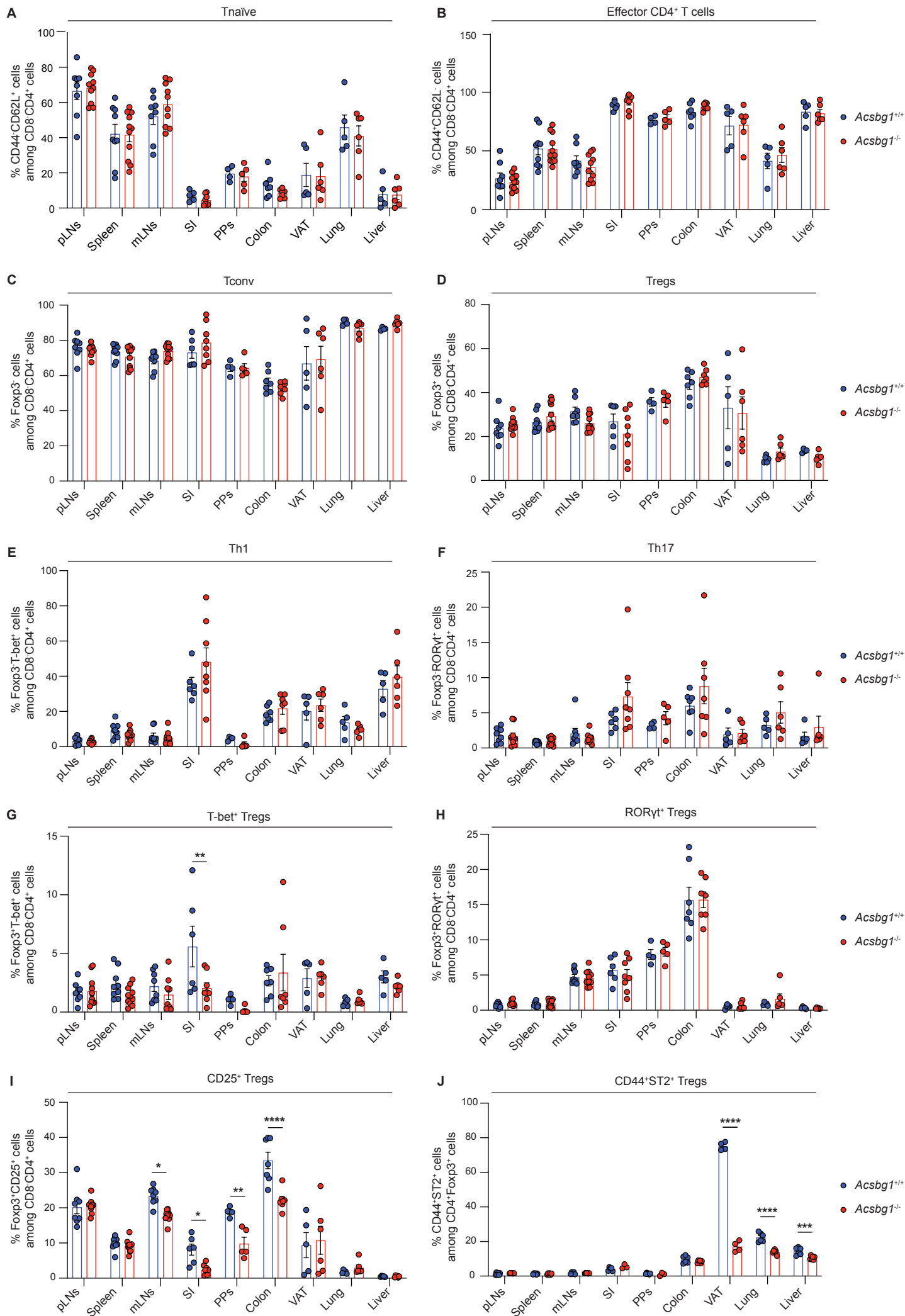
Supplementary Figures 1-11

Acsbg1 maintains intestinal immune homeostasis and controls inflammation by regulating ST2⁺ Tregs

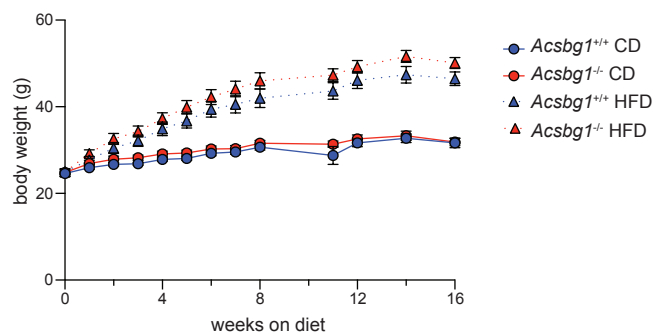
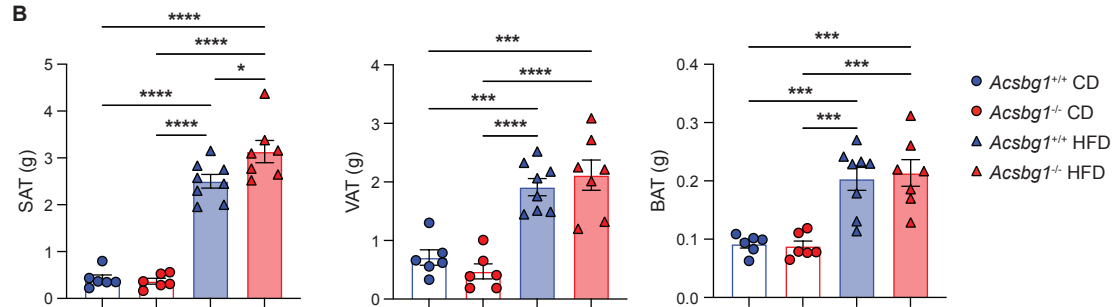
Martina Palatella, Friederike Kruse, Honglei Ji, Alina Langenhagen, Maïke Becker,
Carolin Daniel, Maria Rohm, Jochen Huehn



