



Radiation, inflammation, and immune responses in cancer

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Chronic inflammation has emerged as one of the hallmarks of cancer. Inflammation also plays a pivotal role in modulating radiation responsiveness of tumors. As discussed in this review, ionizing radiation (IR) leads to activation of several transcription factors modulating the expression of numerous mediators in tumor cells and cells of the microenvironment promoting cancer development. Novel therapeutic approaches thus aim to interfere with the activity or expression of these factors, either in single-agent or combinatorial treatment or as supplements of the existing therapeutic concepts. Among them, NF- κ B, STAT-3, and HIF-1 play a crucial role in radiation-induced inflammatory responses embedded in a complex inflammatory network. A great variety of classical or novel drugs including nutraceuticals such as plant phytochemicals have the capacity to interfere with the inflammatory network in cancer and are considered as putative radiosensitizers. Thus, targeting the inflammatory signaling pathways induced by IR offers the opportunity to improve the clinical outcome of radiation therapy by enhancing radiosensitivity and decreasing putative metabolic effects. Since inflammation and sex steroids also impact tumorigenesis, a therapeutic approach targeting glucocorticoid receptors and radiation-induced production of tumorigenic factors might be effective in sensitizing certain tumors to IR.

Keywords: radiation, inflammation, radioresistance, PGHS-2, heat shock proteins, NF- κ B, STAT-3, HIF-1

INTRODUCTION

Chronic inflammation has emerged as one of the hallmarks of cancer impacting any stage of tumorigenesis (Colotta et al., 2009; Grivennikov and Karin, 2010). The persistent expression of inflammatory mediators exert pleiotropic effects on the malignant process. On the one hand, they affect carcinogenesis and malignant transformation, tumor growth, invasion, and metastasis, on the other hand they activate immune effector mechanisms limiting tumor growth. The link between cancer and inflammation can be viewed as consisting of two pathways: an intrinsic and an extrinsic pathway (Mantovani et al., 2008). At the level of the tumor cell, both pathways converge and induce the activation of several transcription factors culminating in the formation of numerous pro-inflammatory molecules that recruit and activate various leukocyte populations into the tumor microenvironment (for a review, see Multhoff et al., 2012). The tumor cell-derived pro-inflammatory molecules now activate the same transcription factors within the cells of the microenvironment and the tumor cells themselves resulting in a more pronounced generation of inflammatory mediators driving a tumor-promoting amplification loop. This amplification mechanism further enhances the impact of inflammatory stimuli within the tumor environment and triggers the manifestation of a cancer-related inflammatory milieu contributing to tumor growth and invasiveness.

NF- κ B provides a mechanistic link between inflammation, carcinogenesis, and tumor radioresistance (Magne et al., 2006; Ben-Neriah and Karin, 2011). NF- κ B is regarded as the key orchestrator controlling the ability of both, preneoplastic and malignant cells,

to resist apoptosis-based tumor surveillance mechanisms activated by DNA damage and chromosomal rearrangement or anti-cancer drugs and radiation, respectively. NF- κ B might also regulate tumor angiogenesis and invasiveness (Karin, 2006), and may contribute to the characteristic radio-/chemoresistance of tumor cells (Fahy et al., 2004; Singh and Khar, 2006; Antoon et al., 2011; Chaturvedi et al., 2011). In conjunction with STAT-3 and HIF-1, NF- κ B serves as a modulator of the expression of several factors promoting cancer development. Novel therapeutic approaches thus aim to interfere with the activity or expression of these factors, either in single-agent or combinatorial treatment or as supplements of the existing therapeutic concepts. Noteworthy, targeting the pro-inflammatory signaling pathways for tumor radiosensitization represents a promising novel therapeutical approach in cancer. A great variety of classical or novel drugs including nutraceuticals have the capacity to interfere with the inflammatory network in cancer and are progressively tested for tumor radiosensitization. Accumulating evidence over the last few years indicate that most chemotherapeutic agents and radiation therapy activate NF- κ B (Wang et al., 1996; Sandur et al., 2009; Li and Sethi, 2010). Thus, NF- κ B blockage has been recognized as a promising tool in increasing radiosensitivity of tumors. Apart from NF- κ B, STAT-3, HIF-1, and PGHS-2 are further inflammatory factors crucially involved in radioresistance of tumors. Thus, interrupting the inflammatory network in cancer by targeting these molecules may be a promising radiosensitization approach in cancer therapy. **Table 1** provides an overview on natural and (semi-)synthetic compounds that are considered as putative radiosensitizers.

Table 1 | Natural and (semi-)synthetic compounds as putative radiosensitizers und their targets.

Compound	Source/systematic name	Target
Anacardic acid	<i>Anacardium occidentale</i> (cashew nuts)	IKK, NF- κ B
Berberine	<i>Berberis aristata</i> (Indian barberry, tree turmeric)	NF- κ B
Butein	<i>Rhus verniciflua</i> (Chinese lacquer tree)	NF- κ B
Caffeic acid phenethyl ester	Honeybee propolis	GSH, NF- κ B
Celecoxib	4-[5-(4-methylphenyl)-3-(trifluoromethyl) pyrazol-1-yl]benzenesulfonamide	PGHS-2, NF- κ B
Cepharanthine	<i>Stephania cepharantha</i> Hayata	NF- κ B, STAT3
Crotopoxide	<i>Kaempferia pulchra</i> (peacock ginger)	TAK-1
Curcumin	<i>Curcuma longa</i> (turmeric)	Akt, IKK, NF- κ B
Daidzein, genistein	<i>Glycine max</i> (soy bean)	STAT3, HIF-1 α
Deguelin	<i>Derris trifoliata</i> (threeleaf derris)	Hsp90, HIF-1 α
EGCG	<i>Camellia sinensis</i> (green tea)	NF- κ B
Emodin	<i>Rheum rhabarbarum</i> (Rhubarb), <i>Aloe vera</i>	HIF-1
Erufosine	Alkylphosphocholine (synthetic phospholipid analog)	Akt
Ethaselen	1,2-[bis(1,2-benzisoselenazolone-3(2H)-ketone)] ethane; BBSKE	Thioredoxin reductase, NF- κ B
Flavopiridol	Semi-synthetic flavonoid based on an extract from the Indian plant, <i>Dysoxylum binectariferum</i>	CDKs, cyclin D1, Rb, Bcl-2
Geldanamycin	Naturally occurring ansamycin antibiotic from <i>Streptomyces hygroscopicus</i>	Hsp90
KNK437	Benzylidene lactam compound	Hsp27, Hsp70
Nitidine chloride	<i>Zanthoxylum nitidum</i> (Tez-mui, Tejamool in Assamese; locally called "liangmianzhen")	STAT3
Oleandrin	<i>Nerium oleander</i> (nerium)	Caspase-3
Parthenolide	<i>Tanacetum parthenium</i> (feverfew)	NF- κ B, p53
Piceatannol	Hydroxylated resveratrol analog found in various plants, e.g., <i>Vitis spec.</i>	NF- κ B
Picroliv	<i>Picrorhiza kurroa</i> (katuka)	NF- κ B
Piperine	<i>Piper nigrum</i> (black pepper)	CYP450 enzymes
Plumbagin	<i>Plumbago rosea</i> (Scarlet leadwort)	NF- κ B
Resveratrol	<i>Vitis spec.</i> (grape, red wine)	STAT3, NF- κ B
Silymarin	<i>Silybum marianum</i> (milk thistle)	NF- κ B
Xanthohumol	<i>Humulus lupulus</i> (common hop)	NF- κ B

GENETIC INSTABILITY

Recent observations allow a deeper insight into the molecular and cellular mechanisms linking inflammation and tumorigenesis. Emerging data suggest that genetic destabilization of tumor cells is regarded as a further hallmark of most human cancers contributing to tumor initiation and progression (Colotta et al., 2009). Apart from the production of cytokines, chemokines, proteases and prostanoids, inflammatory cells are able to produce reactive oxygen species (ROS) and reactive nitrogen species (RNS). Leukocytes are the main source of RNS and ROS acting as chemical effectors in inflammation-driven carcinogenesis (Kundu and Surh, 2008). All of these mediators act together in perpetuating and amplifying the inflammatory cascade. As outlined in **Figure 1**, they suppress DNA repair mechanisms leading to an increase in genetic instability termed microsatellite instability (MSI) as a result of mutations or epigenetic alterations of members of the mismatch repair (MMR) family (Hakem, 2008). The MMR system is strongly affected by inflammatory conditions. It has been shown previously that the transcription factor HIF-1 is induced in tumor cells not only by different cytokines and prostaglandins (Jung et al., 2003) but also by ROS and RNS (Sandau et al., 2000). HIF-1 is a heterodimeric transcription factor consisting of a constitutively expressed β -subunit and an oxygen-regulated

α -subunit (Kaelin Jr. and Ratcliffe, 2008). It was proved that HIF-1 plays a pivotal role in hypoxia-induced tumor radioresistance (Moeller and Dewhirst, 2006; Harada, 2011). In this context, our own investigations revealed no correlation between basal HIF-1 α levels and the survival fraction in irradiated tumor cell lines implying that basal HIF-1 α levels in human tumor cell lines obviously do not predict their radiosensitivity under normoxia (Schilling et al., 2012a). Moreover, HIF-1 α has been found as being responsible for genetic instability to down-regulated MMR proteins by inhibiting the MMR proteins MSH-2 and MSH-6, thereby decreasing levels of the MSH-2/MSH-6 complex, MutS α , which recognizes base mismatches. HIF-1 α displaces the transcriptional activator c-Myc from Sp1 binding to repress MutS α expression in a p53-dependent manner (Koshiji et al., 2005). Chang et al. (2002) observed that hydrogen peroxide inactivates members of the MMR family at the protein level. From this observation the authors speculate that inactivation of the MMR function in response to oxidative stress may be responsible for the low-frequency MSI (MSI-L) seen in non-neoplastic and cancer tissues associated with chronic inflammation.