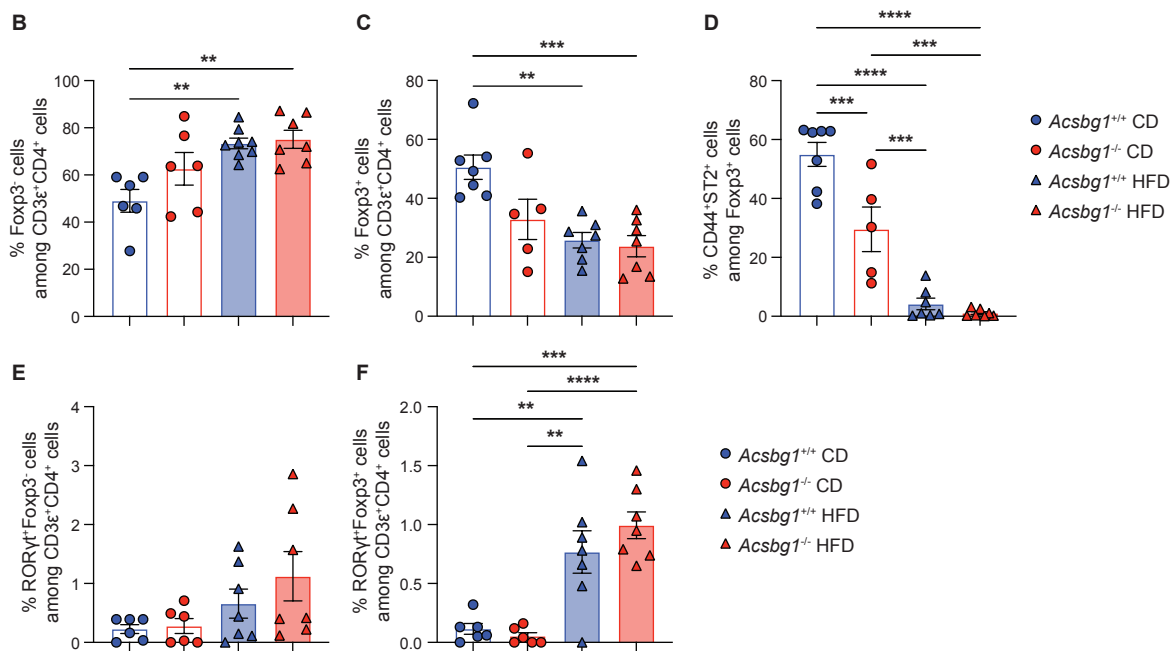
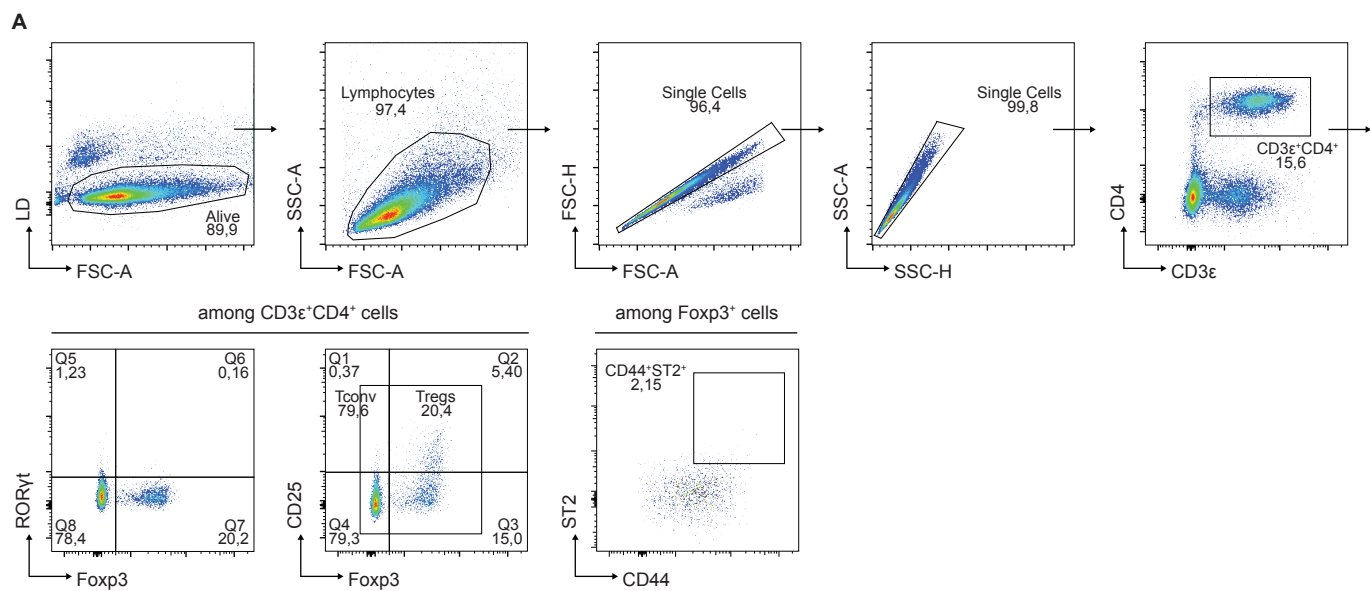
Supplementary Figure 1. *Acsbg1* deficiency affects the composition of CD4⁺ T cell subsets in adult mice. **(A)** Gating strategy for immunophenotyping. Live/dead discrimination was performed using the LIVE/DEAD Fixable Blue Dead Cell Stain. Live cells were gated according to the forward scatter (FSC) and side scatter (SSC) area parameters. Doublet exclusion was performed using the height (H) versus width (W) parameters of FSC and SSC, respectively, and autofluorescent (AF) cells were excluded. CD45R⁻CD3⁺ cells were gated to identify T cells. CD45R⁻CD3⁺ T cells were then further subdivided into CD4⁺ and CD8⁺ T cells. According to the expression of Foxp3, CD4⁺ T cells were subclassified into Foxp3⁺ Tregs and Foxp3⁻ Tconv. CD25 expression was also evaluated in both Foxp3⁺ Tregs and Foxp3⁻ Tconv. According to the expression of Foxp3 in combination with that of the master transcription factors T-bet or RORγt, T-bet⁺ Th1 and RORγt⁺ Th17 cells, RORγt⁺ Tregs and T-bet⁺ Tregs were gated within the CD4⁺ T cell population. The CD4⁺ Tnaïve and effector/memory subpopulations were also analyzed for the expression of CD44 and CD62L. In addition, CD44⁺ST2⁺ cells were gated among the Foxp3⁺ Tregs. **(B-H)** Summary graphs of immunophenotyping performed in pLNs, spleen, mLNs, SI, PPs, colon, VAT, lung and liver of adult 7-10 week old *Acsbg1*^{-/-} mice and *Acsbg1*^{+/+} littermate controls. **(B)** Frequency of total CD4⁺ cells among CD3⁺ T cells. Frequencies of **(C)** CD44⁻CD62L⁺ naïve, **(D)** CD44⁺CD62L⁻ effector, **(E)** Foxp3⁻ Tconv, **(F)** Foxp3⁺T-bet⁺ Th1, **(G)** T-bet⁺ Tregs (Foxp3⁺T-bet⁺) and **(H)** RORγt⁺ Tregs (Foxp3⁺RORγt⁺) among CD8⁻CD4⁺ T cells. Each symbol represents a single mouse. Data are presented as mean ± SEM. Results were pooled from 6 independent experiments with 4-10 mice per experimental group. Significance of differences between means was assessed by ordinary two-way ANOVA with Šídák's multiple comparison test (mean ± 95% CI; **P*<0.05, ***P*<0.01, ****P*<0.001).



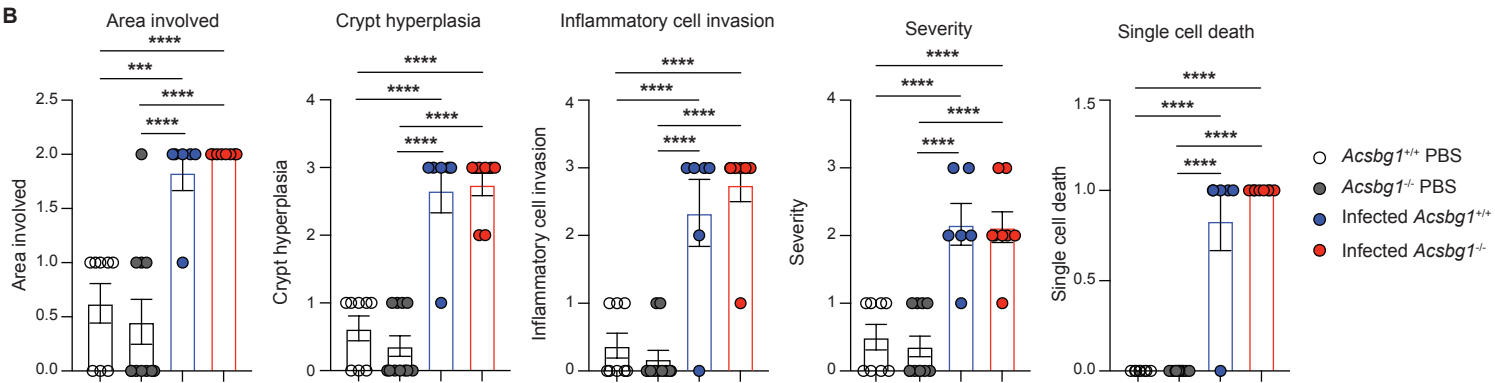
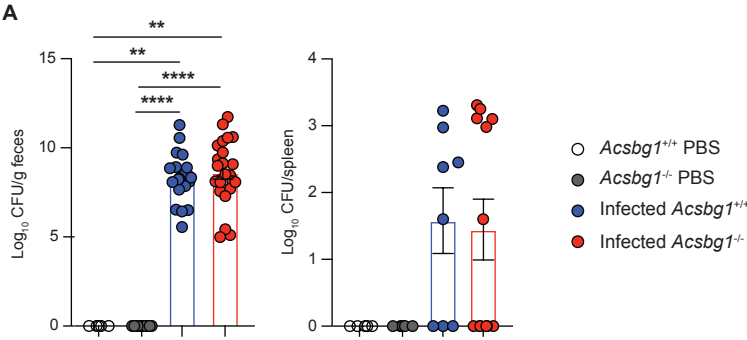
Supplementary Figure 2. Immunological changes induced by *Acsbg1* deficiency persist in aged mice. Summary plots of immunophenotyping performed in pLNs, spleen, mLNs, SI, PPs, colon, VAT, lung and liver of >80 week old *Acsbg1*^{-/-} mice and *Acsbg1*^{+/+} littermate controls. Frequencies of **(A)** CD44⁺CD62L⁺ naïve, **(B)** CD44⁺CD62L⁻ effector, **(C)** Foxp3⁻ Tconv, **(D)** Foxp3⁺ Tregs, **(E)** Foxp3⁻T-bet⁺ Th1 cells, **(F)** Foxp3⁻RORγt⁺ Th17 cells, **(G)** T-bet⁺ Tregs (Foxp3⁺T-bet⁺), **(H)** RORγt⁺ Tregs (Foxp3⁺RORγt⁺) and **(I)** CD25⁺Foxp3⁺ Tregs among CD8⁻CD4⁺ T cells. **(J)** Frequency of CD44⁺ST2⁺ cells among Tregs. Each symbol represents a single mouse. Data are presented as mean ± SEM. Results were pooled from 6 independent experiments with 4-10 mice per experimental group. Significance of differences between means was assessed by ordinary two-way ANOVA with Šídák's multiple comparison test (mean ± 95% CI; **P*<0.05, ***P*<0.01, ****P*<0.001).