Chromosomal instability can also be the result of the deleterious action of inflammatory mediators culminating in abnormal chromosomal segregation and aneuploidy. The deleterious actions

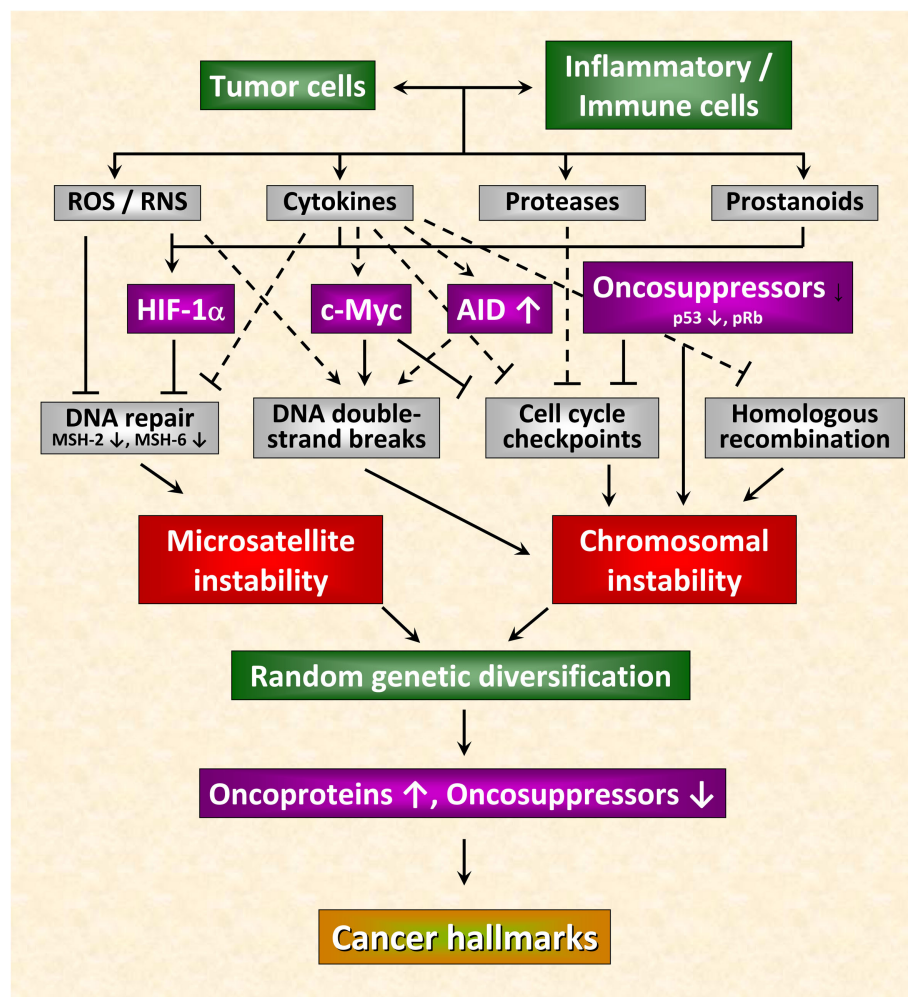


FIGURE 1 | Inflammation-induced molecular pathways causing genetic instability in cancer cells. Genetic destabilization of tumor cells is regarded as a further hallmark of most human cancers contributing to tumor initiation and progression. Apart from the production of cytokines, chemokines, proteases, and prostanoids, inflammatory cells are able to produce reactive oxygen (ROS) and nitrogen species (RNS). All of these mediators act together in perpetuating and amplifying the inflammatory cascade. On the one hand, they suppress DNA repair mechanisms leading to microsatellite

instability. On the other hand, they can cause chromosomal instability culminating in abnormal chromosomal segregation and aneuploidy. These inflammatory mediators induce DNA double-strand breaks, affect function of mitotic checkpoint molecules and dysregulate homologous recombination of DNA double-strand break repair leading to random genetic diversification of tumor cells. Cancer cells harboring the optimal combination of activated oncoproteins and inactivated oncosuppressor proteins will develop the malignant phenotype (figure adapted from Colotta et al., 2009; for details see text).

of inflammatory mediators include direct or indirect induction of DNA double-strand breaks (Karanjawala et al., 2002; Mills et al., 2003), defective mitotic checkpoints (Rajagopalan et al., 2003; Menssen et al., 2007), and dysregulated homologous recombination of DNA double-strand break repair (Saintigny et al., 2001; Hakem, 2008; Plo et al., 2008). Further critical molecules that affect genetic stability comprise activation-induced cytidine deaminase (AID; Endo et al., 2008), c-Myc (Vafa et al., 2002), phospho-retinoblastoma protein pRb (Pickering and Kowalik, 2006), and p53 (Tomasini et al., 2008). By causing microsatellite as well as chromosomal instability these molecules induce random genetic diversification of tumor cells. As already discussed by Colotta et al. (2009), cancer clones harboring the optimal combination

of activated oncoproteins and inactivated oncosuppressors will develop the malignant phenotype (Figure 1).

CELL-CELL INTERACTIONS

In the tumor microenvironment, an intensive interaction between tumor cells and infiltrating immune cells occurs. The latter comprise macrophages, dendritic cells (DC), T cells as well as NK cells with macrophages and T cells as being the most frequent ones (Luster et al., 2005). Inflammatory mediators secreted by tumor and immune cells have been found to play a dual role in tumor development. On the one hand, they promote tumor development and survival of tumor cells, on the other hand they exert surveillance mechanisms against tumor cells (Ben Baruch, 2006; Kim et al.,

2006b). In case of a predominance in anti-tumor immunity, tumor cells are eradicated whereas a predominance in surveillance mechanisms provides cancer cells with an immunosuppressive network extending the immune evasion and promoting tumor progression and metastasis (Hadden, 2003). Among the inflammatory mediators secreted by tumor and immune cells TNF, IL-6, and IL-17 act as crucial players in developing chronic inflammation resulting in immune escape and acceleration of tumor progression and metastasis. Upon activation, myeloid cells produce pro- and anti-inflammatory mediators not only affecting growth, survival, and invasiveness of tumor cells but also controlling functional activities of Th1, NK, Treg, and Th17 cells (Lin and Karin, 2007). TRAIL, a member of the TNF superfamily and the product of activated T and NK cells, directly induces apoptosis in numerous cancer cells (LeBlanc and Ashkenazi, 2003) thus playing a crucial role in tumor surveillance mechanisms. Regulatory T cells (Treg) function as key components for regulating anti-tumor immunity (Yamaguchi and Sakaguchi, 2006). Treg specifically suppress the cytotoxicity of expanded CD8⁺ cytotoxic T cells (Chen et al., 2005) and induce release of IL-17 from Th17 cells acting as key players in chronic inflammation (Mangan et al., 2006). IL-17-mediated effects on the inflammatory response involve recruitment of immune cells (Park et al., 2005), induction of pro-inflammatory factors (IL-1, IL-6, TNF) as well as promotion of angiogenesis and tumor growth (Numasaki et al., 2003). Development of Th17 cells is stimulated by IL-6, IL-23, TGF- β , and TNF released from activated myeloid cells. Anti-inflammatory IL-10 inhibits tumor progression and development by blocking synthesis of IL-6, IL-12, and TNF via NF- κ B inhibition (Moore et al., 2001). Furthermore, the anti-tumor activity of Treg is mediated by IL-10 released from Treg themselves (Erdman et al., 2003). IL-23 belonging to the IL-12 family of cytokines enhances the production of IFN- γ and IL-12 by activated T cells, induces IL-17 release from Th17 cells and

promotes inflammation at its final stage (Cho et al., 2006). Moreover, IL-23 has been found to up-regulate expression of MMP-9, to increase angiogenesis and to decrease CD8⁺ T cell recruitment to tumors possibly providing the basis for the development of a tumor-promoting environment (Langowski et al., 2006). IL-12, also released from antigen-presenting cells (APC) after stimulation with IL-23, is a further member of the IL-12 family of cytokines and harbors anti-tumor activities in particular via stimulation of Th1- and CTL-mediated immune responses (Trinchieri, 2003; Langowski et al., 2006). **Table 2** summarizes the cellular and molecular outcomes based on interactions between various cell types in the tumor microenvironment.