A**B**

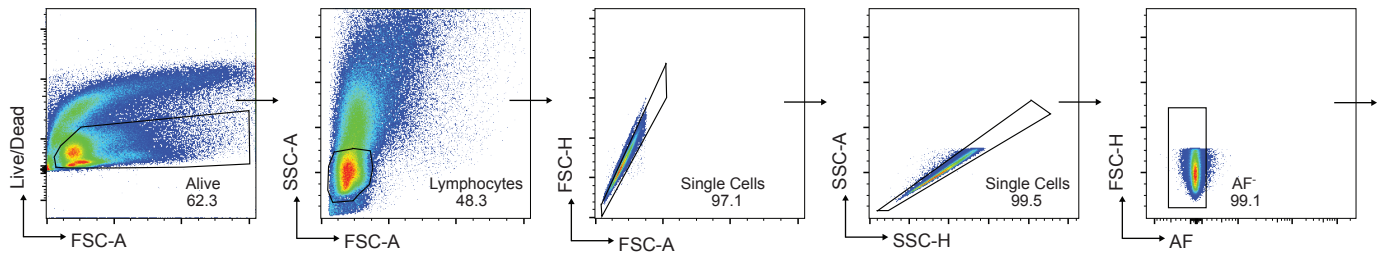
Supplementary Figure 3. *Acsbg1* deficiency increases SAT mass under HFD without affecting overall body weight. Male *Acsbg1*^{+/+} and *Acsbg1*^{-/-} mice were challenged as described in Figure 4. **(A)** Body weight was monitored for 16 weeks. **(B)** Then, mice were sacrificed, and SAT, VAT, and BAT were collected and weighed. Each symbol represents a single mouse. Data are presented as mean \pm SEM. Data are pooled from 2 independent experiments with 5-14 mice per experimental group. Significance of differences between means was assessed by ordinary two-way ANOVA with Tukey's multiple comparison test (mean \pm 95% CI; *P<0.05, **P<0.01, ***P<0.001).



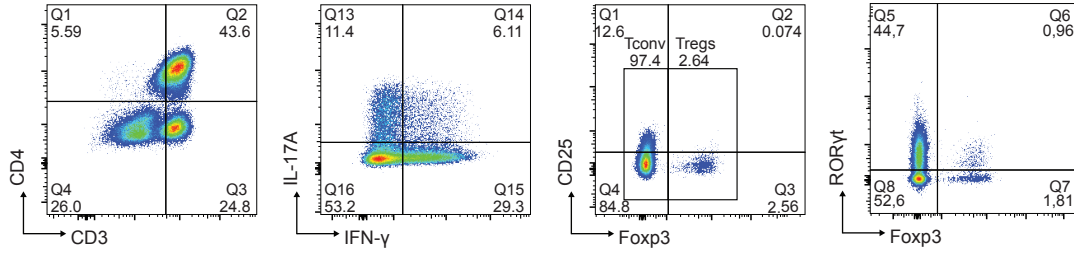
Supplementary Figure 4. HFD induces Tconv expansion and reduction of CD44⁺ST2⁺ Tregs in VAT, independent of *Acsbg1* genotype. Male *Acsbg1*^{+/+} and *Acsbg1*^{-/-} mice were challenged as described in Figure 4. **(A)** Representative gating strategy used to identify Th17, Tconv, Tregs, RORγt⁺ and CD44⁺ST2⁺ Tregs. **(B-F)** Summary graphs of immunophenotyping performed in VAT. Frequencies of **(B)** Tconv, **(C)** Tregs among CD3e⁺CD4⁺ cells. **(D)** Frequencies of CD44⁺ST2⁺ cells among Tregs. Frequencies of **(E)** Foxp3⁺RORγt⁺ Th17 cells, **(F)** RORγt⁺ Tregs among CD3e⁺CD4⁺ cells. Each symbol represents a single mouse. Data are presented as mean ± SEM. Results were pooled from 6 independent experiments with 5-8 mice per experimental group. Significance of differences between means was assessed by ordinary two-way ANOVA with Šídák's multiple comparison test (mean ± 95% CI; **P*<0.05, ***P*<0.01, ****P*<0.001).



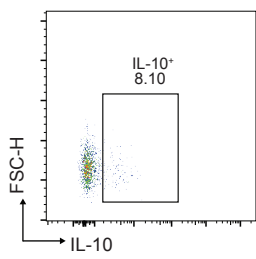
Supplementary Figure 5. Comparable bacterial burden at 7 dpi and colitis at 14 dpi in *Acsbg1*^{-/-} mice and *Acsbg1*^{+/+} mice following *C. rodentium* infection. *Acsbg1*^{-/-} mice and *Acsbg1*^{+/+} littermate controls were infected as described in Figure 4. Mice were sacrificed at either 7 or 14 dpi. **(A)** CFU of bacteria recovered from feces (left) and spleen collected at 7 dpi (right). **(B)** At 14 dpi, colon inflammation was assessed by histological analysis after H&E staining. Scatter plots with bars show the area involved, crypt hyperplasia, inflammatory cell invasion, severity of inflammation and single cell death. Each symbol represents a single mouse. Data are presented as mean ± SEM. Results were pooled from **(A)** 9 independent experiments (n = 4-13 biological replicates per group in total). Significance of differences between means was assessed by ordinary two-way ANOVA with Tukey's multiple comparison test (mean ± 95% CI; **P*<0.05, ***P*<0.01, ****P*<0.001).



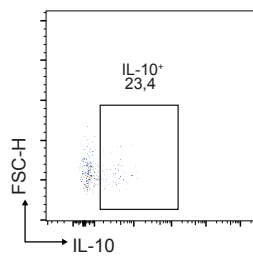
among CD3⁺CD4⁺ cells



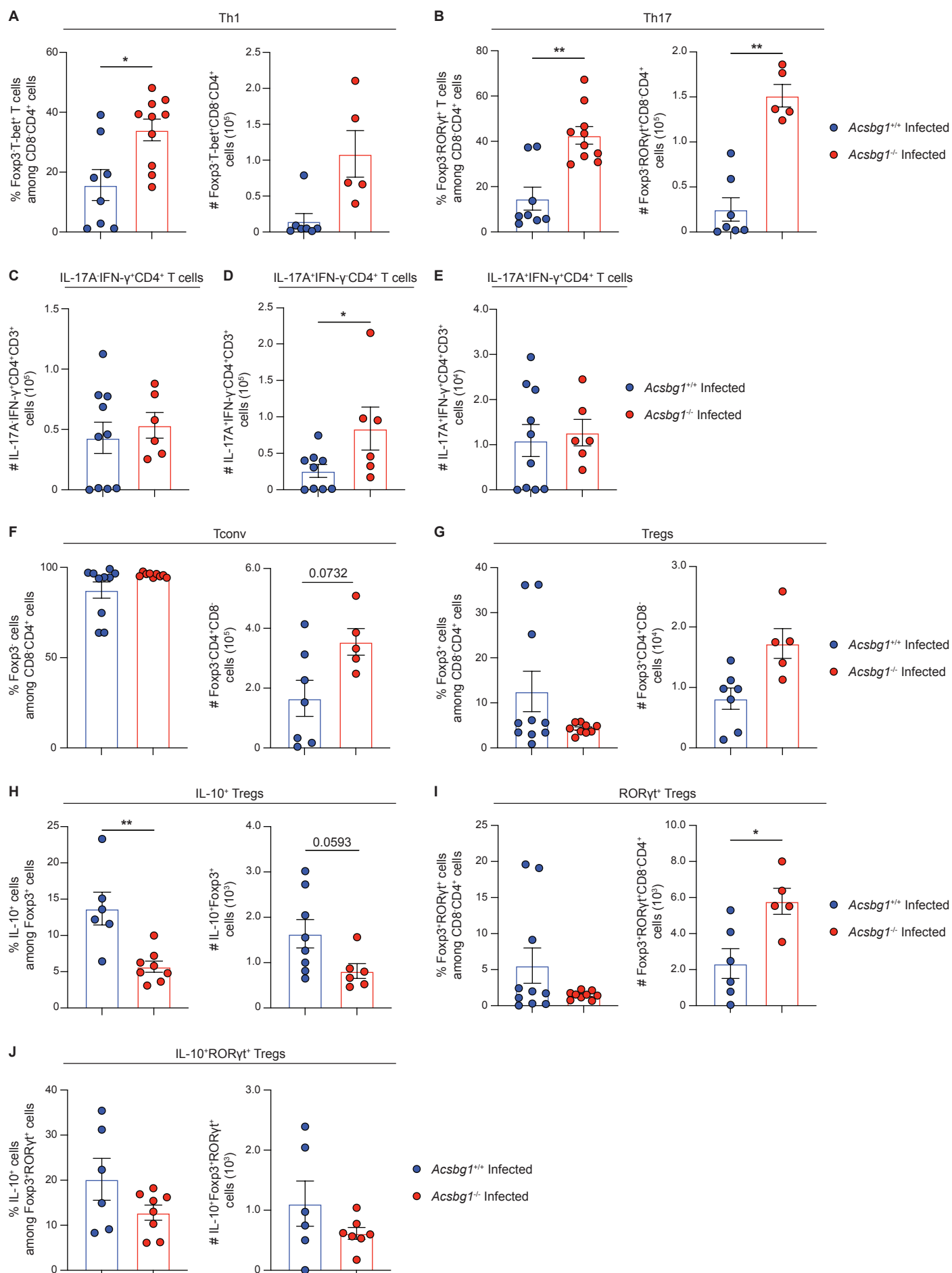
among Fc γ 3⁺ cells



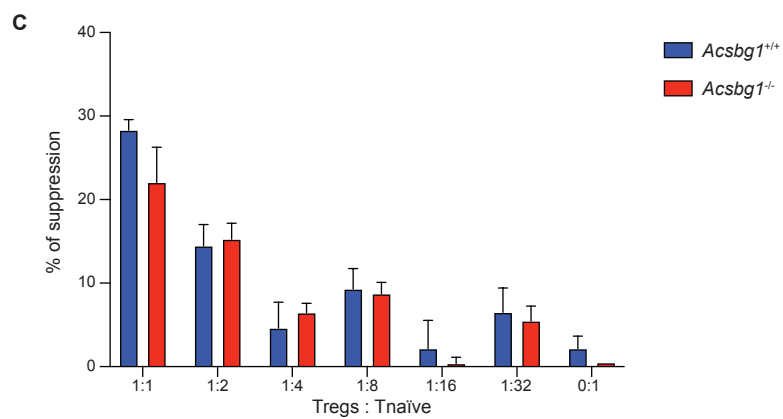
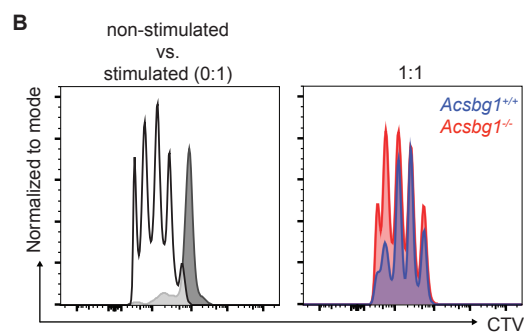
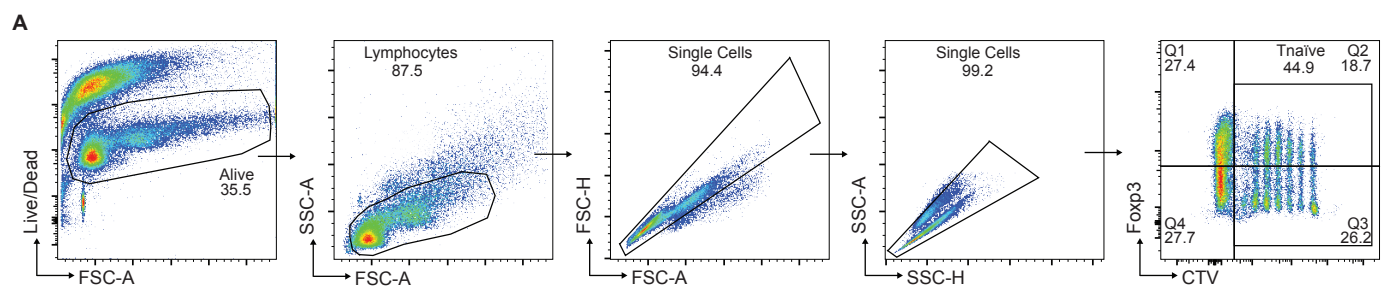
among ROR γ t⁺Fc γ 3⁺ cells