SEX STEROIDS

An increasing number of data currently reveal the close relationship between the two classical pathways in tumor progression: inflammation and gonadal hormones. Since the discovery of the hormone dependency of mammary carcinoma in the 1890s, it has become clear that gonadal steroids play a crucial role in the pathogenesis of breast and prostate cancer. The Scottish surgeon George Thomas Beatson was about the first who showed that oophorectomy in a premenopausal woman with breast cancer led to a complete remission (Beatson, 1896a,b) highlighting the role of sex steroids in tumor pathogenesis. However, more recent investigations revealed an astonishing effect of gonadal hormones on tumorigenesis. Female somatic cells including tumor cells express receptors for sex steroid hormones such as estrogen and progesterone affecting growth of hormone-dependent breast cancer cells (Henderson and Canellos, 1980a,b). Women are well known as being less susceptible to tumors at organ sites not representing classical targets for gonadal hormones including the liver. Thus, hepatocellular carcinoma (HCC), the most common liver cancer, occurs mainly in men. The same gender disparity is seen in mice

Table 2 | Effects of cell–cell interactions in the tumor microenvironment.

Effector	Molecular/cellular outcome	Physiology/pathophysiology
IL-6, IL-10, TNF	Enhancement of tumor cell growth	Chronic inflammation
IL-10	Anti-inflammatory, blockage of IL-6, IL-12, TNF synthesis via NF- κ B inhibition	Tumor suppression
IL-12	Activation of CD8 ⁺ CTL and NK cells, expression of cytotoxic mediators (IFN, TRAIL, TGF- β)	Anti-tumor effect
IL-17	Induction of pro-inflammatory mediators (IL-1, IL-6, TNF)	Chronic inflammation, tumor progression
IL-23	Induction of IL-12/IFN- γ release from activated T cells, TNF/IL-12 from APC, IL-17 from Th17 cells, MMP-9 up-regulation, decrease of CD8 ⁺ CTL recruitment, increase in angiogenesis	Chronic inflammation, tumor progression
TGF- β	Enhancement of tumor cell invasiveness and angiogenesis, inhibition of NK cells, CTL, macrophages	Tumor progression
	Anti-inflammatory effects on T cells, tumor suppressor/cytotoxic activity	Anti-tumor effect
TRAIL	Induction of apoptosis	Tumor suppression
Treg cells	IL-10 release from Treg, suppression of CD8 ⁺ CTL	Anti-tumor effect
	Induction of IL-17 release from Th17	Chronic inflammation
TNF	Promotion of angiogenesis and metastasis, impairment of immune surveillance via T cell and macrophage blockage	Tumor progression, chronic inflammation
	Destruction of tumor vasculature and induction of necrosis	Anti-tumor effect
IL-23, TGF- β , IL-6, TNF	Th17 cell development	Chronic inflammation
IL-6, TNF, TGF- β	Impact on stromal cells and metastasis	Tumor progression
IL-17, TNF	Impact on endothelial cells, increase in angiogenesis	Tumor growth

given a chemical carcinogen, diethylnitrosamine (DEN) resulting in liver parenchymal damage followed by activation of Kupffer cells (KC; Naugler et al., 2007). As demonstrated in this study, DEN induces NF- κ B-dependent production of pro-inflammatory and growth-promoting IL-6 in KC via IL-1 and TLR signaling cascades, respectively, finally leading to tumor development. Estrogen inhibits activation of NF- κ B and blocks secretion of IL-6 from KC thus protecting against liver cancer in females.

Prostate cancer is an androgen-dependent cancer whose susceptibility to gonadal hormones is regulated by selective androgen-receptor modulators (SARM) attenuating the proliferative properties of androgens on tumor cells. Pro-inflammatory IL-1 derived from tumor cells or cells of the microenvironment reverses the properties of SARM from being inhibitory to activatory (Zhu et al., 2006). This de-repression effect requires TGF- β -activated kinase 1 (TAK-1)-binding protein (TAB) 2 (=TAB-2; Takaesu et al., 2000). At the molecular level, IL-1 signaling induces phosphorylation of TAB-2. TAB-2 acts as a sensor for inflammatory signals by serving as a molecular beacon for recruitment of MEKK1, which in turn mediates dismissal of the nuclear receptor co-repressor (N-CoR) holoco-repressor complex from the androgen-receptor and permits de-repression of androgen and estrogen receptor target genes. According to Zhu et al. (2006), this strategy might have come into notice in order to trigger reversal of gonadal hormone-dependent repression of a limited cohort of target genes in response to inflammatory signals linking inflammatory, and nuclear receptor ligand responses to essential reproductive functions. Treatment of prostate cancer by androgen deprivation either by suppression of testicular androgen production or by the use of pharmacological SARM such as flutamide or bicalutamide remains the standard systemic therapy. It has been shown previously that bicalutamide does not function as an androgen-receptor (AR) antagonist by preventing AR binding to DNA but instead stimulates the assembly of a transcriptionally inactive receptor on DNA (Masiello et al., 2002). Recent reports demonstrate that AR can also bind to co-repressor proteins, including N-CoR, and that this binding is enhanced in the presence of bicalutamide indicating that co-repressor binding could further contribute to the *in vivo* antagonist activity of bicalutamide (Cheng et al., 2002; Shang et al., 2002; Yoon and Wong, 2006). Interestingly, the group of Hollenberg clearly demonstrated that the AR/N-CoR interaction is not enhanced by AR antagonists used currently for the treatment of prostate cancer, but can be markedly enhanced by mifepristone (RU486) *in vitro* (Hodgson et al., 2005). RU486 can thus be considered as a novel AR antagonist that will likely have novel activities *in vivo*. However, clinical trials of RU486 or related drugs are needed to determine whether these may be more efficacious than currently available AR antagonists in the treatment of prostate cancer, particularly advanced androgen-independent prostate cancer. A recent phase II study was conducted to assess the efficacy of mifepristone as an AR antagonist in patients with castration-resistant prostate cancer (CRPC). In this study, RU486 showed only limited activity in patients with CRPC, but stimulated a marked increase in adrenal androgens (Taplin et al., 2008). From these findings the authors hypothesized that inhibition of glucocorticoid receptors by mifepristone might lead to an increase in adrenocorticotrophic hormone followed by an increase in adrenal androgens, and that their conversion by

tumor cells to testosterone and DHT might have limited the efficacy of mifepristone. As stated by Taplin et al. (2008), a therapeutic approach that combines mifepristone with a second drug harboring a complementary mechanism might be effective in blocking the compensatory rise in adrenal androgens seen in patients with CRPC. These data clearly demonstrate the impact of inflammation and gonadal hormones in tumor progression thus consolidating the fundamental work of Rudolf Virchow and George Thomas Beatson in the field of tumor-associated inflammation.

INFLAMMATION AND RADIATION

Several lines of evidence indicate that inflammation plays a pivotal role in modulating radiation responsiveness of tumors. Radiation treatment is obviously a two-edged sword. On the one hand, sublethal doses of ionizing radiation (IR) induces a nuclear DNA damage response. On the other hand, they trigger a cellular damage response in tumors by inducing pro-inflammatory pathways predominantly mediated via activation of NF- κ B, the central linker between inflammation, carcinogenesis, and radioresistance. Apart from NF- κ B activation, radiation activates/up-regulates the expression of immediate early genes encoding for, e.g., c-Fos, c-Myc, c-Jun (Hong et al., 1997) as well as TNF (Zhou et al., 2001), GM-CSF (Akashi et al., 1992), PGHS-2 (Steinauer et al., 2000), and ICAM-1 (Son et al., 2006). Radiation also induces activation of receptor tyrosine kinase pathways (Fedrigo et al., 2011) and mitochondria-associated responses (Aykin-Burns et al., 2011). As demonstrated by Valerie et al. (2007), radiation-induced activation of plasma membrane receptors occurs via generation of ionizing events in the liquid phase of the cytosol that are amplified, possibly via mitochondria, generating large amounts of ROS and RNS that inhibit protein tyrosine phosphatase (PTPase) activities. Moreover, radiation activates acidic sphingomyelinase and increases the production of ceramide. Inhibition of PTPases leads to activation of non-receptor and receptor tyrosine kinases (RTK) including epidermal growth factor receptor (EGFR) and the activation of down-stream signal transduction pathways (Goldkorn et al., 1997; Szumiel, 2008). Radiation-induced ceramide was found to promote membrane-associated receptor activation by facilitating the clustering of receptors within lipid rafts (Maziere et al., 2001; Galabova-Kovacs et al., 2006). As a consequence, activated RTK induce down-stream pro-survival pathways (e.g., Akt) that might act as promising targets in enhancing radiosensitivity of tumors.

Furthermore, various inflammatory mediators are reported as being up-regulated during radiation responses. In human glioblastoma cells, exposure to gamma-irradiation stimulated release of IL-6 and IL-8 into culture supernatants (Pasi et al., 2010). Radiation and chemotherapy led to a remarkable increase in the production of these cytokines in human oral carcinoma cells (Tamatani et al., 2004). IR induced a tremendous increase in IL-1, IL-6, and GM-CSF production by human lung cancer cells (Zhang et al., 1994). A similar effect was observed in patients with head and neck cancer where increased IL-6 and IL-8 levels can be detected after chemoradiotherapy (Meirovitz et al., 2010). Elevated IL-6 serum levels are also reported in patients with locally advanced non-small cell lung cancer undergoing concurrent chemoradiation therapy (Wang et al., 2010).