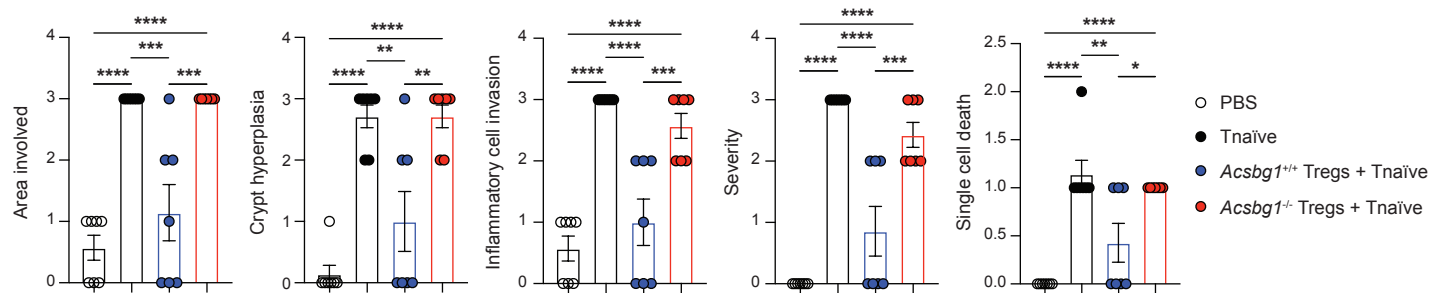
Supplementary Figure 6. Flow cytometry analysis of colon samples following *C. rodentium* infection. *Acsbg1*^{-/-} mice and *Acsbg1*^{+/+} littermate controls were infected as described in Figure 3. At 14 dpi, mice were sacrificed and colon samples were collected and analyzed by flow cytometry. **(A)** Gating strategy for cytokine detection. Isolated cells were stimulated *in vitro* with PMA, ionomycin and brefeldin A for 4 h at 37°C. Live/Dead discrimination was performed using the LIVE/DEAD Fixable Blue Dead Cell stain. Live cells were gated according to the forward scatter (FSC) and side scatter (SSC) area parameters. Doublet exclusion was then performed using the height (H) versus width (W) parameters of FSC and SSC, respectively. Among the single cells, CD3⁺CD4⁺ cells were gated to identify CD4⁺ T cells. IL-17A⁻IFN- γ ⁺, IL-17A⁺IFN- γ ⁻ and IL-17A⁺IFN- γ ⁺ cells were gated to identify single- or double-producing cells among CD4⁺ T cells. According to the expression of Foxp3, CD4⁺ T cells were subdivided into Foxp3⁺ Tregs and Foxp3⁻ Tconv cells. IL-10⁺ cells were gated to identify IL-10-producing cells among Foxp3⁺ Tregs and ROR γ t⁺Foxp3⁺ Tregs.