Some concern had been raised about the effect of IR on the expression of the pleiotropic cytokine TNF. The majority of the studies points to an activatory action of IR. For instance, Chendil et al. (2004) reported on a radiation-induced up-regulated TNF protein in prostate cancer cells PC-3 leading to an increase in NF- κ B activity followed by an induction of Bcl-2 protein. Furthermore, IR persistently induced NF- κ B DNA-binding activity and NF- κ B-dependent TNF transactivation and secretion (Veeraraghavan et al., 2011a). In contrast, three-dimensional conformal blocking radiation therapy did not alter serum levels of TNF in patients with prostate cancer (Lopes and Callera, 2011). It is interesting to note that in an immunocompetent animal model of pancreatic cancer the combination of the radio inducible TNF-expressing adenovector Ad.Egr-TNF with IR resulted in significant anti-tumor effects mediated by the immune system (Meng et al., 2010). As demonstrated in this study, Ad.Egr-TNF/IR therapy contributed to local tumor control through TNF production in the tumor microenvironment. TNF induced expression of the known potent immune regulator IFN- β that, in turn, stimulated the production of chemokines leading to the recruitment of CD8⁺ T cells to the tumor. Several clinical and preclinical studies with Ad.Egr-TNF/IR have suggested that this local approach suppresses the growth of distant metastases (Moral and Tomillero, 2008). However, a Phase III trial comparing Ad.Egr-TNF (TNFeradeTM) along with standard of care therapy (defined as infusion 5-FU and radiation therapy, followed by gemcitabine or gemcitabine/erlotinib maintenance therapy) versus standard of care therapy in the treatment of locally advanced, unresectable pancreatic cancer failed. Since the interim analysis did not provide sufficient evidence of the clinical effectiveness of TNFerade, the supplying company, GenVec, announced the discontinuation of the trial in 2010. From these observations one can hypothesize that up-regulated TNF probably enhances the impact of tumorigenic stimuli within the tumor or the tumor microenvironment thereby forcing a critical amplification mechanism in tumor-associated inflammation triggered by pro-inflammatory mediators.

It should be kept in mind that radiation therapy represents an efficient local anti-cancer approach leading to elimination of both, tumor cells and cells of the tumor microenvironment such as endothelial cells and tumor-induced suppressor T cells (North, 1984). IR also affects function of immune cells culminating in homing of APC and effector T cells (Ganss et al., 2002). According to the study of Ganss et al. (2002), the combination of irradiation and adoptive tumor-specific T cell therapy ensures antigen-driven tumor cell eradication with anti-angiogenic effects on tumor endothelium. It has been shown previously that sublethal doses of IR stimulates anti-tumor T cell responses and up-regulates MHC class I/II expression in melanoma cells rendering the cells more sensitive to T cell recognition (Abdel-Wahab et al., 1996), obviously through tumor-specific antigen presentation by DC (Ciernik et al., 1999). Clinical phase I/II trials are ongoing and will shed light on the efficiency of low-dose single phase fraction radiotherapy on tumor-infiltrating T cells responses in patients with liver metastasis derived from colorectal cancer (Reissfelder et al., 2011) and primarily operable pancreatic cancer (Timke et al., 2011).

PGHS-2 INHIBITION

PGHS-2, the rate-limiting enzyme involved in converting arachidonic acid to prostanoids, has emerged as another crucial NF- κ B-dependent pro-inflammatory mediator in tumorigenesis. Aberrant up-regulation of PGHS-2 is frequently observed in various pre-cancerous and malignant tissues. Because most of the PGHS-2-induced effects are mediated by its product PGE₂, down-regulation of prostaglandins in tumor tissues by PGHS-2 inhibition blocks several neoplastic pathways restricting tumor growth. Radiation is known to induce inflammation and NF- κ B consequently up-regulating/activating PGHS-2. In this context, PGHS-2 inhibitors have been tested for their anti-tumor efficiency in combination with radiation or chemotherapy. It was found that these inhibitors exert promising anti-cancer effects in a variety of human tumor cells and increase the sensitivity of tumor cells toward chemotherapy and/or radiation therapy. For instance, the PGHS-2 inhibitor NS398 enhanced the radiosensitivity of radioresistant esophageal cancer cells CSC-like Eca109R50Gy most likely by down-regulating the expression of β -catenin as well as inhibiting activation of Akt and inducing apoptosis (Che et al., 2010, 2011). NS398 was also found to radiosensitize human melanoma cells through G2/M arrest of the cell cycle, predominantly via necrotic mechanisms (Johnson et al., 2008). A further PGHS-2 inhibitor, celecoxib, increased radiation-induced cell death, and clonogenic kill of prostate cancer cells *in vitro* providing a rationale for clinical evaluation of celecoxib in combination with irradiation in prostate cancer patients (Handrick et al., 2009). Celecoxib also enhanced radiosensitivity of bronchial and colon carcinoma cells by inhibiting EGFR-mediated mechanisms of radioresistance independent of PGHS-2 activity implying that PGHS-2 inhibition might ameliorate the therapeutic outcome of radiation therapy even in patients with PGHS-2-independent tumor radioresistance (Dittmann et al., 2008). In line with this observation, targeting PGHS-2 by different pharmacological inhibitors led to radio enhancement of human glioma cells in the absence of the PGHS-2 protein (Kuipers et al., 2007). Nimesulide is another PGHS-2-selective inhibitor that has been found to increase the efficacy of radiation therapy in non-small cell lung cancer cells possibly via suppression of NF- κ B-mediated, radiation-induced cytoprotective genes (Grimes et al., 2006).

However, selective PGHS-2 inhibitors have come under scrutiny because of reports suggesting an increased cardiovascular risk associated with their use (Solomon et al., 2008). The thitherto used high therapeutic concentrations of these drugs may contribute to a pro-thrombotic state in patients with higher risk for serious cardiovascular events. A novel approach to overcome the limitations associated with the toxicity of PGHS-2 inhibitors might be the combination of pharmacological PGHS-2 inhibitors at low doses with naturally occurring compounds such as the catechin EGCG which is a promising chemopreventive agent derived from green tea (Cerella et al., 2010; Hårdtner et al., 2012). Of note, nutraceuticals such as plant-derived polyphenols have been studied intensively for their potential chemopreventive properties and have been found as being pharmacologically safe. These compounds comprise genistein, curcumin, resveratrol, silymarin, caffeic acid phenethyl ester, flavopiridol, emodin, green tea polyphenols (e.g., EGCG), piperine, oleandrin, ursolic

acid, and betulinic acid. These phytochemicals sensitize tumor cells to chemotherapeutic agents and radiation therapy by inhibiting pathways responsible for treatment resistance (Garg et al., 2005; Nambiar et al., 2011). Among them, curcumin derived from the rhizomes of *Curcuma longa* has been identified to improve the anti-tumor effects of IR by blocking NF- κ B pathways, down-regulating anti-apoptotic Bcl-X_L and survivin as well as increasing G2/M phase arrest in the cell cycle distribution in Burkitt's lymphoma cells (Qiao et al., 2012). Curcumin also potentiates radiation therapy-induced cell death by targeting radiation therapy-induced NF- κ B activation in pancreatic cancer cells (Veeraraghavan et al., 2011b).

STAT-3 AND HIF-1 INHIBITION

Apart from NF- κ B and PGHS-2, STAT-3 is a further inflammatory molecule crucially involved in radioresistance of tumors. Since enhanced radioresistance of cancer cells is additionally related to radiation-induced activation of the JAK/STAT pathway, inhibition of STAT by, e.g., phytochemicals might sensitize tumors to radiation therapy. It has been shown previously that STAT-3-mediated radiosensitization obviously occurs via down-regulation of anti-apoptotic survivin (Kim et al., 2006a). In this context, resveratrol, a polyphenolic phytoalexin, selectively targets numerous cell signaling pathways and decreases clonogenic survival primarily via an apoptotic mechanism. In melanoma cells, resveratrol inhibits STAT-3 and NF- κ B-dependent transcription, culminating in suppression of c-FLIP and Bcl-X_L expression, while activating the MAPK and the ATM-Chk2-p53 pathways (Johnson et al., 2008). Resveratrol also up-regulates TRAIL promoter activity and induces TRAIL surface expression in some melanoma cell lines, resulting in a rapid apoptosis development (Johnson et al., 2008). As also demonstrated in this study, sequential treatment of melanoma cells, first with gamma-irradiation to up-regulate TRAIL receptor surface expression, and then with resveratrol to suppress anti-apoptotic proteins c-FLIP and Bcl-X_L and induce TRAIL surface expression, dramatically up-regulated apoptosis in some melanoma cell lines. Nitidine chloride, a natural phytochemical alkaloid derived from *Zanthoxylum nitidum*, was identified as a potent STAT-3 signaling inhibitor suppressing angiogenesis and growth of human gastric cancer (Chen et al., 2012a). From these data one can hypothesize that phytochemicals in combination with IR may play a significant role in enhancing the therapeutic efficacy of cancer treatment.

As already mentioned, HIF-1, the key mediator in hypoxia signaling pathways, is crucially involved in hypoxia-induced tumor radioresistance. This obviously includes radiation-induced activation of HIF-1 (Moeller et al., 2004; Harada et al., 2009a,b), HIF-1-dependent induction of VEGF and protection of endothelial cells from radiation-induced cytotoxicity by VEGF (Gorski et al., 1999) as well as delivery of oxygen and nutrients to tumor cells by radioprotected tumor blood vessels (Zeng et al., 2008). N-Myc down-stream-regulated gene 2 (*NDRG2*) was recently identified in cervical cancer cells as a new HIF-1 target gene acting down-stream of HIF-1 to promote radioresistance via suppression of radiation-induced Bax expression (Liu et al., 2010). Therefore, it would be reasonable to study the efficacy of HIF-1 and *NDRG2* blockage as radiosensitizer for tumor therapy. Strategies

to over-come radioresistance of hypoxic tumor cells comprise, among others, hyperbaric oxygenation, gene therapy approaches, fractionated radiotherapy, radiosensitization by mimicking the effect of molecular oxygen using nitroimidazole derivatives as well as suppression of hypoxic tumor cell radioresistance by HIF-1 inhibitors (for a review, see Harada, 2011). As described previously, administration of the HIF-1 inhibitor YC-1 to hypoxic cobalt-treated cells derived from squamous-cell carcinoma of the larynx effectively inhibited HIF-1 α expression, and enhanced the sensitivity of cells to radiation, decreasing the surviving fraction to that of normoxic cells (Moon et al., 2009). YC-1 was found to reduce the number of tumor lesions after tumor cell inoculation in nude mice (Shin et al., 2007). Compared to radiation therapy alone, inhibition of radiation-induced HIF-1 activation by YC-1 led to a significant reduction in tumor cell growth (Harada et al., 2009b).

Another HIF-1 inhibitor, acriflavine, was found to inhibit tumor growth and angiogenesis in a xenograft tumor model for human prostate cancer through blockage of HIF-1 dimerization (Lee et al., 2009). Since the same agent blocked lung metastasis in an orthotopic breast cancer model (Wong et al., 2012), it would be of interest to test the efficacy of this HIF-1 inhibitor in increasing radioresistance of certain tumors. Interestingly, inhibition of Hsp90 function by 17-allylamino-17-demethoxygeldanamycin or deguelin, a novel natural inhibitor of Hsp90, suppressed increases in HIF-1 α /Hsp90 interaction and HIF-1 α expression in radioresistant lung cancer cells (Kim et al., 2009). Hsp90 interacts with HIF-1 α in competition with receptor of activated protein C kinase 1 (RACK-1) and inhibits oxygen-independent degradation of HIF-1 α (Semenza, 2007). The study by Kim et al. (2009) also demonstrated that the combined treatment of radiation with deguelin significantly decreased the survival and angiogenic potential of radioresistant lung cancer cells *in vitro* and inhibited tumor growth and angiogenesis *in vivo*.

Even phytochemicals have the capacity to inhibit radiation-induced HIF-1 activation. Pre-treatment of prostate cancer cells with soy isoflavones inhibited Src/STAT-3/HIF-1 α activation by radiation and nuclear translocation of HIF-1 α . These findings correlated with decreased expression of APE1/Ref-1 and DNA-binding activity of HIF-1 α and NF- κ B (Singh-Gupta et al., 2009). Apurinic/aprimidinic (AP) endonuclease 1/redox factor-1 (APE1/Ref-1) is a multifunctional protein involved in DNA repair that also functions as a redox activator of cellular transcription factors. Emodin, a natural anthraquinone enriched in the traditional Chinese herbal medicines and novel small HIF-1 inhibitor, was found to improve efficacy of chemotherapeutic drugs by inhibiting transactivation of HIF-1 without impairing mRNA expression and stability of HIF-1 α protein in prostate cancer cells (Huang et al., 2008). Another approach to HIF-1 blockage might be the use of cell-permeable HIF-1 antagonists (Shi et al., 2007). As mentioned before, HIF-1 contributes to tumor radioresistance by up-regulating survivin expression under hypoxic conditions. Moreover, in hypoxic tumor cells the HIF-1 signaling pathway is activated and could be further enhanced by radiation, thereby providing survival signals to adjacent vascular endothelial cells by up-regulation of VEGF and basic fibroblast growth factor (bFGF) and resulting in tumor radioresistance through vascular

radioprotection. Thus, HIF-1 antagonists might decrease tumor angiogenesis and sensitize tumor cells to radiotherapy.

Targeting HIF-1 for radiosensitization can also be achieved by inhibiting its up-stream mediators. In this respect, the PI3K/Akt/mTOR pathway can be considered as the most relevant target in sensitizing tumors to radiation therapy. Mutations in this pathway can be found in various human cancers and have been found to up-regulate HIF-1 α expression (Zhong et al., 2000). The PI3K/Akt/mTOR pathway also affects the NF- κ B-mediated expression of PGHS-2 (Yang et al., 2009) and radiation-induced MMP-9 expression (Cheng et al., 2006). Several studies demonstrated that specific blockage of this pathway by certain inhibitors such as erufosine (Rudner et al., 2010), LY294002 (Nakamura et al., 2005; Kao et al., 2007), RAD001 (Albert et al., 2006), and rapamycin (Majumder et al., 2004) ensures efficient radiosensitization of distinct tumors cells.

NF- κ B INHIBITION

In recent years NF- κ B inhibition by synthetic compounds as well as nutraceuticals of different sources has been approved for tumor radiosensitization. In particular the use of nutraceuticals became a popular approach due to the broad anti-tumor and anti-inflammatory properties in conjunction with low toxicity risks of these compounds (Deorukhkar et al., 2007; Deorukhkar and Krishnan, 2010). How these drugs block NF- κ B activation is becoming increasingly apparent. Among them, curcumin has emerged as one of the best studied plant-derived polyphenols. In a phase II clinical trial curcumin showed beneficial effects in patients with advanced pancreatic cancer (Dhillon et al., 2008). Curcumin down-regulated expression of NF- κ B, PGHS-2, and phosphorylated STAT-3 in peripheral blood mononuclear cells from these patients. Furthermore, curcumin has been shown to suppress TNF-mediated NF- κ B activation by inhibiting inhibitor of kappaB α (I κ B α) in human myeloid ML-1a cells (Singh and Aggarwal, 1995). Curcumin also confers radiosensitizing effects in prostate cancer cells by inhibiting TNF-mediated NF- κ B activation resulting in Bcl-2 protein down-regulation and concomitant activation of cytochrome c and caspase-9 and -3 (Chendil et al., 2004). More recently, curcumin was found to sensitize colorectal cancer cells to radiotherapy by suppressing radiation-induced NF- κ B activation via inhibition of radiation-induced phosphorylation and degradation of I κ B α , inhibition of inhibitor of kappaB kinase (IKK) activity, and inhibition of Akt phosphorylation (Sandur et al., 2009) consequently leading to down-regulation of several tumorigenic factors in colorectal cancer xenografts in nude mice (Kunnumakkara et al., 2008). Apart from its IKK-inhibitory capacity, curcumin also blocks p65 phosphorylation and acetylation and represses the p300/CREB-binding protein (CBP) HAT activity-dependent transcriptional activation from chromatin (Balasubramanyam et al., 2004). Similarly, anacardic acid (6-pentadecylsalicylic acid) derived from traditional medicinal plants, such as cashew nuts, has been identified to inhibit NF- κ B activation, to suppress the activation of I κ B α kinase that led to abrogation of phosphorylation and degradation of I κ B α and to inhibit acetylation and nuclear translocation of p65 (Sung et al., 2008). The same study demonstrated that down-regulation of the