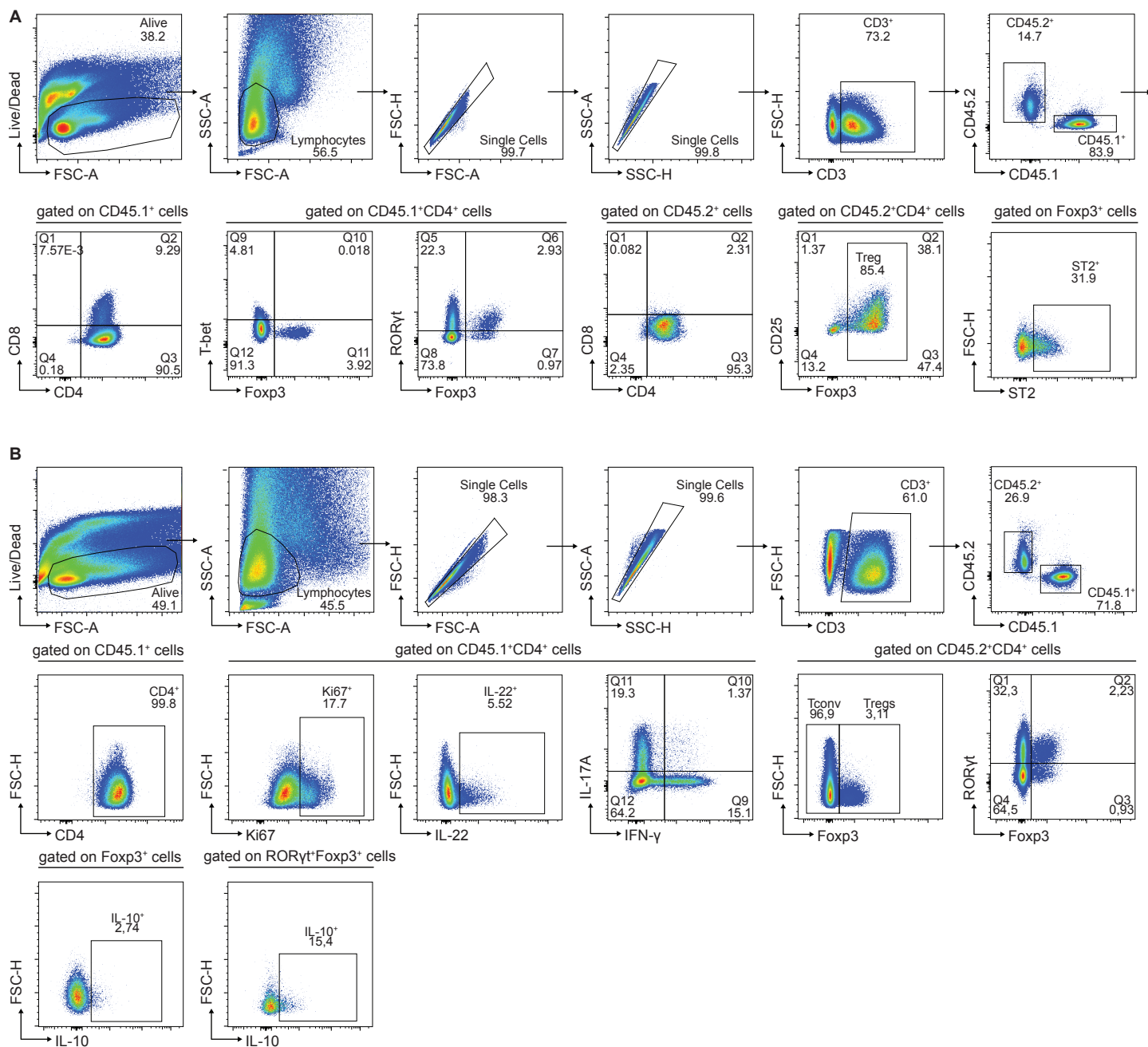
Supplementary Figure 7. *Acsbg1* deficiency exacerbates *C. rodentium* infection by affecting Th1 cells, Th17 cells and Tregs. *Acsbg1*^{-/-} mice and *Acsbg1*^{+/+} littermate controls were infected as described in Figure 3. At 14 dpi, mice were sacrificed and colon samples were collected and analyzed by flow cytometry. **(A-J)** Summary plots showing frequencies and absolute numbers of **(A)** Foxp3⁻T-bet⁺ Th1 cells and **(B)** Foxp3⁻RORγt⁺ Th17 cells among CD8⁻CD4⁺ T cells. Absolute numbers of **(C)** IL-17A⁻IFN-γ⁺, **(D)** IL-17A⁺IFN-γ⁻ and **(E)** IL-17A⁺IFN-γ⁺ cells among CD3⁺CD4⁺ T cells. Frequencies and absolute numbers of **(F)** Foxp3⁻ Tconv and **(G)** Foxp3⁺ Tregs among CD8⁻CD4⁺ T cells. Frequencies and absolute numbers of **(H)** IL-10⁺ cells among Foxp3⁺ Tregs. **(I)** Frequencies and absolute numbers of RORγt⁺ Tregs among CD3⁺CD4⁺ T cells. **(J)** Frequencies and absolute numbers of IL-10⁺ cells among RORγt⁺ Tregs. Each symbol represents a single mouse. Data are presented as mean ± SEM. Results were pooled from 4 independent experiments with 6-12 mice per experimental group. Significance of differences between means was assessed by non-parametric Mann-Whitney test (mean ± 95% CI; **P*<0.05, ***P*<0.01, ****P*<0.001).



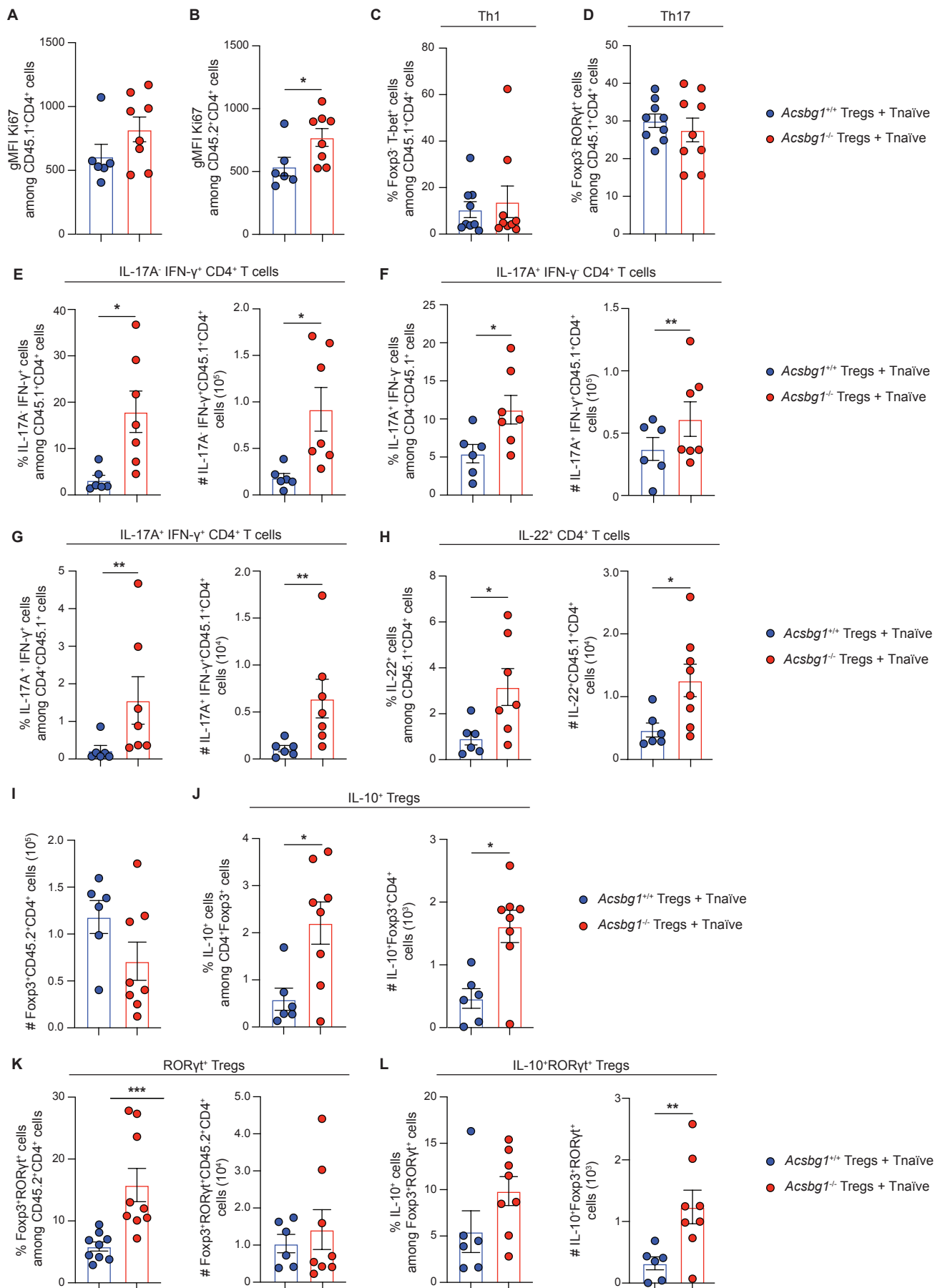
Supplementary Figure 8. *Acsbg1* deficiency moderately reduces the suppressive capacity of Tregs *in vitro*. (A) Exemplary gating strategy to identify Tnaïve from the *in vitro* suppression assay. (B) Histogram plots showing the distribution of CTV-labeled Tnaïve with (white) and without (grey) stimulus as well as co-cultures with *Acsbg1*^{+/+} (blue) or *Acsbg1*^{-/-} (red) Tregs (ratio 1:1). (C) Bar graph summarizing the percentage of suppression of CTV-labelled effector T cells in the indicated Treg/Tnaïve co-cultures, including also non-stimulated (ns) Tnaïve. Data are presented as mean ± SEM. Symbols represent technical replicates of 2-3 independent experiments. Significance was assessed by ordinary two-way ANOVA with Šidák's multiple comparison test (mean ± 95% CI; ***p* < 0.01 and ****p* < 0.001).