p300 HAT gene by RNA interference abrogated the effect of anacardic acid on NF- κ B suppression. Further nutraceuticals including the soy isoflavone genistein (Raffoul et al., 2006), the isoquinoline alkaloid berberine from medicinal plants such as *Berberis aristata*, *Coptis chinensis*, *Coptis japonica*, *Coscinium fenestratum*, and *Hydrastis canadensis* (Pandey et al., 2008), piceatannol (3,3',4,5'-trans-trihydroxystilbene), a naturally occurring hydroxylated analog of resveratrol found in various plants (Son et al., 2010), the principal prenylated flavonoid xanthohumol from *Humulus lupulus* (Harikumar et al., 2009), and the polyphenol butein (3,4,2',4'-tetrahydroxychalcone) from *Rhus verniciflua* Stokes (Pandey et al., 2007) have been identified as blocking NF- κ B by direct interaction with IKK β on cysteine 179 residue. NF- κ B inhibition by direct interaction with one of its subunits occurs in the presence of numerous phytochemicals including sesquiterpene lactones (Garcia-Pineros et al., 2001). In radiation-resistant human CGL1 cells, parthenolide, a major active component of the herbal medicine feverfew (*Tanacetum parthenium*), enhanced radiosensitivity through NF- κ B inhibition and apoptosis induction via p53 stabilization, induction of pro-apoptotic Bax, and phosphorylation of pro-apoptotic Bid (Mendonca et al., 2007). The radiosensitization effect of parthenolide is enhanced in the presence of tumor suppressor protein PTEN (phosphatase and tensin homolog deleted on chromosome 10), in part, by suppressing the absolute amount of activated p-Akt in human prostate cancer cells (Sun et al., 2007). The group of Aggarwal recently found out that crotepoixide (a substituted cyclohexane diepoxide), isolated from *Kaempferia pulchra* (peacock ginger), inhibited activation of TGF- β -activated kinase (TAK)-1, which led to suppression of I κ B α kinase, abrogation of I κ B α phosphorylation and degradation, nuclear translocation of p65, and suppression of NF- κ B-dependent reporter genes encoding for anti-apoptotic (Bcl-2, Bcl-X_L, IAP1/2, Mcl-1, survivin, TRAF-1), pro-apoptotic (Bax, Bid), pro-inflammatory (PGHS-2), proliferation- (cyclin D1, c-Myc), invasion- (ICAM-1, MMP-9), and angiogenesis-promoting (VEGF) factors (Prasad et al., 2010). Moreover, Tamatani et al. (2007) analyzed the effects of radiation therapy in combination with cepharanthine on NF- κ B activation and expression of its down-stream effector molecules in human oral squamous-cell carcinoma cells. Cepharanthine is a bisocoumarin alkaloid extracted from the roots of *Stephania cepharantha* Hayata, and is widely used in Japan for the treatment of patients with leucopenia, nasal allergy, and venomous snakebites. The authors could show that treatment of cancer cells with cepharanthine combined with exposure to IR enhanced radiosensitivity via NF- κ B inhibition and concomitant down-regulation of IL-6, IL-8, and anti-apoptotic proteins such as cellular inhibitor of apoptosis protein (cIAP)-1 and -2. Moreover, the pleiotropic effects of cepharanthine also includes inhibition of STAT-3 in the human osteosarcoma cell line SaOS2 (Chen et al., 2012b). These examples highlight the crucial role of naturally occurring compounds in targeting inflammatory signaling pathways for sensitizing tumors to radiation therapy. Beside them, a variety of further compounds have been identified to enhance radiosensitivity via inhibition of NF- κ B including celecoxib (Raju et al., 2005), pitavastatin (Tsuboi et al., 2009), docosahexaenoic acid (Zand et al., 2008), and the novel organoselenium thioredoxin reductase inhibitor ethaselen (Wang et al., 2011).

TUMOR CELL RE-POPULATION

It is commonly accepted that aggressive radio-/chemotherapy often results in a negative selection toward highly aggressive tumor clones. In recent days, the induction of radiation-induced apoptosis, the principal purpose of radiation therapy, has come under scrutiny due to reports suggesting that dying tumor cells use the apoptotic pathway to stimulate the re-population of tumors subjected to radiation therapy. The process of re-population was originally discovered in the second half of the twentieth century and is commonly accepted as playing a crucial role in radio-/chemotherapy (Hermens and Barendsen, 1969; Stephens et al., 1978). Interestingly, the group of Li from Aurora (USA) demonstrated that radiation-induced apoptosis comprises caspase-3 cleavage and concomitant activation of Ca^{2+} -independent phospholipase A₂ (iPLA₂) followed by PGE₂ release promoting tumor cell re-population *in vitro* and *in vivo* (Huang et al., 2011). The relevance of this mechanism for the caspase-mediated tumor cell re-population was confirmed by caspase-3 determination in different cancer patients. Herein, elevated expression levels of activated caspase-3 in tumor tissues correlated with poor clinical outcome in patients with head and neck cancer as well as advanced stage breast cancer. However, the contribution of macrophages and the subsequently generated clearance-related anti-inflammatory milieu to radiation-induced tumor cell re-population and poor therapeutic outcome should be taken into consideration. As discussed by Tauber and co-workers, apoptotic manifestations such as externalization of phosphatidylserine, bleb formation, and DNA fragmentation are crucially involved in macrophage activation and depend on caspase-3 activation (Jänicke et al., 1998; Coleman et al., 2001; Sebbagh et al., 2001). The authors conclude that caspase-3-positive apoptotic cells can recruit more macrophages and are more efficiently internalized by the phagocytes (“silent clearance”) culminating in a strong anti-inflammatory and growth-promoting phagocyte response via release of clearance-associated cytokines, e.g., PGE₂ than their caspase-3-negative counterparts. However, further preclinical and clinical studies using certain caspase inhibitors are required to strengthen this hypothesis. In this context, the novel caspase inhibitor GS-9450 was found to down-regulate caspase-3 expression on peripheral T cells from chronically HCV-infected patients in a phase II clinical trial (Arends et al., 2011). It would therefore be of interest to study a putative beneficial effect of a combinatorial treatment with caspase inhibitors and radiation therapy on the clinical outcome of patients with various advanced cancer types.

PATTERN RECOGNITION RECEPTORS IN RADIOIMMUNITY

It is well known that chronic inflammation induced by non-infectious agents can also contribute to carcinogenesis and act as a driving force in tumorigenesis. Several factors such as growth factors, oncoproteins, and toxins can affect the host via an activation of pattern recognition receptors (PRR) interacting with exogenous pathogen-associated molecular patterns (PAMP; Kawai and Akira, 2011). Apart from PAMP, the same receptor superfamily also recognizes endogenous “alarmins” both of them comprising the group of danger-associated molecular patterns (DAMP; Bianchi, 2007). Receptor ligation leads to activation of inflammatory cells and initiation of host responses that tend to

eradicate invading microorganisms (Akira et al., 2006; Karin et al., 2006). Not surprisingly, inadequate pathogen elimination, recurring tissue injury, prolonged inflammatory signaling, and failure of anti-inflammatory mechanisms can all culminate in chronic inflammation promoting cancer development.

Signaling via PRR also makes an impact on radiation responses. It has been shown previously that both, low (0.075 Gy) and high (2 Gy) doses of IR causes sustained stimulation of IL-12 and IL-18 secretion by mouse macrophages (Shan et al., 2007) with concomitant activation of NF- κ B accompanied by elevated cytoplasmic MyD88 levels and an up-regulated surface expression of CD14 and TLR-4/MD-2 (Shan et al., 2007), the latter acting as LPS sensor. From these findings the authors hypothesized that IR can stimulate the secretion of IL-12 and IL-18 presumably via activation of the Toll signaling pathway in macrophages. A detailed description of the TLR-4 signaling pathway is visualized schematically in **Figure 2**.

Interestingly, RP105 (radioprotective 105 kDa), a TLR-related molecule, was recently identified on human B cells and DC (Fugier-Vivier et al., 1997) and characterized as being similar to TLR-4 in that the extracellular leucine-rich repeats associate with MD-1, an MD-2-like molecule (Miyake et al., 1995; Fugier-Vivier et al., 1997). MD-2 directly binds to lipid A, the active center of lipopolysaccharide (LPS), leading to dimerization of TLR-4/MD-2 (Ohto et al., 2011). An antibody raised against surface-bound RP105 was found to drive B cell proliferation and protection from subsequent radiation- or dexamethasone-induced apoptosis (Miyake et al., 1994). Studies by Divanovic et al. (2005) demonstrated that: (a) RP105 is a specific inhibitor of TLR-4 signaling in human embryonal kidney cells HEK293; (b) RP105/MD-1 interacts directly with TLR-4/MD-2, thus abolishing the LPS-binding capacity of the complex; (c) RP105 regulates TLR-4 signaling in DC and macrophages; and (d) RP105 regulates *in vivo* responses to LPS supporting the assumption that RP105 acts as a physiological negative regulator of TLR-4 responses. This brief overview makes it clear that, on the one hand, pro-inflammatory responses to radiation and TLR signaling enhance the impact of tumorigenic factors in the tumor and the tumor microenvironment and, on the other hand, might represent pivotal target structures in radiation therapy. As discussed by Schae and McBride (2010), radiation-induced DAMP signaling via TLR-2/-4 has emerged as a critical component in affecting the outcome of anti-cancer therapies. In this context, radiation was found to induce secretion of the prototypical DAMP, the high-mobility-group box 1 (HMGB-1) “alarmin” protein from dying tumor cells as a prerequisite for the development of a tumor antigen-specific T cell immunity mediated by an interaction of HMGB-1 with TLR-4 on DC (Apetoh et al., 2007). The same study revealed that patients with breast cancer who carry a TLR-4 loss-of-function allele relapsed more quickly after radiotherapy and chemotherapy than those carrying the normal TLR-4 allele implying a clinically relevant immunoadjuvant pathway triggered by tumor cell death. An intriguing novel finding comprises the reduction of metastatic ability and MMP-9 expression in MGC-803 gastric cancer cells by silencing of the HMGB-1 expression using an HMGB-1-specific RNAi lentiviral vector (Song et al., 2012). As also shown in this study, HMGB-1 silencing decreased cell proliferation and sensitized cells to