Supplementary Figure 9. Enhanced colitis in mice receiving *Acsbg1*^{-/-} Tregs compared to *Acsbg1*^{+/+} Tregs. Indicated Tregs and Tnaïve were adoptively co-transferred in a 1:4 ratio into sex- and age-matched *Rag2*^{-/-} mice by i.p. injection. Control groups received the same amount of Tnaïve alone or PBS. Eight weeks after transfer, colon inflammation was assessed by histological analysis. Scatter plots with bars show the area involved, crypt hyperplasia, inflammatory cell invasion, severity of inflammation and single cell death. Each symbol represents a single mouse. Data are presented as mean ± SEM. Results were pooled from 4 independent experiments with 7 mice per experimental group. Significance of differences between means was assessed by ordinary two-way ANOVA with Tukey's multiple comparison test (mean ± 95% CI; **P*<0.05, ***P*<0.01, ****P*<0.001).



Supplementary Figure 10. Gating strategies for immunophenotyping of mice from adoptive transfer colitis experiments. (A) Exemplary gating strategy used to identify different CD4⁺ T cell subsets among CD45.1⁺CD4⁺ T cells, and Foxp3⁺ Tregs among CD45.2⁺CD4⁺ T cells. **(B)** Exemplary gating strategy used to identify Ki67⁺, IL-22⁺, IL-17A⁺, IFN- γ ⁺ and IL-17A⁺IFN- γ ⁺ cells among CD45.1⁺CD4⁺ T cells, as well as IL-10⁺ cells among Tregs and ROR γ t⁺ Tregs.



Supplementary Figure 11. *Acsbg1*^{-/-} Tregs fail to attenuate T-cell mediated intestinal inflammation. Tnaïve (Lin⁻CD4⁺hCD2⁻CD25⁻CD62L^{high}) were sorted from Foxp3^{hCD2} mice (CD45.1), whereas Tregs (Lin⁻CD4⁺CD25^{high}) were sorted from *Acsbg1*^{-/-} mice or *Acsbg1*^{+/+} littermate controls (CD45.2). Tregs and Tnaïve were adoptively co-transferred in a 1:4 ratio into sex- and age-matched *Rag2*^{-/-} mice by i.p. injection. Control groups received the same amount of Tnaïve alone or PBS. Eight weeks after adoptive transfer, mice were sacrificed and colon samples analyzed by flow cytometry. gMFI of Ki67 among **(A)** CD45.1⁺CD4⁺ and **(B)** CD45.2⁺CD4⁺ T cells. Frequencies of **(C)** Th1 and **(D)** Th17 cells among CD45.1⁺CD4⁺ T cells. Frequencies and absolute cell numbers of **(E)** IFN γ ⁺IL-17A⁻, **(F)** IFN γ ⁺IL-17A⁺, **(G)** IFN γ ⁻IL-17A⁺ and **(H)** IL-22⁺ cells among CD45.1⁺CD4⁺ T cells. **(I)** Absolute numbers of Tregs among CD45.2⁺CD4⁺ cells. **(J-K)** Frequency and absolute numbers of **(J)** IL-10⁺ and **(K)** ROR γ t⁺ cells among Tregs. **(L)** Frequency and absolute number of IL-10⁺ cells among ROR γ t⁺ Tregs. Each symbol represents a single mouse. Data are presented as mean \pm SEM. Results were pooled from 2 independent experiments with 9 mice per experimental group. Significance of differences between means was assessed by non-parametric Mann-Whitney test (mean \pm 95% CI; **P*<0.05, ***P*<0.01, ****P*<0.001).