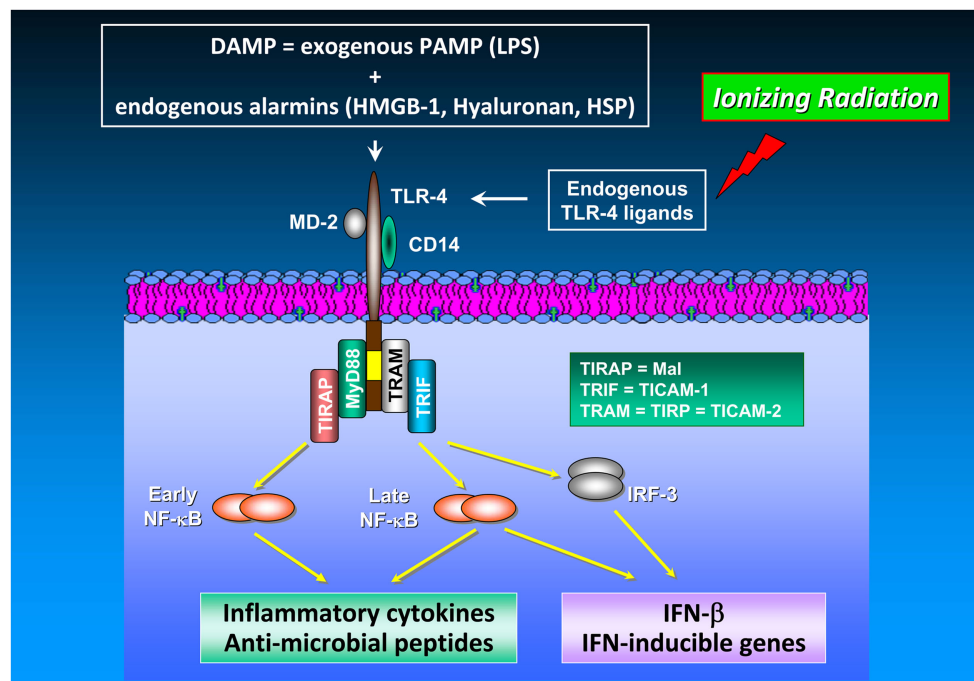


FIGURE 2 | TLR-4 signaling in cancer. The TLR-4/MD-2 receptor complex recognizes and binds exogenous PAMP (e.g., endotoxins such as LPS) as well as endogenous alarmins (HMGB-1, hyaluronan, heat shock proteins). Release of DAMP into the extracellular space is achieved by a number of different mechanisms including (i) leakage from necrotic cells, (ii) increased synthesis and post-translational modification in response to inflammation, and (iii) degradation of inactive precursors into TLR-mimetic degradation products in inflammatory environments (Mencin et al., 2009). TLR-4 induces two distinct signaling pathways controlled by the TIRAP/MyD88 and TRAM/TRIF pairs of adaptor proteins, which elicit the production of pro-inflammatory cytokines and type I interferons, respectively. The cytosolic adapter molecules mentioned above comprise myeloid differentiation protein 88 (MyD88), Toll/IL-1R resistance domain-containing adapter inducing IFN- β (TRIF), TIR domain-containing adapter protein (TIRAP), and TRIF-related adaptor molecule (TRAM). TIRAP is also termed Mal (MyD88 adaptor-like), TRIF is also known as Toll/IL-1R homology domain-containing adaptor molecule 1 (TICAM-1), whereas TRAM is alternatively entitled TIR-containing protein (TIRP) and TICAM-2, respectively. TLR-4-mediated signal transduction occurs via MyD88-dependent and MyD88-independent (i.e., TRAM/TRIF-dependent)

pathways. Both, MyD88-dependent and MyD88-independent pathways induce expression of genes involved in pro-inflammatory and anti-microbial responses (Akira and Takeda, 2004). In TLR-4 signaling, MyD88 up-regulates inflammatory cytokines via NF- κ B activation. Moreover, the MyD88-independent pathway does not only induce inflammatory gene expression in an NF- κ B-dependent manner but also up-regulates type I interferon expression via the transcription factor IRF3. NF- κ B activation and subsequent inflammatory cytokine production are mediated by different mechanisms and kinetics in the MyD88-dependent and the MyD88-independent pathway: NF- κ B activation in the MyD88-dependent pathway is an early event occurring with fast kinetics whereas NF- κ B activation via the MyD88-independent pathway represents a late event occurring with slower kinetics. Unlike TLR-4 signaling in immune cells which has been found to enhance anti-tumor immunity by, e.g., IL-12/IFN- γ up-regulation and promotion of DC maturation and function, TLR-4 signaling in cancer cells increases their tumorigenic capacity under certain circumstances (Oblak and Jerala, 2011). Noteworthy, HMGB-1 which is released from irradiated tumor cells functions as an endogenous TLR-4 ligand leading to the development of a tumor antigen-specific T cell immunity mediated by an interaction of HMGB-1 with TLR-4 on DC.

apoptosis implying HMGB-1 as being a potential target for the therapeutic intervention of certain cancers such as gastric cancer.

TARGETING OF HEAT SHOCK PROTEINS IN RADIOTHERAPY

A promising approach in cancer therapy also might be targeting heat shock proteins (HSP), a class of proteins which are induced under physiologic stress to promote cell survival in the face of endogenous or exogenous injury. Compared to normal cells, tumors frequently have elevated basal Hsp70 levels which are further enhanced in response to a number of pathological and environmental stresses such as nutrient deficiency, hypoxia, heavy metals, irradiation, and/or chemotherapeutic agents. Also normal cells show an increase in the synthesis of Hsp70 following stress in order to mediate protection against lethal damage and to maintain protein homeostasis. Screening of nearly 1,000 primary human

tumor biopsies and the corresponding normal tissues revealed that human carcinomas, but none of the tested normal tissues, frequently present Hsp70 on their cell surface (Multhoff et al., 1995; Multhoff, 2007). A membrane Hsp70-positive tumor phenotype has been found to be associated with a significantly decreased overall survival in tumor patients. Therefore, the expression of this molecule could serve as a negative prognostic marker (Pfister et al., 2007).

Apart from their intracellular localization, Hightower and Guidon Jr. (1989) reported on an ER/Golgi-independent release of Hsp70 from viable cells with intact cell membranes already in the late 1980s. Extracellular HSP are considered as molecules with immunomodulatory functions (Pockley and Multhoff, 2008; Pockley et al., 2008) either as cross-presenters of immunogenic peptides (Srivastava, 1997; Asea et al., 2000) or in a peptide-free

version as chaperokines (Asea et al., 2002) or stimulators of innate immune responses (Multhoff et al., 1995). Despite these well-documented immunological functions, the mechanisms of HSP export are still controversially discussed since cytosolic HSP lack a consensus signal for secretion. However, apart from Hsp70, other molecules lacking a secretory signal such as IL-1 α , IL-1 β , and HMBG-1 are also found outside of cells (Nickel and Seedorf, 2008; Eder, 2009). Hsp70 also has been found to be located on the cell surface although lacking a transmembrane domain (Multhoff and Hightower, 1996). Membrane Hsp70 might help to maintain stability of tumor cells and thus might protect tumors from lethal damage induced by environmental stress (Horvath et al., 2008; Horvath and Vigh, 2010). The fundamental work of the group of Antonio De Maio has demonstrated an interaction of members of the HSP70 family with artificial membranes containing phosphatidylserine (PS; Arispe and De Maio, 2000; Arispe et al., 2002; Vega et al., 2008; De Maio, 2011). Our group reported on an interaction of Hsp70 with the sphingolipid globotriaosylceramide (Gb3) in the plasma membrane of non-stressed human gastrointestinal stromal tumors (Gehrmann et al., 2008). Gb3 is found in cholesterol-rich microdomains, also termed as lipid rafts, which serve as signal transduction platforms. Following irradiation or hypoxia-induced stress Hsp70 was found to be associated predominantly with PS outside of lipid rafts in the plasma membrane of tumor cells (Schilling et al., 2009). These data indicate that environmental stress might result in a re-organization of the lipid bilayer and might modulate the interaction of Hsp70 with lipid components. Surprisingly, only tumors but not the corresponding normal tissues were found as being membrane Hsp70-positive using the IgG1 mouse monoclonal antibody cmHsp70.1. In contrast, other Hsp70-specific antibodies failed to bind to membrane Hsp70 on viable tumor cells (Stangl et al., 2011). The discovery that neither high/low salt concentrations nor changes in the pH were able to release Hsp70 from the plasma membrane of tumor cells (Gehrmann et al., 2008; Vega et al., 2008) confirmed our hypothesis that in tumor cells Hsp70 is an integral membrane protein which can associate with raft (Gb3) and non-raft (PS) lipid components.

We recently observed an increased surface expression of Hsp70 in colorectal tumor cells after IR alone or in combination with hyperthermia (HT) while the amount of extracellular Hsp70 was only increased when HT was given additionally (Schildkopf et al., 2011). Moreover, a high up-regulation of the co-stimulation molecule CD80 and the chemokine receptor CCR7 on DC was measured after contact with supernatants of X-ray plus HT-treated cells. This was dependent on extracellular Hsp70. Combined treatments further led to significantly increased phagocytosis rates of macrophages and DC and increased pro-inflammatory cytokine (IL-8, IL-12) secretion. From these findings we conclude that X-ray combined with HT induces Hsp70-dependent activation of immune cells and might generate a tumor microenvironment beneficial for cure.

HSP over-expression in tumor cells plays a pivotal role in tumorigenesis by inhibiting apoptosis and senescence. Recent studies indicate an involvement of HSP such as Hsp70/Hsp72 and Hsp90 in the recognition of PAMP by binding to TLR-4 within lipid rafts (Triantafyllou and Triantafyllou, 2004; Wheeler et al., 2009). Since extracellular residing Hsp70 acts as a danger signal for

the immune system (Matzinger, 1998), this stress protein has been added to the list of “alarmins” comprising the group of DAMP together with PAMP, hyaluronan, and other HSP members. Consequently, developing means of abrogating HSP expression may provide a way to render cancer cells more susceptible to radiation or chemotherapy. Various attempts are underway to target these proteins, particularly small HSP, in developing potent radiation and chemotherapy sensitizers (Guttmann and Koumenis, 2011). For instance, Hsp27 has been found as being implicated in the resistance to chemotherapy in several types of cancers. The group of Moriaki analyzed the effects of a gemcitabine treatment in pancreatic cancer cells (Nakashima et al., 2011). Gemcitabine is an anti-tumor drug and currently considered to be the standard of care for the treatment of advanced pancreatic cancer, but the clinical outcome is still not satisfactory. It was shown that gemcitabine suppressed growth of pancreatic cancer cells by inducing apoptosis. Gemcitabine also caused activation of p38 mitogen-activated protein kinase (MAPK), MAPK-activated protein kinase 2 (MAPKAPK-2) with concomitant serine phosphorylation of Hsp27 at position 15, 78, and 82 without affecting total Hsp27 levels. From these results the authors conclude that the phosphorylation status of Hsp27 obviously plays a pivotal role in gemcitabine-induced growth suppression of pancreatic cancer. Of Note, the HSP inhibitor KNK437, a benzylidene lactam compound, was observed to dramatically reduce expression of Hsp27 in gemcitabine-resistant pancreatic cancer cells KLM1-R and to enhance the *in vitro* anti-tumor cytotoxic effect of gemcitabine on KLM1-R compared to single-agent gemcitabine (Taba et al., 2011). KNK437 also sensitizes prostate cancer cells to the apoptotic effect of hyperthermia by down-regulating heat-induced Hsp70 mRNA expression (Sahin et al., 2011). Moreover, in patients with locally advanced squamous-cell esophageal cancer neoadjuvant radiochemotherapy (NRCT) led to a decreased expression of Hsp16.2, Hsp90, and heme-binding protein 2 (SOUL), and an increased Bax/Bcl-2 ratio was found in the responding tumors (Farkas et al., 2011).

Also Hsp90 may represent a potentially attractive target for specific molecular anti-cancer agents, because Hsp90 expression is up-regulated in tumors as compared with normal tissues, which implies that tumor cells might be preferentially affected by Hsp90-targeted therapies (Ferrarini et al., 1992). In this context, geldanamycin (GA), a naturally occurring ansamycin antibiotic, along with its clinically used analogs such as 17-allylamino-17-demethoxygeldanamycin (17-AAG), has been evaluated in pre-clinical and clinical trials for its significant anti-tumor properties. These agents disrupt Hsp90 association with client proteins by occupying the nucleotide-binding site of Hsp90 (Grenert et al., 1997; Prodromou et al., 1997; Stebbins et al., 1997), thereby preventing binding of Hsp90 with ATP and profoundly affecting the composition of Hsp90-containing multimolecular chaperone complexes (Obermann et al., 1998; Maloney and Workman, 2002). As demonstrated by the group of Gius, treatment of two human cervical carcinoma cell lines (HeLa, SiHa) with geldanamycin and 17-AAG resulted in cytotoxicity and, when combined with IR, enhanced the radiation response. In addition, mouse *in vivo* models using 17-AAG at clinically achievable concentrations yielded results that paralleled the *in vitro* radiosensitization studies of both

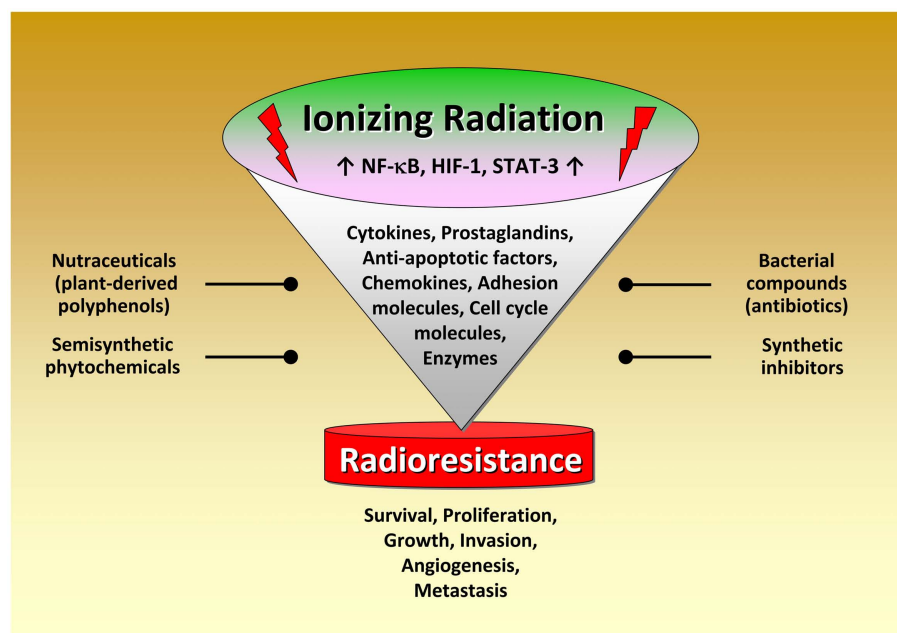


FIGURE 3 | Radiation-induced activation of inflammatory pathways in tumor cells. Schematic simplified representation of the complex intracellular mechanisms leading to radioresistance. Exposure to ionizing radiation leads to activation of several transcription factors modulating the expression of numerous factors promoting cancer development. Novel therapeutic approaches thus aim to interfere with the activity or expression of these factors, either in single-agent or

combinatorial treatment or as supplements of the existing therapeutic concepts. Noteworthy, targeting the pro-inflammatory signaling pathways for tumor radiosensitization represents a promising novel therapeutical approach in cancer. A great variety of classical or novel drugs including nutraceuticals have the capacity to interfere with the inflammatory network in cancer and are considered as putative radiosensitizers.

single and fractioned courses of irradiation (Bisht et al., 2003). We recently analyzed the effects of the novel Hsp90 inhibitor NVP-AUY922 compared to 17-AAG on the HIF-1 α /HIF-2 α expression in combination with radiosensitivity in lung cancer cell lines under normoxic and hypoxic conditions (Schilling et al., 2012b). NVP-AUY922 is a synthetic, isoxazole/resorcinol-based second generation Hsp90 inhibitor exhibiting an enhanced metabolic stability and a tighter binding to Hsp90 compared to 17-AAG (Brough et al., 2008). As given in our study, both inhibitors reduced basal and hypoxia-induced HIF-1 α levels in EPLC-272H lung carcinoma cells. However, despite a down-regulation of HIF-1 α upon Hsp90 inhibition, sensitivity toward irradiation remained unaltered in EPLC-272H cells under normoxic and hypoxic conditions. In contrast, treatment of H1339 lung carcinoma cells with NVP-AUY922 and 17-AAG resulted in a significant up-regulation of their initially high HIF-1 α levels and a concomitant increase in radiosensitivity indicating the ability of an HIF-1 α -independent radiosensitization of normoxic and hypoxic H1339 lung cancer cells via Hsp90 inhibition. From these observations it can be concluded that treatment strategies combining HSP targeting and radiochemotherapy appear to be a high potential therapeutic benefits for cancer patients.

CONCLUDING REMARKS

Although radiation therapy, alone or in combination with chemotherapy, is the primary treatment for several tumors, radioresistance dramatically attenuates radiocurability. Several lines of evidence indicate that inflammation plays a pivotal role

in modulating radiation responsiveness of tumors. As discussed in this review, exposure to IR leads to activation of several transcription factors modulating the expression of numerous factors promoting cancer development. Novel therapeutic approaches thus aim to interfere with the activity or expression of these factors, either in single-agent or combinatorial treatment or as supplements of the existing therapeutic concepts. Among them, NF- κ B, STAT-3, and HIF-1 play a crucial role in radiation-induced inflammatory responses. A great variety of classical or novel drugs including nutraceuticals have the capacity to interfere with the inflammatory network in cancer and are considered to function as putative radiosensitizers (Figure 3). Thus, targeting the inflammatory signaling pathways induced by IR offers the opportunity to improve the clinical outcome of radiation therapy by enhancing radiosensitivity and decreasing putative metabolic effects. Since inflammation and sex steroids also impact tumorigenesis, a therapeutic approach targeting glucocorticoid receptors, and radiation-induced production of tumorigenic factors might be effective in sensitizing tumor cells to IR in certain cases.

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